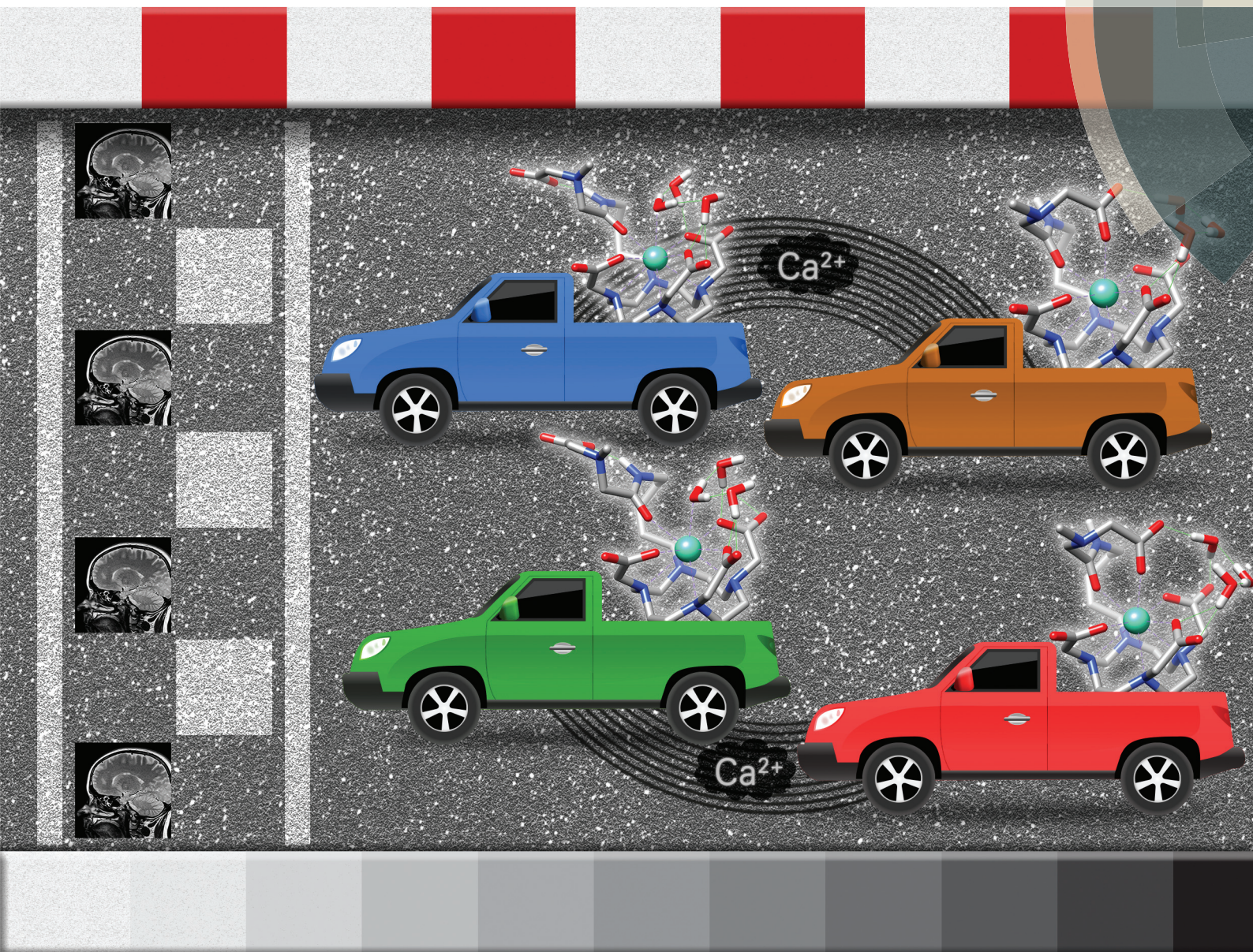


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Goran Angelovski *et al.*

Investigations into the effects of linker length elongation on the behaviour of calcium-responsive MRI probes



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Investigations into the effects of linker length elongation on the behaviour of calcium-responsive MRI probes†

Liam Connah,^a Vincent Truffault,^b Carlos Platas-Iglesias^{id}^c and Goran Angelovski^{id}^{*a}

Understanding the relationship between chemical structure and the effectiveness of bioresponsive magnetic resonance imaging (MRI) contrast agents can offer help to identify key components required for the future development of such probes. Here, we report the development and characterisation of two novel monomeric bifunctional chelators, **L**¹ and **L**², whose paramagnetic metal complexes can serve as calcium-responsive contrast agents. Specifically, relaxometric titrations, luminescence lifetime measurements, high resolution NMR and diffusion experiments, as well as density functional theory (DFT) calculations were carried out to assess the behaviour of each system. Minor structural differences between the probes resulted from the extension of the linker between the macrocyclic lanthanide chelator and the acyclic Ca-binding moiety. Relaxometric titrations of both systems, **GdL**¹ and **GdL**², showed an increase in *r*₁ and *r*₂ relaxivity upon Ca²⁺ addition, with the derivative bearing the longer linker showing a greater overall change. The hydration states of the europium analogues were assessed revealing a higher initial hydration state for **EuL**². Diffusion ordered NMR spectroscopy revealed negligible changes in the diffusive properties of both systems upon the addition of Ca²⁺, while NMR studies of the Y³⁺, Yb³⁺ and Eu³⁺ analogues provided further insights into the structural behaviour of the linker unit in both the unsaturated and Ca-saturated states. DFT calculations supported the different coordination modes of the studied paramagnetic complexes in the presence and absence of Ca²⁺. Overall, our findings demonstrate the impact of subtle changes to the structure of such probes, affecting a range of properties and their coordination behaviour.

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Introduction

Magnetic resonance imaging (MRI) is an essential diagnostic imaging technique used widely in clinics to provide three dimensional soft tissue images at high spatial resolution. In recent years, MRI has been a primary focus in the field of molecular imaging due to attempts to develop methods for the dynamic visualization of biological processes.^{1–4} The advancement of these so-called functional methods and the ability to monitor such changes can provide crucial information such as the function of tissues or organs in pathological states. Typically, MRI can be performed with or without the addition

of a contrast agent (CA) unlike other imaging techniques. The inclusion of such a CA in MRI can provide greater specificity through the shortening of the *T*₁ and *T*₂ relaxation times of the protons in the immediate vicinity of the CA. Differing from conventional CAs, ‘smart’ or bioresponsive smart contrast agents (SCAs) are suitable for functional MRI (fMRI) studies and have been applied to monitor different environmental states as their MR properties fluctuate depending on their microenvironment. Many SCAs have been developed for a variety of functions including the monitoring of pH, specific metal ions (Ca²⁺, Zn²⁺) and enzymes.^{1,5,6} Typically, they are based on cyclen scaffolds complexed with paramagnetic ions, and possess protonable groups (*i.e.* pH sensors), an additional specific chelator for a desired analyte (*e.g.* Ca²⁺, Zn²⁺) or a moiety which can be affected by enzyme activity.⁷ As the mechanism of action, usually SCAs are either considered as ‘off-on’ or ‘on-off’ systems, meaning they alter their relaxometric properties to provide a significant difference between two different environmental states.⁵ One of the preferred targets is Ca²⁺ due to its significance in biological processes.^{8,9}

^aMR Neuroimaging Agents, Max Planck Institute for Biological Cybernetics, 72076 Tuebingen, Germany. E-mail: goran.angelovski@tuebingen.mpg.de

