Targeting the *Mycobacterium tuberculosis* transpeptidase LdtMt2 with cysteine-reactive inhibitors including ebselen†

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The L,D-transpeptidases (Ldts) are promising antibiotic targets for treating tuberculosis. We report screening of cysteine-reactive inhibitors against LdtMt2 from *Mycobacterium tuberculosis*. Structural studies on LdtMt2 with potent inhibitor ebselen reveal opening of the benzisoselenazolone ring by a nucleophilic cysteine, forming a complex involving extensive hydrophobic interactions with a substrate-binding loop.

Tuberculosis (TB) is the leading cause of death associated with a single infectious agent worldwide.1 Whilst current TB therapies can be effective, the treatment duration, the need for co-administration of multiple drugs, side effects, and limited drug availability in developing countries hinders successful treatment in many highly-affected regions.2 In recent years multi-drug resistant (MDR) and extensively-drug resistant (XDR) strains of the causative agent, *Mycobacterium tuberculosis*, have emerged.2 There is thus a pressing need for improved and inexpensive TB therapies which target resistant strains and which can shorten treatment duration.

The cell wall peptidoglycan of most Gram-negative bacteria consists primarily of 4 → 3 peptide cross-links between meso-diaminopimelate (meso-Dap) and D-alanine residues (i.e., meso-Dap-D-Ala cross-links). The formation of these 4 → 3 cross-links is catalysed by the n,D-transpeptidases (or penicillin-binding proteins; PBPs), which employ a nucleophilic serine residue. The PBPs are the primary targets of the β-lactam antibacterials, which are of immense clinical importance, but which, despite promise,3–5 have not been developed for clinical use against TB. In *M. tuberculosis*, the cell wall contains high levels of 3 → 3 (meso-Dap–meso-Dap) cross-links (approx. 80% at stationary phase), which are formed by the L,D-transpeptidases (Ldts) (Fig. S1, ESI†).6 Evidence has emerged that LdtMt2, in particular, plays an important role in the virulence of *M. tuberculosis*, as disruption of the *ldtMt2* gene results in altered morphology and inhibition of colony growth.7 Mechanistically, the Ldts differ from the PBPs through their use of a nucleophilic cysteine rather than serine during catalysis.8

Several β-lactam antibiotics, in particular carbapenems, have been shown to inhibit the activity of LdtMt2 and to have anti-TB activity.3,5,10–12 Such LdtMt2 inhibition involves covalent modification of the nucleophilic cysteine (Cys354), which reacts with β-lactam antibiotics to give (a) stable acyl–enzyme complex(es) (Fig. 1A).5,10,11 Despite the promise associated with the treatment of TB using β-lactams, their application is hindered by cost, stability, and delivery issues (due in part to the need to target *M. tuberculosis* present in macrophages). There is therefore interest in developing alternative ways of inhibiting the Ldts and, more generally, mycobacterial transpeptidases (including PBPs). As targeting nucleophilic cysteine residues is a validated method for inhibitor development for human intracellular targets,13–15 we were interested in exploiting such an inhibition strategy for the treatment of TB. Here we report the application of a fluorescence-based assay for LdtMt2 for the identification of cysteine-reactive reagents, including the drug candidate ebselen,16 as promising inhibitors of the Ldts.

Due to the limitations associated with spectrophotometric assays for the Ldts (e.g., poor sensitivity and high protein requirements),13,10 we devised an alternative fluorescence-based assay for screening LdtMt2 inhibitors (manuscript in preparation).17 Although a number of fluorogenic probes selective for small molecule thiols (e.g., cysteine) are reported,9,18,19 to our knowledge they have not been applied with the Ldts for assay development. We chose to focus on the 2,4-dinitrobenzenesulfonyl fluorescein probe 1 (Fig. 1B), due to the ease of synthesis and the strong fluorescent signal associated with the fluorescein fluorophore.9

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We applied this assay to study cysteine-reactive reagents as potential LdtMt2 inhibitors (Fig. 1C). Commonly used methods for the covalent modification of cysteine include thiol-reactive groups as seen in the haloacetyl reagent iodoacetamide (2) and the Michael acceptor maleimide (3). Moreover, sulphur- and selenium-based compounds are known for their ability to interact with cysteine residues, therefore, compounds 4–11 (including ebselen) were included in the screen. The penem faropenem (11), a known inhibitor of LdtMt2, was used as a positive control.

