



Cite this: *Soft Matter*, 2018, 14, 3998

Probing nano-scale viscoelastic response in air and in liquid with dynamic atomic force microscopy

Federica Crippa,^a Per-Anders Thorén,^b Daniel Forchheimer,^b Riccardo Borgani,^b Barbara Rothen-Rutishauser,^a Alke Petri-Fink^{ac} and David B. Haviland^{*b}

We perform a comparative study of dynamic force measurements using an Atomic Force Microscope (AFM) on the same soft polymer blend samples in both air and liquid environments. Our quantitative analysis starts with calibration of the same cantilever in both environments. Intermodulation AFM (ImAFM) is used to measure dynamic force quadratures on the same sample. We validate the accuracy of the reconstructed dynamic force quadratures by numerical simulation of a realistic model of the cantilever in liquid. In spite of the very low quality factor of this resonance, we find excellent agreement between experiment and simulation. A recently developed moving surface model explains the measured force quadrature curves on the soft polymer, in both air and liquid.

Received 19th January 2018,
Accepted 17th April 2018

DOI: 10.1039/c8sm00149a

rsc.li/soft-matter-journal

1. Introduction

Applications in biology such as stiffness measurements on cancer cells or characterization of extremely soft gels^{1–5} require an understanding of interfacial properties in an aqueous environment. Not only the surface topography but also the viscoelasticity of biomimetic scaffolds affects the interaction between cells and their environment, in phenomena such as cellular adhesion, differentiation and migration.^{6–8} An atomic force microscope (AFM)⁹ is well suited to study such viscoelastic responses, and various methods have been used to investigate single cells^{10,11} and intracellular compartments such as cytoskeleton and nucleus^{12,13} and to compare physiological and pathological conditions.^{1,14}

The AFM is a versatile tool for the micro- and nano-scale characterization of surfaces, with applications from mapping of topography¹⁵ to the quantitative measurements of contact, electrostatic or magnetic forces between the tip and the surface.^{16–19} The AFM is frequently performed in air and in the vacuum where the cantilever's mechanical resonance has a high quality factor, but there are many applications of quantitative AFM in liquid environments where the quality factor is relatively low.^{20–23} A liquid environment is challenging for quantitative AFM due to the large inertial and viscous forces acting on the cantilever

body, which complicate calibration and call into question the standard model of transduction between the tip–surface force and beam deflection – the starting points for quantitative surface analysis with the AFM.

Elastic response, or the tip–surface force resulting from static deformation of a cell or a substrate, is typically inferred from the quasi-static force–distance curve. Such curves are measured under the assumption that the tip–surface force is linearly dependent on cantilever deflection (Hooke's law) when the probe is moved toward and away from the surface at sufficiently low (and constant) velocity.²⁴ Viscous response, or the tip–surface force that depends on velocity, requires a dynamic method of measurement.^{25–27} With dynamic methods, care must be taken to understand and calibrate the frequency-dependent relationship between the tip–surface and cantilever deflection, which changes radically from an air to liquid environment.

The dynamic method used in this study is called Intermodulation AFM (ImAFM).²⁵ A special multi-frequency lockin amplifier captures many Fourier coefficients of the motion (*i.e.* both amplitude and phase at many frequencies) which are the result of frequency mixing (intermodulation) of two or more discrete frequency components in the drive waveform. Our work reports the first experimental results and the analysis of ImAFM in liquid. In spite of the low quality factor and significant hydrodynamic load of the liquid, we find that ImAFM can reconstruct both the viscous and elastic forces due to the interaction between the tip and the surface. We show that force reconstruction using the so-called dynamic force quadrature curves²⁸ works remarkably well in liquid. We verify

^a Adolphe Merkle Institute, University of Fribourg, BioNanomaterials Group, Chemin des Verdiers 4, CH-1700 Fribourg, Switzerland. E-mail: federica.crippa@unifr.ch

^b KTH Royal Institute of Technology, Nanostructure Physics, Albanova, SE-10791 Stockholm, Sweden. E-mail: pathoren@kth.se

^c University of Fribourg, Department of Chemistry, Chemin du Musée 8, CH-1700 Fribourg, Switzerland



its accuracy by reconstructing the known force quadrature curves from simulated data using a realistic model for the cantilever oscillating in liquid. We further demonstrate the reconstruction of the unknown force quadrature curves from experimental data with a well-known test sample in liquid. Our study serves as a validation of the ImAFM method in liquid, setting the stage for future investigations of liquid-to-soft-solid interfaces more relevant to polymeric or biological applications.

II. Experiment

ImAFM has been thoroughly explained in our previous publications.^{28–31} Fig. 1 and its caption describe the basic idea behind the technique. ImAFM can be performed with almost any AFM, using a hardware and software kit commercially available from Intermodulation Products AB.^{32,43} The AFM used here is the NanoWizard[®]3 from JPK instruments³³ which is well suited to work in liquid. The cantilever is acoustically actuated with a small piezo shaker mounted below the cantilever base (JPK, direct drive module). We used a standard gold coated tapping mode cantilever (PPP-NCHAuD Nanosensors[™], NanoWorld AG).