^bMax Planck Institute for Developmental Biology, 72076 Tuebingen, Germany

^cCentro de Investigaciones Científicas Avanzadas (CICA) and Departamento de Química, Facultad de Ciencias, Universidade da Coruña, 15071 A Coruña, Spain

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These include a substantial involvement in critical cellular signalling events and an association with various pathologies including ischemia, Alzheimer's disease and epilepsy.^{10,11} Abilities to track Ca^{2+} fluctuations could prove useful in monitoring and understanding these pathological processes. Furthermore, due to the involvement of Ca^{2+} in neural activity, the development of Ca^{2+} sensing MRI probes could lead to the mapping of brain activity in a direct manner. Many attempts to develop Ca-responsive SCAs have been pursued over recent years resulting in the production of a variety of molecules ranging from smaller monomeric compounds to larger nano-sized derivatives.^{3,12,13} The desire to understand the relationship between SCA structure and relaxometric enhancement has brought about a series of studies investigating such effects, thus providing significant insights into the behaviour of these systems.^{14,15} Specifically, the interaction of the carboxylates from the Ca-binding chelate with the paramagnetic metal ion or the roles of the two stereoisomers, square antiprismatic (SAP) and twisted square antiprismatic (TSAP), of 1,4,7,10-tetraazacyclododecane-1,4,7-tris(methylenecarboxylic) acid (DO3A) macrocyclic chelators have revealed interesting coordination properties of these responsive systems. Furthermore, the influence of the distance between the MR reporting and bioresponsive units has shown to be significant in determining the relaxometric behaviour. Previous comparisons of bismacrocylic Ca-responsive SCAs with either an ethyl or propyl linker between the DO3A MR reporting unit and the EGTA-derived Ca^{2+} chelator revealed such an effect.¹⁶ The propyl derivative showed an increased relaxometric enhancement (*i.e.* increase in longitudinal r_1 relaxivity in the presence of Ca^{2+}) versus the ethyl version with slightly different starting relaxivities and hydration states of the lanthanide complex. A more advanced coordination study with ethyl and propyl linker SCA model compounds demonstrated further the significant impact of the linker distance on the coordination properties of the complex.¹⁷ Here, it was evaluated that the shorter ethyl linker is likely to be too close to the paramagnetic metal centre hindering the interaction of the carboxylate group with Gd^{3+} .

Recently we reported a study where a series of G4 polyamidoamine (PAMAM) dendrimers conjugated to monomeric SCAs exhibited significant differences in the observed relaxometric properties. Amongst the studied nano-sized conjugates with different monomers, those with butyl and pentyl linkers connecting the common DO3A- and EGTA-derived chelators showed the greatest changes and outlook for potential future application.¹⁸ Intrigued by these results, we embarked on a detailed structural study of these two newly developed monomeric systems. Firstly, we prepared different pairs of diamagnetic and paramagnetic complexes, which allowed the initiation of different types of coordination chemistry studies. Through the use of a variety of techniques such as 1D and 2D NMR, luminescence lifetime measurements, diffusion ordered NMR spectroscopy (DOSY), density functional theory (DFT) calculations and proton relaxometric titrations, we aimed to characterize the newly developed probes and gain an

insight into how subtle structural differences impact on their relaxometric and coordination behaviour.

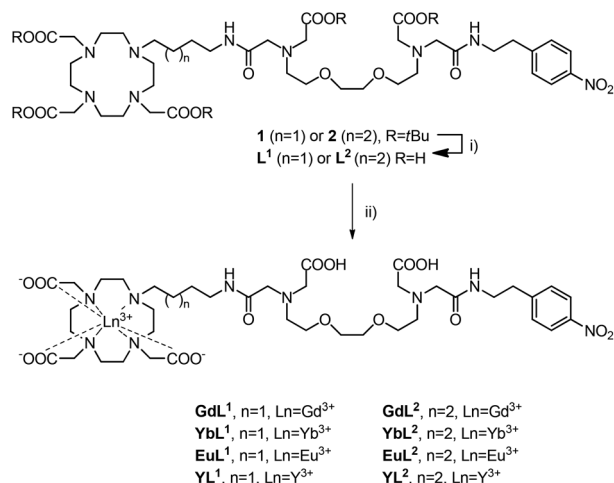
Results and discussion

Synthesis and complexation of ligands

Protected nitro compounds **1** and **2** were synthesized following previously reported procedures.¹⁸ Subsequent deprotection with formic acid at 60 °C overnight yielded ligands **L¹** and **L²** (Scheme 1). Complexation reactions of each ligand with either Yb^{3+} , Eu^{3+} , Gd^{3+} or Y^{3+} were all carried out under identical conditions. Specifically, the ligands **L¹** and **L²** were dissolved in water and the pH adjusted to 7 through the addition of a 0.1 M NaOH solution. A slight excess of the required lanthanide chloride hydrate was added while maintaining the pH at 7 through further addition of 0.1 M NaOH. After the final addition, the reaction mixture was stirred overnight at room temperature. The removal of excess lanthanide was achieved by repeated treatment of the solution with Chelex® before filtering and lyophilizing to yield the final complexes, **LnL¹** and **LnL²** ($\text{Ln} = \text{Gd}^{3+}$, Yb^{3+} , Eu^{3+} or Y^{3+}).

Relaxometric titrations

The relaxometric behaviour of both systems was assessed with proton longitudinal and transverse relaxometric titrations with **GdL¹** and **GdL²**. Each measurement was conducted with an identical initial concentration of Gd^{3+} (2.5 mM) at 25 °C and 7 T. The T_1 and T_2 relaxation times were recorded and the r_1 and r_2 relaxivities were calculated as a function of $[\text{Ca}^{2+}]$. For **GdL¹** with the shorter butyl linker, the starting r_1 was found to be $2.49 \text{ mM}^{-1} \text{ s}^{-1}$, which is slightly lower than that of the propyl derivative and other SCAs developed by us previously (Fig. 1a).^{13,19} Upon saturation with Ca^{2+} , r_1 increases to $4.96 \text{ mM}^{-1} \text{ s}^{-1}$ (99% increase). Concurrently, the change in r_2 over the same $[\text{Ca}^{2+}]$ range was found to increase to the same



Scheme 1 Synthesis of **L¹**, **L²** and the subsequent diamagnetic and paramagnetic complexes **LnL^{1,2}**. Reagents and conditions: (i) HCO_2H , 60 °C. (ii) LnCl_3 hydrate, H_2O , pH 7, RT, followed by Chelex®, RT.



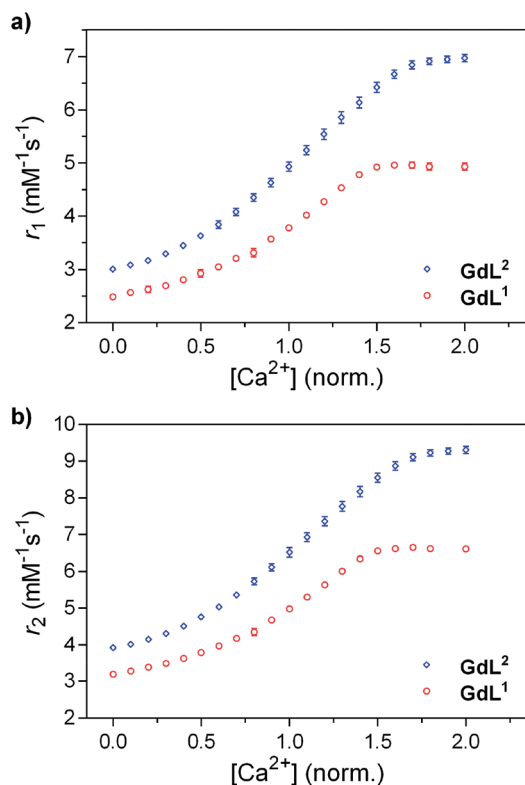


Fig. 1 Relaxometric titration curves for **GdL¹** and **GdL²** with Ca^{2+} : (a) longitudinal (r_1) and (b) transverse (r_2) titrations. Conditions: $[\text{Gd}^{3+}] = 2.5 \text{ mM}$, $\text{pH} = 7.4$ (50 mM HEPES), 25°C , 7 T .

relative extent, specifically from 3.19 to $6.62 \text{ mM}^{-1} \text{ s}^{-1}$ (Fig. 1b).

