

Cite this: *Food Funct.*, 2019, 10, 4751

A single-enzyme system for starch digestibility screening and its relevance to understanding and predicting the glycaemic index of food products†

Cathrina H. Edwards, * Noriane Cochetel, Lauren Setterfield, Natalia Perez-Moral and Frederick J. Warren 

There is currently great interest in increasing provisions of healthier carbohydrate foods, particularly those that possess a low Glycaemic Index (GI) when measured *in vivo*. The metabolic response to many starch-rich foods is driven largely by differences in the rate and extent of starch amylolysis. Enzyme-kinetic parameters obtained from high-throughput *in vitro* amylolysis assays therefore have potential for rapid prediction of GI for starch-rich foods. The aim of this study was to evaluate the usefulness of a starch digestibility screening method and resulting enzyme-kinetic parameters in comparing and predicting the GI of a range of carbohydrate-rich foods. Starch-rich foods ($n = 20$) with GI ranging from 36 to 81 were digested by porcine pancreatic α -amylase for 90 min under a fixed enzyme–substrate ratio (4 U/10 mg starch) at 37 °C on a rotary mixer. Starch digestion progress was determined by quantification of reducing sugar concentration in aliquots collected throughout the incubation. Indices of starch digestibility (C_{20} , C_{60} , C_{90} , HI, C_{∞} , and k) were obtained and compared with GI values. Digestibility curves revealed differences in the starch amylolysis for the broad range of foods tested. *In vitro* starch digestibility indices were significantly correlated ($p < 0.01$) with GI, with the exception of the rate constant, k . Out of all the indices tested, C_{90} and C_{∞} were the most strongly correlated with *in vivo* rankings for GI of matched food products ($T_b = 0.596$, $p < 0.001$ and $T_b = 0.599$, $p < 0.01$, respectively), however the digestibility plots obtained for some of the more slowly digested foods were linear over 90 min meaning that C_{∞} and k could not be obtained from first order kinetic analysis. C_{90} was most strongly correlated with the absolute GI values ($r = 0.724$, $p < 0.001$). Overall starch digestibility profiles reflected differences in starch amylolysis for food with varying GI, and C_{90} provided the best indication of absolute and relative GI values across all product categories. The *in vitro* starch digestibility screening method shows potential for rapid prediction of GI values and is recommended for early stage food product development and for mechanistic studies.

Received 22nd March 2019,
Accepted 20th May 2019

DOI: 10.1039/c9fo00603f

rsc.li/food-function

Introduction

Starch-rich foods are a major source of dietary energy and therefore play an important role in influencing public health. The metabolic responses to starch-rich foods can be vastly different and are a critical consideration for the design of healthier foods. For instance, it has been shown that consumption of starch-rich foods with a low Glycaemic Index (GI) and high amounts of Resistant Starch (RS) are associated with reduction in Type 2 Diabetes and Cardiovascular Disease risk and could play an important role in the dietary management of these diseases.¹ Given the alarming prevalence of chronic diet-related

diseases, increasing the supply of consumer-acceptable carbohydrate foods that possess these (and other) desirable properties should be a priority. Although *in vivo* studies remain the gold standard for testing physiological outcomes, *in vitro* methodologies play an important role in rational product design.

The *in vivo* ‘Glycaemic Index methodology’ for ranking carbohydrate foods according to the glycaemic response that they evoke is well-established² and many foods have already been indexed.³ The resource-demanding nature of human studies does however impose some restriction on the number of food products that can be tested, and inter- and intra-individual variations (although relevant) means that human studies lack the sensitivity to detect subtle changes that may be of importance to product concept development. Thus, high-throughput *in vitro* methodologies that rank products by predicted GI would aid rational design of interventions for *in vivo*

Quadram Institute Bioscience, Norwich Research Park, NR4 7UA Norwich, UK.

E-mail: Cathrina.edwards@quadram.ac.uk; Tel: +44 (0)1603 251466

†Electronic supplementary information (ESI) available. See DOI: 10.1039/c9fo00603f



testing while providing for additional mechanistic insight to inform formulation of functional food products.

The glycaemic response to starch-rich food is strongly influenced by the rate and extent of starch digestion by α -amylase in the upper gastrointestinal tract.⁴ *In vitro* methods that simulate digestive conditions can therefore potentially provide a means of predicting relative differences in GI of various food. Digestion models that replicate the biochemical conditions (enzymes, electrolytes, pH, mixing *etc.*) of oral, gastric and intestinal digestion^{5,6} are increasingly popular, however the methodological complexity and large number of reagents required prevents the use of these models for high-throughput screening. Moreover, for predicting the glycaemic response to starch-rich foods, this level of complexity may not always be required; indeed, good agreement between *in vitro* starch hydrolysis indices and glycaemic responses has been achieved previously using simpler protocols.^{7,8} One limitation of these previous methods however is the tendency to pulverise, grind or homogenise foods prior to analysis; such treatments breakdown food structures (such as plant cells, tissues, intracellular networks) that are now known to be of great importance with regard to influencing digestion kinetics and measurement of Type 1 resistant starch (RS1) (*i.e.* starch that is resistant to digestion because it is physically inaccessible to digestive enzymes).^{9–11} It is therefore important that *in vitro* tests are assessed based on their performance across a broad range of food products with different structures and susceptibility to digestion.

We have previously reported on the value of an enzyme-kinetic approach that is routinely used in our laboratory for mechanistic digestibility studies of starch-rich food materials.^{12,13} When combined with a controlled and mechanistically-informed approach to sample preparation, this methodology may enable rapid comparison of starch susceptibility to digestion and prediction of GI in a range of carbohydrate foods. The aim of the present study was to apply this *in vitro* starch amylolysis methodology to a broad range of food products and evaluate the use of *in vitro* starch digestibility indices for rapid prediction of GI values.

Materials & methods

Selection and preparation of food materials

Commonly consumed starch-rich foods were selected from various product categories (pulses, tubers, cereals, pasta, rice, bread and biscuits) to cover a range of Glycaemic Index (GI) values, as reported by Atkinson *et al.* 2008.³ An overview of the food products, starch content and method of preparation is provided in Table 1 and further details of food composition is provided in **OSM1**. The nutrient composition data was obtained from nutrition declaration information provided on food products (according to EU regulation no. 1169/2011) with the exception of two products where the on pack information was not representative of how the product was prepared, and therefore food composition reference values were used.¹⁴ The starch

content was calculated by subtracting sugars from the available carbohydrate value. Values reported on an 'as weighed' basis were corrected to account for the moisture content (determined on a Moisture Balance, VWR International) at the time of weighing for digestibility analysis. It is noteworthy that large errors are commonly observed for calculated and direct enzymic determination of total starch, and that this is a likely source of error in both the *in vitro* assays and the *in vivo* studies. Wherever possible, the source and method of food preparation for *in vitro* analysis matched the description provided in the Glycaemic Index tables, and any discrepancies are disclosed within these tables.