A. Calibration

We first calibrate the cantilever's dynamic stiffness in air using the Sader method with the analytical expression for

the hydrodynamic damping as a function of the Reynolds number,³⁴

$$\text{Re} = \frac{\rho b^2 \omega_0}{4\mu} \quad (1)$$

where b is the width of the beam, $\omega_0 = 2\pi f_0$ is the measured resonant frequency, and ρ and μ are the room-temperature density and the dynamic viscosity of air respectively. The resonance frequency $f_{0,\text{air}} = 268$ kHz and the quality factor $Q_{\text{air}} = 516$ are determined by fitting the power spectral density of the cantilever fluctuations. Applying this method in air ($\rho_{\text{air}} = 1.18$ [kg m⁻³], $\mu_{\text{air}} = 1.86 \times 10^{-5}$ [kgm s⁻¹] and $\text{Re}_{\text{air}} = 24$), we find $k_{\text{air}} = 23.7$ N m⁻¹. With this determination of k we use the fluctuation–dissipation theorem to determine the magnitude of displacement fluctuations,³⁵ thereby calibrating the inverse optical lever responsibility $\text{invOLR}_{\text{air}} = 55.2$ [nm V⁻¹].

After performing our measurements in air, we calibrate the cantilever in liquid. We completely submerge the cantilever in a 100 μ L drop of liquid buffer solution and leave it to equilibrate for a few minutes. We then re-calibrated using the same procedure ($\rho_{\text{liquid}} = 998.2$ [kg m⁻³], $\mu_{\text{liquid}} = 0.001$ [kgm s⁻¹] and $\text{Re}_{\text{liquid}} = 186$). The resonance frequency decreases by a factor of about 2, $f_{0,\text{liquid}} = 132$ kHz, and the quality factor is drastically lowered to $Q_{\text{liquid}} = 8$. The stiffness found by using the Sader method in liquid, $k_{\text{liquid}} = 26.8$ N m⁻¹, differs from that in air by about 13%. The inverse responsivity is $\text{invOLR}_{\text{liquid}} = 23.3$ [nm V⁻¹].

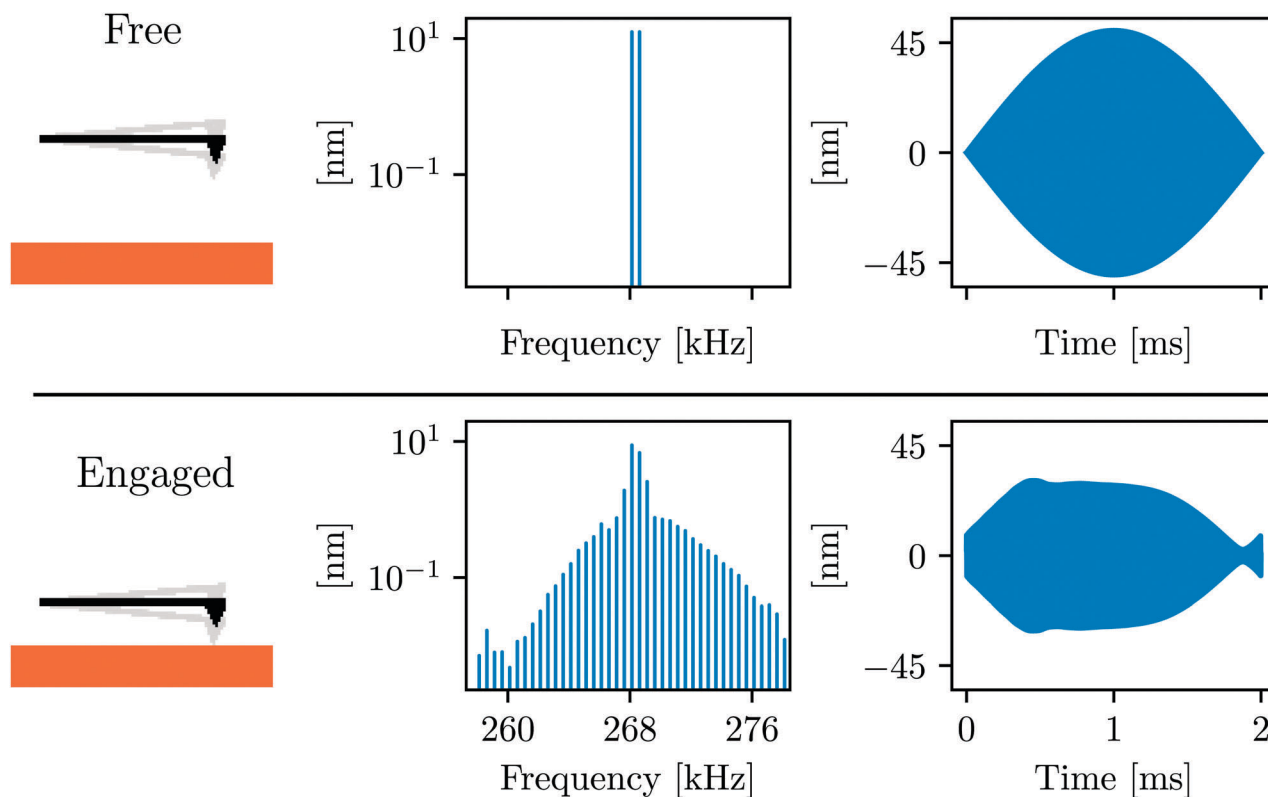


Fig. 1 Schematic illustrating the working principle of ImAFM: the cantilever is acoustically excited with two frequencies around its first flexural resonance. In the time domain, driving at two closely spaced frequencies gives a beating waveform, or slow modulation of a rapid oscillation. When engaging the sample surface many intermodulation products of the two drive tones are created around resonance, describing the distortion of the slow modulation. Conservative and dissipative force quadratures are reconstructed from the analysis of this distortion.^{28–30}



