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Understanding the interdependence of operating parameters in microbial electrosynthesis: a numerical investigation†

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This study describes and evaluates a dynamic computational model for a two chamber microbial electrosynthesis (MES) system. The analysis is based on redox mediators and a two population model, describing bioelectrochemical kinetics at both anode and cathode. Mass transfer rates of the substrate and bacteria in the two chambers are combined with the kinetics and Ohm's law to derive an expression for the cell current density. The effect of operational parameters such as initial substrate concentration at the anode and cathode and the operation cycle time on MES performance is evaluated in terms of product formation rate, substrate consumption and coulombic efficiency (CE). For a fixed operation cycle time of 3 or 4 days, the anode and cathode initial substrate concentrations show linear relationship with product formation rate; however MES operation with a 2 day cycle time shows a more complex behaviour, with acetic acid production rates reaching a plateau and even a slight decrease at higher concentrations of the two substrates. It is also shown that there is a trade-off between product formation rate and substrate consumption and CE. MES performance for operation with cycle time being controlled by substrate consumption is also described. Results from the analysis demonstrate the interdependence of the system parameters and highlight the importance of multi-objective system optimization based on targeted end-use.

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1 Introduction

Microbial electrosynthesis (MES) is a type of bioelectrochemical device which involves electricity driven microbial reduction of carbon dioxide into a variety of useful multi-carbon compounds.^{1–3} While the MES setup typically involves the combination of a chemical anode with a biocatalysed cathode (biocathode), systems with both biocatalysed anode and cathode are also being investigated.^{1,4–7} When a biocatalysed anode (bioanode) is used in MES, wastewater or sludge hydrolysates which typically contain a multitude of organic substances or contaminants can be used as a substrate at the anode.^{1,4,6} At the bioanode the organics/contaminants in the waste stream function as the electron source and undergo oxidation to produce electrons and protons, which can then be subsequently used for microbial reduction at the cathode. The external power required for microbial oxidation of

wastewater organics at the bioanode is lower than that required for water splitting at the abiotic anode.⁷ Bioanode based MES has the potential to produce high value organic chemicals from wastewater streams and CO₂, thus allowing recovery of useful energy in the form of chemicals from waste. It also allows the use of low-cost carbon materials as electrodes in comparison with chemical anodes, which typically require costly electrodes for water electrolysis.^{1,4,5}

Several products, including methane, acetic acid, propanol, butanol and ethanol, can be microbially synthesised from CO₂ reduction at different potentials.^{1,3,8} To date, acetic acid (AA) has been the most reported product in the literature.^{9–12} AA is a versatile intermediate for producing several chemicals, including vinyl acetate monomers, esters and acetic anhydride, for the manufacture of synthetic fibers, textiles, inks, and pesticides.¹³ Current industrial production methods of AA such as methanol carbonylation, dubbed the 'Monsanto process', acetaldehyde oxidation, oxidation of *n*-butane, fermentation of hydrocarbons, and ethane direct oxidation all use raw materials extracted from fossil fuels.^{14,15} Consequently, they are resource-intensive and environmentally damaging. The plausibility of MES to replace these industrial systems necessitates the optimization of MES and efficient quantification of the production rates.

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Significant progress has been made in the past decade towards understanding the physico-electrochemical and biological mechanisms of the process, which has helped in improving the product yields.^{16–18} However the performance is still far behind from making a strong business case for large scale implementation.^{3,16,18} Thorough understanding of the relationships among the various parameters and their dynamic processes is important to make this technology more efficient.^{2,19,20} Computational models can complement experimental studies towards MES optimization and expedite the development of this technology. However past research on MES has extensively focused on experimental studies, and barring a few numerical investigations, modeling and simulation remain mostly neglected.^{19–24}

Pinto *et al.*²² proposed a multi-population dynamic model of a microbial electrolysis cell (MEC) and derived an expression for the hydrogen production rate at the chemical cathode based on MEC current and specific cathode efficiency. The bacterial population at the anode is assumed to consist of fermentative, electricigenic, methanogenic acetoclastic, and methanogenic hydrogenophilic microorganisms, whose metabolic activity and growth rates are described using multiplicative Monod kinetics. This model expressed in ordinary differential equations is based on the assumption that a single hydrolysis and fermentation step of complex organic matter conversion to acetate can be used to describe the anaerobic degradation of wastewater. Though this model is not derived for MES directly and does not consider any biocatalyst at the cathode, it serves as a good starting point for more comprehensive numerical models for different microbial electrochemical cells.²²

Kazemi *et al.*²³ used the conductive biofilm approach and modeled the biocathode in an MES to study the bioelectrosynthesis of acetate by CO₂ reduction. The rate of microbial reduction at the cathode is described using a Nernst–Monod expression assuming that the biofilm matrix conducts electrons from the cathode to the biofilm bacteria. The biomass is classified into two types, active (bacteria responsible for CO₂ reduction) and inert (inactive conductive components such as EPS and nanowires). An expression for overpotential is derived from electron balance and Ohm's law, combining the electron flux in the conductive matrix with electron consumption (from CO₂ reduction) and electron generation (due to self-oxidation of biomass) terms. This model predicts the biofilm dynamics, substrate concentration and current density profiles; however the analysis is limited to the cathode and does not account for the influence of electrode kinetics and mass transfer losses at the anode.²³

In a previous study by the authors,²⁴ a dynamic model also based on the conductive biofilm approach was used to study formic acid synthesis in a bioelectrochemical system (BES). Current density and local overpotential along the biofilm depth were correlated with the conductivity of the biofilm matrix. However this model was based on bioanode kinetics alone and assumed a non-limiting cathode. To the authors' knowledge, thus far, a comprehensive dynamic simulation (considering both anode and cathode kinetics) of microbially catalyzed electrosynthesis of AA from CO₂ reduction *via* MES is yet to be performed.

In the present study, for the first time, a comprehensive one dimensional dynamic model has been developed for a two chamber microbial electrosynthesis system. The modeling framework is based on a two-population model and accounts for the bioelectrochemical kinetics at both the (bio)electrodes, which are combined to the mass balance of substrates and bacteria in the respective electrode chambers. Potential losses are derived and are used to calculate the MES current and product formation rate. The dynamic model applicability for optimization studies is illustrated by examining the effect of operating parameters on MES performance.

2 Methodology

The mathematical modeling framework has been developed based on a two chamber MES, assuming both electrodes are catalyzed using bacteria. The experimental setup as described by Modestra and Mohan⁴ has been used for reference. The anode chamber which is inoculated with untreated anaerobic bacteria may consist of several different types of microbial populations.⁴ For this analysis, the bacteria in the anode are classified into two types, primary anodic bacteria (the 'exoelectrogenic' bacteria that consume the substrate (COD) and release electrons which can be transferred to the anode) and secondary anodic bacteria (which can include fermentative/methanogenic bacterial types that decompose the substrate into smaller compounds but do not transfer any free electrons in the process). The cathode chamber is inoculated with selectively enriched homoacetogenic bacteria which mainly reduce CO₂/bicarbonate to AA.⁴ However the presence of small quantities of butyric and propionic acids in the final product suggests the presence of other bacterial types that can either reduce CO₂/bicarbonate or elongate AA to produce higher carbon (C3 and C4) compounds.⁴ Thus in the model, the bacterial population at the cathode is also classified into two types, primary cathodic bacteria (such as the enriched homoacetogenic bacteria in this case, that reduce CO₂/bicarbonate to acetic acid) and secondary cathodic bacteria (chain elongators or other bacterial types that are capable of producing higher carbon compounds by direct CO₂ reduction or elongation of C2 compounds). Here it should be noted that the terms 'primary' and 'secondary' are being used in a generalized framework and the specific sub-type of bacteria in each of these categories will depend on the individual systems. Some MES systems with pure culture may not have any secondary bacteria at all. Also, for MES with a chemical anode, there will be no anodic bacteria.

In addition to the above description, the following assumptions are made in deriving the dynamic model:

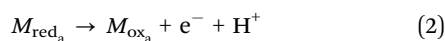
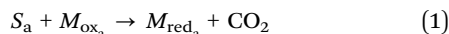
- Substrates in both anolyte and catholyte are perfectly mixed.
- Microbial populations in the biofilms of the anode and cathode chambers are uniformly distributed.
- pH and temperature are strictly controlled.
- Gases produced during oxidation and reduction reactions in the anode and cathode remain dissolved in the respective bulk solutions.



- Activation overpotential at the anode is negligible.
- Extracellular electron transfer to and from the electrodes is assumed to take place using redox mediators.

2.1 Material balance in the anode chamber

In the anode chamber, the primary microbial population ($x_{p,a}$) consumes the substrate (S_a) and in this process the oxidized form of the redox mediator (M_{ox_a}) is also converted into its reduced form (M_{red_a}). This reduced mediator exchanges the electron with the anode and also releases a proton while it regains its oxidized form. The above sequence of reactions between the anode substrate and the redox mediators can be described conceptually as in eqn (1) and (2).