In preparation for *in vitro* testing, the food samples were freshly cooked (if applicable) according to manufacturer instructions and ground to achieve a controlled particle size reduction. In general, dry products (*i.e.*, biscuits, bread, breakfast cereals) were ground to 'crumbs' and sieved to select a standard particle size fraction (250–500 μm). This size range was chosen as it provides a reasonable yield of representative particles in which a high proportion of the starch granules are exposed on the particle surface. Cooked products were prepared according to manufacturer instructions and analysed immediately (within 30 min) of cooking. Canned products were drained and then ground to a loosely formed paste. For this particular study, these conditions were selected in an effort to preserve food structures (*e.g.*, plant cells and RS1) that are known to reach the intestinal lumen. Variables such as particle size, cooking and cooling conditions, were standardised and controlled to enable good reproducibility.

Starch digestibility assay

The principles of the starch digestibility assay have been described previously.^{12,13} The method is described below and is similar to that performed by Edwards 2018,¹⁵ with minor modifications to the volumes and time-points used. An important feature of this procedure is that the starch-amylase ratio has been kept constant for all food products tested such that any differences observed reflect the starch digestibility of the food rather than their variable starch contents.

In brief, freshly prepared food materials (see section 2.1 for source and preparation) were weighed into 15 mL corning tubes and suspended in 10 mL phosphate buffered saline (PBS, pH 7.4) so that each tube contained 100 ± 2 mg starch. Sample tubes were mixed for 20 min at 37 °C on a Stuart SB3 rotary mixer (20 rpm, 30° angle) inside an incubator (E24 Excella, New Brunswick Scientific) to equilibrate. Before starting the assay, a 200 μL 'blank' aliquot of each sample was taken into a 1.5 mL microcentrifuge tube containing 200 μL 'stop solution' (0.3 M Na_2CO_3 , pH 9). The 0.3 M Na_2CO_3 , referred to as 'stop solution', inactivates the amylase activity in the aliquots and is used to promptly stop the amylolysis at each time point. To start the assay, porcine pancreatic α -amylase (EC 3.2.1.1 supplied in DFP-treated suspension of 2.9 M NaCl containing 2 mM CaCl_2 , A6255, Sigma-Aldrich Co. Ltd, Poole, UK) prepared in PBS (pH 7.4 at 37 °C) was added to achieve an activity of 4 U mL^{-1} in the digestion mixture (*i.e.*



Table 1 Source and preparation of food materials

Sample ^a	Method of preparation	Starch g per 100 g ^b	Comparator ^c
Biscuits			
Fullkorn digestive (United Biscuits, Sweden) ^d	Ground with pestle and mortar and sieved to collect 0.25–0.5 mm particles	39.0	Average of Digestives (UK) and Digestives (Canada). REF 630; 631
Water cracker (United Biscuits, UK)		72.3	Average of water cracker (Canada) and water cracker (Arnotts, AU). REF 740; 741
Cornish wafer (United Biscuits, UK) ^d		55.7	Average of Cream cracker (LU Triumpho, Brazil), and Jatz plain salted cracker biscuits (Arnotts, AU). REF 718; 720
Oatcake (Nairn's UK)		55.5	Average of Oat biscuit (UK); (China); Nairn's ginger; Oatmeal (Canada); Highland Oatmeal (Weston, AU). REF 676; 677; 678; 679; 641
Rich tea biscuit (Lyons, UK)		53.4	Average of Rich Tea (UK) and Rich Tea (Canada). REF 699; 700
Breads			
Bread; sunflower & barley (Vogel's UK)	Blended into bread crumbs	37.4	Sunflower and barley bread (Vogel's, UK). REF 111
Bread; wholemeal (Tesco, UK)		34	Mean of 10 studies on wholemeal (whole wheat) wheat flour bread. REF 234–243
Bread; white (Tesco, UK)		42.8	Mean of 16 studies on white wheat flour breads. REF 172–187
Breakfast cereals			
Cornflakes (Kellogg's, UK)	Ground with pestle and mortar and sieved to collect 0.25–0.5 mm particles	76	Mean of five studies on cornflakes. REF 321–325
Rice pops (Sainsbury's Supermarket Ltd, UK)		70.6	Rice Pops™ (Sainsbury's, UK), with semi-skimmed milk. REF 468
Balance (Sainsbury's Supermarket Ltd, UK)		69.6	Balance™ breakfast cereal (Sainsbury's, UK). REF 304
Pasta			
Spaghetti (Napolina)	Added to slightly salted boiling water for 9–11 min, ground with pestle & mortar	30.5 ^c	Boiled in 0.7% salted water for 11 min. REF 1375
Potatoes			
King Edward Potato (Sainsbury's Supermarket Ltd, UK)	Peeled, cut, added to boiling water, covered and simmered 15–20 min moderate heat, mashed	16.7	King Edward potato, peeled, quartered, boiled 15 min in unsalted water (UK). REF 1639
Charlotte potatoes (Sainsbury's Supermarket Ltd, UK)		16.7	Charlotte, peeled, quartered, boiled 15 min (UK). REF 1634
Pulses			
Chickpea-canned in water (Sainsbury's Supermarket Ltd, UK)	Drained, ground with pestle and mortar	16	Chickpeas, canned, drained, Edgell's™ brand. REF 1096
Brown Lentils-canned in water (Napolina)		9.9	Lentils, brown, canned, drained, Edgell's™ brand. REF 1113
Butter beans-canned in water (Sainsbury's Supermarket Ltd, UK)		14.3	Butter beans, canned, drained, Edgell's™ brand. REF 1089
Rice			
Long grain white rice-easy cook, parboiled (Sainsbury's Supermarket Ltd, UK)	Added to boiling water, 15–18 min moderate heat, drained well, ground with pestle and mortar	24.9	Long grain, parboiled 10 min cooking time Uncle Ben's, Masterfoods (Belgium). REF 522
Vegetables			
Sweet potatoes (Sainsbury's Supermarket Ltd, UK)	Peeled, cut, added to boiling water, covered and simmered 8–10 min moderate heat, mashed	8.9 ^d	Mean of boiled sweet potato. REF 1684; 1685; 1686; 1692
Garden peas-frozen (Birds Eye)	Boiled from frozen; added boiling water to cover, covered and simmered 3 min, ground and blended	2.6	Pea, frozen, boiled (Canada). REF 1611

^a Sample details; manufacturer listed in brackets. ^b Starch content g per 100 g as weighed, sourced from nutrition declaration. ^c McCance & Widdowson entries 11–722. ^d McCance & Widdowson entries 13–551. ^e Comparator products with reference numbers ('REF') as listed in Table A1 'Glycemic index and glycemic load values determined in subjects with normal glucose tolerance: 2008' in Atkinson *et al.* 2008.³



containing 10 mg mL⁻¹ starch). This enzyme–substrate ratio was chosen on the basis of results obtained with previous studies^{12,15} taking into account the range of starch susceptibility to digestion expected for the type of foods included within the study. One unit is defined as the amount of amylase needed to liberate 1.0 mg of maltose from starch in 3 min at pH 6.9 at 20 °C. Tubes were promptly returned to the mixer in the incubator after addition of amylase and incubated at 37 °C in the mixer for the duration of the digestion. After 5, 10, 15, 20, 25, 30, 45, 60, and 90 min, aliquots (200 μL) were collected from the digestion mixture into an equal volume of the stop solution. Aliquots were centrifuged at 15 000g for 5 min (Haraeus Pico, Thermo Scientific) to exclude any starch remnants and the supernatants stored at -20 °C for subsequent analysis of starch amylolysis products.