As a test-sample we used a well-studied polymer blend of polystyrene and low-density polyethylene (PS-LDPE, Bruker's *HarmoniX* sample). The polymers are well-separated in two domains, with PS forming a homogeneous background and LDPE forming circular domains scattered around the sample. A significantly different mechanical response is expected in the two domains, as the bulk Young's moduli for PS and LDPE are $E_{\text{nom}}^{\text{PS}} \approx 2\text{--}4\text{ GPa}$ and $E_{\text{nom}}^{\text{LDPE}} \approx 0.3\text{ GPa}$ respectively. Indeed the phase images in both air and liquid show a sharp contrast when scanning an area around a LDPE domain (see Fig. 2).

In the following, we examine the interaction force between the tip and the liquid–solid interface, comparing the measurements at the individual position in each domain, marked with an \times in Fig. 2. In order to isolate and examine only the tip–surface force, it is first necessary to subtract background forces due to squeezing of the dense and viscous liquid films between the cantilever body and the sample surface. We used the method of Borgani *et al.*³⁶

B. Force quadrature curves

At different positions we characterize the interaction between the tip and the surface by measuring dynamic force quadratures.

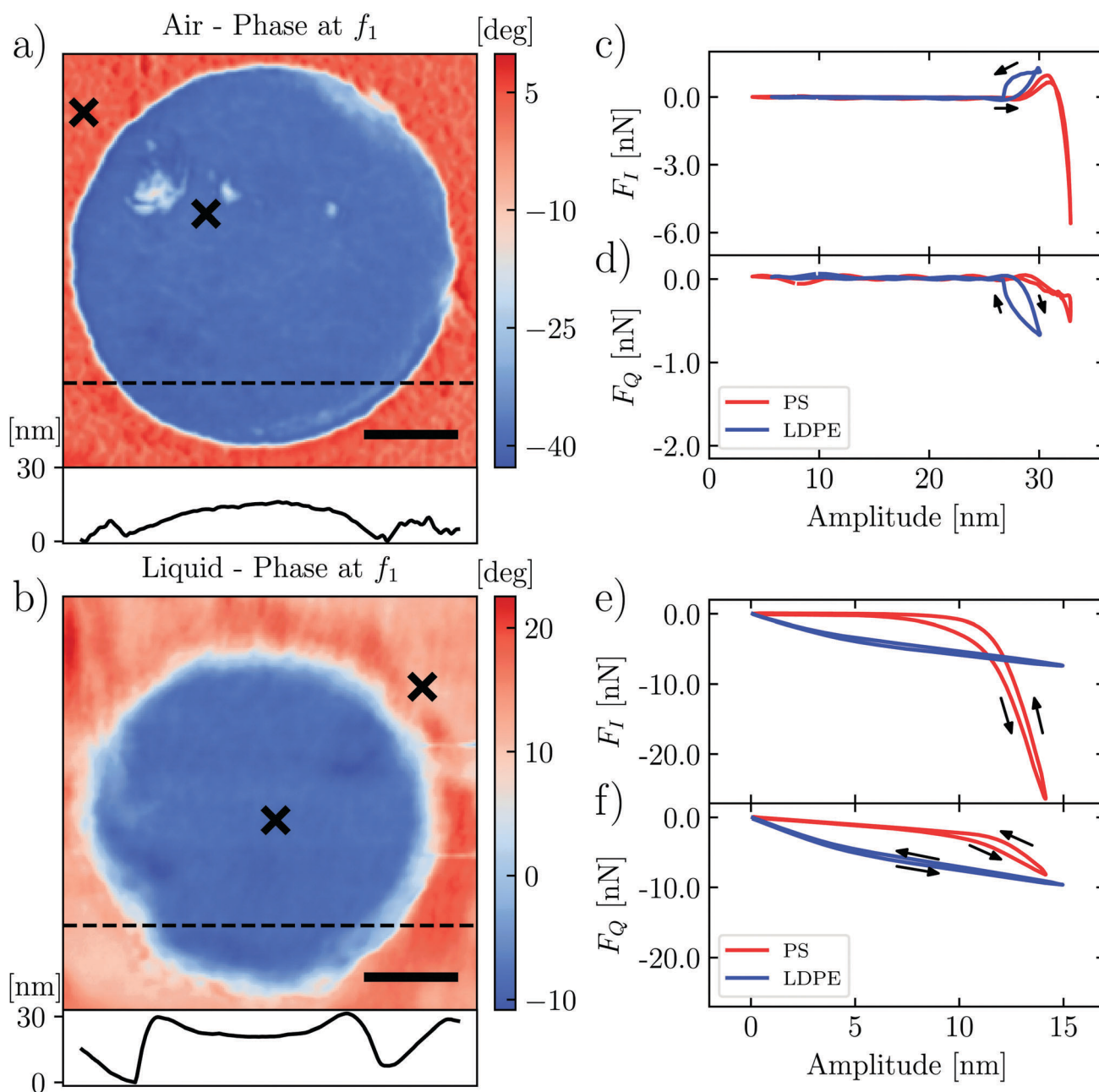


Fig. 2 LDPE domain in the PS matrix: the images in air (a) and in liquid (b) display the response phase at the first drive frequency of ImAFM. The scale bar is 250 nm. Below each image, the height profile at the dashed line is shown. The conservative force quadrature $F_T(A)$ and the dissipative force quadrature $F_Q(A)$ are shown at positions marked with an \times on PS (red) and LDPE (blue) in air (c and d) and in liquid (e and f). Note that the LDPE domains are not necessarily the same.



This method was introduced by Platz *et al.* and it has been well described in previous publications.^{28,29} We measure two force curves at each pixel of the image: the curve $F_I(A)$ shows the conservative part of the tip–surface interaction and the curve $F_Q(A)$ shows the dissipative part. These curves are determined independent of any assumed model of the interaction, but unlike the more familiar force–distance curves, they do not display the instantaneous force at some tip positions. Rather, they are the weighted integrals of force over a single oscillation cycle having a particular amplitude A . The force curve $F_I(A)$ shows the integrated force that is in-phase with the cantilever oscillation, and the curve $F_Q(A)$ shows the integrated force which is a quadrature to the motion or in-phase with the velocity.

Fig. 2(c)–(f) show the force quadratures $F_I(A)$ and $F_Q(A)$ measured at one point on the stiffer PS region (red) and at one point on the softer LDPE region (blue), in both air and liquid. In air (Fig. 2(c) and (d)) we see the curves with typical shapes: the stiffer PS shows $F_I(A)$ reaching a large negative value, indicating that the force was dominantly repulsive for oscillation cycles with the largest amplitude. In contrast, $F_I(A)$ on the softer LDPE is always positive up to the largest amplitude, corresponding to a dominantly attractive force. We also see hysteresis in the $F_I(A)$ and $F_Q(A)$ curves, and a larger negative value of $F_Q(A)$ corresponding to a greater energy loss in the oscillation cycle. In the next section, we explain these features as resulting from adhesion and viscous damping, giving rise to slow relaxation of an up-lifted surface.