In comparison, **GdL²** gave an initial r_1 of $3.01 \text{ mM}^{-1} \text{ s}^{-1}$, which is in line with that of previously investigated SCAs (Fig. 1a).¹³ However, r_1 reached a value of $6.97 \text{ mM}^{-1} \text{ s}^{-1}$ (131% increase) upon saturation with Ca^{2+} . The change in r_2 was determined to be from 3.92 to $9.31 \text{ mM}^{-1} \text{ s}^{-1}$ (138% increase). Concurrently, both r_2/r_1 ratios remained constant at the studied Ca^{2+} concentrations, indicating that the formation of the **Ca-GdL^{1,2}** complex is likely not followed by any size changes between the paramagnetic species in the absence and presence of Ca^{2+} (Fig. S1 in ESI†).¹⁹ It can also be observed that the initial relaxivity values in the absence of Ca^{2+} are lower for **GdL¹** than the other complexes studied, indicating a lower hydration state of the complex (see below) and a changed coordination environment of the carboxylate from the EGTA-derived component. Clearly, the linker length between the MR reporting moiety and the bioresponsive unit has significant effects on the relaxometric behaviour of the SCA.

High resolution ^1H NMR studies

In order to understand and observe how the addition of Ca^{2+} impacts the structure and coordination environment of both derivatives, we conducted a series of high-resolution 1D and 2D NMR experiments at 800 MHz . Specifically, a series of diamagnetic and paramagnetic complexes were synthesised and

studied at 20 mM concentration and $\text{pD} 7.0$. ^1H -NMR, COSY and ^{13}C -HSQC of the complexes were recorded, the peaks were assigned and the behaviour of specific groups in the absence and presence of Ca^{2+} was followed.

We first assigned the relevant peaks that exhibited changes at different conditions (*i.e.* absence or presence of Ca^{2+}), excluding the resonances of the methylene groups neighbouring the amines of the EGTA-derived chelator. These were not observable in the recorded spectra, likely due to exchange processes related to the partial deprotonation of the amine groups at the studied pD .¹⁵ Furthermore, we noticed that the differences between the two states (without Ca^{2+} /with Ca^{2+}) exist only in the linker between the MR reporting and bioresponsive moiety; hence more detailed analysis followed only for this structural region. The ^{13}C -HSQC spectra of the diamagnetic derivatives (**YL¹** and **YL²**) were the most informative, with the obtained results then being compared to the paramagnetic complexes **EuL^{1,2}** and **YbL^{1,2}**. For the butyl derivative, **YL¹**, the closest (**A₁**) and furthest (**D₁**) methylene units to the macrocycle ring show a minor shift upon the addition of Ca^{2+} (Fig. 2). On the other hand, the 'inner' methylene units of the linker (**B₁** and **C₁**) show significant changes upon Ca^{2+} binding. Moreover, for both groups a single major signal is observable in the unsaturated state, which splits into two clear signals with equal intensity when saturated with Ca^{2+} . Exchanging Y^{3+} for paramagnetic ions (Eu^{3+} , Yb^{3+}) shows the same trends. This time, the signal splitting for **B₁** and **C₁** methylene groups upon Ca^{2+} binding is accompanied with the

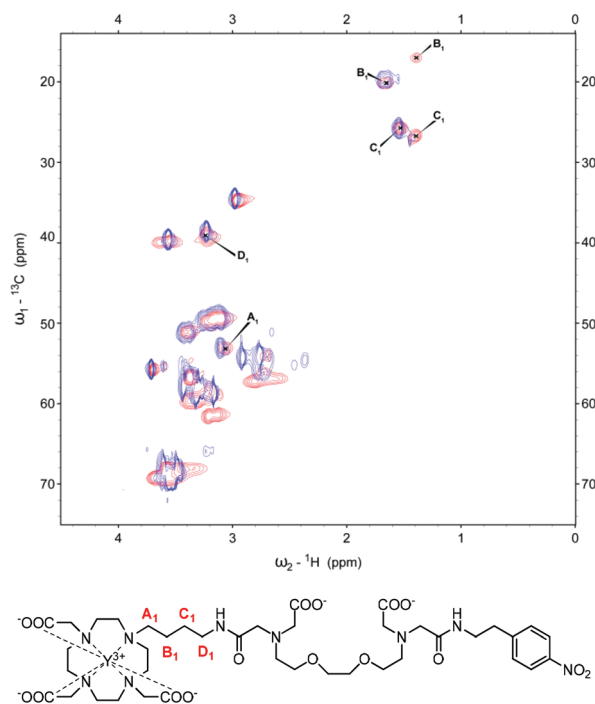


Fig. 2 ^{13}C -HSQC spectrum of **YL¹** with the signals of interest assigned pre- (blue) and post- Ca^{2+} (red) addition (top). Chemical structure of **YL¹** with the methylene units of interest labelled (bottom).



partial or complete signal loss of the **A**₁ methylene in **EuL**¹ and **YbL**¹, respectively (Fig. S2 in ESI†).

Similarly for **YL**², the CH₂ unit closest to the macrocyclic ring (**A**₂) shows a minor shift upon the addition of Ca²⁺ (Fig. 3). Moving further away from the macrocyclic ring, one signal is observable for the **B**₂ methylene group prior to Ca²⁺ addition, which then splits to form a new additional signal with lower intensity upon Ca²⁺ binding. **C**₂ methylene shows similar splitting behaviour to **B**₂ when Ca²⁺ binds, however both new signals are present with similar intensities. Moving towards the amide unit, the **D**₂ methylene group initially presents as two spots which then merge towards one with the addition of Ca²⁺, while the furthest methylene group in the linker, **E**₂, experiences a minor change. As in the case for the complexes with the butyl linker, analysis of paramagnetic complexes reveals loss of the signal for the **A**₂ methylene group in both **EuL**² and **YbL**² upon Ca²⁺ binding (Fig. S2 in ESI†). Obviously, the difference in the observed spectra of the studied peaks across the metals originates from their different paramagnetic effects. For example, the signals for **A**₁/**A**₂ methylene groups, which are observable in **YL**¹⁻² after the addition of Ca²⁺, disappear in the cases of Ca²⁺ addition to **EuL**¹⁻² and **YbL**¹⁻². We assume that a combination of the paramagnetic relaxation effect (shortening of *T*₁ and *T*₂ relaxation times) due to the proximity of the **A**₁/**A**₂ groups to the lanthanide and the increased conformational exchange leads to the overall signal loss that cannot be detected *via* NMR anymore.