Reducing sugar analysis (PAHBAH assay)

The PAHBAH assay¹⁶ was subsequently used to determine the concentration of reducing sugars produced from starch amylolysis in aliquots recovered at the various time points. Stored supernatants were appropriately diluted (typically 1:10) in deionised water, and 100 μL of the diluted sample transferred to a 1.5 mL Eppendorf® safe-lock™ tube, to which was added 1000 μL freshly prepared ‘PAHBAH working reagent’ (250 mg *p*-hydroxybenzoic acid hydrazide dissolved in 4.75 mL of 0.5 M HCL, and made up to 50 mL with 0.5 M NaOH). These tubes were vortex mixed and secured in a Nalgene® floating rack and incubated at 100 °C for 5 min, then equilibrated for 10 min at room temperature before transfer to cuvettes and absorbance measurement ($\lambda = 405$ nm) in a spectrophotometer (Biochrom Libra). Standards containing known concentrations of maltose (0–900 μM) were prepared in 1.5 mL Eppendorf® safe-lock™ and reacted with PAHBAH reagent as described above, and these were included with every analysis. It is noteworthy that this assay can also be performed on a 96-well plate as described in ref. 15. Reducing sugar concentration in each sample was expressed as maltose equivalents by reference to the standard curve.

Data processing

Data obtained from the PAHBAH analysis was further processed to correct for endogenous reducing sugars present within the sample prior to addition of α -amylase.

For ease of interpretation, data expressed in maltose equivalent concentration units was converted to show the corresponding percentage of starch digested during amylolysis according to eqn (1):

$$(\text{Starch amylolysis } \%)_t = \frac{[\text{maltose}]_t}{[\text{maltose}]_{\text{substrate}}} \times 100 \quad (1)$$

in which $[\text{maltose}]_t$ is the maltose equivalent concentration (after baseline correction) measured in the liquid phase of the reaction mixture at a time point t , and $[\text{maltose}]_{\text{substrate}}$ is the theoretical maltose equivalent concentration that would be present at the start of the reaction, assuming that all starch within the food sample can be converted to maltose.

It is noteworthy that the conversion (eqn (1)) assumes that all amylolysis of starch yields maltose, and that this approach does not precisely account for minor products of starch amylolysis (glucose, α -limit dextrins and maltodextrins) thereby resulting in a net underestimation of total starch amylolysis. Expressing starch amylolysis relative to a highly digestible, high glycaemic index reference may provide absolute values that relate to GI (see Hydrolysis Index in section 2.5).

Starch digestibility and indices of glycaemic starch

Starch digestibility curves showing starch amylolysis progress over time were produced for each food material and used to derive the various starch digestibility indices described below and summarised in Fig. 1.

The enzyme-kinetic parameters C_∞ , which represent the product concentration at the reaction end-point, and k , which is the digestibility rate constant, can be used to describe starch amylolysis according to a first-order reaction (eqn (2)). These values were obtained using Logarithm of slope analysis, as described previously.^{12,13}

$$C_t = C_\infty(1 - e^{-kt}) \quad (2)$$

in which the C_t is the product concentration at a given time, t , C_∞ is product concentration at the reaction end-point, and k is the digestibility rate constant.

The terms C_{20} , C_{60} , and C_{90} represent the extent of starch digested after 20, 60, 90 min and were obtained from the starch digestibility data. The selection of specific time points for comparison is rather arbitrary, yet this approach has been used previously to describe starch digestibility. In the present study the amylase-starch ratio has been kept constant for all samples to enable direct comparisons.

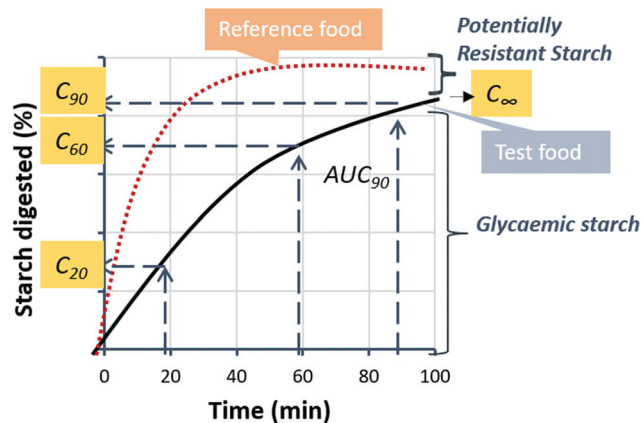


Fig. 1 Schematic of starch digestibility indices. One starch digestibility curve provides for rapid insight into glycaemic and potentially resistant starch fractions. The indices C_{20} , C_{60} and C_{90} correspond to the proportion of starch digested by α -amylase after 20, 60 and 90 min and are obtained directly from the starch digestibility curve. C_∞ is the proportion of starch that is digested at the endpoint of the reaction and is obtained together with the rate constant (k ; not shown) by applying logarithm of slope analysis to digestibility data. The incremental area under the curve (iAUC) is used in the calculation of Hydrolysis Index.



The Hydrolysis index (HI) is the area under the digestibility curve up to 90 min (iAUC90) of the test sample expressed relative to a highly digestible reference food as shown in eqn (3). In this case, boiled King Edward potato was used as the reference food as this gave consistently the highest starch digestibility value out of all the foods examined.

$$HI = \frac{iAUC90_{\text{test}}}{iAUC90_{\text{reference}}} \times 100 \quad (3)$$

Statistical analyses

Digestibility curves were plotted using SigmaPlot 14.0 software, and fitted to the first-order equation using non-linear regression. The Area Under Curves was obtained using the Area Below Curves Macro which is available from the Toolbox supplied within the Sigma Plot software. All values are presented as means of assays performed in triplicate with error bars as 95% CIs or SEM as specified in text and figure legend. *In vitro* data was compared statistically with published values from *in vivo* studies of matched food products taken from Atkinson *et al.* 2008,³ using values for healthy subjects with glucose (GI = 100) as the reference. Sources of *in vivo* data are fully referenced and details of any discrepancies between products are highlighted in the results table (Table 1). Statistical analyses were performed using IBM SPSS Statistics version 22. Pearson's correlation test (co-efficient denoted r) was used to test the correlation between absolute GI values and in *in vitro* indices,

whereas Kendalls Tau b (co-efficient denoted T_b) test was used to test the rank correlation between food ranked based on *in vitro* starch digestibility indices and *in vivo* Glycaemic Index data.