The shapes of $F_I(A)$ and $F_Q(A)$ in liquid (Fig. 2(e) and (f)) are distinctly different from those in air. In liquid we do not see attractive force (adhesion) at any amplitude, consistent with quasi-static force measurements. For the stiffer PS surface (red curves), we see that $F_I(A) = 0$ for $A < 7$ nm, which we interpret as the tip not yet touching the surface when the oscillation amplitude is low. However, in contrast to air, we now see significant $F_Q(A)$ in this low amplitude regime. We may explain this observation as additional damping force when the liquid is compressed between the tip and the surface, as opposed to that when the liquid is compressed between the cantilever body and the surface. The squeeze-film damping by the cantilever body has been compensated for by subtracting off linear background forces.³⁶ We also see hysteresis in both $F_I(A)$ and $F_Q(A)$ measured in liquid, which we explain in the next section using a moving surface model.

In comparison to liquid, the cantilever oscillation is larger in air (Fig. 2(c) and (d) and Fig. 2(e), (f)) and the probe works further from the surface. Hence we see a larger region of amplitude A , where the force quadratures are zero. This region of zero force could be reduced by decreasing the cantilever drive force, so that the free amplitude is smaller. However, it is not always possible to achieve stable imaging with lower drive, because the stored energy in the cantilever oscillation can be insufficient to break free of the adhesion experienced by the tip when it touches the soft surface.

For the stiffer PS surface, we can compare elastic response in air and liquid by simply comparing the slopes of $F_I(A)$ in the high amplitude regime, where we find $\frac{dF_I}{dA} \approx 6 \text{ N m}^{-1}$ in both cases.

To relate this stiffness to a modulus, we use the method of Forchheimer *et al.* and fit the parameters of the Derjaguin–Müller–Toporov (DMT) model to the measured intermodulation spectrum.³⁰ Assuming a tip radius of 10 nm, we obtain the reduced moduli (*i.e.* the geometric mean of indenter modulus and surface modulus) $E_{\text{PS,air}}^* = 8.1 \pm 3 \text{ GPa}$ and $E_{\text{PS,liquid}}^* = 6.4 \pm 2 \text{ GPa}$. For the softer LDPE region, this type of analysis is not appropriate. We explain both the shape and magnitude of $F_I(A)$ and $F_Q(A)$ on this soft surface in the next section.

The force quadrature curves in Fig. 2 are shown at one position, but they are measured at every pixel while scanning. We can study the surface and liquid forces in more detail by measuring force quadrature curves while slowly approaching and retracting the probe at one point on the surface. Fig. 3 shows such a measurement in liquid when the free oscillation amplitude at drive frequency f_1 was 10 nm. The other drive frequency f_2 had the same amplitude, resulting in a beating waveform with a maximum amplitude of 20 nm, or 40 nm peak-to-peak. Fig. 3a plots the response amplitude at f_1 versus the change in probe height Δh , where the latter is measured with the internal z -sensor of the AFM scanner. We see that close to the surface, but well before touching, the amplitude at f_1 has dropped to about 8 nm, due to squeeze-film forces acting on the cantilever body.