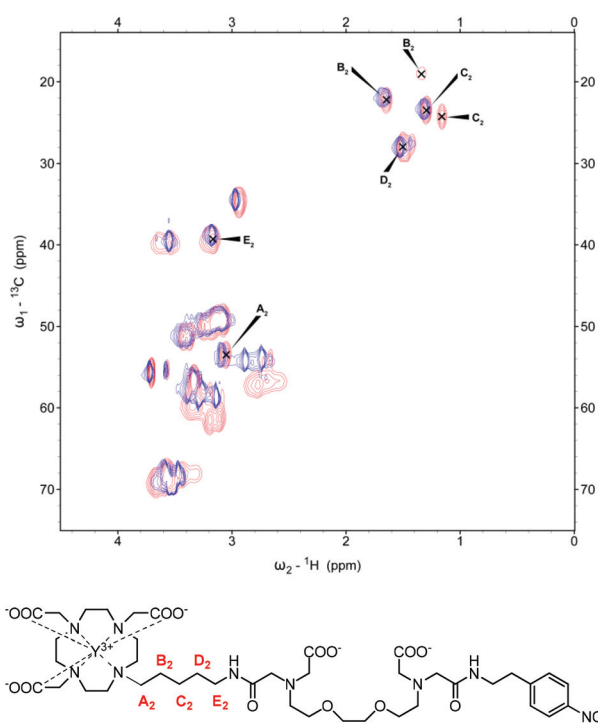


Fig. 3 ¹³C-HSQC spectrum of **YL**² with the signals of interest assigned pre- (blue) and post-Ca²⁺ (red) addition (top). Chemical structure of **YL**² with the carbon units of interest labelled (bottom).

More importantly, the signal splitting and shifting of peaks seen in the **B**_{1,2}, **C**_{1,2} and **D**₂ methylene groups suggest a substantial change in their environment between both the starting and saturated Ca²⁺ states. As these units are not in direct contact and should not participate directly in the binding of the added Ca²⁺, the movement of such signals indicates a change in the coordination behaviour of the complex. The mechanism of action for these Ca²⁺-responsive SCAs has previously been investigated.¹⁵ Specifically, in the absence of Ca²⁺, the closest carboxylate of the EGTA-derived moiety binds to the metal centre restricting water access. In the presence of Ca²⁺, this carboxylate preferentially binds to Ca²⁺ thus increasing the access of water to the complex. The results obtained here are in line with this theory, as it can be envisaged that the 'off-state' will present in a conformation which can be considered as a ring-like structure through which the EGTA-derived carboxylate is coordinated to the lanthanide of the macrocycle. Subsequent Ca²⁺ addition triggers the breaking of this arrangement due to its higher affinity for Ca²⁺ *vs.* Gd³⁺ to form a new structure/conformation, which is highlighted through the shifting and splitting of signals in the ¹³C-HSQC spectra. The observed changes of the signals from the linker methylene units indicate a noteworthy change in the coordination environment around the macrocyclic chelator between the two states and further evidences a conformational change in the responsive mechanism previously described. Moreover, the existence of two signals upon addition of Ca²⁺ supports the presence of two SAP and TSAP stereoisomers (see below). Since the signals of the C=O group of the amide could not be observed in any of the performed NMR experiments, we could not draw any conclusion on its fate, and whether this carbonyl does or does not coordinate to the metal centre. Indeed, the luminescence lifetime experiments revealed values that suggest an equilibrium of non- and mono-hydrated species (see below). Combined with the absence of NMR signals, our findings suggest that the carbonyl interacts with the metal ion throughout in either of the studied conditions (with or without added Ca²⁺).

The ¹H NMR spectra of the **EuL**¹ and **EuL**² complexes show paramagnetically shifted signals in the range ~–20 to 28 ppm (Fig. 4). The signals of the most shifted axial protons are observed between 20 and 28 ppm, a range that is typical of square antiprismatic coordination (SAP) around the metal ion.^{20–23} The signals due to the axial protons of the twisted square antiprismatic (TSAP) isomers are usually observed around 10–15 ppm. The resonances due to the TSAP isomer are barely visible in the case of **EuL**², while for **EuL**¹ the population of the TSAP isomer can be estimated to be 10–15%.

Interestingly, the NMR spectra of the **EuL**¹ and **EuL**² complexes are very similar, which indicates that they possess comparable magnetic anisotropies and thus coordination environments around the metal ion.²⁴ Furthermore, the spectra are characterised by rather well resolved signals, which indicates fairly rigid conformations in the Ca-free conditions and likely coordination of both the carboxyl group and the amide pendant arm to the metal ion. Indeed, Eu³⁺ complexes of



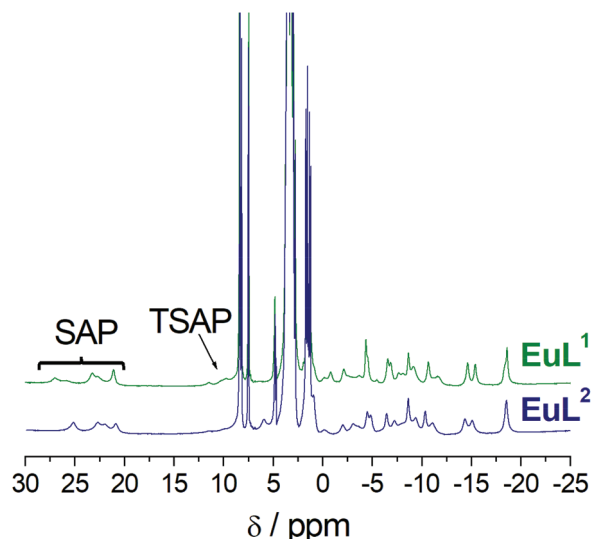


Fig. 4 ^1H NMR spectra of the EuL^1 and EuL^2 complexes recorded in D_2O solution (800 MHz, 25 $^\circ\text{C}$, pD 7).

DO3A derivatives generally give ^1H NMR spectra with broad signals.²⁵ Obviously, the spectra recorded for EuL^{1-2} suggest their atypical coordination mode with the paramagnetic metal ions, which supports the hypothesized existence of the coordination of the carboxyl and amide groups in absence of Ca^{2+} . On the other hand, addition of Ca^{2+} provokes severe broadening of the ^1H NMR resonances. This is in line with a more fluxional behaviour of the macrocyclic ring due to expected decoordination of the carboxylate and possibly partial decoordination or elongation of the coordination bond of the amide group (see below and Fig. S3 in ESI†).

NMR diffusion measurements

The diffusion of molecules can significantly describe their behaviour in solution and could potentially show any aggregation present. Diffusion coefficients for each derivative was therefore assessed prior to and after the addition of Ca^{2+} with DOSY on the Eu^{3+} analogues (EuL^1 and EuL^2) at three different concentrations (20, 10 and 5 mM). The diffusion coefficients obtained for the two systems studied here were both in agreement with similar monomeric species previously studied (Table 1).^{19,26}

Generally, decreasing the complex concentration led to a slight increase in diffusion coefficient which is naturally

expected. Additionally, the diffusion of EuL^2 was slightly slower than that of EuL^1 , which is logical due to the minor difference in their size and greater linker length in EuL^2 . When comparing samples of identical concentrations upon Ca^{2+} addition (2 equiv.), similar, or a slight increase in diffusion coefficient was also observed. This is indicative of a change in conformation expected upon Ca^{2+} binding and faster diffusion of these complexes in solution. These tiny observed changes in diffusion along with the absolute values indicate systems which interact with Ca^{2+} ; however, its addition leads to conformational changes that cause slightly faster diffusion of the species in solution for both studied complexes, while likely excluding the formation of aggregates.