Results

Starch digestibility curves are shown for selected products in Fig. 2. Based on GI values³ of the food products shown in Fig. 2, the breakfast cereals (Fig. 2A) and breads (Fig. 2B) would be classes as 'high GI (GI \geq 70) food', rice, spaghetti and charlotte potato (Fig. 2C) would be 'medium GI (GI > 55 and <70) food' and the pulses (Fig. 2D) would be classed as 'low GI (GI \leq 55) food'. Within each of these categories, there are clear differences in the shape of the digestibility profiles of each food product. For instance, comparison of curves obtained for bread and breakfast cereals (all classed as high GI products) highlights how starch digestibility curves can reveal differences in starch digestion kinetics that are not detected in GI values or other digestibility indices. Food products with the lower GI were digested more slowly and some were still within a linear phase of digestion after 90 min. For these linear digestibility plots (observed for butter bean, chickpea, brown lentil, long-grain rice, sunflower-barley bread, and digestive biscuits), the resulting LOS plots (see example Fig. 3C) were characterised by a low rate constant ($k < 0.01$), with the implication that unreliable C_∞ values were calculated (data shown

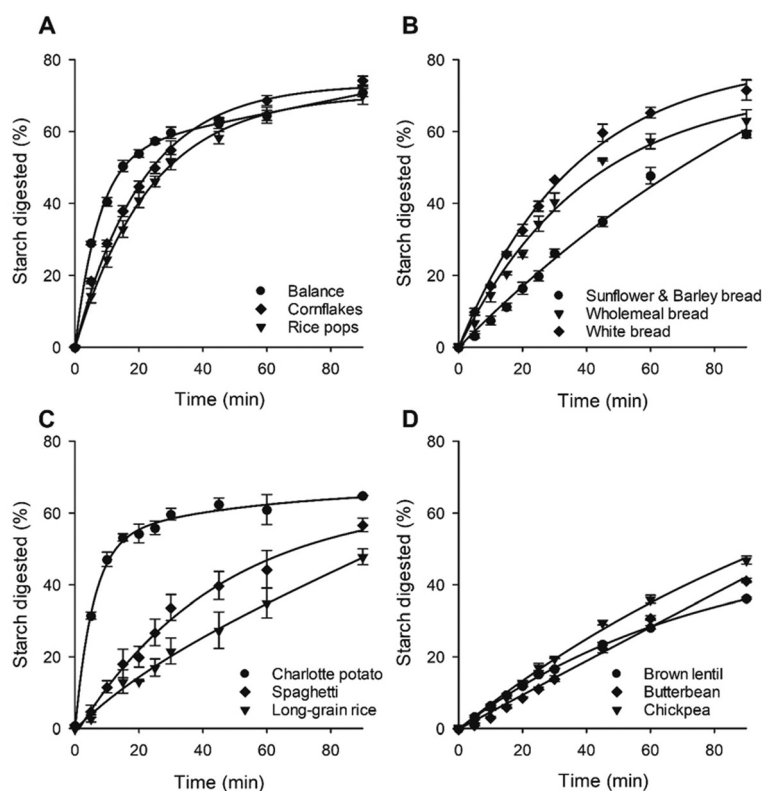


Fig. 2 Starch digestibility curves obtained for breakfast cereals (A); breads (B); potato, spaghetti and rice (C); and pulses (D). Values are means of triplicate analysis with error bars as SEM. Curve fits were obtained by maximum likelihood estimate regression to the first order equation.



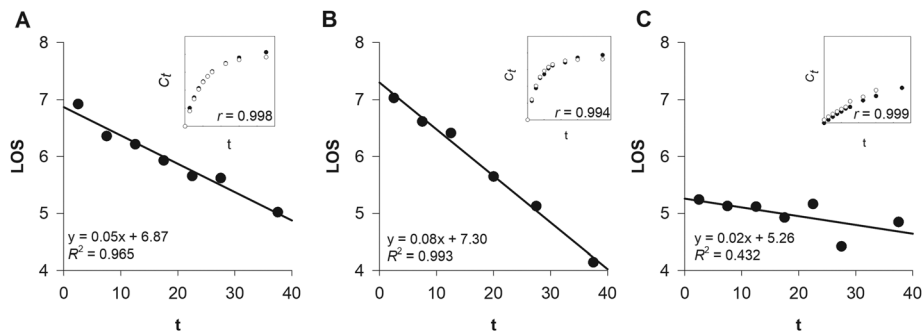


Fig. 3 Logarithm of Slope (LOS) plots for Cornflakes (A), Cream Crackers (B) and Brown Lentils (C). Inserts show calculated (filled circles) and experimental (open circles) data values for maltose concentration (C_t) over time (t). The calculated values are obtained from the first-order equation using the k and C_∞ values obtained from the LOS plot, and r (Pearson's r) is the correlation between predicted and experimental data as shown in the insert. R^2 is the co-efficient of determination obtained from linear regression in the LOS plot.

in parentheses, Table 2), as the digestion progress curves were essentially zero-order, and therefore not appropriate for 1st order kinetic analysis. This was a limitation of the data collected rather than the LOS analysis, and the C_∞ and k values for these 6 samples were therefore excluded from subsequent statistical analysis. For the remaining food products ($n = 14$), the digestibility data followed 1st order kinetics and was suitable for LOS analysis, and the LOS plots (Fig. 3A and B) were used to obtain the kinetic parameters C_∞ and k (see Table 2), which provided a good fit to experimental data collected at time points up to 90 min (Pearson's $r = \geq 0.97$).

An overview of the various starch digestibility indices is shown in Table 2. The *in vitro* indices obtained directly from the starch digestibility curves (C_{20} , C_{60} , C_{90} and HI) and the C_∞

obtained from LOS analyses (Fig. 3) were all significantly correlated ($p < 0.01$) with literature GI values, both in terms of rank (Kendalls Tau b test) and absolute values (Pearson's correlation). Scatter plots showing the relationship between *in vivo* and *in vitro* values are shown in OSM2. Out of all the indices compared, C_{90} was most strongly correlated with the absolute GI values ($r = 0.724$, $p < 0.001$). C_{90} and C_∞ were the most strongly correlated with *in vivo* rankings for GI of matched food products ($T_b = 0.596$, $p < 0.001$ and $T_b = 0.599$, $p < 0.01$, respectively). The only *in vitro* index that did not correlate with *in vivo* values was the rate constant, k obtained from LOS analysis. The value of k seemed to be inversely related with absolute GI values, however the relationship was not statistically significant ($r = -0.590$, $p > 0.1$).

