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Deswelling behaviour of ionic microgel particles from low to ultra-high densities†

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The swelling of ionic microgel particles is investigated at a wide range of concentrations using a combination of light, X-ray and neutron scattering techniques. We employ a zero-average contrast approach for small-angle neutron scattering experiments, which enables a direct determination of the form factor at high concentrations. The observed particle size initially decreases strongly with the particle concentration in the dilute regime but approaches a constant value at intermediate concentrations. This is followed by a further deswelling at high concentrations above particle overlap. Theory and experiments point at a pivotal contribution of dangling polymer ends to the strong variation in size of ionic microgels, which presents itself mainly through the hydrodynamics properties of the system.

Introduction

Colloids are a popular model system to study various phenomena in condensed matter physics^{1–6} as length and time scales are significantly larger than for atomic systems. The latter provides access to a variety of structural and dynamic properties using methods such as neutron, X-ray and light scattering together with different optical microscopy techniques. The phase behaviour of spherical colloids has been extensively investigated with respect to number density, charge and external fields.^{1–3,7–9} While research mainly focused on (charged) hard sphere particles historically, soft colloidal particles have demonstrated a surprisingly rich and intriguing phase behaviour. Thus, both the theoretical and experimental focus has increasingly shifted towards soft repulsive colloids such as neutral poly(*N*-isopropylacrylamide), PNIPAM, microgels.^{10–13}

PNIPAM microgels are spherically-shaped polymeric networks with a radially inhomogeneous mass distribution. The particle softness can be controlled by varying the amount of added crosslinker during synthesis. Moreover, several properties of

these particles can also easily be adjusted *in situ*, which originates from the multi-responsive nature of the polymer network towards different external stimuli. The most prominent one is the temperature; the initial hydrophobicity of the polymers vanishes below 32 °C and yields a swift transition from dense colloidal spheres to swollen microscopic hydrogels. Furthermore, the intrinsic soft interactions between the swollen particles enable packings to effective volume fractions, ϕ_{eff} , far above hard sphere close packing, ϕ_{cp} , where the centre-to-centre distance, a_{ss} , can be smaller than the particle diameter. This has significant consequences for the resulting structural and dynamic properties of the system. Interesting equilibrium and non-equilibrium phases have been observed in theoretical as well as experimental studies of these systems.^{11,14,15}

Ionic microgels contain charged groups which can be used to control inter-particle forces by means of pH or salt concentration. The interplay between the electrostatics and the soft-repulsive potential arising from the elasticity of the backbone could result in a very rich and complex phase behaviour.^{16,17} Attempts have been made to directly measure the (pH-dependent) pair potential in charged microgel systems,¹⁸ and it has been suggested that ionic microgels at low concentrations interact *via* an effective pair potential that crosses over from Yukawa-like, at large separation distances, to a soft-core interaction at shorter distances.^{19–23} A recent theoretical study on ionic microgels close to infinite dilution shows a significantly stronger repulsion upon deswelling at increased temperatures,²⁴ which can be explained by the expulsion of counterions from the core upon deswelling.

Experiments on dilute ionic microgel suspensions have shown that the temperature-induced swelling behaviour is highly dependent on the ionic strength of the solvent.²⁵ Specifically, in deionized

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† Electronic supplementary information (ESI) available: Determination of the ZAC solvent, information on the measured pH, as well as the effect of salt on particle size is included. Fits to the light-scattering data using an additional Ornstein–Zernike like contribution, as referred to in the main text, is included as well. See DOI: 10.1039/c8sm00390d

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systems the collapse temperature of the particles can shift several degrees Celsius.²⁶ It has also been shown that ionic microgels shrink at higher particle concentrations,²⁷ which is believed to be due to a decreasing ion-osmotic pressure gradient between the inside and the outside of the particles. The latter effect is absent in neutral PNIPAM colloids, for which a reduction in size is only observed at concentrations above overlap.²⁸

To understand the complex interactions between particles in charged systems, a detailed understanding of the concentration-induced deswelling behaviour is pivotal. Attempts have been made by Romeo *et al.* to determine the deswelling of charged microgels upon raising the particle concentrations.²⁹ In contrast to Holmqvist *et al.*²⁷ these authors, however, reported on the absence of counter-ion induced deswelling at effective volume fractions below one.

To elucidate some of these seemingly contradictory findings, we have resorted to a combined experimental and theoretical study of the deswelling behaviour of ionic PNIPAM particles from very low concentrations to concentrations far above particle overlap. The experimental studies have been performed using a combination of light, neutron and X-ray scattering techniques. At high concentrations, small-angle neutron scattering (SANS), in combination with the zero average contrast (ZAC) approach, enables us to study single-particle properties, such as size and shape, regardless of their interactions. At concentrations that are too low for SANS, we rely on an analysis of static and dynamic structure factors from light and X-ray scattering. This combined analysis indicates a non-trivial relation between particle size and particle concentration. The experimental data can be analysed using traditional microgel models such as the fuzzy-sphere model, yet we also include a bottom-up approach that accounts for the heterogeneous cross-linker distribution in the microgel core as well as simulations of the dangling polymer ends. The observed deswelling behaviour observed with different experimental techniques may be explained by large differences between the static and hydrodynamic size of ionic microgels.

Experimental

Material

Ionic hydrogenated, Hm, and deuterated, Dm, PNIPAM particles were synthesised by precipitation polymerisation in the presence of surfactant in order to control the final size of the particles. Particles were synthesised in 85 g of water using *N*-isopropylacrylamide (NIPAM, Acros Organics) or d7-*N*-isopropylacrylamide (dNIPAM, Polymer Source) as the monomeric unit. *N,N*-Methylenebis(acrylamide) (BIS, Sigma-Aldrich) was used as a crosslinker and sodium dodecyl sulfate (SDS, Duchefa Biochemie) as a surfactant. In order to incorporate internal titratable charges, acrylic acid or deuterated acrylic acid (AA or dAA, Acros or Polymer Source) diluted in 10 g of water was added to the reaction mixture. Potassium persulfate (0.05 g dissolved in 5 g water, Sigma Aldrich) was used as initiator. Except for NIPAM, that was re-crystallised in hexane prior to use,

Table 1 Added chemicals in the particle synthesis. The particle name Dm corresponds to the use of deuterated monomer and acrylic acid. R_h is the hydrodynamic radius obtained from light scattering close to infinite dilution at a pH of 3.2

Particle name	NIPAM (g)	BIS (g)	AA (g)	SDS (g)	R_h (nm, 20 °C)
Hm	1.43	0.112	0.079	0.065	102
Dm	1.52	0.112	0.083	0.062	115

all chemicals were used as received. The exact amount of the chemicals used can be looked up in Table 1. When deuterated particles were synthesised, the monomeric unit and the acrylic acid used were deuterated. The reactions were carried out in a three-necked round bottom flask under a constant flow of argon in combination with constant magnetic stirring at 250 rpm. The temperature was stabilised at 70 °C before the injection of initiator took place. The suspensions were left to react for 4 h at 70 °C and thereafter left to cool down over night in room temperature under constant stirring. The suspensions were filtered and then dialysed for 2 weeks against deionised water until the conductivity was found to be below 1 $\mu\text{S cm}^{-1}$. The suspensions were thereafter freeze-dried in order to prepare samples with well-controlled weight fractions.