Although compounds 2–11 were all observed to inhibit LdtMt2 (Table 1 and Fig. S2, ESI†), substantial variations in their activities were observed. IC50 values were dependent on incubation time, consistent with a covalent inactivation mechanism. Ebselen (4) was the most potent inhibitor identified, with an IC50 of 0.36 μM with no pre-incubation, and an IC50 of 0.143 μM following 60 minutes of pre-incubation; both IC50 values are significantly smaller than those obtained for the positive control faropenem (11) under similar conditions (i.e., IC50 of 0.686 μM following a 60 minute pre-incubation).

Ebselen has been shown to be an effective cysteine-targeting reagent in previous studies, inhibiting the enzymes γ-butyrobetaine hydroxylase and JMJD2A through interaction with cysteine residues. In addition, ebselen has been in late stage clinical development as a treatment for stroke, and is currently being investigated for the treatment of bipolar disorder and hearing loss. The other selenium-containing compounds tested, i.e., 5–7, were significantly less active than ebselen (Table 1). The sulphur-based compounds 8–10 were also not as potent as ebselen, but following a 1 hour pre-incubation, all manifested substantial inhibition; in particular, thiram (8) was the most potent of these compounds, and showed inhibition comparable with that of faropenem (11). The activities of both iodoacetamide (2) and maleimide (3) were highly dependent on the length of the pre-incubation period, indicating slow reaction with LdtMt2.

Having shown that the inhibitory activity of compounds 2–11 is time dependent, we investigated their interaction with LdtMt2 using protein mass spectrometry (MS; Table S1, ESI†). Mass spectra obtained for LdtMt2 treated with a 10-fold excess of compounds 2–11 showed the formation of adducts consistent with reaction with a single molecule of inhibitor under these conditions. Maleimide (3) and ebselen (4) appeared to react with LdtMt2 without fragmentation, while reaction with iodoacetamide (2), PhSeBr (5) and PhSeCl (6) was accompanied with the loss of the corresponding halide. PhSe(O)OH (7) reacted to form a similar adduct as 5 and 6. The molecular weights of the adducts formed with thiram (8), aldrithiol (9) and PX-12 (10) indicated that these inhibitors likely react via disulphide exchange.

We then used high-throughput mass spectrometry to examine the rate of adduct formation for LdtMt2 with two equivalents of inhibitor (Fig. S3 and S4, ESI†). While ebselen (4) fully reacted with the enzyme within 1 min of addition, the other selenium-containing compounds investigated (i.e., 5–7) reacted much more slowly (Fig. S3, ESI†). Complete reaction between LdtMt2 and aldrithiol (9) was observed within 10 minutes. Although thiram and the other sulphur-based compounds (8–10) reacted more slowly, complete reaction was observed in all cases within 45 minutes (within detection limits). Similar results were obtained with iodoacetamide (2). In agreement with the high degree of time dependence observed for it in the dose–response analysis, maleimide (3) reacted relatively slowly, with complete reaction not being observed by 82 minutes. The covalent

### Table 1 The inhibitory activity of compounds 2–11 with LdtMt2

<table>
<thead>
<tr>
<th>Compound</th>
<th>IC50 (μM, mean ± SD)</th>
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<tbody>
<tr>
<td>2 Iodoacetamide</td>
<td>129 ± 14</td>
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<tr>
<td>3 Maleimide</td>
<td>158 ± 9</td>
</tr>
<tr>
<td>4 Ebselen</td>
<td>0.36 ± 0.02</td>
</tr>
<tr>
<td>5 PhSeBr</td>
<td>11.8 ± 1.3</td>
</tr>
<tr>
<td>6 PhSeCl</td>
<td>62.8 ± 2.2</td>
</tr>
<tr>
<td>7 PhSe(O)OH</td>
<td>309 ± 7</td>
</tr>
<tr>
<td>8 Thiram</td>
<td>7.01 ± 0.29</td>
</tr>
<tr>
<td>9 Aldrithiol</td>
<td>22.1 ± 0.7</td>
</tr>
<tr>
<td>10 PX-12</td>
<td>27.2 ± 0.5</td>
</tr>
<tr>
<td>11 Faropenem</td>
<td>1.69 ± 0.17</td>
</tr>
</tbody>
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a The impact of inhibitors on LdtMt2 was tested without pre-incubation, or with 10 min or 60 min pre-incubation prior to addition of fluorogenic probe 1.
LdtMt2 crystals were soaked with ebselen, with the conformation and a C-terminal catalytic domain (Fig. 2A). The overall fold consists of two N-terminal immunoglobulin fold-related domains and a C-terminal catalytic domain (Fig. 2A). The overall fold of LdtMt2 in our structure aligns well with reported LdtMt2 structures with a root-mean-square-deviation of 0.70 Å for backbone Cα atoms compared to PDB entry 5D7H.