Table 2 Starch digestibility indices^{a,b,c}

	GI	C_{20}	C_{60}	C_{90}	HI	C_∞	k
Cereal; cornflakes	81 ± 3	45 ± 1.5	69 ± 1.4	74 ± 1.2	89 ± 1	70	0.05
Cereal; rice pops	80 ± 6	41 ± 2.1	65 ± 2.2	70 ± 2.5	80 ± 2	67	0.048
Bread; white	75 ± 2	33 ± 1.6	65 ± 1.5	72 ± 2.8	72 ± 8	79	0.029
Potato; King Edward	75 ± 10	56 ± 3	75 ± 0.6	80 ± 3.4	100 ± 2	77	0.056
Bread; wholemeal	74 ± 2	26 ± 1	57 ± 2.1	63 ± 3	59 ± 10	68	0.03
Cereal; balance	74 ± 5	54 ± 1.1	65 ± 1.4	71 ± 1.1	92 ± 2	60	0.095
Biscuit; water	71 ± 10	18 ± 1.3	40 ± 1.9	47 ± 1.6	48 ± 4	71	0.015
Bread; sunflower-Barley	70 ± 10	16 ± 1.7	48 ± 2.3	59 ± 1	54 ± 2	(126)	(0.007)
Rice; long grain	68 ± 6	13 ± 0.1	35 ± 4.2	48 ± 2.2	40 ± 5	(89)	(0.007)
Potato; charlotte	66 ± 5	54 ± 2.6	61 ± 4.2	65 ± 0.1	78 ± 10	62	0.219
Biscuit; cream	60 ± 8	49 ± 0.9	67 ± 3	70 ± 0.7	90 ± 2	65	0.082
Pasta; spaghetti	59 ± 15	20 ± 2.9	44 ± 5.3	57 ± 1.9	38 ± 17	68	0.019
Vegetable; sweet potato	57 ± 5	25 ± 1	31 ± 2	32 ± 0.9	41 ± 4	29	0.075
Biscuit; oatmeal	53 ± 5	45 ± 1	57 ± 0.4	60 ± 0.5	73 ± 6	53	0.086
Vegetable; garden pea	51 ± 6	24 ± 1.1	35 ± 1	42 ± 0.2	62 ± 5	40	0.043
Biscuit; digestive	49 ± 6	15 ± 1	45 ± 0.4	53 ± 1.7	43 ± 5	(138)	(0.006)
Biscuit; tea	48 ± 5	19 ± 1.2	43 ± 0.6	50 ± 1.7	52 ± 1	73	0.018
Pulse; brown lentil	42 ± 5	12 ± 0.2	28 ± 0.5	36 ± 0.3	34 ± 1	(45)	(0.015)
Pulse; chickpea	38 ± 3	13 ± 0.9	36 ± 1.1	47 ± 1.1	42 ± 1	(105)	(0.007)
Pulse; butterbean	36 ± 3	9 ± 0.2	31 ± 0.6	41 ± 0.6	34 ± 1	(163)	(0.003)
Correlation with GI ^c	$T_b =$	0.517 ^d	0.581 ^d	0.596 ^d	0.504 ^d	0.599 ^d	0.033
	$r =$	0.573 ^d	0.721 ^d	0.724 ^d	0.641 ^d	0.704 ^d	-0.59

^a Values are mean ± SEM. ^b Numbers in brackets were identified as outliers because the experimental data was not appropriate for LOS analysis and were therefore excluded from statistical analysis. ^c Statistical tests: Kendalls Tau b, T_b and Pearson's correlation, r . ^d Correlation is significant at the 0.01 level (2-tailed).



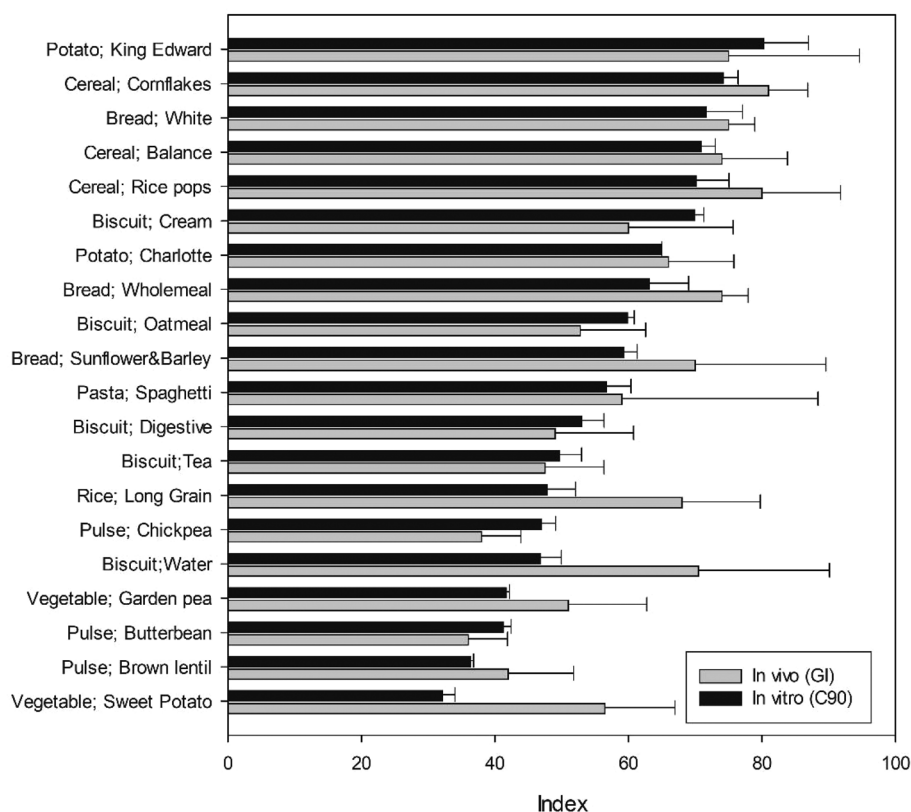


Fig. 4 *In vitro* values for C90 shown alongside *in vivo* GI values. *In vitro* values were obtained based on C₉₀ and *in vivo* values for matched food products ($n = 20$) were taken from the published literature. All values are means shown with error bars as 95% confidence intervals.