Methods

To achieve the lowest possible ionic strength, the samples were kept on ion-exchanging resins (Amberlite IRN 150, Fluka), except for the highly concentrated samples due to their high viscosity. The hydrodynamic radius, R_h , close to infinite dilution, as a function of temperature was obtained by means of dynamic light scattering using a modulated 3D light scattering instrument (LS instruments GmbH, Switzerland), implementing a laser with a wavelength of 660 nm. For these measurements the pH was lowered to 3.2, which renders the particles uncharged and prevents concentration induced deswelling effects. The data were obtained over an angular range of $40^\circ \leq \theta \leq 60^\circ$ with a step size of 10° and analysed by a first-order cumulant analysis. Using static light scattering, and for both the Hm and Dm particle batches separately, we correlated the weight fraction with the number density by following the evolution of the structure peak with increasing particle concentration. This was performed in an angular range of $30^\circ \leq \theta \leq 130^\circ$ and enabled us to make equal number density mixtures of Hm and Dm for the neutron scattering experiments from samples with known weight concentrations.

Small-angle neutron scattering experiments were performed at D11 at Institut Laue-Langevin, ILL, France. The zero-average contrast (ZAC) solvent, *e.g.* the solvent in which Hm and Dm contribute to the scattered intensity equally, was evaluated by a contrast-variation study. The used amounts of H₂O and D₂O were varied for a fixed low number density of Hm and Dm particles, respectively. Due to the strong electrostatic repulsion between particles even in the dilute regime, the structure peaks were cancelled, as confirmed by static light scattering, by addition of 5 mM potassium chloride (KCl). The details of the



determination of the ZAC solvent to be at 48 vol% of H₂O in D₂O can be found in the ESI† (Fig. S.1). A concentration series of equal density mixtures of Hm and Dm particles, was thereafter measured in the obtained ZAC solvent, without added salt, covering a wide span of concentrations at 20 °C in the fully deionised state. Small-angle X-ray scattering (SAXS) experiments were performed on the same samples as measured by SANS at the cSAXS beam line at Paul Scherrer institute, PSI, Switzerland. In the low-concentration regime, static and dynamic structure factors were obtained using the light-scattering instrument mentioned above over an angular range of $30^\circ \leq \theta \leq 150^\circ$ using a step size of 2° .

Results

The hydrodynamic radius as a function of temperature was measured for both sets of uncharged particles at lowered pH (3.2) close to infinite dilution. The similarities in size can be seen in Fig. 1. Furthermore, the characteristic volume phase transition, VPT, at around 32 °C is captured for Hm particles while the VPT temperature for Dm particles is shifted up by approximately 5 °C due to the isotope effect. The absence of a structure factor peak in the SANS results in Fig. 2A demonstrates that the structure factors were adequately cancelled over a broad range of concentrations at 20 °C when the samples were prepared in the ZAC solvent.³⁰ This enables us to proceed and determine the change in size and shape of individual particles. To cover the required q -range two instrumental settings were used, while the data from the two settings can be fitted simultaneously to form factors for microgel particles. We consider the fuzzy-sphere model

$$P(q) = \left[\frac{3(\sin(qR_{\text{HC}}) - qR_{\text{HC}} \cos(qR_{\text{HC}}))}{(qR_{\text{HC}})^3} \exp\left(-\frac{(\sigma q)^2}{2}\right) \right]^2, \quad (1)$$

which fits the experimental data using a hard-core radius R_{HC} , as well as a fuzziness parameter σ that characterizes the width of the smeared particle surface.¹⁰ Consecutively, an empirical relation for the effective particle size $R_{\text{FS}} = R_{\text{HC}} + 2\sigma$ is used to find agreement with measured hydrodynamic radii of PNIPAM particles.

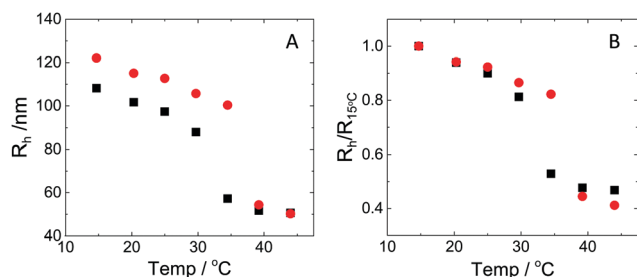


Fig. 1 (A) Hydrodynamic radius as a function of temperature for Hm (squares) and Dm (circles). (B) Hydrodynamic radius normalised by the hydrodynamic radius at 15 °C as a function of temperature for Hm (squares) and Dm (circles). The pH of the solutions were set to 3.2 to avoid concentration-induced deswelling effects.

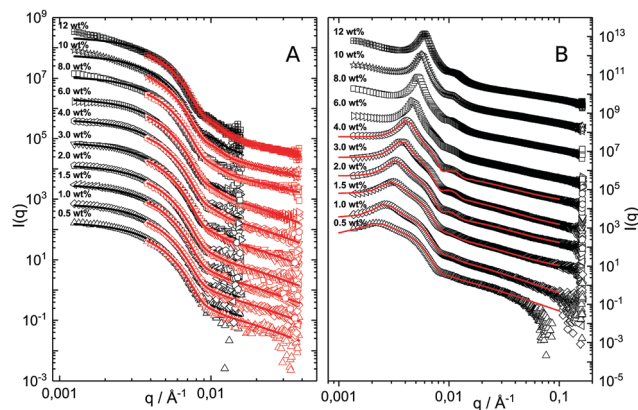


Fig. 2 (A) SANS data obtained as a function of concentration, bottom to top. Solid lines are fits to the data using the fuzzy sphere model.¹⁰ The curves have been offset along the ordinate for clarity. (B) SAXS data obtained using the same samples as in A. The solid lines are fits to the data based on input information from A. The curves are offset along the ordinate for clarity.