Luminescence experiments

Luminescence emission lifetime and steady-state measurements were performed to assess the hydration states of the complexes and further understand the coordination environment of the lanthanide ion. As previously well established for this class of SCAs, the hydration of the inner sphere of the complex is expected to increase upon addition of the analyte, which is commonly the major cause for the enhancement in the relaxivity.^{14,27} However, the hydration change must not be the sole reason for the relaxivity enhancement, as the intramolecular hydrogen bonding or the deprotonation of proximate amines and the consequent changes in the prototopic exchange may additionally influence the amplitude of the relaxivity changes.^{17,28}

Measurements were carried out with the EuL^{1-2} (5 mM) analogues in both H_2O and D_2O and the hydration number (q) was calculated (Table 2) following a previously reported method.²⁹ Consequently, the calculated hydration numbers for both EuL^{1-2} complexes increase upon Ca^{2+} binding, complementary to that previously observed with other DO3A-based Ca-responsive SCAs.^{15,16} The length of the linker impacts the initial inner sphere hydration state of the complex. The shorter butyl derivative, EuL^1 , is less hydrated in the initial

Table 2 q values calculated for EuL^{1-2} with and without Ca^{2+} (2 equiv.)

	No Ca^{2+}			+ Ca^{2+} (2 equiv.)		
	$\tau_{\text{H}_2\text{O}}$ (ms)	$\tau_{\text{D}_2\text{O}}$ (ms)	q	$\tau_{\text{H}_2\text{O}}$ (ms)	$\tau_{\text{D}_2\text{O}}$ (ms)	q
EuL^1	0.726	1.142	0.3	0.584	1.159	0.7
EuL^2	0.652	1.140	0.5	0.539	0.982	0.7

Table 1 Diffusion coefficients calculated for EuL^1 and EuL^2 with and without Ca^{2+} (2 equiv.)

	Diffusion coefficient D ($10^{-10} \text{ m}^2 \text{ s}^{-1}$)					
	No Ca^{2+}			+ Ca^{2+}		
	20 mM	10 mM	5 mM	20 mM	10 mM	5 mM
EuL^1	1.76 ± 0.06	1.97 ± 0.01	1.99 ± 0.04	1.89 ± 0.02	2.01 ± 0.04	2.05 ± 0.06
EuL^2	1.76 ± 0.02	1.90 ± 0.04	1.95 ± 0.04	1.71 ± 0.01	1.91 ± 0.01	1.96 ± 0.02



state compared to that of **EuL**², which explains the initial r_1 relaxivity value observed in the relaxometric titrations previously discussed (Fig. 1). The increased initial hydration state of **EuL**² additionally indicates a lesser interaction between the EGTA-derived carboxylate and **Eu**³⁺ compared to **EuL**¹, possibly resulting from an increased distance between these two moieties. Otherwise, reasons involving an altered conformational arrangement or a change in ratio of the SAP and TSAP isomers could provide possible explanations. The luminescence steady-state emission spectra were in line with the determined q values – the initial emission signal intensity was greater for **EuL**¹, due to its lower hydration (Fig. S4 in ESI†); subsequently, the signal intensities for **EuL**¹ and **EuL**² equalized upon the addition of **Ca**²⁺.

DFT studies

DFT calculations were carried out in an attempt to rationalize the structure of the **GdL**^{1–2} complexes and their response to the presence of **Ca**²⁺ (see computational details in Experimental section). For the sake of computational simplicity, the structures of the complexes were truncated at the carbon of the ethylene bridge closer to the amide group (Fig. 5 and Fig. S5 in ESI†). The simplified models will be denoted as **GdL**¹ and **GdL**². Following previous studies,^{30,31} the optimized structures of the SAP isomers of **GdL**^{1–2} included three explicit

water molecules to improve the description of the **Gd**–O_{water} bonds. Geometry optimizations provided two energy minima for each complex in which either a water molecule or a carboxylate group is coordinating at the apical position of the SAP coordination polyhedron. Coordination of the amide oxygen atom takes place regardless of the length of the spacer or the concomitant coordination of the acetate group of the EGTA moiety at the apical position. The calculated **Gd**–O_{amide} distances are 2.405 and 2.418 Å for the monohydrated ($q = 1$) forms of **GdL**¹ and **GdL**², respectively.

Coordination of the carboxylate group at the apical position results in a significant lengthening of the **Gd**–O_{amide} bonds to 2.454 and 2.492 Å for **GdL**¹ and **GdL**², respectively. The longer **Gd**–O_{amide} distances calculated for the **GdL**² complex reflect a weaker coordination of the amide oxygen atom associated to the longer pentyl spacer. The energy differences between the calculated structures with $q = 1$ and $q = 0$ favour the apical coordination of the carboxylate group by $\Delta E_{\text{ZPE}} = 1.2$ and 0.93 kcal mol^{–1} for **GdL**¹ and **GdL**², respectively. The slightly smaller energy difference calculated for **GdL**² suggests that the higher relaxivity observed for **GdL**² in the absence of **Ca**²⁺ might be related to a higher contribution of the hydrated form of the complex. The DFT results are in line with our previous studies that indicated the coordination of a carboxylate group of the EGTA moiety at the apical position in the absence of **Ca**²⁺, resulting in a non-hydrated species. Addition of **Ca**²⁺ causes the de-coordination of this group, which gets involved in **Ca**²⁺ coordination, yielding a $q = 1$ complex. Thus, our DFT results indicate that extending the length of the spacer connecting the DO3A moiety and the amide group, by introducing butyl or pentyl linkers, does not alter the mechanism for **Ca**²⁺ response. Our results also support the coordination of the amide group both in the absence and in the presence of **Ca**²⁺, in line with the hydration numbers determined from luminescence lifetime measurements, which point to monohydrated complexes after **Ca**²⁺ binding. This was also supported in a number of previous studies by ¹⁷O NMR measurements,^{32–34} which provided hyperfine coupling constants A/h that confirmed the formation of mono-hydrated species. The higher relaxivities observed for the **Ca**²⁺-bound form of **GdL**² compared to **GdL**¹ are therefore not likely related by an increased number of coordinated water molecules, but perhaps to a more important contribution of second-sphere water molecules.

Conclusions

To conclude, we have designed, synthesised and characterised a new pair of **Ca**-responsive SCAs with altered linker lengths between the MR reporting and bioresponsive moieties. Relaxometric, NMR, DOSY, luminescence lifetime and DFT studies were performed to analyse and evaluate the coordination behaviour of both systems. Throughout the studies, the sensitivity of such molecules to discrete structural changes is apparent. As the linker length increased, the initial hydration

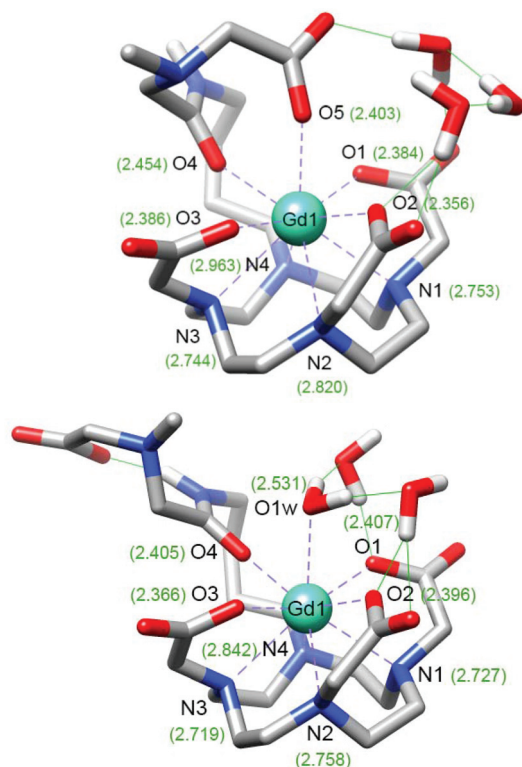


Fig. 5 Optimized structures of the **GdL**¹ system obtained with DFT calculations showing the coordination of the carboxylate group of the side chain (top) or the coordination of a water molecule (bottom). The numbers represent the calculated bond distances (Å) between the metal centre and the ligand donor atoms.