The C₉₀ (which correlated most strongly with GI), obtained *in vitro* is shown alongside *in vivo* GI values for corresponding food products in Fig. 4 and together with all *in vitro* indices in Fig. 5. It is noteworthy that the standard deviation of *in vivo* measurements was greater than those observed *in vitro*, and the *in vitro* values were within the 95% confidence interval of the *in vivo* values, with the exception of long grain rice, chickpea, water biscuit, wholemeal bread and sweet potato. It is plausible that this is due to difference in the food material (*e.g.*, processing, preparation, botanical source) rather than a limitation of the *in vitro* starch digestibility methodology *per se*. The *in vitro* C₉₀ values ranged from 32% to 80% starch digested, and corresponding GI values ranged from 36 to 81.

In terms of nutrient composition (OSM1) starch was the main carbohydrate (mean \pm SD = 60 \pm 18 g starch per 100 g dry weight) in 18/20 foods tested in the present study, the exceptions being sweet potato and garden peas, where the sugar content was higher than the starch content. The fat content was less than 5% (dry weight basis) for 14/20 foods, the exceptions being biscuits which contained up to 29% fat (dry weight basis). Similarly, protein content ranged from 5–26% of dry weight in the tested products, where the pulses and bread products were at the top end of this range. These compositional differences do not seem to systematically influence the reliability of GI predictions.

Discussion

In the present study, we used a relatively simple *in vitro* starch amylolysis assay to obtain indices of starch digestibility (C₂₀, C₆₀, C₉₀, HI, C_∞, and k) for 20 different food products with a known GI. Despite the simplicity of the *in vitro* method, a reasonable correlation was observed between *in vitro* indices and *in vivo* GI values, which demonstrates that the *in vitro* methodology shows potential for predicting relative differences in *in vivo* GI. Overall, the proposed assay methodology was well-suited to the broad range of starch-rich food products tested, and the low standard error and high sensitivity enabled differences in starch digestibility profiles of similar food products to be detected.

Starch amylolysis has long been known to influence glycaemic responses, and several studies have reported on such correlations previously.^{4,7,17,18} Out of all the indices tested in the present study, the extent of starch digested at 90 min (C₉₀) was most strongly correlated with GI and for most foods the *in vitro* value was within the range of *in vivo* values for matched food products. Reasonable predictions were achieved for products from all product categories. Notable exceptions were sweet potato and garden pea, in which the high sugar content (48 and 25 g sugar per 100 g dry weight) is likely to be contributing to the GI observed *in vivo*. The *in vitro* method also seemed to underestimate the GI of water biscuits, however



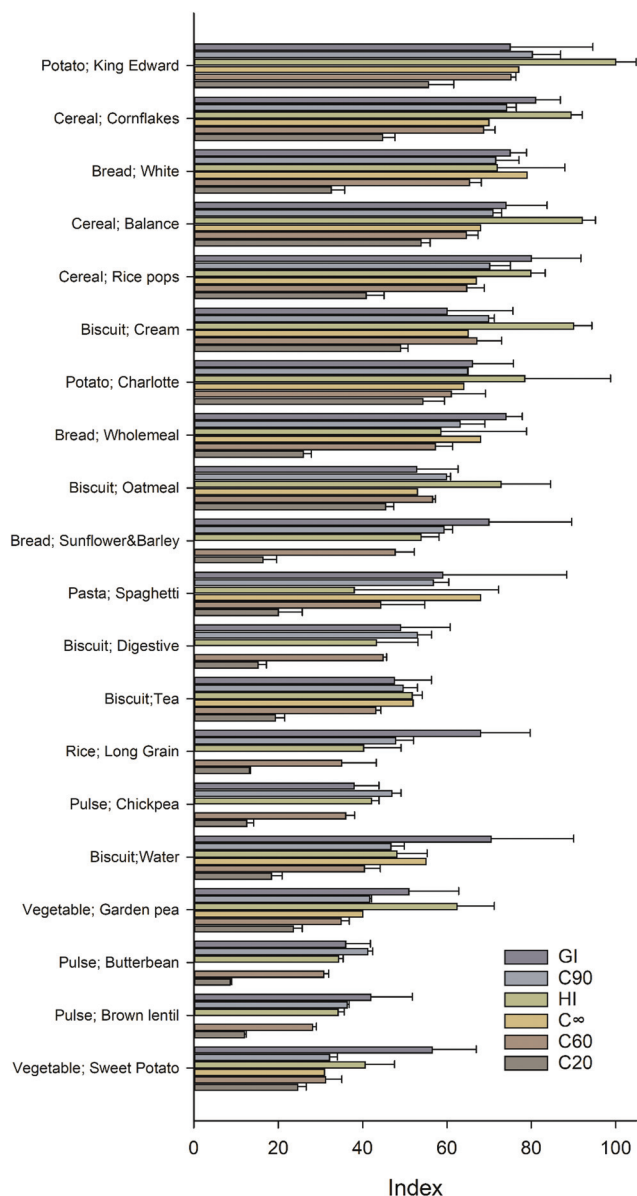


Fig. 5 All *in vitro* indices shown alongside *in vivo* GI values. *In vitro* values for C_{90} , HI, C_{∞} , C_{60} and C_{20} obtained from analysis of starch digestibility data and *in vivo* values for matched food products were taken from the published literature. Values are shown for all 20 food products, with the exception of C_{∞} which could not be obtained for some products (no bar shown for missing values). All values are means shown with error bars as 95% confidence intervals.

the product analysed *in vitro* was from a different manufacturer than those used in the *in vivo* studies, and the products that were compared may have had different characteristics. The GI values for long grain rice and wholemeal bread were also higher than the *in vitro* predictions. These products are particularly susceptible to retrogradation, and a difference in how these foods were handled and prepared for testing could explain this discrepancy. There was however, also a high variability in GI values reported for many of the food products, which may reflect inconsistencies in the characteristics of the

foods used. For example, cooking and cooling profiles are well known to have a major impact on starch structure and digestion kinetics in foods such as spaghetti, rice and potato^{11,19–21} and are a likely source of variability in GI values. A strength of the *in vitro* methodology is that it provides for systematic studies of such parameters and can thereby be used to understand the consequences of these factors, prior to *in vivo* trials.

We previously reported on the value of using LOS analysis of digestibility plots for mechanistic understanding of factors underpinning differences in starch digestibility^{12,13} and have recently applied this technique to examine the effect of particle size, cell wall encapsulation and starch structure on digestion kinetics.^{12,15,22} Within the present study we considered the use of C_{∞} and k (obtained from LOS plots) as a potential predictive index of GI. The theoretical basis for this was that C_{∞} reflects the proportion of starch that has the potential to be digested over an infinite time period and thereby resembles the potential glycaemic load, whereas k provides an indication of the rate at which starch-digestion products become available for absorption. In our dataset, the value of C_{∞} was reasonably correlated with GI values, however the value of the rate constant was not. This suggests that GI cannot be predicted from the rate constant alone, but requires both the rate and extent of digestion to be taken into account.