Meanwhile, the secondary bacterial population ($x_{s,a}$) also consumes the substrate but does not release any free electrons which can be utilized by the redox mediators. Considering a fed-batch operation,⁴ the rate of change of substrate and biomass concentrations can be expressed as in eqn (3)–(5).

$$\frac{dS_a}{dt} = -q_{p,a}x_{p,a} - q_{s,a}x_{s,a} \quad (3)$$

$$\frac{dx_{p,a}}{dt} = \mu_{p,a}x_{p,a} - K_{d,p,a}x_{p,a} \quad (4)$$

$$\frac{dx_{s,a}}{dt} = \mu_{s,a}x_{s,a} - K_{d,s,a}x_{s,a} \quad (5)$$

where S is the substrate concentration ($g\text{-}S\text{ L}^{-1}$), x is the microbial concentration ($g\text{-}x\text{ L}^{-1}$), q is the substrate consumption rate ($g\text{-}S\text{ g}\text{-}x^{-1}\text{ d}^{-1}$), μ is the microbial growth rate (d^{-1}) and K_d is the microbial decay rate (d^{-1}). Subscript 'a' represents quantities in the anode chamber. The subscripts 'p,a' and 's,a' represent the primary and secondary microbial populations at the anode respectively.

The redox mediator in the anode biomass exists in either oxidized (M_{ox_a}) or reduced form (M_{red_a}); however the total mediator concentration M_{total_a} remains constant.

$$M_{total_a} = M_{ox_a} + M_{red_a} \quad (6)$$

As the primary bacteria decompose the substrate, the oxidized form of the redox mediator is transformed into its reduced form, which then transfers electrons to the anode and is converted back to its oxidized form. The transfer of electrons results in current flow and thus the rate of change of oxidized mediator concentration can be expressed as in eqn (7).

$$\frac{dM_{ox_a}}{dt} = -Y_a q_{p,a} + \frac{M_{m_a}}{V_a x_{p,a}} \left(\frac{I_{mes}}{mF} \right) \quad (7)$$

where Y is the dimensionless mediator yield, I_{mes} is the MES current (A), m is the number of electrons transferred per mol of mediator, F is the Faraday constant ($C\text{ mol}^{-1}$), V_a is the working volume of the anode chamber (L), and M_m is the molar mass of the mediator ($g\text{ mol}^{-1}$).

The Monod kinetic model, describing the relationship between growth rate of bacteria and the concentration of the limiting nutrients, is the most commonly used model for describing the bacterial growth and substrate consumption rates.²⁵ The substrate consumption rate of the primary bacteria and the corresponding microbial growth rate are limited by both the substrate and the oxidized mediator concentrations. On the other hand, the substrate consumption rate of the secondary bacteria and the corresponding microbial growth rate are only limited by the substrate concentration. Thus the substrate consumption and growth rates for primary bacteria are represented by multiplicative Monod kinetics (eqn (8) and (9)), while the same for secondary bacteria are described using standard Monod kinetics (eqn (10) and (11)).

$$q_{p,a} = q_{max,p,a} \left(\frac{S_a}{K_{S,p,a} + S_a} \right) \left(\frac{M_{ox_a}}{K_{M_a} + M_{ox_a}} \right) \quad (8)$$

$$\mu_{p,a} = \mu_{max,p,a} \left(\frac{S_a}{K_{S,p,a} + S_a} \right) \left(\frac{M_{ox_a}}{K_{M_a} + M_{ox_a}} \right) \quad (9)$$

$$q_{s,a} = q_{max,s,a} \left(\frac{S_a}{K_{S,s,a} + S_a} \right) \quad (10)$$

$$\mu_{s,a} = \mu_{max,s,a} \left(\frac{S_a}{K_{S,s,a} + S_a} \right) \quad (11)$$

where K_S and K_M are the half saturation Monod constants for the substrate and mediator respectively ($g\text{ L}^{-1}$), and q_{max} and μ_{max} are the maximum substrate consumption rate and maximum growth rate respectively (d^{-1}).

2.2 Material balance in the cathode chamber

In the cathode chamber, the oxidized form of the redox mediator (M_{ox_c}) accepts the electron from the cathode and is converted into its reduced form (M_{red_c}). The primary microbial population at the cathode ($x_{p,c}$) consume the substrate (S_c) and utilize the reduced mediator to produce the main product, AA. In this process the reduced form of the mediator is converted back to the oxidized form. The sequence of reactions between the cathode substrate and the redox mediators, leading to the formation of product (AA), can be described conceptually as in eqn (12) and (13).



The secondary bacterial population at the cathode ($x_{s,c}$) may also consume the substrate. The rate of change of substrate and that of microbial populations at the cathode are represented as in eqn (14)–(16).

$$\frac{dS_c}{dt} = -q_{p,c}x_{p,c} - q_{s,c}x_{s,c} \quad (14)$$

$$\frac{dx_{p,c}}{dt} = \mu_{p,c}x_{p,c} - K_{d,p,c}x_{p,c} \quad (15)$$



$$\frac{dx_{s,c}}{dt} = \mu_{s,c}x_{s,c} - K_{d,s,c}x_{s,c} \quad (16)$$

where subscript 'c' represents quantities in the cathode chamber. The subscripts 'p,c' and 's,c' represent the primary and secondary microbial populations at the cathode respectively.

Similar to the anode chamber, the total mediator concentration in the cathode biomass ($M_{\text{total,c}}$) is the sum of its reduced ($M_{\text{red,c}}$) and oxidized ($M_{\text{ox,c}}$) forms.

$$M_{\text{total,c}} = M_{\text{ox,c}} + M_{\text{red,c}} \quad (17)$$

The rate of change of reduced mediator concentration at the cathode is a function of the substrate consumption rate by primary bacteria and the MES current and is described as in eqn (18).

$$\frac{dM_{\text{red,c}}}{dt} = -Y_c q_{p,c} + \frac{M_{\text{me}} I_{\text{mes}}}{V_c x_{p,c} mF} \quad (18)$$

The substrate consumption and growth rates for primary bacteria at the cathode are represented by multiplicative Monod kinetics, while the same for secondary bacteria are represented using standard Monod kinetics as described in eqn (19)–(22).

$$q_{p,c} = q_{\text{max},p,c} \left(\frac{S_c}{K_{S,p,c} + S_c} \right) \left(\frac{M_{\text{red,c}}}{K_{M,c} + M_{\text{red,c}}} \right) \quad (19)$$

$$\mu_{p,c} = \mu_{\text{max},p,c} \left(\frac{S_c}{K_{S,p,c} + S_c} \right) \left(\frac{M_{\text{red,c}}}{K_{M,c} + M_{\text{red,c}}} \right) \quad (20)$$

$$q_{s,c} = q_{\text{max},s,c} \left(\frac{S_c}{K_{S,s,c} + S_c} \right) \quad (21)$$

$$\mu_{s,c} = \mu_{\text{max},s,c} \left(\frac{S_c}{K_{S,s,c} + S_c} \right) \quad (22)$$

2.3 Ohm's law and voltage losses

As the equilibrium cell voltage in a MES is negative, the reactions at the electrodes are non-spontaneous and thus an external power is required.²⁶ Additionally there are activation (anode and cathode), concentration (anode and cathode) and ohmic voltage losses in the system, which need to be compensated by the external power requirement.^{22,26} Thus the total applied voltage in a MES can be expressed as in eqn (23).

$$-E_{\text{app}} = E_{B_{\text{EMF}}} - \eta_{\text{ohm}} - \eta_{\text{conc}_a} - \eta_{\text{conc}_c} - \eta_{\text{act}_a} - \eta_{\text{act}_c} \quad (23)$$

where E_{app} is the externally applied voltage (V), $E_{B_{\text{EMF}}}$ is the back-electromotive force for the MES (V), and η_{ohm} , η_{act} and η_{conc} represent the ohmic, activation and concentration over-potentials (V).

This analysis assumes a constant supply of redox mediators, which undergo transformation between oxidized and reduced forms as they exchange electrons to or from the electrodes. Considering this, the major factor influencing the concentration overpotential is the substrate concentration at the two electrodes.

The corresponding voltage losses at the anode and cathode can thus be represented as in eqn (24) and (25).

$$\eta_{\text{conc}_a} = \frac{RT}{mF} \ln \left(\frac{S_{a,\text{in}}}{S_a} \right) \quad (24)$$

$$\eta_{\text{conc}_c} = \frac{RT}{mF} \ln \left(\frac{S_{c,\text{in}}}{S_c} \right) \quad (25)$$

where $S_{a,\text{in}}$ and $S_{c,\text{in}}$ represent the initial substrate concentrations in anode and cathode chambers respectively (g-S L^{-1}).