state of **EuL**² was observed to be higher than in **EuL**¹, resulting in a greater starting relaxivity. The subsequent change in relaxivity upon Ca²⁺ addition varied significantly depending on the linker length. Specifically, the overall *r*₁ and *r*₂ relaxivity change for **GdL**² was found to be 131 and 138% respectively; while for the shorter system, **GdL**¹, changes of 99 and 107% were observed for both *r*₁ and *r*₂ respectively. Results from the DOSY experiments, described a system in which negligible change in the diffusive behaviour of the complex between the initial and Ca²⁺ saturated states took place. High-resolution NMR studies provided further insights and validations to the responsive mechanism of these types of Ca-responsive probes previously reported. DFT calculations supported the existence of the postulated Ca-triggered coordination changes evidenced in the above described experiments.

Overall, here we have evaluated the coordination properties of two DO3A-based Ca-responsive systems with increased spacer distance and compared them to the previously employed derivatives for their suitability as scaffolds in the future development of Ca-responsive SCAs. The results of this study have provided important information into how subtle structural changes can impact the responsivity of such systems, aiding in the design of future monomeric SCAs in the pursuit of more effective Ca-responsive probes. Moreover, the investigated systems exhibited affirmative relaxometric properties that can be further exploited, making them promising candidates for the preparation and utilisation of multifunctional SCAs for fMRI applications.

Experimental section

General remarks

Commercially available reagents and solvents were used without further purification. Compounds **1** and **2** were synthesised following published procedures.¹⁸ High resolution mass spectra were recorded on a Bruker Daltonics APEX II (FT-ICR-MS) with an electrospray ionization source. Low resolution mass spectra were recorded on an Agilent ion trap SL 1100 system with an electrospray ionization source. All NMR, diffusion and relaxometric experiments were performed on Bruker Avance III 300 MHz and Avance III 800 MHz spectrometers at 298 K. Processing was performed using TopSpin 2.1 (Bruker GmbH), ACD/SpecManager 9.0 (Advanced Chemistry Development, Inc.) or Sparky 3.115. The NMR spectra were obtained in D₂O, using the deuterium lock frequency. The concentration of Gd³⁺, Eu³⁺ or Yb³⁺ in analysed solutions was determined using the bulk magnetic susceptibility shift (BMS).³⁵ The luminescence lifetime measurements were performed on a QuantaMaster™ 3 PH fluorescence spectrometer from Photon Technology International, Inc. (USA).

Synthetic procedures

General procedure for ligands L¹ and L². Protected compounds **1** or **2** were dissolved in formic acid and the mixture

was stirred at 60 °C overnight. The solvent was then removed under reduced pressure and the resulting residue was redissolved in water and lyophilized to give ligands **L**^{1–2} which were used without further purification (Fig. S6–8 in ESI†).

[4,7-Bis-carboxymethyl-10-(4-{2-[carboxymethyl-(2-{2-(carboxymethyl-{2-(4-nitro-phenyl)-ethylcarbamoyl]-methyl}-amino)-ethoxy]-ethoxy}-ethyl)-amino]-acetylaminobutyl)-1,4,7,10 tetra-aza-cyclododec-1-yl]-acetic acid (L¹). ¹H NMR (300 MHz, D₂O) δ (ppm): 1.45–1.81 (m, 4H), 2.92 (t, *J* = 6.2 Hz, 2H), 2.98–3.94 (br, 44H), 4.02 (s, 2H), 4.12 (s, 2H), 7.43 (d, *J* = 8.3 Hz, 2H), 8.10 (d, *J* = 8.3 Hz, 2H). ¹³C NMR (75 MHz, D₂O) δ (ppm): 20.7, 25.5, 34.6, 38.9, 40.1, 48.7, 49.8, 51.1, 53.6, 54.0, 54.9, 55.2, 55.8, 56.1, 57.0, 57.2, 64.4, 64.6, 69.8 (–CH₂–), 123.7, 130.1, 146.3, 147.6 (ArC), 165.0, 165.3, 169.5, 169.7, 170.3, 174.3 (C=O). **ESI-HRMS:** (*m/z*) [*M* + Ca–H]⁺ calcd for C₄₀H₆₄CaN₉O₁₆⁺, 966.4091, found: 966.4073.

[4,7-Bis-carboxymethyl-10-(5-{2-[carboxymethyl-(2-{2-(carboxymethyl-{2-(4-nitro-phenyl)-ethylcarbamoyl]-methyl}-amino)-ethoxy]-ethoxy}-ethyl)-amino]-acetylaminopentyl)-1,4,7,10 tetra-aza-cyclododec-1-yl]-acetic acid (L²). ¹H NMR (300 MHz, D₂O) δ (ppm): 1.22–1.82 (m, 6H) 2.62–4.20 (br, 50H), 7.44 (d, *J* = 7.9 Hz, 2H), 8.12 (d, *J* = 7.7 Hz, 2H). ¹³C NMR (75 MHz, D₂O) δ (ppm): 22.6, 23.2, 27.7, 34.6, 39.2, 40.1, 48.5, 49.9, 51.3, 53.8, 54.0, 54.9, 55.2, 55.8, 56.0, 56.2, 57.0, 57.2, 64.4, 64.6, 69.8 (–CH₂–), 123.7, 130.1, 146.3, 147.6 (ArC), 165.0, 165.1, 169.4, 169.7, 170.1, 174.5 (C=O). **ESI-HRMS:** (*m/z*) [*M*–3H + Ca][–] calcd for C₄₁H₆₄CaN₉O₁₆[–], 978.4102, found: 978.4113.

General procedure for the synthesis of LnL^{1–2} complexes. Ligands **L**¹ or **L**² were dissolved in water and the pH adjusted to 7 by addition of a 0.1 M NaOH solution. A slight excess of lanthanide chloride (1.1 equiv.) was then added portionwise while maintaining the pH at 7 with 0.1 M NaOH. After the final addition of lanthanide, the reaction was stirred at room temperature overnight. To ensure the removal of excess lanthanide, Chelex® was added to the solution and stirred for 30 min. After this time, the sample was filtered and another portion of Chelex® added to the filtered solution. This was repeated once more (full procedure involved three applications of Chelex®). After the final treatment, the solution was filtered and lyophilised to yield the final complex (Fig. S9–17 in ESI†).

NMR

YL¹. ¹H NMR (300 MHz, D₂O): δ (ppm) 1.33–1.83 (br), 2.23–4.08 (br), 7.46 (d), 8.16 (d). ¹³C NMR (75 MHz, D₂O): δ (ppm) 19.7, 25.8, 34.5, 38.5, 39.5, 46.4, 46.8, 49.3, 49.5, 49.7, 51.1, 51.1, 51.2, 53.0, 54.0, 54.3, 54.3, 54.6, 55.2, 55.3, 55.3, 55.7, 56.2, 56.8, 57.9, 58.2, 58.3, 58.7, 58.7, 65.7, 65.8, 65.9, 67.9, 68.1, 69.5, 69.5 (–CH₂–), 123.8, 130.1, 147.7 (ArC), 171.1, 173.5, 173.6, 173.8, 178.3, 178.5 (C=O).