In a previous work,³¹ we derived an alternative form factor for PNIPAM microgels, which focuses on the heterogeneous density profile of the crosslinked core,

$$P(q) = \left[\int_0^{R_{\text{core}}} \frac{24r^2}{5R_{\text{core}}^3} \left(1 - \frac{r^3}{R_{\text{core}}^3}\right)^{3/5} \frac{\sin(qr)}{qr} dr \right]^2. \quad (2)$$

Here, the size of the crosslinked core R_{core} is the only fit parameter. Dangling polymer chains are assumed to be too low in weight to contribute to the measured scattering intensity $I(q)$. Differences between R_{core} and the hydrodynamic radius of approximately 30% can be attributed to dangling ends which, despite their low density, are known to affect hydrodynamic properties. This will also be discussed later in the text.

For consistency with previous experimental work, we initially based the fitting of the data on the fuzzy-sphere model. We require an additional Lorentzian function to account for polymer network fluctuations,

$$I_{\text{fluct}} = \frac{I_{\text{fluct}}(0)}{1 + \xi^2 q^2}, \quad (3)$$

in which ξ is the length scale over which the fluctuations are correlated. Furthermore, a Gaussian size distribution was included together with the instrumental smearing function, as described elsewhere.¹⁰ The resulting fits are shown as the solid curves in Fig. 2A. For all concentrations, the polydispersity was kept constant at 15% and originates primarily from the difference in R_h between Hm and Dm particles, whereas the two sets of particles have an individual polydispersity of around 5%. Increasing the polydispersity as the concentration increases did not give rise to a better fit of the data. The latter indicates that the particles are not (measurably) anisotropically deformed in the investigated concentration range. For the highest concentrations a slight upturn of the intensity can be observed at low q -values which is most probably due to the presence of a small amount of aggregates. The latter is hard to avoid at these high concentrations.



The cancelled structure peaks in the ZAC approach reappear again in the SAXS analysis shown in Fig. 2B. The size information from SANS were used as input parameters for the fitting procedures performed on the SAXS data. Here, we follow the same fitting procedure for the SAXS data as described by Westermeier *et al.*,³² for all concentrations up to 4 wt%, see solid lines in Fig. 2B. The q -dependent intensity, $I(q)$, of the SAXS data can be described by $I(q) \propto P(q)S(q)$, where $P(q)$ and $S(q)$ are the form and structure factor, respectively. The $S(q)$ was calculated using the modified penetrating background-corrected rescaled mean sphere approximation (MPB-RMSA) scheme used in previous publications.^{27,33} The input for the MPB-RMSA calculation is the number density, n_p , the particle charge, Z_{eff} , salinity, c_{salt} and the total size of the particles, R . Since the primary peak position, q_{max} , of highly repulsive particles scales as $q_{\text{max}} \propto a_s^{-1} \propto n_p^{-1/3}$, see the curve in Fig. 3, we can, from the determination of q_{max} of the SAXS data at different concentrations determine n_p . From this we deduce a relation between n_p and wt% to be $n_p = 5.5 \times 10^{-8}$ wt% for both the Hm and Dm particles, which allows us to determine n_p for every experimental weight concentration.³⁴ It also enables an estimation of the molecular weight of the microgels, which is 1.14×10^8 g mol⁻¹, similar to reported molecular weights in work by Senff and Richtering.³⁵ For comparison with the latter reference, the molar fraction of crosslinkers in the current work can be estimated at 5% with the values given in Table 1. By means of titration, we have determined the number of acid groups per particle to be $N_{\text{acid}} = 1.1 \times 10^5$ for both species, which corresponds well with estimates based on the molar fraction of acrylic acid during synthesis, which yields $N_{\text{acid}} \approx 8 \times 10^4$.

The polydispersity for the SAXS data fits is reduced to 8% which is close to the value one gets from approximating the two particle size distributions with a single Gaussian distribution. The only other fitting parameter is Z_{eff} , which is an effective charge that accounts for charge renormalization,¹⁹ giving a

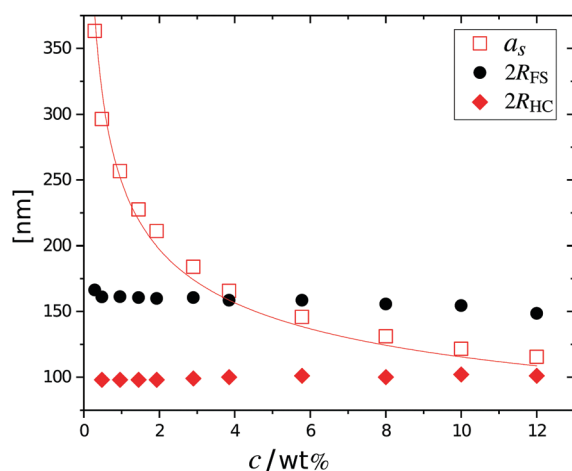


Fig. 3 Total particle size as a function of the concentration. The fuzzy-sphere diameter $2R_{\text{FS}}$ (circles), the associated hard-core diameter $2R_{\text{HC}}$ (diamonds), extracted from SANS, as well as the center-to-center distance, a_s , (open squares) as obtained from SAXS. The curve shows the best fit to that data using $a_s \sim c^{-1/3}$.

value of around $90e$, where e refers to the elementary charge. Note, however, that for concentrations above 4 wt% the MPB-RMSA model can no longer reproduce the experimental $S(q)$. This failure is most likely due to its use of a ‘hard’ core potential instead of a softer core interaction, which becomes particularly important at significant volume fractions. A more in-depth investigation of the influence of the core contribution is not covered by this paper.

The results from the form factor fits of the ZAC measurements are summarized in Fig. 3, diamonds, together with the center-to-center distances extracted from SAXS measurements.

While the fitted center-to-center distance a_s is changing with the expected $\propto n_p^{-1/3}$ relation, the SANS and SAXS-data analysis indicates no change in size except for the largest concentrations. Specifically, the size of the particles seems not to be affected by the concentration until the center-to-center distance approaches the fitted fuzzy-sphere hard-core diameter $2R_{\text{HC}}$ (red squares). At these larger concentrations, the fitted fuzzy-sphere particle size $2R_{\text{FS}}$ is substantially smaller than the measured hydrodynamic radius close to infinite dilution (see Fig. 1). To bridge this gap with results at lower concentrations we therefore need additional analysis.