Voltage is also lost as activation energy due to slow electrochemical kinetics at the electrodes. Such potential loss in MES to drive the electrochemical reactions from equilibrium state is assumed to be far greater at the cathode as compared to the anode and is often the controlling factor of MES performance. Thus the activation overpotential at the anode is neglected while that at the cathode (η_{act_c}) can be expressed as a function of current density using the Butler–Volmer expression. Hamelers *et al.*²⁷ combined the Monod kinetics (describing the biochemical oxidation of the substrate) with the Butler–Volmer electron transfer kinetics to derive a steady state expression for the current density as a function of overpotential in a microbial fuel cell. In this analysis, the voltage balance and Ohm's law are used to derive the dynamic expression for current density. The Butler–Volmer equation is used to describe the polarisation at the cathode. Assuming the symmetry factor or the charge transfer coefficient as 0.5, the Butler–Volmer equation can be re-arranged as a hyperbolic sine function, which provides the expression for activation overpotential as in eqn (26):

$$\eta_{\text{act}_c} = \frac{RT}{\beta_c mF} \sinh^{-1} \left(\frac{I_{\text{mes}}}{A_c i_0} \right) \quad (26)$$

where T is the system temperature (K), β_c is the charge transfer coefficient, A_c is the effective surface area of the cathode (m^2), and i_0 is the exchange current density (A m^{-2}). A similar expression of activation overpotential derived from the standard Butler–Volmer equation has been used in previous studies.^{22,28} It should be also noted that the mass transfer effect on the overpotentials at the two electrodes has already been described in eqn (24) and (25).

Based on Ohm's law, the ohmic voltage drop (η_{ohm}) is directly proportional to the current density, and can be expressed as in eqn (27):

$$\eta_{\text{ohm}} = I_{\text{mes}} R_{\text{ohm}} \quad (27)$$

where R_{ohm} is the internal ohmic resistance of the cell (Ω).

Eqn (27) can be substituted in eqn (23) to obtain an expression for MES current as described in eqn (28).

$$I_{\text{mes}} = \frac{E_{B_{\text{EMF}}} + E_{\text{app}} - \eta_{\text{conc}_a} - \eta_{\text{conc}_c} - \eta_{\text{act}_c}}{R_{\text{ohm}}} \quad (28)$$

Following a similar approach as suggested by Pinto *et al.*,²⁹ the model rigor was further improved by incorporating an expression for R_{ohm} as a function of primary bacteria concentration, described as in eqn (29).

$$R_{\text{ohm}} = R_{\text{min}} + (R_{\text{max}} - R_{\text{min}}) e^{(-K_{\text{R}}(x_{p,a} + x_{p,c}))} \quad (29)$$



where R_{\max} and R_{\min} represent the maximum and minimum internal resistance respectively (Ω), and K_R is the curve steepness factor ($L g^{-1}$).

2.4 Product formation rate

The product formation rate is a direct function of current density. However as mentioned before, the secondary bacteria can also use the primary product to convert it into secondary products. Thus the final production rate of the primary product, Q_p ($\text{mol m}^{-3} \text{d}^{-1}$), can be expressed as in eqn (30).

$$Q_p = \frac{E_c}{V_c} \left(\frac{I_{\text{mes}}}{mF} \right) - Q_{\text{sp}} \quad (30)$$

where E_c is the dimensionless cathode efficiency and Q_{sp} ($\text{mol m}^{-3} \text{d}^{-1}$) is the production rate of secondary products.

Q_{sp} is assumed to be a function of only Q_p and is expressed as in eqn (31).

$$\frac{dQ_{\text{sp}}}{dt} = q_{p_2} Q_p \quad (31)$$

where q_{p_2} is the rate of primary product (AA) elongation to secondary products (C3/C4 compounds) (d^{-1}).

A substantial consumption of AA begins only after reaching a certain threshold concentration, which would differ for different cathode bacterial populations and is described using the rate of primary product elongation (q_{p_2}). It should also be noted that the primary and secondary products can be different for different MES setups and depend mainly on the bacterial strains used at the cathode. For example, in the MES studied by Modestra and Mohan,⁴ the cathode is inoculated with enriched acetogenic bacteria, thus the primary product is AA, whereas Choi *et al.*³⁰ used an acidogenic clostridium strain, *Clostridium tyrobutyricum*, which primarily produces butyrate and acetate is only a secondary product.

3 Results and discussion

The mathematical model for a two chamber MES system as described in Section 2 is generic. It can be used to assess the productivity and performance of wastewater valorization and carbon dioxide reduction in different microbial electrosynthesis systems. The effectiveness of the proposed model has been demonstrated by studying the performance of the MES described by Modestra and Mohan,⁴ under different operating conditions.

3.1 Parameter estimation

First, the model parameters such as the maximum substrate consumption rates, maximum microbial growth rates, yield coefficients, *etc.*, that are specific to the experimental study and the setup are estimated by best-fit regression analysis by comparing the numerical results with experimental values obtained from Modestra and Mohan.⁴ In the simulation based curve fitting method, the objective function (defined in eqn (32)) is minimized to obtain the parameter values.

$$J = \sum_t (y_{\text{exp}(t)} - y_{\text{sim}(t)})^2 \quad (32)$$

where $y_{\text{exp}(t)}$ and $y_{\text{sim}(t)}$ represent the variable values obtained from experiment and numerical simulation respectively at a particular time t . In this case study, the objective function is defined as the difference between measured and predicted current density as a function of time. This function is minimized using the simplex search method based on the Nelder–Mead optimization technique,³¹ a method which is well known for handling unconstrained optimization problems where the gradient is not known or is difficult to find.³² Fig. 1 shows the experimental and fitted values of current density. The predicted values from the numerical fitting algorithm are in fairly good agreement with the experimental data, except in the first 12 hours (0.5 day). The list of all the parameter values obtained from the curve-fitting is provided in the supporting information.

In the current analysis, the Nelder–Mead optimization algorithm is constrained to obtain 90% agreement with the experimental data. Once this condition is met the simulation is stopped and the fitted parameter values are obtained. The constraint helps in obtaining a reasonably good agreement with minimum computational expense. However, it is also responsible for the poor compliance of predicted values during the first 12 hours. The agreement can be improved by continuing the simulation longer; however this would also extend the computation time significantly.

It should also be noted that the reliability of the fitted parameters is largely dependent on the available experimental data. As highlighted, in this analysis, the data are available for only one set of operating conditions, and thus the fitted parameters are conditional to the single objective function and may not be unique. Fitting to experimental data for a range of operating conditions could offer a more unique estimate; however for the current analysis this remains one of the main limitations.

Parameter values which are known to have the largest impact on MES performance such as the maximum substrate consumption and growth rates of the primary bacteria at the anode and cathode are fitted first before obtaining other values. The accuracy of the current predicted values is assessed in the

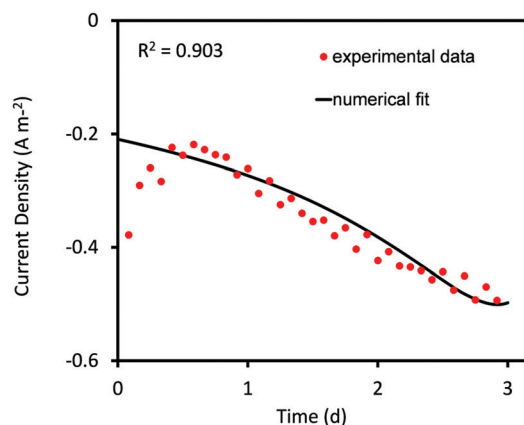


Fig. 1 Predicted values of current density obtained from curve fitting using the Nelder–Mead Simplex algorithm compared to the corresponding experimental values for 3 days of MES operation.



next section, comparing the predicted results for change in cathode substrate concentration and AA production for the same set of operating conditions.

3.2 Validation

Based on the experimental data of AA production,⁴ the threshold value of AA above which its elongation to higher carbon compounds starts is $0.09 \text{ mol m}^{-3} \text{ s}^{-1}$. This is substituted while solving for eqn (30) and (31). Fig. 2 shows the comparison of numerically predicted values of change in cathode substrate concentration and amount of acetic acid produced as a function of time to that observed in the experimental study.⁴

As can be seen in Fig. 2, there is a close agreement between the experimental and predicted values, indicating a reasonably good estimate of the parameters. As expected, the substrate concentration at the cathode almost linearly decreases with time (Fig. 2A). On the other hand, the concentration of AA produced increases linearly for the first 1.8 days, after which it remains almost constant for some time and then slowly starts decreasing. The reason for such a decrease is the consumption of AA for elongation to secondary products (C3/C4 compounds). This utilization of the primary product has been accurately included in the model (eqn (31)), which provides a good agreement between predicted and experimental values, as shown in Fig. 2B. Several experimental studies on MES have reported the formation of different secondary products, the type and concentrations of which vary with the nature of microbial populations used at the cathode.^{30,33} The current model is generic and can be used to analyze different MES focusing on a variety of products using different bacterial strains and substrates. In case where the primary product is not being consumed/elongated to produce secondary products, q_{P_2} in eqn (31) should be equated to zero.