For many of the food products tested, the C_{∞} values were similar to C_{90} , meaning that the amylolysis reaction was nearing completion within the 90 min of the assay. This occurred mainly for the more rapidly digested products with a medium to high GI, and for these products C_{∞} and C_{90} both provided a reasonable estimation of GI. Obtaining reliable estimates of C_{∞} for some slowly digested foods (such as pulses, digestive biscuits and long grain rice) proved more challenging because the digestibility plots were linear and therefore unsuitable for LOS analyses. Although this limitation could be overcome by using a higher enzyme–substrate ratio for *in vitro* assays on low GI foods, it is unclear how relevant C_{∞} values would be to prediction of GI under these circumstances. The value of C_{∞} reflects the endpoint of starch amylolysis if it is allowed to progress over infinite time. However, *in vivo* the duration of starch exposure to pancreatic α -amylase in the intestinal lumen would occur over a limited time period.²³ Thus, C_{∞} might be expected to overestimate the glycaemic potency of starch in some slowly digested products. *In vivo*, the proportion of starch that escapes digestion in the upper gastrointestinal tract would be physiologically classed as resistant starch. Thus, the proportion of starch that has not been digested by α -amylase after a fixed time period can be considered as ‘potentially resistant’ starch,⁷ although the relationship between this *in vitro* indicator of resistant starch and the quantity of resistant starch that enters the colon has yet to be investigated.

In recent years, *in vitro* digestion models have become more advanced to provide a more physiological representation of digestive conditions, and now typically include an oral, gastric and intestinal phase to mimic the changing biochemical conditions (enzymic, pH) encountered during digestive



passage.^{5,24} In the present study, however, there was no oral or gastric phase prior to amylolysis, only one enzyme (porcine pancreatic α -amylase) was used and under a fixed enzyme–substrate ratio. This protocol resembled an enzyme-kinetic experiment rather than a simulation of physiological digestive conditions, with the advantage that low standard errors can be achieved. Overall, this is a user-friendly protocol with potential for high-throughput screening.

Users of sophisticated digestion models may be surprised by the predictive power of the ‘enzyme-kinetic’ approach used in this study. There may be an expectation that the inclusion of proteases and lipases is necessary to obtain a reliable view of starch digestion in real foods. One key consideration is that for many starch-rich staple foods, including most of those tested in the present study, the primary digestible component is starch. The starch digestibility screening method used in the present study is based on the premise that amylolysis is the rate-limiting step in the digestion process, and therefore has a major impact on glucose availability from food. Factors that are known to have a major impact on starch-susceptibility to digestion and thereby GI include processing conditions (*i.e.* which impact on starch molecular organisation)^{25–27} and physical properties of the food matrix (*e.g.*, particle size and permeability to digestive enzymes).^{9,10,15,18,28,29} Some properties (*e.g.*, starch crystallinity) can be reproduced by subjecting food materials to realistic cooking/processing conditions. Particle size and microstructure however is more challenging to reproduce *in vitro*. *In vivo* mastication produces a range of particle size distributions and significant variations exist between individuals and between food materials tested.³⁰ With the view of providing a starch digestibility screening tool and its likely applications, our preferred approach in this study was to control and standardise food structures used in the digestion. Particles were generally ground or sieved to a fixed size to preserve micro-scale structures and plant cells, although it is noteworthy that highly processed and refined starch-rich foods may not contain this level of structural integrity. This approach is more mechanistic than realistic, and users are encouraged to tailor the approach to address their research question.

One limitation of this study is that the *in vitro* and *in vivo* values were not obtained from exactly the same food products. As discussed, subtle differences in food preparation or composition can influence the digestibility and GI of foods. It remains possible that some of the observed discrepancies between *in vitro* and *in vivo* values were due to differences in the characteristics of the foods tested, rather than a limitation of the *in vitro* assay methodology.

Human GI studies are relatively expensive and time-consuming, and do not provide for a large number of products to be tested. Nevertheless, this study provides justification for a future validation study, in which a representative selection of well-characterised foods can be tested in parallel *in vitro* and *in vivo*.

Understanding the critical factors driving the rate of starch digestion in foods will help to inform and improve the design of food preparation protocols for *in vitro* testing and may also

aid researchers in identifying the most suitable protocol for addressing specific research questions. Although the single-enzyme (*i.e.* amylase only) system used in the present study provided reasonable predictions of the glycaemic index, it would not be expected to perform as well for foods where anti-nutritional inhibitors of glucose absorption are rate-limiting, or where co-digestion of fat and/or protein is necessary to expose starch to digestive fluids. It is currently unclear to what extent these mechanisms apply in different food products. Further investigations are needed to improve understanding of the mechanisms and rate-limiting steps that govern digestion of food and metabolic responses and this will require a combination of *in vitro* and *in silico* digestion models and *in vivo* data.

Conclusions

The *in vitro* method presented in this study provides for rapid and accurate comparison of starch digestibility and/or resistance. Starch digestibility indices obtained *in vitro* are significant correlated to GI values for matched food products and show potential for use as predictive indicators of GI. We envisage that the simple protocol presented will serve as a high throughput screening method to precede and complement more advanced *in vitro* and *in silico* digestion models and *in vivo* studies.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

The author(s) gratefully acknowledge the support of the Biotechnology and Biological Sciences Research Council (BBSRC); this research was funded by the BBSRC Institute Strategic Programme Food Innovation and Health BB/R012512/1 and its constituent projects BBS/E/F/000PR10345 (Theme 2, Digestion in the Upper GI Tract) and BBS/E/F/000PR10343 (Theme 1, Food Innovation).

References

- 1 D. J. A. Jenkins, C. W. C. Kendall, L. S. A. Augustin, S. Franceschi, M. Hamidi, A. Marchie, A. L. Jenkins and M. Axelsen, Glycemic index: Overview of implications in health and disease, *Am. J. Clin. Nutr.*, 2002, **76**, 266S–273S.
- 2 F. Brouns, I. Bjorck, K. N. Frayn, A. L. Gibbs, V. Lang, G. Slama and T. M. S. Wolever, Glycaemic index methodology, *Nutr. Res. Rev.*, 2008, **18**, 145–171.
- 3 F. S. Atkinson, K. Foster-Powell and J. C. Brand-Miller, International Tables of Glycemic Index and Glycemic Load Values: 2008, *Diabetes Care*, 2008, **31**, 2281–2283.