Since the flux of neutrons is insufficient to yield data with adequate statistics at low concentrations we turned to light-scattering methods instead. Here the static and dynamic structure factor peaks were fitted simultaneously in order to decouple the size and shape of the particles and their spatial arrangement. Specifically, we performed static (SLS) and dynamic (DLS) light scattering on concentrations below 0.6 wt%, where the primary peak of the structure factor, $S_m(q)$, appears in the q -range of the light scattering measurements. The intensity auto-correlation function, $g^2(q,t) - 1$, was measured throughout the whole light scattering q -range. To investigate the swelling behaviour at low particle concentrations we extract from these measurements both the diffusive function, $D(q)$, and the scattering intensity, $I(q)$, from the light scattering measurements at concentrations between 0.5 and 0.005 wt%. The $I(q)$ and $D(q)$ for 0.3, 0.06 and 0.02 wt% are shown in Fig. 4 where the peak position of $S_m(q)$, q_{max} , and the minima in $D(q)$, $D(q_{\text{max}})$, can be clearly seen. This shows that there are still strong interactions at these low concentrations.

Even though the shape of both $I(q)$ and $D(q)$ looks as expected with respect to earlier SAXS measurements and previous investigations of microgels at low densities,²⁷ the low- q part, $q < q_{\text{max}}$, indicates a substantial increase in $I(q)$ as well as slower dynamics, as can be observed from $D(q)$. We will use the $\delta\gamma$ -approximation as used in ref. 36 and 37 to fit SLS data, $I(q)$, and the DLS data, $D(q)$, simultaneously by calculating $D(q)$ through the hydrodynamic function, $H(q)$, in the same way as in ref. 27. This approach, however, cannot capture our data at low q values while using the polydispersity determined from SAXS and SANS. As can be seen in Fig. 4, full black lines, the fitting deviates at low q values. This deviation is more pronounced with increasing concentrations, A to C to E in Fig. 4, and less pronounced for the dynamic measurements, see B, D and F in Fig. 4. The low- q deviation of the experimental data



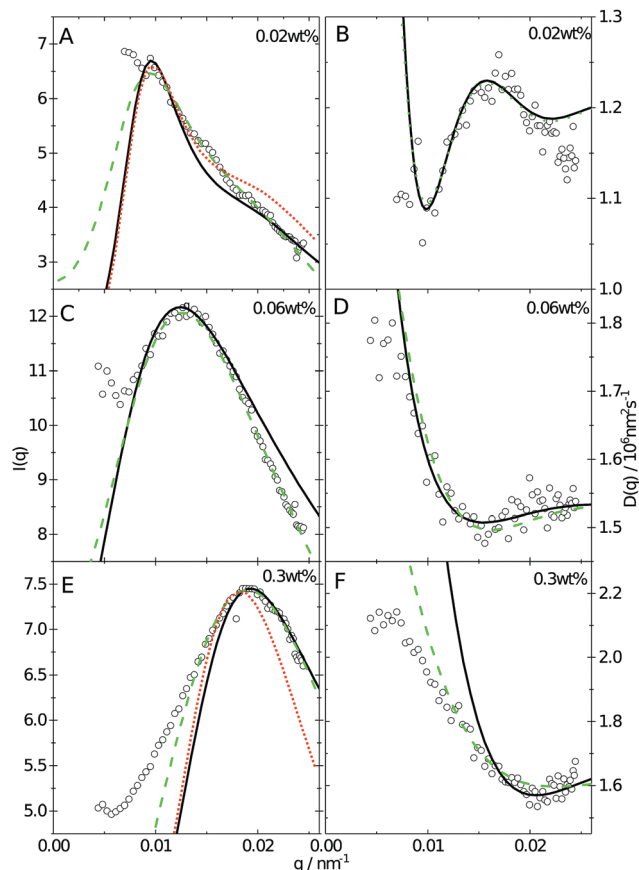


Fig. 4 $I(q)$ as a function of q -vector at concentrations (A) 0.02 wt%, (C) 0.06 wt% and (E) 0.3 wt%. The diffusive function, $D(q)$, constructed from extracted diffusion coefficients as a function of q -vector at a concentration of (B) 0.02 wt%, (D) 0.06 wt% and (F) 0.3 wt%. The full curves and dashed curves show the result of different fitting methods to the data, as described in the main text. The dotted curves in A and E show fits to the SLS data only using a fixed core size $R_{\text{core}} = 70$ nm.

in both the static as well as the dynamic experiments can be largely captured through the addition of a phenomenological Ornstein–Zernike type contribution to the structure factor. Further investigation of this phenomenon could be of interest but is not the scope of this paper. This approach is, therefore, included in the ESI† (Fig. S.3). By ignoring the low- q -range and fitting the $I(q)$ and $D(q)$ data simultaneously for $q \geq q_{\text{max}}$ we recover the swelling trend as found in previous studies of charged microgels in low salinity.²⁷ This suggests that, as the concentration is lowered below 0.5 wt%, the particles are getting increasingly larger.

With the additional light-scattering data the extracted fuzzy-sphere sizes, R_{FS} , throughout the full concentration regime can be plotted. This is shown in Fig. 5, where the inset shows the fitted parameters to the data. At low concentrations a strong decrease of particle size with increasing concentration is found. Yet, the size levels out at concentrations where light scattering and neutron data coincide and remains constant in the SANS regime. At the same time, the polydispersity does not increase with concentration, but rather slightly decreases. This is an indication that no anisotropic particle deformation takes place

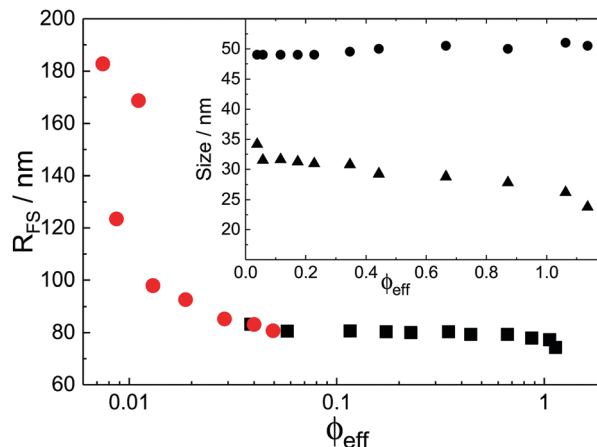


Fig. 5 Total particle size $R_{\text{FS}} = R_{\text{HC}} + 2\sigma$ as a function of volume fraction, ϕ_{eff} , determined from R_{FS} , as extracted from light scattering (circles) and SANS (squares). The inset shows the corresponding fuzzy-sphere parameters R_{HC} (circles) and 2σ (triangles) in the SANS regime.

in the investigated concentration regime. The pH, which is a measure of the osmotic pressure generated by the counterions outside the particles, is found to decrease monotonically with increasing concentration, in line with the decrease of the ion-osmotic pressure difference between the particle interior and its exterior (included as Fig. S.2 in the ESI†). In this analysis, the radii from the light scattering are normalised by dividing by a factor of 1.21, and reflects the frequently reported discrepancy in measured size of fuzzy particles using either dynamic or static methods.¹⁰ Note that a difference in polydispersity-induced discrepancies between weighted averages, as is the case for SANS data, and z -averages, which applies to the DLS data, seems to be insufficient here. We investigated the electrostatic component of this discrepancy in more detail by adding salt to the system. The static size obtained from SANS and static light scattering showed no significant change in particle size whereas the dynamic size significantly decreased, and even coincided with the static size at the highest salt concentration. We refer to Fig. S.4 of the ESI† for these findings.