3.3 Effect of substrate concentrations

The dynamic model is used to study how the initial substrate concentrations at the anode ($S_{a_{in}}$) and at the cathode ($S_{c_{in}}$) influence the MES performance in terms of the primary product (in this case AA) formation rate. For this analysis, the MES is assumed to be operating in fed-batch mode in a cycle of 3 days.⁴ After 3 days, the feed in both the anode and cathode chambers is changed and replenished to initial concentrations. The substrate feed at the anode is wastewater and is expressed in terms of COD concentration.

As can be seen from Fig. 3, the increase in both COD and initial cathode substrate concentrations ($S_{a_{in}}$ and $S_{c_{in}}$) shows an overall increase in the AA production rate (Q_{AA}); there are however some variations from this general behaviour particularly for $S_{c_{in}}$. While the increase in $S_{a_{in}}$ from 1 g L^{-1} to 5 g L^{-1} leads to an increase in Q_{AA} for all values of $S_{c_{in}}$, the increase in $S_{c_{in}}$ from 2.5 g L^{-1} to 7.5 g L^{-1} does not always lead to an increase in Q_{AA} , mainly at lower values of $S_{a_{in}}$ ($< 2.25 \text{ g L}^{-1}$). For smaller $S_{a_{in}}$, Q_{AA} first decreases with increasing $S_{c_{in}}$ up to a certain threshold $S_{c_{in}}$ and then starts increasing with a further increase in $S_{c_{in}}$. This threshold $S_{c_{in}}$ where the relationship between Q_{AA} and $S_{c_{in}}$ changes starts decreasing as $S_{a_{in}}$ increases.

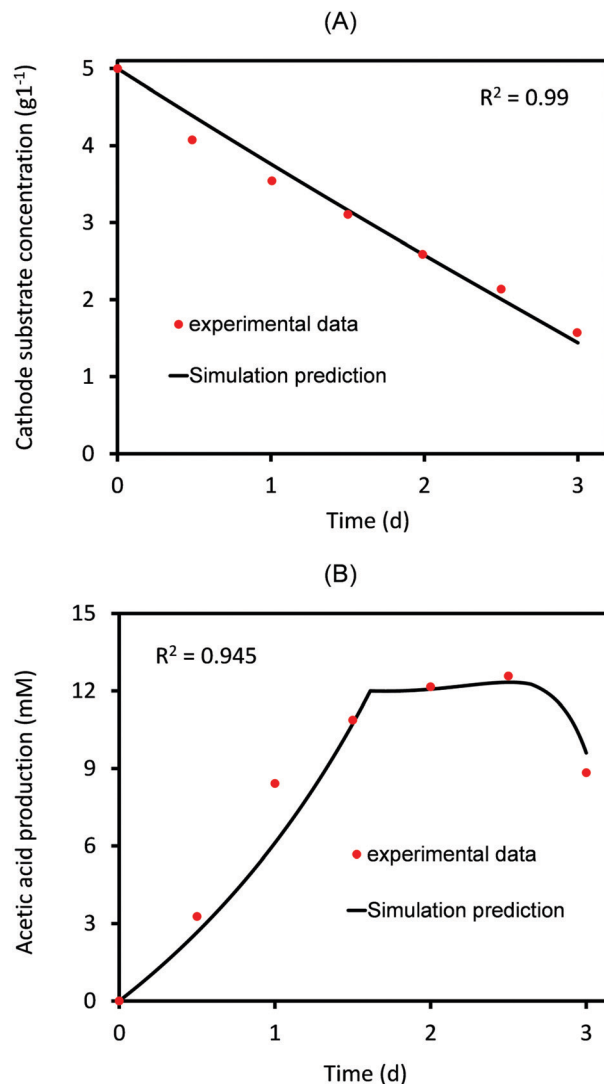


Fig. 2 Predicted values of (A) cathode substrate concentration and the (B) acetic acid produced as a function of time compared to the corresponding experimental values for 3 days of MES operation.

For the lowest $S_{a_{in}}$ used in this analysis, 1 g L^{-1} , Q_{AA} decreases as $S_{c_{in}}$ is increased from 2.5 g L^{-1} to 5 g L^{-1} , and then starts increasing for higher $S_{c_{in}}$ values. For $S_{a_{in}} = 1.25 \text{ g L}^{-1}$, 1.5 g L^{-1} , 1.75 g L^{-1} and 2 g L^{-1} , the threshold $S_{c_{in}}$ value decreases from 4 g L^{-1} to 2.75 g L^{-1} . For all $S_{a_{in}} > 2 \text{ g L}^{-1}$, an increase in $S_{c_{in}}$ leads to a linear increase in the AA production rate!

Another important observation is that Q_{AA} reaches a plateau and shows a slight decrease at high values of both $S_{a_{in}}$ and $S_{c_{in}}$. Thus a slight flattening of the surface plot in Fig. 3 is observed near the highest values of substrate concentrations. For the range of initial substrate concentrations studied in this analysis, the highest $Q_{AA} = 7.75 \text{ mM d}^{-1}$ is obtained for $S_{c_{in}} = 7.25 \text{ g L}^{-1}$ and $S_{a_{in}} = 5 \text{ g L}^{-1}$, whereas the lowest $Q_{AA} = 2.6 \text{ mM d}^{-1}$ is obtained for $S_{c_{in}} = 5 \text{ g L}^{-1}$ and $S_{a_{in}} = 1 \text{ g L}^{-1}$.

Mohanakrishna *et al.*³⁴ used enriched mixed homoacetogenic bacteria as cathodic bio-catalyst and studied the influence of substrate concentration at the cathode on the acetate



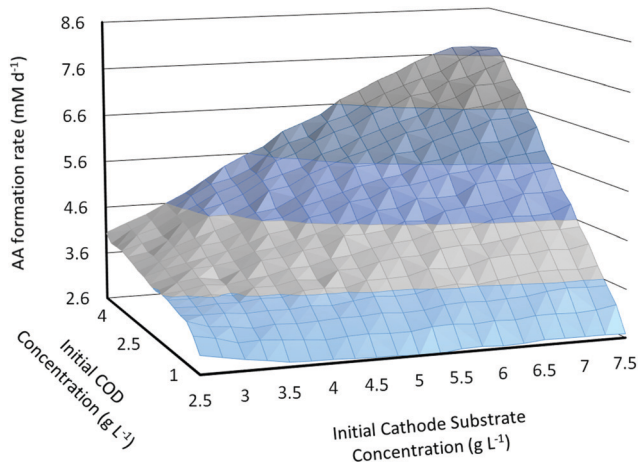


Fig. 3 Predicted AA production rate (mM d^{-1}) as a function of initial substrate concentrations in the anode and cathode chambers.

production rate. They also observed an increase in the acetate production rate as cathode substrate concentration was increased from 1 g L^{-1} to 4 g L^{-1} . Mohanakrishna *et al.*³⁴ suggested that an increase in substrate concentration leads to an increase in substrate availability, change in the biocathode reduction potential and gain in the coulombic efficiency, all of which contribute to improving the acetate production rate at higher substrate concentrations. Increased $S_{\text{a,in}}$ also increases substrate availability for consumption at the bioanode, thus releasing more electrochemical reductive equivalents such as protons and electrons for eventual consumption at the cathode. This increased availability of reductive equivalents helps in improving the AA production rate.

An important factor that would closely determine the MES economical performance is the amount of substrate consumption achieved. It should be noted that with a bioanode, the substrate used in the anode chamber could be wastewater, which is quantified by the COD concentration. The amount of COD degraded/consumed in the bioanode could reduce the subsequent processing required to treat the wastewater. This will have major implications in terms of the energy saved and the final effective cost per kg of product. At the cathode, the amount of substrate ($\text{CO}_2/\text{bicarbonate}$) consumed in each cycle would determine the recycling rate and would also affect the overall cost significantly. An ideal MES operation would provide 100% substrate consumption with a high product formation rate to achieve the best economic performance.