- 4 D. J. A. Jenkins, H. Ghafari, T. M. S. Wolever, R. H. Taylor, A. L. Jenkins, H. M. Barker, H. Fielden and A. C. Bowling, Relationship between rate of digestion of foods and postprandial glycaemia, *Diabetologia*, 1982, **22**, 450–455.
- 5 M. Minekus, M. Alminger, P. Alvito, S. Ballance, T. Bohn, C. Bourlieu, F. Carriere, R. Boutrou, M. Corredig, D. Dupont, C. Dufour, L. Egger, M. Golding, S. Karakaya, B. Kirkhus, S. Le Feunteun, U. Lesmes, A. Macierzanka, A. Mackie, S. Marze, D. J. McClements, O. Menard, I. Recio, C. N. Santos, R. P. Singh, G. E. Vegarud, M. S. Wickham, W. Weitschies and A. Brodkorb, A standardised static in vitro digestion method suitable for food - an international consensus, *Food Funct.*, 2014, **5**, 1113–1124.
- 6 A. Guerra, L. Etienne-Mesmin, V. Livrelli, S. Denis, S. Blanquet-Diot and M. Alric, Relevance and challenges in modeling human gastric and small intestinal digestion, *Trends Biotechnol.*, 2012, **30**, 591–600.
- 7 I. Goñi, A. Garcia-Alonso and F. Saura-Calixto, A starch hydrolysis procedure to estimate glycemic index, *Nutr. Res.*, 1997, **17**, 427–437.
- 8 K. N. Englyst, H. N. Englyst, G. J. Hudson, T. J. Cole and J. H. Cummings, Rapidly available glucose in foods: an in vitro measurement that reflects the glycemic response, *Am. J. Clin. Nutr.*, 1999, **69**, 448–454.
- 9 M. M. L. Grundy, C. H. Edwards, A. R. Mackie, M. J. Gidley, P. J. Butterworth and P. R. Ellis, Re-evaluation of the mechanisms of dietary fibre and implications for macronutrient bioaccessibility, digestion and postprandial metabolism, *Br. J. Nutr.*, 2016, **116**, 816–833.
- 10 J. Singh, A. Dartois and L. Kaur, Starch digestibility in food matrix: a review, *Trends Food Sci. Technol.*, 2010, **21**, 168–180.
- 11 J. Parada and J. M. Aguilera, Review: Starch matrices and the glycemic response, *Food Sci. Technol. Int.*, 2011, **17**, 187–204.
- 12 C. H. Edwards, F. J. Warren, P. J. Milligan, P. J. Butterworth and P. R. Ellis, A novel method for classifying starch digestion by modelling the amylolysis of plant foods using first-order enzyme kinetic principles, *Food Funct.*, 2014, **5**, 2751–2758.
- 13 P. J. Butterworth, F. J. Warren, T. Grassby, H. Patel and P. R. Ellis, Analysis of starch amylolysis using plots for first-order kinetics, *Carbohydr. Polym.*, 2012, **87**, 2189–2197.
- 14 Public Health England (2019) McCance & Widdowson's composition of foods integrated dataset. Available online: <https://www.gov.uk/government/publications/composition-of-foods-integrated-dataset-cofid>.
- 15 C. H. Edwards, M. Maillot, R. Parker and F. J. Warren, A comparison of the kinetics of in vitro starch digestion in smooth and wrinkled peas by porcine pancreatic alpha-amylase, *Food Chem.*, 2018, **244**, 386–393.
- 16 M. Lever, A new reaction for colorimetric determination of carbohydrates, *Anal. Biochem.*, 1972, **47**, 273–279.
- 17 Y. Granfeldt, I. Bjorck, A. Drews and J. Tovar, An in vitro procedure based on chewing to predict metabolic response to starch in cereal and legume products, *Eur. J. Clin. Nutr.*, 1992, **46**, 649–660.
- 18 K. A. Germaine, S. Samman, C. G. Fryirs, P. J. Griffiths, S. K. Johnson and K. J. Quail, Comparison of in vitro starch digestibility methods for predicting the glycaemic index of grain foods, *J. Sci. Food Agric.*, 2008, **88**, 652–658.
- 19 S. Wang and L. Copeland, Molecular disassembly of starch granules during gelatinization and its effect on starch digestibility: A review, *Food Funct.*, 2013, **4**, 1564–1580.
- 20 J. Parada and J. M. Aguilera, In vitro Digestibility and Glycemic Response of Potato Starch is Related to Granule Size and Degree of Gelatinization, *J. Food Sci.*, 2009, **74**, E34–E38.
- 21 M. Stuknyte, S. Cattaneo, M. A. Pagani, A. Marti, V. Micard, J. Hogenboom and I. De Noni, Spaghetti from durum wheat: effect of drying conditions on heat damage, ultrastructure and in vitro digestibility, *Food Chem.*, 2014, **149**, 40–46.
- 22 H. Patel, R. Day, P. J. Butterworth and P. R. Ellis, A mechanistic approach to studies of the possible digestion of retrograded starch by alpha-amylase revealed using a log of slope (LOS) plot, *Carbohydr. Polym.*, 2014, **113**, 182–188.
- 23 P. Layer and G. Gröger, Fate of Pancreatic Enzymes in the Human Intestinal Lumen in Health and Pancreatic Insufficiency, *Digestion*, 1993, **54**(suppl 2), 10–14.
- 24 J. W. Woolnough, J. A. Monro, C. S. Brennan and A. R. Bird, Simulating human carbohydrate digestion in vitro: a review of methods and the need for standardisation, *Int. J. Food Sci. Technol.*, 2008, **43**, 2245–2256.
- 25 R. F. Tester and W. R. Morrison, Swelling and gelatinization of cereal starches. I. Effects of amylopectin, amylose, and lipids, *Cereal Chem.*, 1990, **67**, 551–557.
- 26 C. H. Edwards, F. J. Warren, G. M. Campbell, S. Gaisford, P. G. Royall, P. J. Butterworth and P. R. Ellis, A study of starch gelatinisation behaviour in hydrothermally-processed plant food tissues and implications for in vitro digestibility, *Food Funct.*, 2015, **6**, 3634–3641.
- 27 S. M. Kingman and H. N. Englyst, The influence of food preparation methods on the *in vitro* digestibility of starch in potatoes, *Food Chem.*, 1994, **49**, 181–186.
- 28 S. Dhital, R. R. Bhattarai, J. Gorham and M. J. Gidley, Intactness of cell wall structure controls the in vitro digestion of starch in legumes, *Food Funct.*, 2016, **7**, 1367–1379.
- 29 A. Lovegrove, C. H. Edwards, I. De Noni, H. Patel, E. Sedef, T. Grassby, C. Zielke, M. Ulmius, L. Nilsson, P. J. Butterworth, P. R. Ellis and P. R. Shewry, Role of Polysaccharides in Food, Digestion and Health, *Crit. Rev. Food Sci. Nutr.*, 2017, **57**, 237–253.
- 30 G. M. Bornhorst and R. P. Singh, Bolus formation and disintegration during digestion of food carbohydrates, *Compr. Rev. Food Sci. Food Saf.*, 2012, **11**, 101–118.