YL². ¹H NMR (300 MHz, D₂O): δ (ppm) 0.95–1.93 (br), 2.22–4.30 (br), 7.45 (d), 8.15 (d). ¹³C NMR (75 MHz, D₂O): δ (ppm) 21.9, 23.3, 28.0, 34.5, 38.7, 39.5, 49.2, 49.2, 49.7, 49.9, 51.2, 51.2, 53.5, 54.0, 54.3, 55.3, 55.6, 56.7, 58.0, 58.2, 58.7, 58.7, 67.9, 68.2, 68.5, 69.5, 69.5 (–CH₂–), 123.7, 130.0, 147.7 (ArC), 170.7, 173.2, 173.3, 173.6, 173.7, 177.6, 178.1, 178.3 (C=O).



ESI-LRMS

GdL¹. (*m/z*) [M – H][–] calcd for C₄₀H₆₁GdN₉O₁₆[–], 1081.3, found: 1081.3. [M + Na–2H][–] calcd for C₄₀H₆₀GdN₉NaO₁₆[–], 1103.3, found: 1103.3.

EuL¹. (*m/z*) [M – H][–] calcd for C₄₀H₆₁EuN₉O₁₆[–], 1076.3, found: 1076.3. [M + Na–2H][–] calcd for C₄₀H₆₀EuN₉NaO₁₆[–], 1098.3, found: 1098.3.

YbL¹. (*m/z*) [M – H][–] calcd for C₄₀H₆₁YbN₉O₁₆[–], 1097.4, found: 1097.3. [M + Na–2H][–] calcd for C₄₀H₆₀YbN₉NaO₁₆[–], 1119.3, found: 1119.3.

YL¹. (*m/z*) [M – H][–] calcd for C₄₀H₆₁YN₉O₁₆[–], 1012.3, found: 1012.4. [M + Na–2H][–] calcd for C₄₀H₆₀YN₉NaO₁₆[–], 1034.3, found: 1034.3.

GdL². (*m/z*) [M – H][–] calcd for C₄₁H₆₃GdN₉O₁₆[–], 1095.4, found: 1095.4. [M + Na–2H][–] calcd for C₄₁H₆₂GdN₉NaO₁₆[–], 1117.4, found: 1117.3.

EuL². (*m/z*) [M – H][–] calcd for C₄₁H₆₃EuN₉O₁₆[–], 1090.4, found: 1090.4. [M + Na–2H][–] calcd for C₄₁H₆₂EuN₉NaO₁₆[–], 1112.3, found: 1112.3.

YbL². (*m/z*) [M – H][–] calcd for C₄₁H₆₃YbN₉O₁₆[–], 1111.4, found: 1111.4. [M + Na–2H][–] calcd for C₄₁H₆₂YbN₉NaO₁₆[–], 1133.4, found: 1133.3.

YL². (*m/z*) [M – H][–] calcd for C₄₁H₆₃YN₉O₁₆[–], 1026.3, found: 1026.3. [M + Na–2H][–] calcd for C₄₁H₆₂YN₉NaO₁₆[–], 1048.3, found: 1048.3.

Relaxometric titrations

Proton longitudinal and transverse relaxometric titrations with Ca²⁺ were performed at 7.0 T, 298 K and pH 7.4 (50 mM HEPES buffer) using inversion recovery (*T*₁) and Car–Purcell–Meiboom–Gill (*T*₂) pulse sequences. To a solution of **GdL^{1–2}** (2.5 mM), a CaCl₂ solution of known concentration was added stepwise and *T*₁ and *T*₂ measurements were performed after each addition. The longitudinal and transverse relaxivities, *r*₁ and *r*₂, were then calculated using eqn (1), where *T*_{1,obs} is the measured *T*₁ (*i* = 1) or *T*₂ (*i* = 2), *T*_{*i*,d} is the diamagnetic contribution from the solvent and [Gd] is the actual concentration of Gd³⁺. A total of three independent measurements were recorded for each Gd³⁺ complex and the results averaged at each point for the titration to give the final relaxivity values.

$$\frac{1}{T_{i,\text{obs}}} = T_{i,\text{d}} + r_i \times [\text{Gd}]. \quad (1)$$

High resolution NMR experiments

All high resolution NMR spectra were acquired on Bruker AVIII-800 spectrometer, processed with TopSpin 2.1 (Bruker GmbH), and analyzed with Sparky 3.115.³⁶ For every sample (20 mM [Ln³⁺]), a ¹H-NMR spectra was measured, as well as a DQF-COSY and a ¹³C-HSQC. A standard DQF-COSY³⁷ was acquired with 4096 and 128 complex points in *t*₂ and *t*₁, respectively, performing 256 scans per increment.

The ¹³C-HSQC was recorded at a resolution of 1024 (*t*₂) × 128 (*t*₁) complex points, using 64 scans per increment. All

experiments were recorded at 298 K, using a relaxation delay of 1.5 s. The water suppression was achieved *via* pre-saturation for the ¹H-NMR and the COSY, while the ¹³C-HSQC uses gradient selection (echo/*anti*-echo).

NMR diffusion measurements

Samples of **EuL^{1–2}** with and without Ca²⁺ (2 equiv.) were dissolved in D₂O and the pD adjusted to 7.0. Diffusion coefficient determination was performed using 2D – Diffusion Ordered NMR Spectroscopy (DOSY). Measurements were performed at 298 K with 20, 10 and 5 mM [Eu³⁺] samples (δ*t* = 2 ms, Δ*T* = 250 ms). Data analysis was conducted with TopSpin 2.1 using 16 linear points with a 5–95% gradient strength. The diffusion coefficients were calculated through the analysis of individual peaks in the aromatic region. Each peak used in the determination provided the diffusion coefficient that was within an error of 0.05 × 10^{–10} m² s^{–1}. The reported diffusion coefficients in Table 1 are the averages of each of the peaks from the three independent measurements with their standard deviation.

Luminescence lifetime measurements

Luminescence lifetime measurements were performed with **EuL^{1–2}** at 5 mM [Eu³⁺] in D₂O and H₂O with and without Ca²⁺ (2 equiv.) (298 K, pH 7.4, HEPES). The Eu³⁺ ion was directly excited and the emission intensity was recorded with a 10 μs resolution. The excitation and emission slits were set at 5 nm. In total, three independent measurements each with 25 scans were performed to obtain the data set. The obtained curves were fitted with a first order exponential decay with an *r*² = 0.99. The resulting *q* values were then calculated using eqn (2).

$$q = 1.2 \times (\tau_{\text{H}_2\text{O}}^{-1} - \tau_{\text{D}_2\text{O}}^{-1} - 0.25). \quad (2)$$

DFT computational details. The DFT study presented in this work was performed employing the hybrid meta generalized gradient approximation (hybrid *meta*-GGA), with the TPSSH exchange–correlation functional³⁸ and the Gaussian09³⁹ program package. The large-core approximation with the quasirelativistic effective core potential proposed by Dolg *et al.*⁴⁰ and the associated [5s4p3d]–GTO valence basis set was used for Gd, while the standard 6–31G(d,p) basis set was used for all other atoms. Solvent effects were included by using the integral equation formalism variant of the polarizable continuum model (IEFPCM),⁴¹ with solute cavities defined by the universal force field (UFF)⁴² radii scaled by a factor of 1.1. Calculations of analytical second derivatives were used to confirm that the optimized geometries correspond to local energy minima on the potential energy surface and to estimate the zero point energies (ZPE).

Conflicts of interest

There are no conflicts to declare.