When the tip begins to touch the surface around $\Delta h = 28$ nm, the reconstructed force quadratures (orange curves in Fig. 3(b) and (c) reveal the onset of the tip–surface force). As expected, moving the probe closer to the surface shifts this onset to a lower amplitude. The evolution of these curves shows that the repulsive conservative force at full amplitude (the largest negative $F_I(A)$) goes through the maximum and begins to decrease as we move very close to the surface. At small oscillation amplitudes, the tip is most likely never breaking free from the surface. Therefore, the non-monotonic behaviour of $F_I(A)$ may be associated with changing from the intermittent contact to the constant contact regimes of measurement. In contrast, the magnitude of the dissipative force is always increasing. However, the dissipated energy at the maximum amplitude $W_{\text{peak}} = 2\pi A_{\text{max}} F_{Q,\text{max}} \approx -282 \text{ aJ}$ does not change with probe height Δh (see the dashed line in Fig. 3c). This measurement demonstrates that, contrary to intuition, the work done by the cantilever on the sample surface in tapping mode AFM does not depend on how heavily the probe is engaged (oscillation amplitude).

The force quadrature curves are reconstructed from intermodulation spectra taken at the probe heights marked with an \times of the corresponding color in Fig. 3(a). Two examples of such spectra are shown in Fig. 3(d). Here we see the spectrum at the ‘lift’ height $\Delta h = 35$ nm (gray) with no distinguishable intermodulation above the noise level, verifying that the response is highly linear. In contrast, at an engaged height $\Delta h = 23$ nm (green), we see strong intermodulation near resonance, well above the noise level. In Fig. 3(e) we see the same two spectra plotted over a much wider frequency range to display all nonlinear responses appearing near the integer multiples of the center frequency, nf_0 , where $f_0 = \frac{1}{2}(f_1 + f_2)$.