LdtMt2 crystals were soaked with ebselen, and the structure of the complex was solved by molecular replacement (Table S2, ESI†). Consistent with previous reports, the structure of LdtMt2 consists of two N-terminal immunoglobulin fold-related domains and a C-terminal catalytic domain (Fig. 2A). The overall fold of LdtMt2 in our structure aligns well with reported LdtMt2 structures with a root-mean-square-deviation of 0.70 Å for backbone Cα atoms compared to PDB entry 5D7H.

Consistent with previous reports, the structure of LdtMt2 contains a catalytic active-site loop region (lid) of the catalytic domain (residues 300–323) including a two-stranded β-sheet encompassing residues 300–323) including Met303, Tyr308, Tyr318, and Thr320, contribute to the hydrophobic pocket around Cys354 (Fig. 2A). Apparent pi-stacking between the phenol ring of Tyr318 and the ebselen-derived phenyl ring proximal to Cys354 (ring A in Fig. 2A) is present in conformations I and II; this interaction has also been observed in the complex derived from LdtMt2 and 5,5′-dithiobis(2-nitrobenzoic acid). The position of the ebselen-derived phenyl ring distal to Cys354 (ring B) depends on the conformation of ebselen (Fig. 2A); in conformation I, ring B appears to pi-stack with His352, while in conformation II, it interacts primarily with Val333.

The position of the active-site lid (residues 300–323; Fig. 2A) is apparently altered by ebselen, with its precise orientation appearing to depend on the conformation of ebselen (i.e., I or II; Fig. 2B). Previous work has suggested that modification of Cys354 by an inhibitor (e.g., carbapenems, penems) leads to conformational changes in the lid which stabilize the inhibitor-enzyme complex, thereby contributing to inhibitor potency.

A comparison of our structures with reported LdtMt2 complex structures reveals variations in the conformation of the lid depending on the nature of the modification to Cys354 (Fig. 2C). It appears that the hydrophobic residues of the lid...
(e.g., Tyr308, Met303, Tyr318, Thr320) can adjust to accommodate the group bonded to Cys354, apparently to optimise hydrophobic interactions. These results imply that the lid is conformationally dynamic, with the precise structure observed being related to the nature of modification of the nuclophilic Cys354. Thus, it seems likely that there is considerable scope for induced fit during catalysis and inhibition of LdtM2, and by implication other Ldts.

The combined results demonstrate the potential of non-β-lactam compounds to inhibit LdtM2 via reaction with its nuclophilic cysteine, the probable mechanism for most, if not all of the compounds investigated. There is likely very considerable scope for application of this general mechanism for Ldt inhibition, perhaps building on efforts to target cysteine nucleophiles in proteases, for chemical biology purposes. Of the cysteine-targeting reagents, ebselen was found to be the most potent against LdtM2. Crystallographic analyses indicate that this potency relates to hydrophobic interactions involving the active-site lid of LdtM2, and the conformation of this lid appears to depend on the nature of the modification to the nuclophilic Cys354. Whilst ebselen is almost certainly non-selective, there is clear potential for optimisation of the general approach. Overall, we hope the results presented here will help enable and inspire efforts to explore targeting the Ldts for TB treatment.

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Conflicts of interest

There are no conflicts to declare.

Notes and references

17 M. de Munnik, C. T. Lohans, G. W. Langley, C. Bon, J. Brem and C. J. Schofield, unpublished work.