For a fed-batch system, substrate consumption is quantified per cycle of operation, whereas in a continuous system this would be determined by the flow rate of anolyte/catholyte and residence time of substrates in each electrode chamber. Fig. 4 shows the prediction of substrate consumption in one cycle, for the range of anode and cathode initial substrate concentrations used in Fig. 3. As can be seen from Fig. 4, the percentage of substrate consumed in both anode and cathode decreases with the increase in the respective substrate concentrations. This is expected as under a given cycle time, the amount of each substrate consumed is limited and when a higher concentration

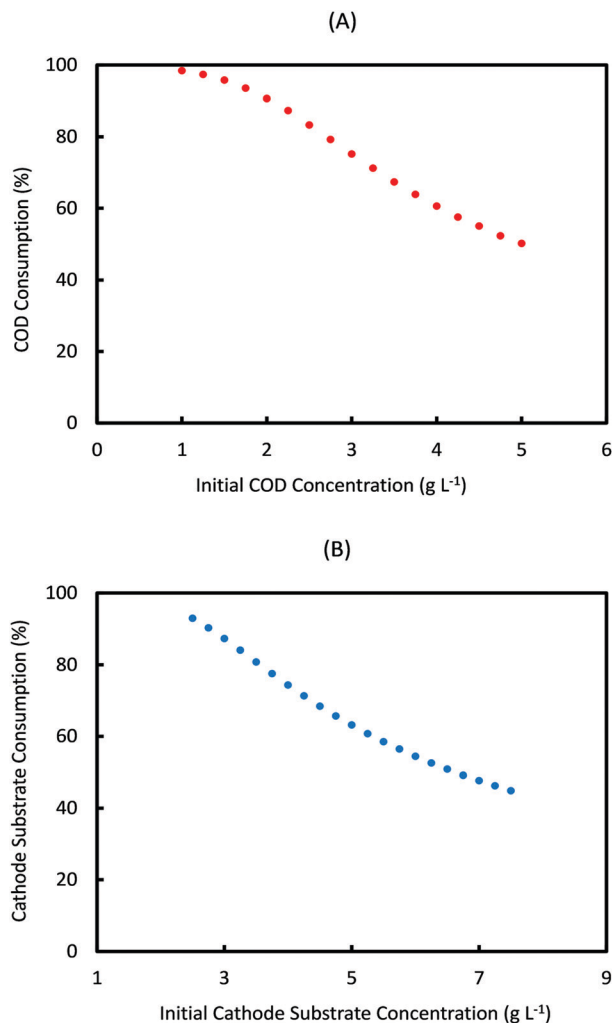


Fig. 4 Simulation prediction of percentage substrate consumption at the end of each operational cycle (3 days) for different initial concentrations of (A) anode and (B) cathode substrates in the respective electrode chambers.

feed is used, more substrate will remain unconsumed. However these data are important to assess the performance of MES, by comparing with the product formation rate at these concentrations.

Based on Fig. 4, the percentage of anode substrate consumed in one cycle decreases from 100% at $S_{\text{a,in}} = 1 \text{ g L}^{-1}$ to 50% at $S_{\text{a,in}} = 5 \text{ g L}^{-1}$. Similarly the percentage of cathode substrate consumed decreases from 93% at $S_{\text{c,in}} = 2.5 \text{ g L}^{-1}$ to 45% at $S_{\text{c,in}} = 7.5 \text{ g L}^{-1}$. It should also be noted that change in $S_{\text{a,in}}$ had a negligible influence on cathode substrate consumption, and similarly change in $S_{\text{c,in}}$ had almost no effect on anode substrate consumption. For the specific initial substrate concentrations ($S_{\text{c,in}} = 7.25 \text{ g L}^{-1}$ and $S_{\text{a,in}} = 5 \text{ g L}^{-1}$) which show the maximum AA production rate (7.75 mM d^{-1}), the anode and cathode substrate consumption achieved is only 50% and 46% respectively. On the other hand, the consumption achieved for initial substrate concentrations ($S_{\text{c,in}} = 5 \text{ g L}^{-1}$ and $S_{\text{a,in}} = 1 \text{ g L}^{-1}$) which predict the minimum AA production rate (2.6 mM d^{-1}) is 98.5% and 63% respectively!



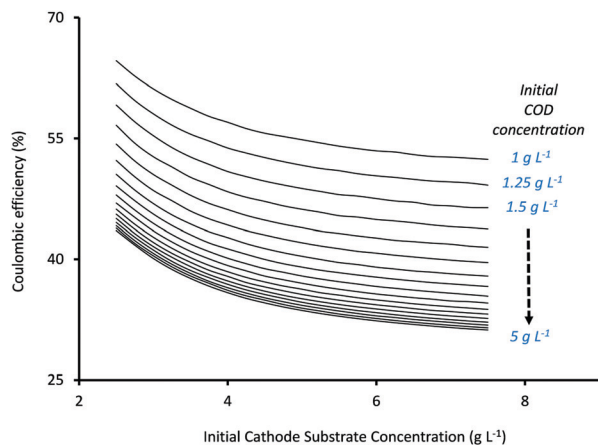


Fig. 5 Predicted coulombic efficiency (%) as a function of initial substrate concentrations in the anode and cathode chambers.

The third important factor that determines MES performance is the percentage of total charge (Coulombs) that is recovered as products or the coulombic efficiency (CE) of the system. Fig. 5 shows the variation in CE (%) as a function of initial substrate concentrations at the anode and cathode. While the x -axis in Fig. 5 represents $S_{c,in}$, the individual curves refer to different $S_{a,in}$, moving from 1 g L^{-1} for the top curve to 5 g L^{-1} for the bottommost curve. As can be seen from Fig. 5, CE decreases with the increase in both $S_{a,in}$ and $S_{c,in}$, and for both, the decrease slows down at higher values as CE reaches a plateau. A similar trend of CE decreasing with increasing $S_{c,in}$ has been previously described in the numerical study by Kazemi *et al.*²³

Mohanakrishna *et al.*³⁴ also studied the effect of $S_{c,in}$ on CE and found it to vary in a rather non-linear fashion. They observed that CE initially decreased from 45.55% to 39.56% as the initial substrate concentration (HCO_3^-) was increased from 1 g L^{-1} to 1.5 g L^{-1} . However with a further increase in HCO_3^- concentration to 2 g L^{-1} and 2.5 g L^{-1} , an average CE of 40.37% and 56.25% respectively, was observed. Their hypothesis was that the increase in bicarbonate concentration increased the conductivity of the catholyte which thereby improved the use of hydrogen for acetate production. However they mentioned that CE showed fluctuations in the different cycles of operation with different HCO_3^- concentrations and highlighted the need for more comprehensive studies.³⁴

Based on the current analysis, the maximum CE of 64.63% was observed for $S_{a,in} = 1 \text{ g L}^{-1}$ and $S_{c,in} = 2.5 \text{ g L}^{-1}$, while the lowest CE (31.2%) was observed at the highest substrate concentrations of $S_{a,in} = 5 \text{ g L}^{-1}$ and $S_{c,in} = 7.5 \text{ g L}^{-1}$. The increase in substrate concentration also results in a simultaneous increase in substrate availability for secondary bacteria at both anode and cathode, and depending on the competition between the individual rates of substrate consumption of the different bacteria, it can result in reduced charge (Coulombs) recovery at the cathode (or lower CE). Thus the relationship between substrate concentrations and CE becomes very specific for each system (depending on the type of substrate and the bacterial inoculum used in the MES) and cannot be generalized. For

the current analysis, a reduction in CE at higher substrate concentrations (which show better product formation rates) will result in poor energy efficiency and will effectively drive up the total operational cost of the system.

By comparing Fig. 3–5, it can be seen that there is a trade-off between product formation rate, and substrate consumption and coulombic efficiency. Similar relationship between the performance factors has been previously shown for microbial fuel cells, with power density increasing with substrate concentration but COD consumption and CE decreasing at higher concentrations.^{35,36} Selecting the specific initial concentrations will depend on the targeted end-use and the economic implications of the different options. The current dynamic model will serve as a good starting point for shortlisting a range of $S_{a,in}$ and $S_{c,in}$ values for improved performance under given operating conditions. The data from the dynamic model can be integrated with techno-economic and life cycle assessment studies for further sustainability analysis, as shown in our previous study.²⁴

3.4 Effect of operation cycle time

In addition to substrate concentration, another important parameter that can have a significant influence on the performance of fed-batch MES systems is the operation cycle time (Ct). For the analysis so far, Ct was fixed as 3 days. Similar to flow rate in continuous systems, Ct determines the residence time of substrates in the fed-batch systems. Reducing or increasing Ct would potentially introduce changes in the AA production rate as well as the substrate consumption rates and CE. Fig. 6A and B show the change in the AA production rates as a function of different initial substrate concentrations for Ct of 2 days and 4 days, respectively. As can be seen from these figures, the effect of increasing initial substrate concentrations on Q_{AA} for Ct of 4 days (Fig. 6B) is very similar to that observed for Ct of 3 days (described in Section 3.3, Fig. 3); however the effect is very different when the Ct is reduced to 2 days (Fig. 6A). It should also be noted that the range of Q_{AA} observed with 4 days Ct for different substrate concentrations is relatively much lower than that observed for either 2 or 3 days of operation cycle time.

For a cycle time of 2 days (Fig. 6A), Q_{AA} first shows a linear increase with the increase in $S_{a,in}$ or $S_{c,in}$; however such linear relationship is only valid at smaller values of substrate concentrations. For all $S_{c,in} > 3.25 \text{ g L}^{-1}$, when $S_{a,in}$ is increased from 1 g L^{-1} to 5 g L^{-1} , Q_{AA} first increases linearly and then reaches a plateau followed by a slight decrease. Similarly for all $S_{a,in} > 1.25 \text{ g L}^{-1}$, when $S_{c,in}$ is increased from 2.5 g L^{-1} to 7.5 g L^{-1} , Q_{AA} increases linearly but soon reaches a plateau after a certain threshold $S_{c,in}$ and then begins to decrease with a further increase in $S_{c,in}$.