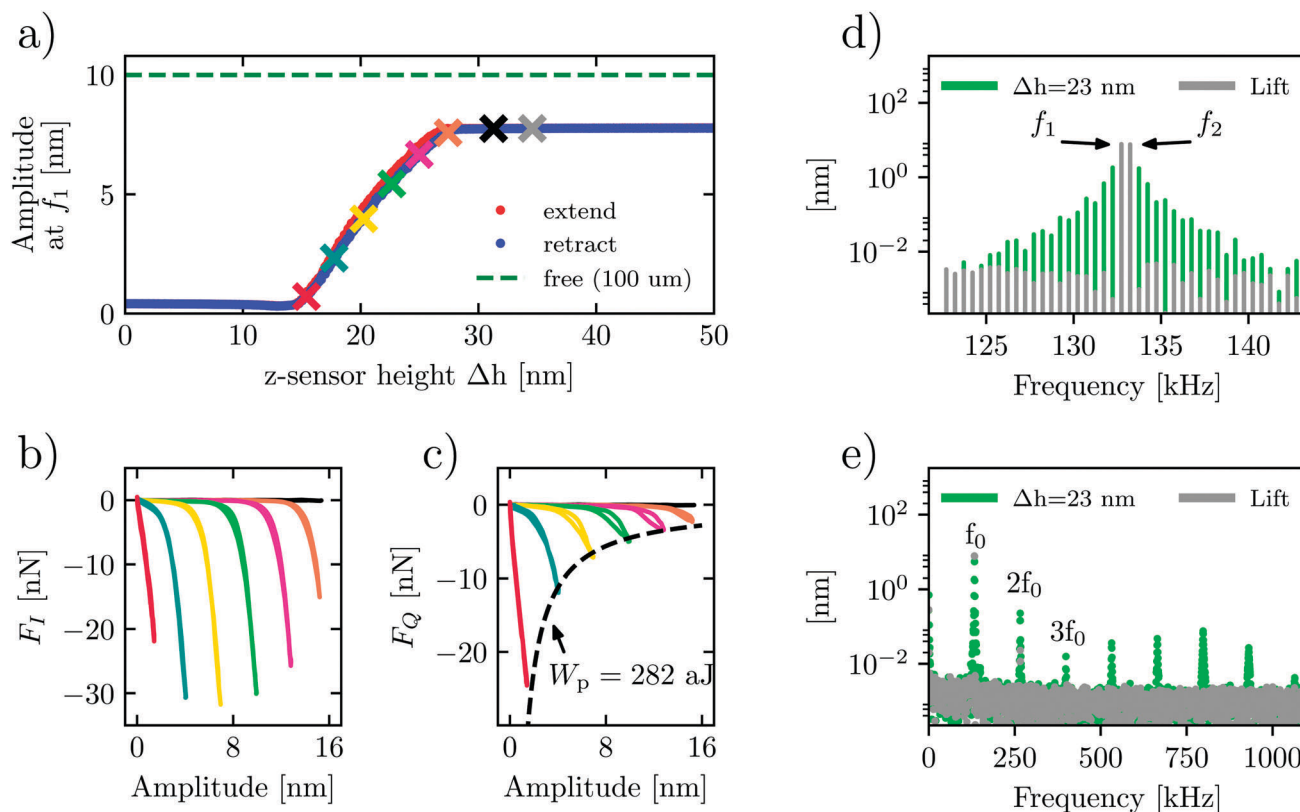


Fig. 3 Approach–retract measurements on PS in liquid. (a) The response amplitude at the first drive plotted against the extension of the probe as measured by the internal height sensor of AFM scanners. At the points marked with a colored x, we show the force quadrature curves $F_I(A)$ and $F_Q(A)$ in (b) and (c) respectively. (d) The measured intermodulation spectrum close to f_0 at the lifted position (gray) and at $h = 23$ nm (green). The same spectra are shown in (e) over a larger frequency range, where the response near higher harmonics is seen.

For the engaged case (green spectrum, $\Delta h = 23$ nm), one can see significant response at these higher harmonics and intermodulation products. We note that the response at higher frequencies around $5f_0$ to $7f_0$ is enhanced as a result of excitation of the second flexural eigenmode.²¹ While this higher-frequency nonlinear response is clearly observed above the detector noise due to the large dynamic range of the measurement, it is still a factor of 100 below the dominant response near f_0 . At the lift height none of this high-frequency nonlinear response is observed, apart from a small remnant near $2f_0$.

III. Theory and simulation

To validate our experimental methods and to better understand the nature of soft visco-elastic interfaces in liquid, we performed numerical simulations of the cantilever dynamics. We first simulate the cantilever dynamics assuming a particular model for the tip–surface interaction. From this simulated data we can verify the reconstruction of actual force quadrature curves using only the intermodulation spectrum near the resonance. In the second set of simulations, we apply a more realistic moving surface model to explain in detail the shape and hysteresis seen in the experimental force quadrature curves as resulting from the viscoelastic nature of the sample and liquid surroundings.

A. Force quadrature reconstruction with low Q

Force quadrature reconstruction is based on the idea that the dominant response is located in a narrow frequency band around resonance. The data given in Fig. 3(e) show significant response at higher frequencies well above f_0 , clearly bringing this assumption into question. Nevertheless, the simulations described below show that reasonable reconstruction accuracy can be achieved when neglecting the high-frequency nonlinear response.

We simulate the dynamics of a model cantilever with two flexural eigenmodes,^{37,38} where the total deflection from its equilibrium is $d = d_1 + d_2$. The equations of motion read,

$$\frac{1}{\omega_1^2} \ddot{d}_1 + \frac{1}{Q_1 \omega_1} \dot{d}_1 + d_1 = \frac{1}{k_1} (F_{\text{drive}}(t) + F_{\text{ts}}(d, \dot{d})), \quad (2)$$

$$\frac{1}{\omega_2^2} \ddot{d}_2 + \frac{1}{Q_2 \omega_2} \dot{d}_2 + d_2 = \frac{1}{k_2} (F_{\text{drive}}(t) + F_{\text{ts}}(d, \dot{d})) \quad (3)$$

where $d_{1(2)}$, $\omega_{1(2)}$, $Q_{1(2)}$, and $k_{1(2)}$ are the deflection, resonance frequency, quality factor and stiffness of the first (second) eigenmode respectively. The parameters for the first eigenmode are the calibrated experimental values, and for the second eigenmode we used the scaling relations described by Garcia.³⁹ The two modes are coupled through a nonlinear viscoelastic tip–surface



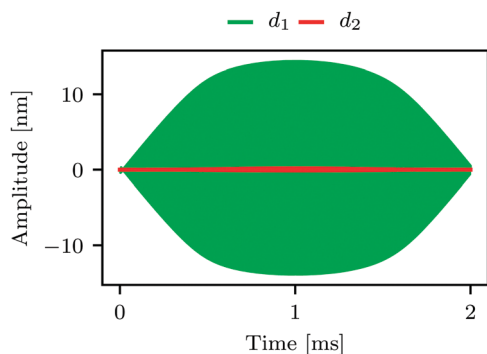


Fig. 4 Simulated motion of the first eigenmode $d_1(t)$ (green) and second eigenmode $d_2(t)$ (red). At this timescale, corresponding to one pixel in an AFM image, one cannot see the rapid oscillations, only the smooth envelope of the motion. The influence of $d_2(t)$ appears to be weak using this linear vertical scale, but the Fourier transforms shown in Fig. 5(c) and (d) show significant response around the integer multiples of f_0 .

force that depends on the total deflection and velocity. We used a Kelvin–Voigt-type model,

$$F_{\text{ts}}(d, \dot{d}) = F_c e^{-d/l_c} + \dot{d} F_d e^{-d/l_d}, \quad (4)$$

where the characteristic length scale l_c describes the range of a repulsive conservative force, and l_d is the range of a position-dependent viscous damping force. This phenomenological force captures the general shape of the force quadrature curves for a stiff surface, but it cannot reconstruct the hysteresis observed in the experiment.