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Notes and references

- 1 D. V. Hingorani, A. S. Bernstein and M. D. Pagel, *Contrast Media Mol. Imaging*, 2015, **10**, 245–265.
- 2 M. Carril, *J. Mater. Chem. B*, 2017, **5**, 4332–4347.
- 3 M. C. Heffern, L. M. Matosziuk and T. J. Meade, *Chem. Rev.*, 2014, **114**, 4496–4539.
- 4 G. L. Davies, I. Kramberger and J. J. Davis, *Chem. Commun.*, 2013, **49**, 9704–9721.
- 5 C. S. Bonnet and E. Toth, *Chimia*, 2016, **70**, 102–108.
- 6 D. Parrott, W. S. Fernando and A. F. Martins, *Inorganics*, 2019, **7**, 18.
- 7 J. Wahsner, E. M. Gale, A. Rodríguez-Rodríguez and P. Caravan, *Chem. Rev.*, 2019, **119**, 957–1057.
- 8 C. Grienberger and A. Konnerth, *Neuron*, 2012, **73**, 862–885.
- 9 G. W. Zamponi, *ACS Chem. Neurosci.*, 2017, **8**, 2583–2585.
- 10 G. G. Somjen, *Ions in the brain: normal function, seizures, and strokes*, Oxford University Press, New York, Oxford, 2004.
- 11 I. Dal Pra, A. Chiarini, R. Pacchiana, E. Gardenal, B. Chakravarthy, J. F. Whitfield and U. Armato, *Curr. Neuropharmacol.*, 2014, **12**, 353–364.
- 12 E. L. Que and C. J. Chang, *Chem. Soc. Rev.*, 2010, **39**, 51–60.
- 13 G. Angelovski, *Acc. Chem. Res.*, 2017, **50**, 2215–2224.
- 14 J. L. Major, R. M. Boiteau and T. J. Meade, *Inorg. Chem.*, 2008, **47**, 10788–10795.
- 15 P. Kadjane, C. Platas-Iglesias, P. Boehm-Sturm, V. Truffault, G. E. Hagberg, M. Hoehn, N. K. Logothetis and G. Angelovski, *Chem. – Eur. J.*, 2014, **20**, 7351–7362.
- 16 G. Angelovski, P. Fouskova, I. Mamedov, S. Canals, E. Toth and N. K. Logothetis, *ChemBioChem*, 2008, **9**, 1729–1734.
- 17 S. Gündüz, S. Vibhute, R. Botár, F. K. Kálmán, I. Tóth, G. Tircsó, M. Regueiro-Figueroa, D. Esteban-Gómez, C. Platas-Iglesias and G. Angelovski, *Inorg. Chem.*, 2018, **57**, 5973–5986.
- 18 L. Connah and G. Angelovski, *Biomacromolecules*, 2018, **19**, 4668–4676.
- 19 S. Gündüz, T. Savić, R. Pohmann, N. K. Logothetis, K. Scheffler and G. Angelovski, *ACS Sens.*, 2016, **1**, 483–487.
- 20 C. Kumas, W. S. Fernando, P. Y. Zhao, M. Regueiro-Figueroa, G. E. Kiefer, A. F. Martins, C. Platas-Iglesias and A. D. Sherry, *Inorg. Chem.*, 2016, **55**, 9297–9305.
- 21 T. Mani, G. Tircso, O. Togao, P. Zhao, T. C. Soesbe, M. Takahashi and A. D. Sherry, *Contrast Media Mol. Imaging*, 2009, **4**, 183–191.
- 22 C. M. Fisher, E. Fuller, B. P. Burke, V. Mogilireddy, S. J. A. Pope, A. E. Sparke, I. Dechamps-Olivier, C. Cadiou, F. Chuburu, S. Faulkner and S. J. Archibald, *Dalton Trans.*, 2014, **43**, 9567–9578.
- 23 T. Mani, A. C. L. Opina, P. Y. Zhao, O. M. Evbuomwan, N. Milburn, G. Tircso, C. Kumas and A. D. Sherry, *J. Biol. Inorg. Chem.*, 2014, **19**, 161–171.
- 24 A. W. J. Poh, J. A. Aguilar, A. M. Kenwright, K. Mason and D. Parker, *Chem. – Eur. J.*, 2018, **24**, 16170–16175.
- 25 O. A. Blackburn, M. Tropicano, L. S. Natrajan, A. M. Kenwright and S. Faulkner, *Chem. Commun.*, 2016, **52**, 6111–6114.
- 26 J. Henig, I. Mamedov, P. Fouskova, E. Tóth, N. K. Logothetis, G. Angelovski and H. A. Mayer, *Inorg. Chem.*, 2011, **50**, 6472–6481.
- 27 W. H. Li, G. Parigi, M. Fragai, C. Luchinat and T. J. Meade, *Inorg. Chem.*, 2002, **41**, 4018–4024.
- 28 E. Boros, R. Srinivas, H. K. Kim, A. M. Raitsimring, A. V. Astashkin, O. G. Poluektov, J. Niklas, A. D. Horning, B. Tidor and P. Caravan, *Angew. Chem., Int. Ed.*, 2017, **56**, 5603–5606.
- 29 A. Beeby, I. M. Clarkson, R. S. Dickins, S. Faulkner, D. Parker, L. Royle, A. S. de Sousa, J. A. G. Williams and M. Woods, *J. Chem. Soc., Perkin Trans. 2*, 1999, 493–503.
- 30 D. Esteban-Gomez, A. de Blas, T. Rodriguez-Blas, L. Helm and C. Platas-Iglesias, *ChemPhysChem*, 2012, **13**, 3640–3650.
- 31 M. Regueiro-Figueroa and C. Platas-Iglesias, *J. Phys. Chem. A*, 2015, **119**, 6436–6445.
- 32 K. D. Verma, A. Forgács, H. Uh, M. Beyerlein, M. E. Maier, S. Petoud, M. Botta and N. K. Logothetis, *Chem. – Eur. J.*, 2013, **19**, 18011–18026.
- 33 F. Oukhatar, H. Meudal, C. Landon, N. K. Logothetis, C. Platas-Iglesias, G. Angelovski and É. Tóth, *Chem. – Eur. J.*, 2015, **21**, 11226–11237.
- 34 F. Oukhatar, S. V. Eliseeva, C. S. Bonnet, M. Placidi, N. K. Logothetis, S. Petoud, G. Angelovski and É. Tóth, *Inorg. Chem.*, 2019, DOI: 10.1021/acs.inorgchem.1029b00887.
- 35 D. M. Corsi, C. Platas-Iglesias, H. van Bekkum and J. A. Peters, *Magn. Reson. Chem.*, 2001, **39**, 723–726.
- 36 T. D. Goddard and D. G. Kneller, *SPARKY 3*, University of California, San Francisco, ver.
- 37 A. E. Derome and M. P. Williamson, *J. Magn. Reson.*, 1990, **88**, 177–185.
- 38 J. M. Tao, J. P. Perdew, V. N. Staroverov and G. E. Scuseria, *Phys. Rev. Lett.*, 2003, **91**, 146401.
- 39 *Gaussian 09, Revision B.01*, Gaussian, Inc., Wallingford CT, ver., 2009.
- 40 M. Dolg, H. Stoll, A. Savin and H. Preuss, *Theor. Chim. Acta*, 1989, **75**, 173–194.
- 41 J. Tomasi, B. Mennucci and R. Cammi, *Chem. Rev.*, 2005, **105**, 2999–3093.
- 42 A. K. Rappe, C. J. Casewit, K. S. Colwell, W. A. Goddard and W. M. Skiff, *J. Am. Chem. Soc.*, 1992, **114**, 10024–10035.