The threshold value where the increase in Q_{AA} with $S_{a,in}$ or $S_{c,in}$ reaches a plateau increases at higher values of $S_{c,in}$ or $S_{a,in}$ respectively. This leads to the unique trough observed at higher values of $S_{a,in}$ and $S_{c,in}$ in the surface plot shown in Fig. 6. Compared to a 3 day operation cycle, where the AA production rate varies from 2.6 mM d^{-1} to 7.74 mM d^{-1} , the range of AA production rates for a 2 day operation cycle (5.7 mM d^{-1} to 7.76 mM d^{-1})



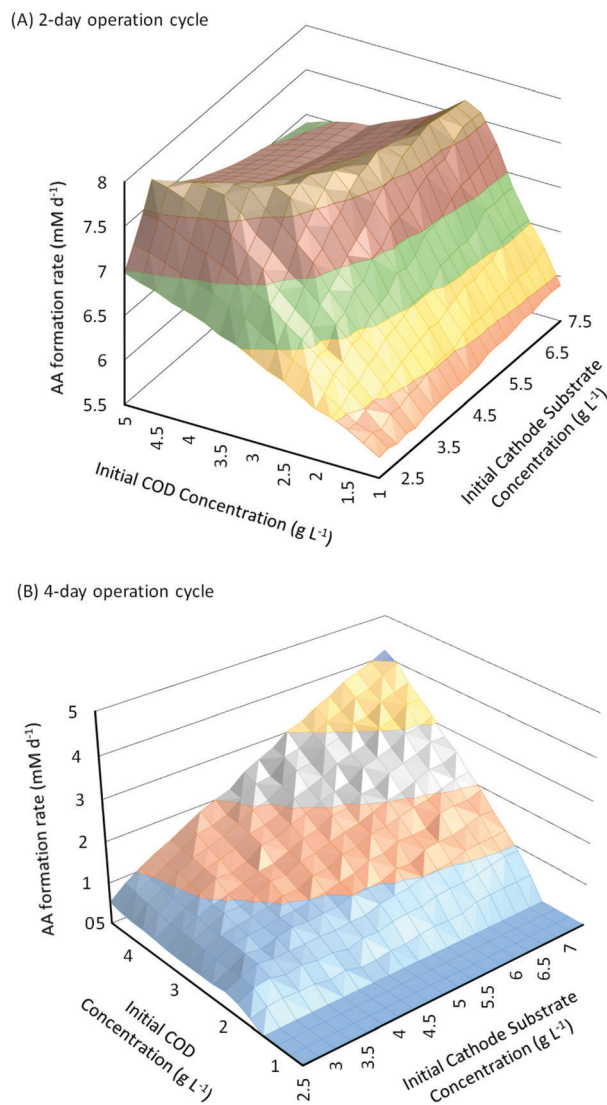


Fig. 6 Predicted AA production rate (mM d^{-1}) as a function of initial substrate concentrations in the anode and cathode chambers for (A) 2 day and (B) 4 day operation cycles.

is much smaller between different substrate concentrations. However this range also suggests that AA production rates are relatively high with Ct of 2 days even at low values of $S_{\text{a,in}}$ and $S_{\text{c,in}}$. Maximum Q_{AA} with Ct of 2 days (7.76 mM d^{-1}) is observed for $S_{\text{a,in}} = 5 \text{ g L}^{-1}$ and $S_{\text{c,in}} = 3.25 \text{ g L}^{-1}$.

When the operation cycle time is increased to 4 days (Fig. 6B), both $S_{\text{a,in}}$ and $S_{\text{c,in}}$ show a linear relationship with Q_{AA} , but the product formation rate is substantially reduced. It should also be noted that Q_{AA} is negligible (~ 0) for $S_{\text{a,in}} < 2 \text{ g L}^{-1}$. This occurs because a longer cycle time allows a higher conversion of AA into secondary products. Thus until Q_{AA} reaches a sufficiently high value (which is obtained for $S_{\text{a,in}} \geq 2 \text{ g L}^{-1}$), all the AA formed is consumed before it can be recovered at the end of the cycle. Higher Ct and in effect higher residence time is also responsible for the overall reduction in Q_{AA} (maximum of 4.3 mM d^{-1} obtained when $S_{\text{a,in}} = 5 \text{ g L}^{-1}$ and $S_{\text{c,in}} = 7.5 \text{ g L}^{-1}$) observed for a 4 day operation cycle.

For the range of values of $S_{\text{a,in}}$ and $S_{\text{c,in}}$ studied in Fig. 6, the amount of substrate consumed and the coulombic efficiency as a function of initial substrate concentrations are shown in Fig. 7 and 8, respectively, for a cycle time of 2 days and 4 days. As can be seen from Fig. 7, percentage substrate consumption for Ct of 2 days is relatively smaller than that observed for 4 days Ct for same values of $S_{\text{a,in}}$ and $S_{\text{c,in}}$. This is expected as Ct decides the residence time of the substrate, and a higher residence time allows for higher consumption of substrates.

Also, as can be seen from Fig. 8, CE for cycle times of 2 and 4 days follows the same trend with substrate concentrations as observed for a cycle time of 3 days (Fig. 5). It should be noted that CE clearly shows a linear relationship with Ct, with the highest CE $\sim 90\%$ being observed for lowest substrate concentrations when Ct is 4 days (Fig. 8B) while the lowest CE $\sim 28\%$ is obtained at highest substrate concentrations when Ct is reduced to 2 days (Fig. 8A).

Comparing the results observed in Fig. 6 with that in Fig. 7 and 8, it can be seen that the specific substrate concentrations which provide the highest AA production rate with a 2 day Ct (7.76 mM d^{-1}) show anode and cathode substrate consumption of just 33% and 60% respectively and CE of 31.5%. Thus even though reducing the cycle time can help to produce higher quantities of the product, the overall operation may not be economically feasible given the poor substrate consumption rates and the low coulombic efficiency. Optimum operating conditions (initial substrate concentrations and cycle time) can be found by selecting the parameter values that provide sufficiently high product formation rates without compromising on the substrate consumption rates. The specific criterion can be decided based on the targeted end-use. For the current system, Ct of 2 days and initial anode and cathode substrate concentration values of 1.75 g L^{-1} and 2.5 g L^{-1} , respectively, provide an AA production rate of 6.02 mM d^{-1} (which is only $\sim 20\%$ smaller than the highest product formation rate observed) and anode and cathode substrate consumption of 75% and 71% respectively with a coulombic efficiency of $\sim 40\%$. These operating conditions can be used to obtain sufficiently high product formation and substrate consumption rates with average coulombic efficiency. If product formation or COD consumption is one of the major goals, specific values that provide highest Q_{AA} or anode substrate consumption could be selected respectively. Predictions obtained from the current model, in terms of operating diagrams (plots described in Fig. 3–8), can be effectively used to obtain the optimum range of operation for a given MES system based on the specific end-use.

3.5 Cycle time controlled by substrate consumption

Ct determines the specific time when the feed is changed in the two electrode chambers. As the amount of substrate consumed is determined by the specific type of bacteria at the anode and cathode and their initial concentrations, the effective rates of consumption are different in the two chambers. As can be seen from Fig. 4 and 7, this often leads to one or both the substrates remaining unconsumed which then adds up to the recycling



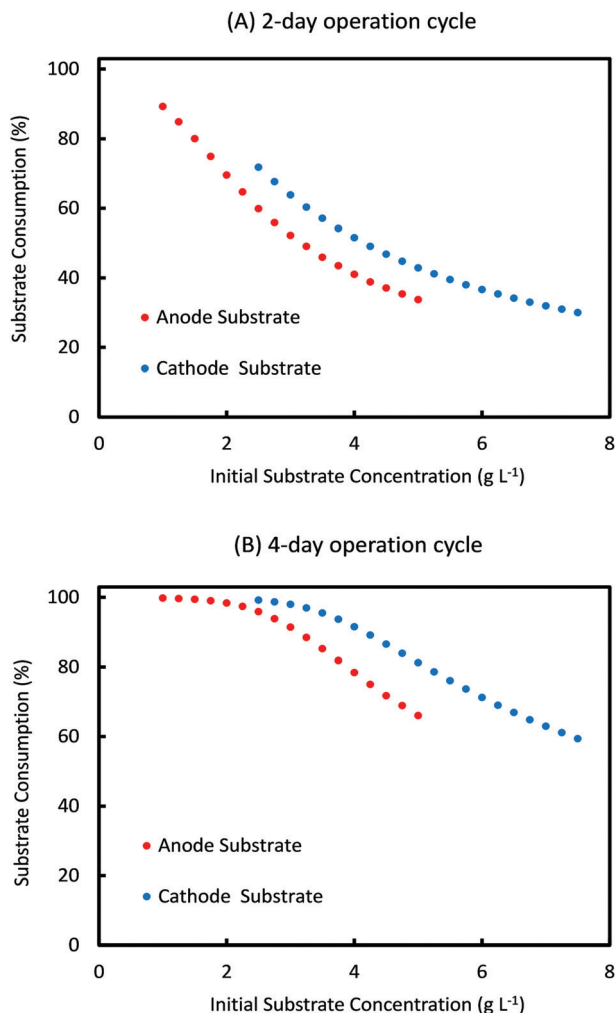


Fig. 7 Simulation prediction of percentage substrate consumption for different initial concentrations of anode and cathode substrates for a cycle time of (A) 2 days and (B) 4 days.

cost or may even result in increasing waste. If C_t is not fixed but is determined by the amount of substrate consumption, it can lead to increased utilization of each substrate and reduced waste. Such a mechanism can be easily implemented with the help of specific sensors which monitor the concentration of substrate in each chamber and accordingly control the replenishment. New feed can be introduced once a certain minimum consumption (for *e.g.* 90 or 95%) has been achieved. However while such a mechanism reduces substrate loss or recycling costs, the installation of the new sensors and the control valves will add to the capital investment. A techno-economic analysis (TEA) of the system can help to determine the difference in operating costs the new mechanism would entail. From the dynamic analysis perspective, it is interesting to understand the effect of a non-fixed C_t (which is only governed by substrate consumption) on the MES performance.