We apply the two-frequency drive force used in ImAFM

$$F_{\text{drive}}(t) = F_1 \cos(2\pi f_1 t) + F_2 \cos(2\pi f_2 t) \quad (5)$$

with both frequencies closely spaced and centered at resonance. We simulate the steady-state motion $d(t)$ by numerical

integration of eqn (2) and (3) using the Python library *scipy.integrate.ode*⁴⁰ which implements an implicit Adams method. The simulated motion of the first and second eigenmodes is plotted in Fig. 4 over one period of the characteristic beating waveform. On this timescale we cannot see rapid sinusoidal oscillation, but one can see the oscillation envelope for each eigenmode. We note that the second mode is only weakly excited such that the total motion is dominated by the first eigenmode.

The force quadratures reconstructed from this simulation are shown in Fig. 5(a) and (b) (red curves) where they can be compared with the actual force quadratures (black curves). We emphasize that the actual curves are calculated directly from the force model, not from the simulated dynamics. The reconstructed force quadratures determined from the simulated dynamics using only the Fourier coefficients of the total deflection $\hat{d}(\omega)$ near f_0 (i.e. response at 2 drives and 38 intermodulation products). Fig. 5(c) shows an expanded view of the 20 center peaks located at the shaded region in Fig. 5(d). As with the experiment, the simulation shows significant response at higher frequencies which is enhanced near the second eigenmode (compare Fig. 3(e) and 5(d)).

Comparing the actual and reconstructed force quadratures (red vs. black curves in Fig. 5(a) and (b)), it is clear that the reconstruction works quite well. The good agreement is due to the fact that the higher frequency spectral components are relatively small, with most of the spectral power contained in the components near f_0 . One can imagine alternative methods of force reconstruction using the higher frequency response. However, any such method would require an additional calibration of the second eigenmode parameters including its invOLR, where the latter calibration depends strongly on the position of the laser spot on the cantilever.

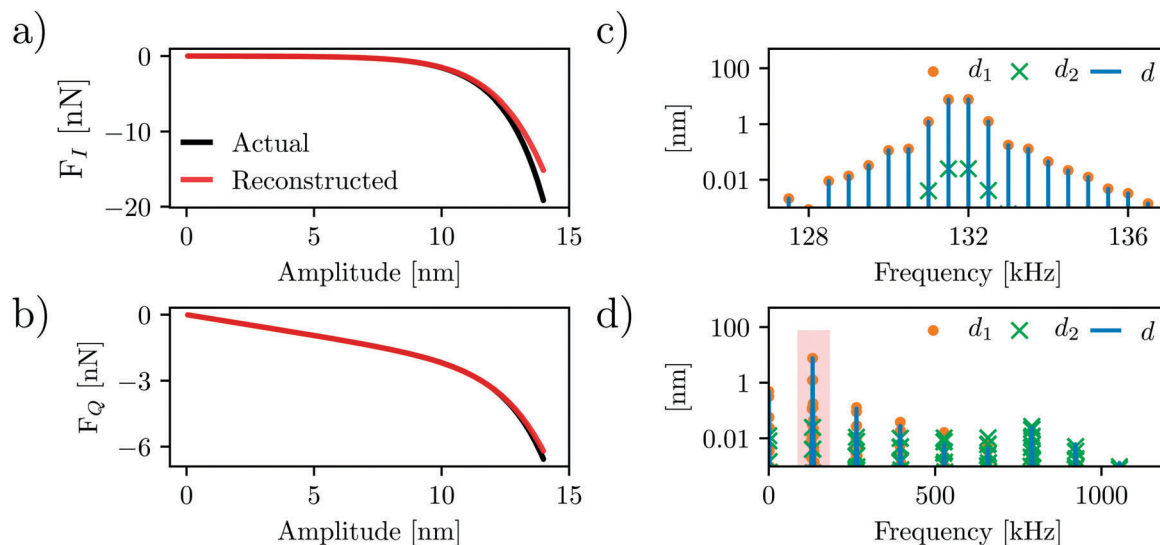


Fig. 5 Comparison of the actual and reconstructed force quadrature curves. (a and b) show the force quadrature curves $F_I(A)$ shown in (a) and $F_Q(A)$ respectively. The actual curves (black) are calculated directly from the force model such as eqn (4), whereas the reconstructed curves (red) are determined from the simulated nonlinear dynamics, using only frequency components of the motion near resonance. (c) shows the spectral content of the tip motion for each eigenmode near resonance, and (d) shows the same over a wider frequency range.



B. Viscoelastic response of a soft surface

The force quadrature curves on very soft materials cannot be explained in terms of simple force models such as eqn (4) which treat the interaction as a function of tip position and velocity only. In our previous publications^{31,41} we described a simple model which accounts for the motion of the viscoelastic surface, and we demonstrated that it explains the force quadrature curves on many different soft materials measured in air. The total force needed to deform the soft surface is modeled with two stiffnesses k_s and k_v , and two viscous relaxation times τ_s and τ_v ,

resulting from interfacial and volumetric contributions respectively. In Fig. 6 we show that this model also works exceptionally well for LDPE, both in air and in liquid.

The force quadratures in air are shown in Fig. 6(a) and (b) where we see excellent agreement between experiment and simulation of the moving surface model. All model parameters are given in the table and described in the caption of Fig. 6. The simulated tip motion and surface motion are shown in Fig. 6(c) and (d) where one sees that with each tip, the adhesion force lifts the surface, which does not fully relax before the next tip.