Theory and modeling

While a neutral microgel particle swells as a result of steric repulsions within the polymer backbone, the introduction of charged groups into the network can induce even larger swelling ratios. This additional swelling may be understood from the osmotic pressure, which should be the same inside and outside the network. Inside the microgel, this pressure has an ionic component resulting from the counterions that accompany the charged groups,³⁸ Π_{ion} , as well as a contribution from the net osmotic pressure generated by the hydrogel backbone itself, Π_{gel} . Since the ion concentration outside the microgel is much smaller than inside, the increased ion-osmotic pressure inside should thus be accompanied by a negative contribution from the hydrogel backbone, which is accomplished by additional swelling of the microgel. This effect is more pronounced



at low concentrations of microgels, as well as low salinities, as this renders the counterion concentration outside the gel small, and the gradient in ion-osmotic pressure, therefore, large. Generally, swelling a hydrogel (far) beyond its uncharged volume requires the ion-osmotic pressure to be of the same magnitude as the hydrogel bulk modulus K , which has been measured to be approximately 70 kPa for NIPAM hydrogels.^{39–41} The acidic groups, however, do not deprotonate much further once the pH reaches below the pK, which is ≈ 4.2 for acrylic acid. This yields a counterion pressure of approximately 4 kPa inside the particles, for which we use the estimated number of 1.1×10^5 of acid groups per particle. Under normal circumstances the counterions would not be able to swell the hydrogel core beyond its 'uncharged' volume. Although that would explain the appearance of a plateau in particle size at intermediate densities, it would not explain the continued swelling at very low densities.

It is tempting to assume that the re-entrant swelling at low concentrations could be a result of the heterogeneous crosslinking of the hydrogel core, as this produces a network that is more loosely crosslinked on the outside. In previous work the associated core-density profile $\phi(r)$ for neutral particles was derived from first principles,

$$\phi(r) = \phi(0) \left(1 - \frac{r^3}{R_{\text{core}}^3} \right)^{3/5} \quad (4)$$

Here, we modify this approach by including an additional swelling pressure due to the partial charging of the acrylic acid groups. Our approach involves solving the Poisson–Boltzmann equation for the electrostatic potential $\Psi(r)$ within a spherical cell model,^{42,43} in which the cell radius R_{cell} is set by the available volume per particle in the system, *i.e.* $4/3\pi R_{\text{cell}}^3 = n_p^{-1}$. Charge neutrality dictates that the electric field $E(r) = -\psi'(r)$ must be screened at the boundary of this cell, *i.e.* $E(R_{\text{cell}}) = 0$. The charged species in the cell are the deprotonated acrylic acid with local concentration $\rho_A(r)$, as well as the counterions (protons) with local concentration $\rho_{H^+}(r)$. Note that we here assume that the acrylic acid is only present inside the particle core, *i.e.* $r < R_{\text{core}}$, where its deprotonation can be calculated *via* $[A^-][H^+]/[AH] = 10^{-\text{pK}}$. Here, $[AH]$ is the concentration of protonated (uncharged) acrylic acid and brackets indicate molar concentrations as usual. For later reference, any charges in dangling polymer ends outside the core are estimated to contribute only insignificantly to the overall electric field, and are therefore ignored here. The local concentration of acidic groups, $\rho_A(r) + \rho_{\text{AH}}(r)$, is assumed to scale linearly with the local monomer density $\rho_{\text{gel}}(r)$. Therefore, $\rho_{\text{AH}}(r) + \rho_A(r) = \rho_{\text{gel}}(r) \cdot N_{\text{acid}}/N_{\text{monomers}}$, where $N_{\text{acid}}/N_{\text{monomers}}$ is the number fraction of acrylic acid groups. The Poisson part of the PB equation is $\nabla^2 e\psi(r)/kT = -4\pi\lambda(\rho_{H^+}(r) - \rho_A(r))$, with $\lambda = 0.7$ nm the Bjerrum length and e the elementary charge, while the counterion density is $\rho_{H^+}(r) = \rho_s \exp(-e\psi(r)/kT)$. Here, ρ_s is a reservoir concentration of counterions that can be chosen arbitrarily small for deionized systems. The deprotonation equilibrium is applied to relate $\rho_A(r)$ to $\rho_{H^+}(r)$ and, therefore, $\psi(r)$. Solving the Poisson–Boltzmann equations proceeds by choosing a trial value for $\psi_0 \equiv \psi(0)$

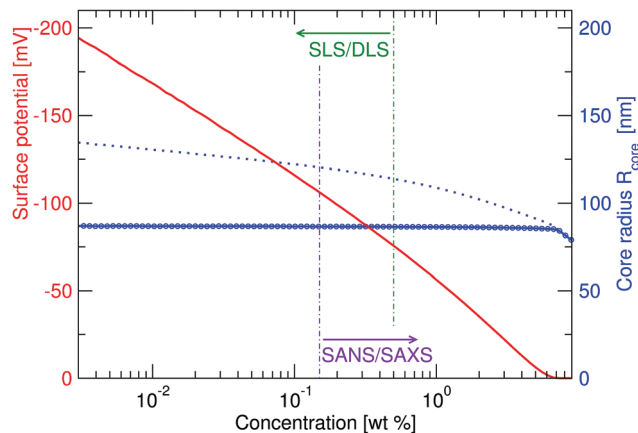


Fig. 6 Calculated charge-induced swelling profile (blue) for a heterogeneously crosslinked core that has a swollen radius $R_{\text{core}} = 70$ nm in the absence of charge. While the full blue curve shows results for a bulk modulus $K = 7 \times 10^4$ Pa, the dotted line also shows results for an unrealistically low bulk modulus $K = 7 \times 10^3$ Pa. The full curve (red) corresponds to the electrostatic surface potential of the former (w.r.t. the Donnan potential) and demonstrates a strong dependency of this variable on the particle concentration.