Fig. 9 shows the simulation predictions of the AA production rate as a function of different initial substrate concentrations at the anode and cathode while keeping the initial concentration

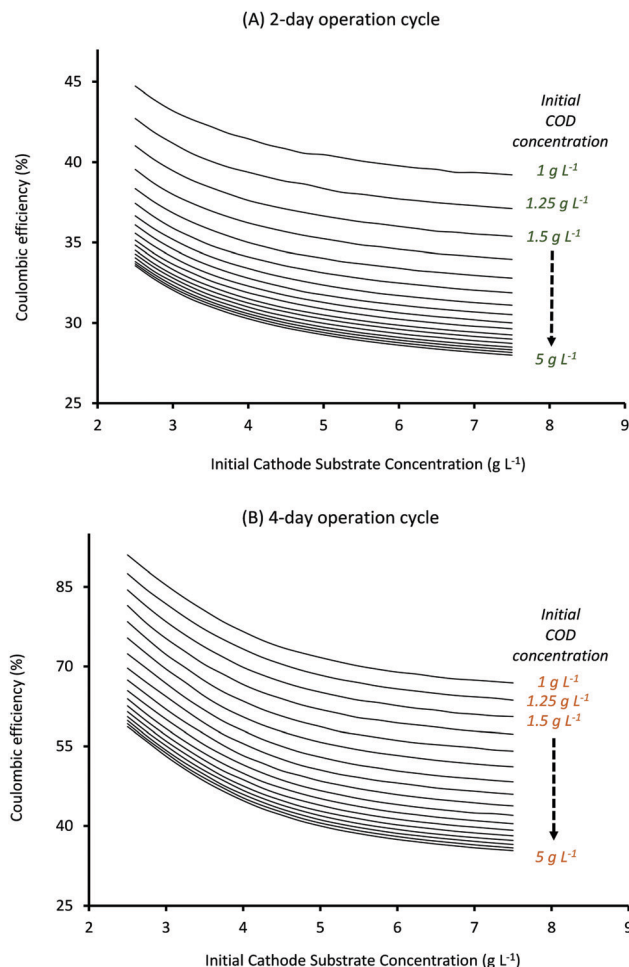


Fig. 8 Predicted AA production rate (mM d^{-1}) as a function of initial substrate concentrations in the anode and cathode chambers for (A) 2 day and (B) 4 day operation cycles.

of other substrate fixed. For this analysis, the substrate feed in each chamber is changed after achieving 95% consumption. As can be seen from Fig. 9, there is no specific pattern observed for Q_{AA} as a function of either $S_{a_{in}}$ or $S_{c_{in}}$. For the anode substrate, Q_{AA} first decreases as $S_{a_{in}}$ is increased from 1 g L^{-1} to 2 g L^{-1} and then remains constant up to $S_{a_{in}} = 4 \text{ g L}^{-1}$. For higher $S_{a_{in}}$ values ($> 4 \text{ g L}^{-1}$), Q_{AA} shows a linear increase with $S_{a_{in}}$. This nonlinear pattern as seen in Fig. 9A is actually expected because the AA production rate is a function of cell current density, which undergoes fluctuation every time either anode or cathode feed is changed. Since the feed change time is not fixed, as opposed to the results described in Sections 3.3 and 3.4, the current density profile is no longer a smooth periodic curve. This difference is highlighted in Fig. 10, which compares the profile of current density as a function of time for a MES cell with fixed C_t of 3 days to a MES cell where C_t is not fixed, but the feed is replenished after achieving 95% substrate consumption in respective chambers. As can be seen from Fig. 10, for the MES cell where C_t is not fixed, the current density profile is no longer smooth and periodic but undergoes arbitrary fluctuations which occur due to feed changes.



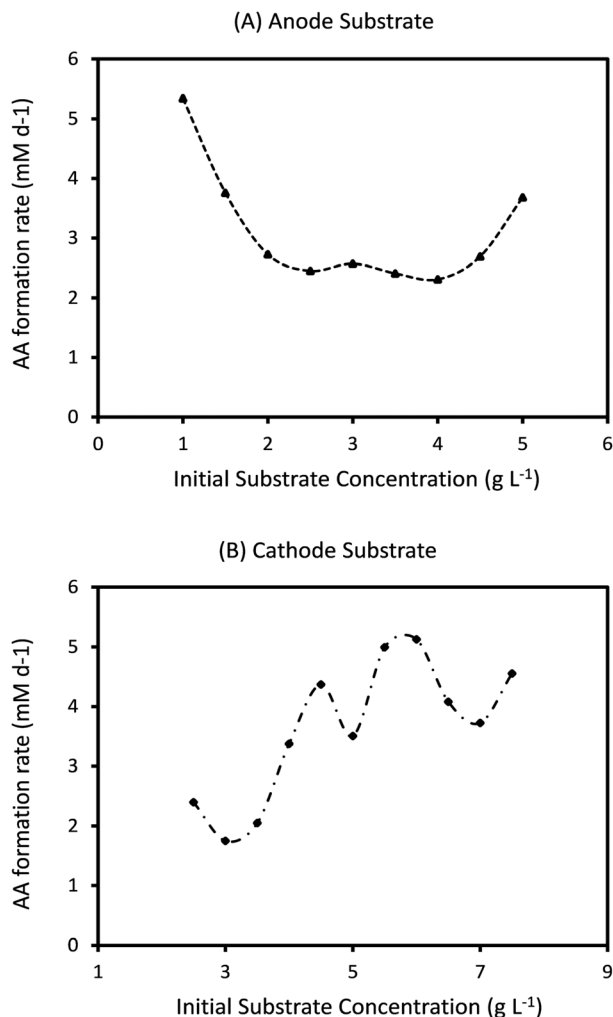


Fig. 9 Predicted AA production rate (mM d^{-1}) as a function of different initial substrate concentrations at (A) anode (with fixed $S_{\text{c,in}} = 5 \text{ g L}^{-1}$) and (B) cathode (with fixed $S_{\text{a,in}} = 1.5 \text{ g L}^{-1}$).

The results shown in Fig. 9A and B refer to the average values of Q_{AA} obtained after 20 cycles of operation. For the specific conditions described in Fig. 9A, the highest $Q_{\text{AA}} = 5.3 \text{ mM d}^{-1}$ is obtained for $S_{\text{a,in}} = 1 \text{ g L}^{-1}$, which is much lower than the maximum Q_{AA} observed for MES operated with fixed Ct of 2 or 3 days. Also, Q_{AA} observed under the operating conditions ($S_{\text{a,in}} = 1 \text{ g L}^{-1}$ and $S_{\text{c,in}} = 5 \text{ g L}^{-1}$) which provide the maximum production rate with a non-fixed Ct (Fig. 9A) is close to 100% higher than that observed when Ct is fixed at 3 days and about 10% lower than that with Ct of 2 days. For the cathode substrate, as can be seen in Fig. 9B, with increasing $S_{\text{c,in}}$, Q_{AA} shows sinusoidal variation with an upward trend. The maximum $Q_{\text{AA}} = 5.1 \text{ mM d}^{-1}$, observed at $S_{\text{c,in}} = 6 \text{ g L}^{-1}$, is even lower than that obtained with changing $S_{\text{a,in}}$ (Fig. 9A). However under the same operating conditions ($S_{\text{a,in}} = 1.5 \text{ g L}^{-1}$ and $S_{\text{c,in}} = 6 \text{ g L}^{-1}$) that provide maximum Q_{AA} when Ct is not fixed (Fig. 9B), Q_{AA} is $\sim 52\%$ lower when Ct is fixed at 3 days but about 25% higher when Ct is reduced to 2 days.