(noting that $\psi'(0) = 0$ due to symmetry), and numerically calculating the corresponding $\psi'(R)$. This process repeats until the ψ_0 is found that is in agreement with the boundary condition $\psi'(R_{\text{cell}}) = 0$. We have included the resulting electric field profiles at various concentrations in the ESI.† The resulting ion pressure, as well as the electric field,⁴⁴ constitutes an additional local contribution to the osmotic pressure compared to the swelling of a neutral microgel network. We refer to earlier work³¹ for the numerical method that can be used for calculating the resulting gel-density profiles $\rho_{\text{gel}}(r)$. Since this gel density couples back to the PB calculations through the acrylic-acid profile, we solve for $\rho_{\text{gel}}(r)$ and $\psi(r)$ simultaneously. The full (red) curve in Fig. 6 gives an estimate of the resulting (not renormalized) zeta potential of the microgels by plotting the difference between the electrostatic potential at the microgel surface $\Psi(R_{\text{core}})$ and the Donnan potential (the cell-surface potential). The latter shows a strong increase upon lowering the microgel concentration. Also the associated counterion profiles demonstrate a strong dependency on concentration. Despite this, the heterogeneous crosslinking triggers no significant swelling of the network, apart from an increase of 6% in radius which is already observed for neutral particles. The dotted curve in Fig. 6, therefore, recovers the plateau in size, which extends to infinite dilution. For the reader's interest we also include results for an artificially low bulk modulus of 7 kPa, as shown by the dotted curve. As expected, softer microgels will swell more upon lowering the particle concentration. Although the resulting swelling profile agrees qualitatively with theoretical work on fully deprotonated microgels by Denton *et al.*,⁴⁵ this curve also fails to reproduce the observed plateau at intermediate particle concentrations as observed in Fig. 5.

While the theoretical model above considers the swelling of the crosslinked core of the particle, the presence of dangling polymer ends has been ignored so far. In previous work we



estimated the number of dangling ends in the corona of PNIPAM microgels to be of the order of thousands per particle.³¹ Although these chains constitute a minority of the total polymer mass, we have found indications that their ‘dangling’ nature may have a strong effect on the diffusive behaviour of the microgels. To separate this dynamics property from the ‘static’ information (*i.e.* form factor) we turn back to the light scattering data and re-fit the static data (SLS) only using the form factor from the heterogeneously-crosslinked core model described in eqn (2). The shape of this form factor is fully determined by the core size only which facilitates fitting in the limited q -range that was available. The SLS data are now refitted using the PBRMSA scheme. The experimentally determined number density is used when only fitting the SLS and the size of the particles in the PBRMSA scheme is kept the same as the radius in eqn (2). The only input from the $D(q)$ is to superimpose the minima. The latter approach improves on the earlier fit with the fuzzy-sphere model, as is demonstrated by the green and the black curves in Fig. 4A, C and E, corresponding to the heterogeneous-core fit and the fuzzy-sphere fit respectively. The open circles in the inset of Fig. 7 show this refitted core radius throughout the light-scattering regime. For convenience, we use the effective volume fraction ϕ_{eff}^* here, which results from a ‘plateau’ radius of 80 nm (as shown by the solid line). To compare these sizes with the SANS results we re-fit the resulting core-density profiles with a fuzzy-sphere model, as indicated by the open squares in the main plot. This shows an extension of the plateau towards much lower concentrations and dramatically reduces the increase in size at the lowest concentrations. Note that a small increase in core size remains to be the best fit to the experimental data at these very low concentrations and one cannot rule out a minor core swelling beyond what is predicted by the network model. Specifically, the dotted curve in Fig. 4A and E shows the effect of forcing a

constant core size of $R_{\text{core}} = 70$ nm, while the dashed curves demonstrate the optimal fit within the q -range. Investigations on other charged microgels in this concentration regime may elucidate this further.

With the model and the new interpretation of the data suggesting that the crosslinked core largely resists additional swelling induced by the acrylic acid, we shift our focus to the behaviour of the dangling polymer chains. Due to the sharp decrease in crosslinker density towards ‘completion’ of the particle during synthesis, it may be estimated³¹ that there are a few tens of dangling chains with a very long contour length (>150 nm), while there may be hundreds of shorter chains (*e.g.* with a contour length >30 nm). To investigate whether the electric field generated by the core is strong enough to induce significant stretching of these dangling ends we perform Monte-Carlo simulations. Here, the dangling polymers are modeled as simple bead chains grafted on a single solid gel core. The bead-bead interactions were included through a standard hard-core interaction of monomers with a diameter of 0.26 nm, as our results did not seem to rely strongly on the choice of a more physical bead-bead potential such as a Lennard-Jones. The chain stiffness, *i.e.* the Kuhn length of PNIPAM,⁴⁶ is included through a rotational-dependent bond energy given by $k_{\theta}(1 - \cos \theta_i)$ as in ref. 47, in which we set $k_{\theta} = 7.5kT$ as the rigidity parameter. Here, $\cos \theta_i = \hat{\mathbf{a}}_i \cdot \hat{\mathbf{a}}_{i-1}$, where $\hat{\mathbf{a}}_i$ is the unit vector in the direction of bond i . Note that the acrylic acid is also incorporated in the dangling polymer ends, which induces a repulsive interaction with the core. Following the standard Langmuir adsorption model⁴⁸ one finds the cation-density dependent free energy of an acidic group $f(r)/kT = \log(1 + K/\rho_+(r))$, where $\rho_+(r)$ is the local counterion concentration that we obtain from cell model calculations. Note that $f(r)$ reduces to the electrostatic energy of charged (uncharged) groups for high (low) values of K , when the Boltzmann relation $\rho_+(r) \sim \exp(-e\Psi(r)/kT)$ is used. Note that the beads in simulations may extend beyond the maximum radius in the cell calculations of $f(r)$. We therefore set $f(r) = f(R_{\text{cell}})$ for $r > R_{\text{cell}}$. In simulations, the radius-dependent free energy of the beads was chosen proportional to $f(r)$ to take account for the fact that the acidic groups constitute 9% of the monomers in the chain here. Cation profiles from the cell-model calculations were used to calculate the $f(r)$ locally. In this way, the different microgel concentrations can be simulated independently using their associated profiles of $f(r)$ that result from cell-model calculations at each concentration.

In Fig. 8, we show resulting dangling-end density profiles from simulations. The results of this approach indicate that the polymer density profile close to the core remains unaffected by variations of the microgel concentration. At the same time, the low-density tail of the profile demonstrates a strong response on changing their concentration. Interestingly, at intermediate and low microgel concentrations the longer chains seem to favour a porcupine-like configuration, indicated by a nonzero polymer density profile that extends over much longer distances than in the neutral case. On the other hand, for the

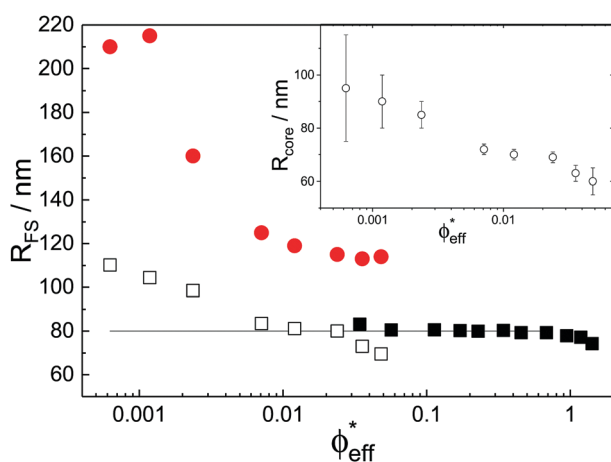


Fig. 7 Total particle radius R_{FS} as a function of effective volume fraction ϕ_{eff}^* . The open squares represent effective fuzzy-sphere radii as described in the text, which are calculated from the fitted core size R_{core} to the SLS data only. These data points are plotted in the inset. The solid squares represent fits to SANS data as before. The extracted hydrodynamic radius from DLS is represented by the solid circles.



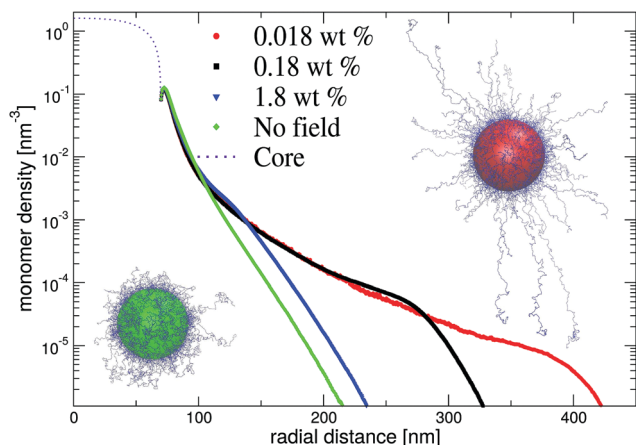


Fig. 8 Monte-Carlo results for the dangling-end density profiles at various densities. We use counterion profiles from cell-model calculations, which serve as an external field. The snapshots of the simulations show configurations at high salt (no field/green) as well as at low particle concentrations $c = 0.018$ wt% (red).

larger weight fraction $c \approx 1$ wt% the profile displays the same characteristics as without electric fields. This observed behaviour is confirmed by snapshots of the simulations, which clearly shows the porcupine-like configuration at lower weight fractions. We conclude that the latter state, which minimizes the electrostatic-energy configuration of the dangling ends, can be reached in experiments despite the associated entropic penalty of chain stretching. The strong deionization may be pivotal for the occurrence of the porcupine state as this permits the electric field to extend relatively far from the core before being screened.

Configurational changes in the dangling ends will unlikely be visible in static (light) scattering data due to their minor contribution to the total scattered intensity of the microgels. The dangling-end density profiles may, however, set the diffusive behaviour of the particles. We thus turn back to the extracted diffusion coefficients as shown in Fig. 4B, D, and F. Contrary to our earlier approach, this time we enable the dynamic radius to vary from the SLS radius. Specifically, the parameter d_0 is fitted independently of other fit parameters in order to superimpose the calculated $D(q)$. Releasing the restriction of matching the static and the hydrodynamic size, we now also choose to not correct the latter by scaling with a factor of 1.21 as before. Following this approach, a very large discrepancy between the hydrodynamic size and the static size emerges, as the filled circles in Fig. 7 indicate. We conclude that the charge-induced swelling at low density predominantly expresses itself as a hydrodynamic effect, which likely results from a configurational change in the dangling ends induced by an increased electrostatic repulsion from the core. Note that this hydrodynamic size does not correspond to a microscopic property of the particle that can be measured directly in a static scattering or microscopy experiment, yet it enables interpreting the hydrodynamic friction coefficient as an effective sphere size. However, we currently lack a full quantitative model that would allow us to calculate the additional contribution to the

overall friction coefficient of the microgels that results from the extended configuration of the dangling polymer chains, and thus a quantitative calculation of the hydrodynamic radius remains rather difficult for now. Although Fig. 8 shows that significant stretching of the longest chains sets in at intermediate volume fractions, the exact relation between hydrodynamic size and concentration will require a more detailed approach towards the hydrodynamic effect of the dangling ends. From the experimental side it would be interesting to probe the properties of the dangling ends *in situ* to elucidate their effect on hydrodynamic properties of the microgel in both its neutral and charged state.

Conclusion

The deswelling behaviour of soft microgel particles is an extensively discussed topic in the soft-matter community. The intrinsic particle softness and the responsive nature of the polymer network towards external stimuli rises questions about whether the particles interpenetrate or compress upon increasing the number density. Here we report our results on the deswelling behaviour of ionic PNIPAM particles spanning a large concentration regime. For this purpose we use a combination of the analysis of static and dynamic structure factors obtained with static and dynamic light scattering and small-angle X-ray scattering, and the zero average contrast approach in small-angle neutron scattering. The ZAC experiments enable us to cancel the structure factor contribution to the total scattering intensity, giving direct access to the form factors. The structure factors are later recovered by means of small-angle X-ray scattering. We demonstrate that a significantly increased apparent size of the particles at very low concentrations can be attributed to the presence of charge in the polymer backbone. At intermediate concentrations the size remains, however, constant up to the point where particles are significantly overlapping. Then, at concentrations above shell overlap the particle size is again found to decrease without any sign of particle deformation. The plateau in the particle size as observed with SANS at intermediate concentrations could not be explained by modelling the swelling of the heterogeneously crosslinked hydrogel core. Instead, we have found that the dangling ends may be pivotal for the observed increase in size. Specifically, the charge-induced configurational changes of those polymers in favour of a 'porcupine' state may increase the hydrodynamic size while leaving the static scattering profiles largely unaffected. The latter is also consistent with discrepancies between SANS and DLS measurements, indicating a large discrepancy between the static size and the hydrodynamic size of ionic microgels. There remain, however, a few open questions. We currently lack a model to explain the additional low- q scattering contribution that we observe at very low concentrations. While some of the observed features still need further work to explain them quantitatively, a comparison between the characteristic inter-particle distance at high effective volume fractions and the measured particle radii clearly