Overall the results show the complex interdependence between the different operating parameters and their influence

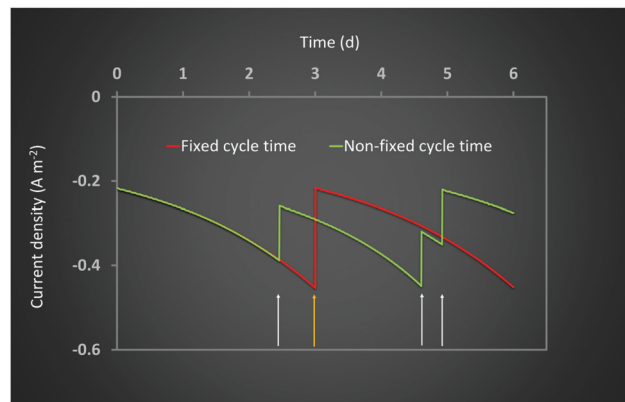


Fig. 10 Prediction of current density profiles during MES operation (A) with a fixed cycle time of 3 days and (B) with a non-fixed cycle time where the substrate is replenished in the electrode chambers after 95% consumption has been achieved. For both the cases, $S_{\text{a,in}}$ and $S_{\text{c,in}}$ were fixed at 1.5 g L^{-1} and 5 g L^{-1} respectively. The white arrows indicate feed change for MES with non-fixed cycle time, whereas the yellow arrow represents the same for MES with fixed cycle time.

on the MES performance. Based on the current analysis, it can be seen that before an optimum set of conditions can be selected, it is important to correlate the effect of any given parameter as a function of other dependent parameters, and also how it affects the different performance factors. The problem becomes more daunting when along with the design and operational parameters, the biological factors (different for various bacterial strains) also need to be accounted.

The proposed dynamic model presented above has wide applicability in parametric studies to understand the influence of different parameters on the MES performance and can be used for multi-objective optimization. Based on the targeted end-use, specific constraints such as product formation rate above a certain value, substrate consumption rate greater than a fixed percentage, specific coulombic efficiency, initial substrate concentrations above or below a specific value depending on the feed, limitations of growth rate or substrate consumption rates depending on the inoculum, *etc.*, can be included in the model to determine the optimum operating conditions for MES design and operation for the particular end-use.

The inclusion of bioelectrochemical kinetics and potential losses at both anode and cathode makes this model unique and an important advancement over previous modeling studies of microbial electrosynthesis systems. The one-dimensional nature of the model, though restrictive by not accounting for the spatial dynamics of the biofilm and its interaction with the substrate and the electrodes, does provide fast computation without being overly simplistic.

4 Conclusion

In this study, a detailed mathematical model is presented for the wastewater valorization and carbon dioxide reduction to carboxylic acids in a two chamber microbial electrosynthesis



system. The dynamic model is used to study the effect of initial substrate concentrations of anode and cathode and the operation cycle time on the rate of product formation, substrate consumption and coulombic efficiency. While an increase in initial substrate concentrations generally improved the product formation rates, it reduced the substrate consumption as well as CE. On the other hand reducing the operation cycle time favored product formation rates but reduced the substrate consumption and CE. The dynamic model is also used to evaluate MES performance when cycle time is not fixed but is controlled by substrate consumption rate. To conclude, the proposed model can be used for effective multi-objective optimization of the different operating parameters of MES to achieve optimum performance based on the targeted end use. Integration of these dynamic simulation results with techno-economic and life cycle analysis can provide a holistic understanding of the overall sustainability of MES.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 K. Rabaey and R. A. Rozendal, *Nat. Rev. Microbiol.*, 2010, **8**, 706–716.
- 2 J. Sadhukhan, J. R. Lloyd, K. Scott, G. C. Premier, H. Y. Eileen, T. Curtis and I. M. Head, *Renewable Sustainable Energy Rev.*, 2016, **56**, 116–132.
- 3 Y. Jiang, H. D. May, L. Lu, P. Liang, X. Huang and Z. J. Ren, *Water Res.*, 2019, **149**, 42–55.
- 4 J. A. Modestra and S. V. Mohan, *J. CO₂ Util.*, 2017, **20**, 190–199.
- 5 S. Cheng, D. Xing, D. F. Call and B. E. Logan, *Environ. Sci. Technol.*, 2009, **43**, 3953–3958.
- 6 Y. Gong, A. Ebrahim, A. M. Feist, M. Embree, T. Zhang, D. Lovley and K. Zengler, *Environ. Sci. Technol.*, 2012, **47**, 568–573.
- 7 Y. Xiang, G. Liu, R. Zhang, Y. Lu and H. Luo, *Bioresour. Technol.*, 2017, **233**, 227–235.
- 8 J. Desloover, J. B. Arends, T. Henebel and K. Rabaey, *Biochem. Soc. Trans.*, 2012, **40**, 1233–1238.
- 9 K. P. Nevin, T. L. Woodard, A. E. Franks, Z. M. Summers and D. R. Lovley, *mBio*, 2010, **1**, e00103.
- 10 C. W. Marshall, D. E. Ross, E. B. Fichot, R. S. Norman and H. D. May, *Appl. Environ. Microbiol.*, 2012, **78**, 8412–8420.
- 11 J. A. Modestra, B. Navaneeth and S. V. Mohan, *J. CO₂ Util.*, 2015, **10**, 78–87.
- 12 K. P. Nevin, S. A. Hensley, A. E. Franks, Z. M. Summers, J. Ou, T. L. Woodard, O. L. Snoeyenbos-West and D. R. Lovley, *Appl. Environ. Microbiol.*, 2011, **77**, 2882–2886.
- 13 F. Wagner, in *Encyclopedia of Chemical Technology*, ed. K. Othmer, John Wiley & Sons, 2000, pp. 63–64.
- 14 V. H. Agreda, *Acetic acid and its derivatives*, CRC Press, 1992.
- 15 P. Pal and J. Nayak, *Sep. Purif. Rev.*, 2017, **46**, 44–61.
- 16 Y. Jiang and R. J. Zeng, *Bioresour. Technol.*, 2018, **269**, 503–512.
- 17 H. Nie, T. Zhang, M. Cui, H. Lu, D. R. Lovley and T. P. Russell, *Phys. Chem. Chem. Phys.*, 2013, **15**, 14290–14294.
- 18 F. Harnisch and C. Urban, *Angew. Chem., Int. Ed.*, 2018, **57**, 10016–10023.
- 19 D. Recio-Garrido, M. Perrier and B. Tartakovsky, *Chem. Eng. J.*, 2016, **289**, 180–190.
- 20 S. Gadkari, S. Gu and J. Sadhukhan, *Chem. Eng. J.*, 2018, **343**, 303–316.
- 21 S. Luo, H. Sun, Q. Ping, R. Jin and Z. He, *Energies*, 2016, **9**, 111.
- 22 R. Pinto, B. Srinivasan, A. Escapa and B. Tartakovsky, *Environ. Sci. Technol.*, 2011, **45**, 5039–5046.
- 23 M. Kazemi, D. Biria and H. Rismani-Yazdi, *Phys. Chem. Chem. Phys.*, 2015, **17**, 12561–12574.
- 24 M. Shemfe, S. Gadkari, E. Yu, S. Rasul, K. Scott, I. Head, S. Gu and J. Sadhukhan, *Bioresour. Technol.*, 2018, **255**, 39–49.
- 25 N. S. Panikov, *Microbial Growth Kinetics*, Springer Science & Business Media, 1995.
- 26 B. Korth and F. Harnisch, in *Bioelectrosynthesis*, ed. F. Harnisch and D. Holtmann, Springer International Publishing, Cham, Switzerland, 2019, pp. 273–325.
- 27 H. V. Hamelers, A. Ter Heijne, N. Stein, R. A. Rozendal and C. J. Buisman, *Bioresour. Technol.*, 2011, **102**, 381–387.
- 28 M. Esfandyari, M. A. Fanaei, R. Gheshlaghi and M. A. Mahdavi, *Int. J. Hydrogen Energy*, 2017, **42**, 21198–21202.
- 29 R. Pinto, B. Srinivasan, S. Uiot and B. Tartakovsky, *Water Res.*, 2011, **45**, 1571–1578.
- 30 O. Choi, Y. Um and B.-I. Sang, *Biotechnol. Bioeng.*, 2012, **109**, 2494–2502.
- 31 J. A. Nelder and R. Mead, *Comput. J.*, 1965, **7**, 308–313.
- 32 L. Surhone, M. Timpledon and S. Marseken, *Nelder–Mead Method: Simplex Algorithm, Optimization (Mathematics), Numerical Analysis, Unimodal Function, Heuristic, John Nelder, Simplex, Polytope*, Betascript Publishing, 2010.
- 33 A. K. Mostafazadeh, P. Drogui, S. K. Brar, R. D. Tyagi, Y. Le Bihan and G. Buelna, *J. Environ. Chem. Eng.*, 2017, **5**, 940–954.
- 34 G. Mohanakrishna, K. Vanbroekhoven and D. Pant, *React. Chem. Eng.*, 2018, **3**, 371–378.
- 35 Y. Feng, X. Wang, B. E. Logan and H. Lee, *Appl. Microbiol. Biotechnol.*, 2008, **78**, 873–880.
- 36 T. H. Sleutels, L. Darus, H. V. Hamelers and C. J. Buisman, *Bioresour. Technol.*, 2011, **102**, 11172–11176.