indicate that significant particle interpenetration must occur to explain these data. This raises questions about previous interpretations of rheological data, and further studies are needed on systems comparable to the one studied here.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 P. N. Pusey and W. V. Megen, *Nature*, 1986, **320**, 340.
- 2 E. B. Sirota, H. D. Ou-Yang, S. K. Sinha, P. M. Chaikin, J. D. Axe and Y. Fujii, *Phys. Rev. Lett.*, 1989, **62**, 1524.
- 3 Y. Monovoukas and A. P. Gast, *J. Colloid Interface Sci.*, 1989, **128**, 533.
- 4 A. van Blaaderen and P. Wiltzius, *Science*, 1995, **270**, 1177.
- 5 E. R. Weeks, J. C. Crocker, A. C. Levitt, A. Schofield and D. A. Weitz, *Science*, 2000, **287**, 627.
- 6 E. Zaccarelli, C. Valeriani, E. Sanz, W. C. K. Poon, M. E. Cates and P. N. Pusey, *Phys. Rev. Lett.*, 2009, **103**, 135704.
- 7 U. Dassanayake, S. Fraden and A. van Blaaderen, *J. Chem. Phys.*, 2000, **112**, 3851.
- 8 A. Yethiraj and A. van Blaaderen, *Nature*, 2003, **421**, 513.
- 9 A.-P. Hynninen and M. Dijkstra, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2005, **72**, 051402.
- 10 M. Stieger, W. Richtering, J. S. Pedersen and P. Lindner, *J. Chem. Phys.*, 2004, **120**, 6197.
- 11 J. C. Pámies, A. Cacciuto and D. Frenkel, *J. Chem. Phys.*, 2009, **131**, 44514.
- 12 D. M. Heyes and A. C. Branka, *Soft Matter*, 2009, **5**, 2681.
- 13 L. Berthier, A. J. Moreno and G. Szamel, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2010, **82**, 060501.
- 14 H. Senff and W. Richtering, *J. Chem. Phys.*, 1999, **111**, 1705.
- 15 D. Paloli, P. S. Mohanty, J. J. Crassous, E. Zaccarelli and P. Schurtenberger, *Soft Matter*, 2012, **9**, 3000.
- 16 D. Gottwald, C. N. Likos, G. Kahl and H. Löwen, *Phys. Rev. Lett.*, 2004, **92**, 068301.
- 17 P. S. Mohanty and W. Richtering, *J. Phys. Chem. B*, 2008, **112**, 14692.
- 18 J. K. Cho, Z. Meng, L. A. Lyon and V. Breedveld, *Soft Matter*, 2009, **5**, 3599.
- 19 A. R. Denton, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2003, **67**, 011804.
- 20 T. Eckert and W. Richtering, *J. Chem. Phys.*, 2008, **129**, 124902.
- 21 J. Crassous, M. Siebenbürger, M. Ballauff, M. Drechsler, O. Henrich and M. Fuchs, *J. Chem. Phys.*, 2006, **125**, 204906.
- 22 Z. Meng, J. K. Cho, S. Debord, V. Breedveld and L. A. Lyon, *J. Phys. Chem. B*, 2007, **111**, 6992–6997.
- 23 J. Riest, P. Mohanty, P. Schurtenberger and C. N. Likos, *Z. Phys. Chem.*, 2012, **226**, 711.
- 24 A. Moncho-Jorda, *J. Chem. Phys.*, 2013, **139**, 064906.
- 25 D. Caprile-Gonzalez, B. Sierra-Martin, A. Fernandez-Nieves and A. Fernandez-Barbero, *J. Phys. Chem. B*, 2008, **112**, 12195.
- 26 K. Kratz, T. Hellweg and W. Eimer, *Colloids Surf., A*, 2000, **170**, 137.
- 27 P. Holmqvist, P. S. Mohanty, G. Nägele, P. Schurtenberger and M. Heinen, *Phys. Rev. Lett.*, 2012, **109**, 048302.
- 28 P. S. Mohanty, S. Nöjd, K. van Gruijthuijsen, J. J. Crassous, M. Obiols-Rabasa, R. Schweins, A. Stradner and P. Schurtenberger, *Sci. Rep.*, 2017, **7**, 1487.
- 29 G. Romeo, L. Imperiali, J. W. Kim and A. Fernandez-Nieves, *J. Chem. Phys.*, 2012, **136**, 124905.
- 30 L. Willner, O. Jucknischke, D. Richter, J. Roovers, L.-L. Zhou, P. M. Toporowski, L. J. Fetters, J. S. Huang, M. Y. Lin and N. Hadjichristidis, *Macromolecules*, 1994, **27**, 3821.
- 31 N. Boon and P. Schurtenberger, *Phys. Chem. Chem. Phys.*, 2017, **19**, 23740.
- 32 F. Westermeier, B. Fischer, W. Roseker, G. Grübel, G. Nägele and M. Heinen, *J. Chem. Phys.*, 2012, **137**, 114504.
- 33 M. Heinen, P. Holmqvist, A. J. Banchio and G. Nägele, *J. Chem. Phys.*, 2011, **134**, 044532.
- 34 J. Liu, H.-J. Schöpe and T. Palberg, *Part. Part. Syst. Charact.*, 2000, **17**, 206.
- 35 H. Senff and W. Richtering, *Colloid Polym. Sci.*, 2000, **278**, 830–840.
- 36 C. W. J. Beenakker and P. Mazur, *Physica A*, 1983, **120**, 388.
- 37 C. W. J. Beenakker, *Physica A*, 1984, **128**, 48.
- 38 M. M. Hedrick, J. K. Chung and A. R. Denton, *J. Chem. Phys.*, 2015, **142**, 034904.
- 39 B. R. Saunders and B. Vincent, *J. Chem. Soc., Faraday Trans.*, 1996, **92**, 3385.
- 40 P. A. L. Fernandes, S. Schmidt, M. Zeiser and T. Hellweg, *Soft Matter*, 2010, **6**, 3457.



- 41 S. Schmidt, M. Zeiser, T. Hellweg, C. Duschl, A. Fery and H. Möhwald, *Adv. Funct. Mater.*, 2010, **20**, 3235.
- 42 T. L. Hill, *Statistical Mechanics*, Addison-Wesley, 1960.
- 43 H. Wennerström, B. Jönsson and P. Linse, *J. Chem. Phys.*, 1982, **76**, 4665.
- 44 P. Das and S. Bhattacharjee, *Langmuir*, 2005, **21**, 4755–4764.
- 45 A. R. Denton and Q. Tang, *J. Chem. Phys.*, 2016, **145**, 164901.
- 46 K. Kubota, *Polym. J.*, 1990, **22**, 1051.
- 47 R. Auhl, R. Everaers, G. Grest, K. Kremer and S. Plimpton, *J. Chem. Phys.*, 2003, **119**, 12718–12728.
- 48 I. Langmuir, *J. Am. Chem. Soc.*, 1918, **40**, 1361–1403.