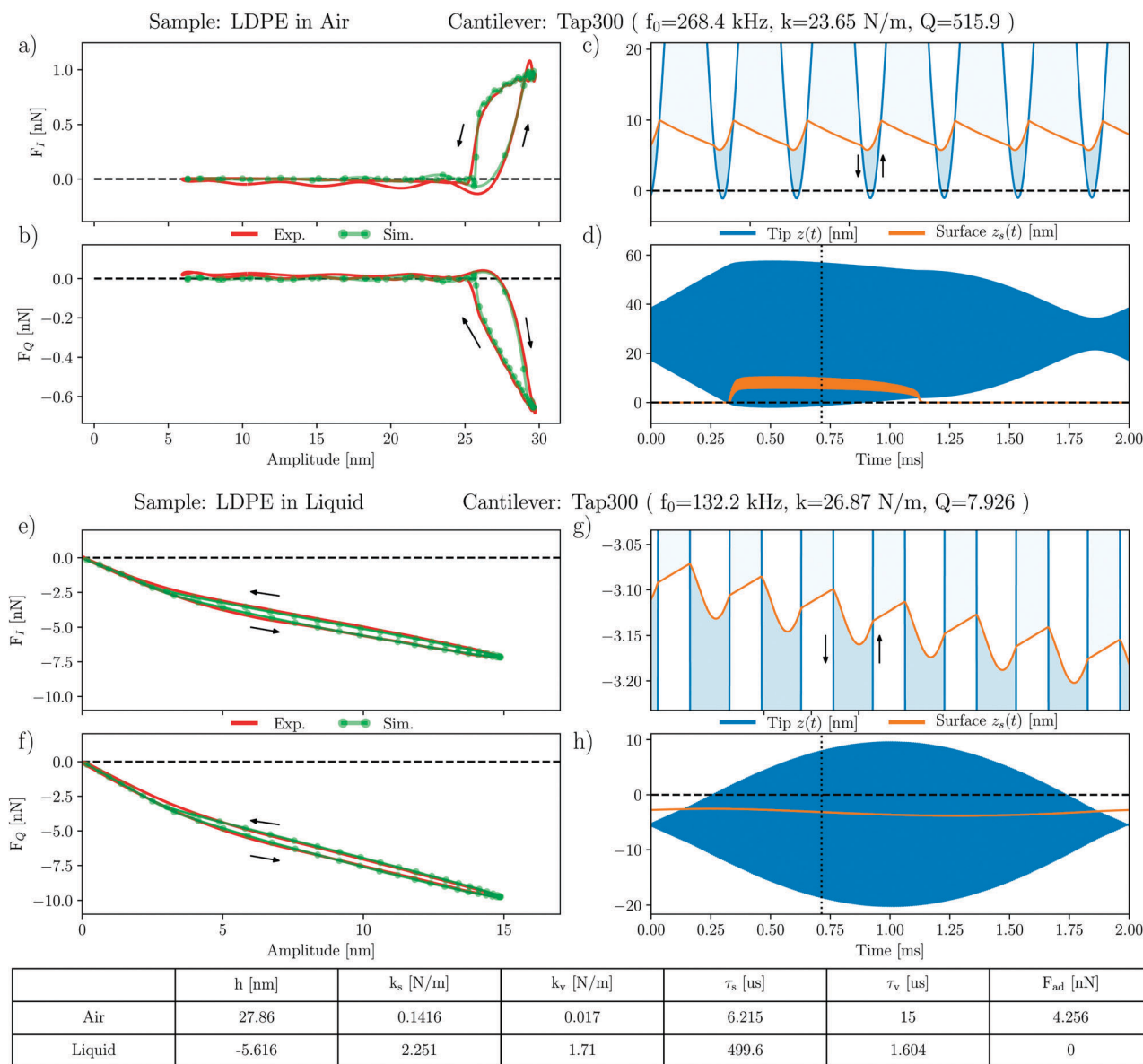


Fig. 6 Simulations of the moving surface model on LDPE in air and liquid. (a–d) show measurements and simulations on LDPE in air, and (e)–(h) show the same but in liquid. The same cantilever was used in both air and liquid, with the respective calibration parameters given in the figure. Model parameters were found by fitting the simulated and measured intermodulation spectral components of the force near resonance, resulting in excellent agreement between theory and experiment. The model parameters are: h – the equilibrium position of the tip, where the instantaneous tip position is $z(t) = h + d(t)$ and the equilibrium position of the surface is defined to be $z_s = 0$. k_s – stiffness giving a force proportional to the deformation of the surface from its equilibrium position. k_v – stiffness giving a force proportional to the penetration of the tip below the surface. $\tau_s = \eta_s/k_s$ – viscoelastic relaxation time where η_s is the viscous damping coefficient describing the force proportional to the rate of change in the surface position. $\tau_v = \eta_v/k_v$ the viscoelastic relaxation time where η_v is the viscous damping coefficient describing the force proportional to the rate of change in tip penetration.



The result is a time-average lifting of the surface, giving rise to the hysteresis observed in the force quadrature curves.

Excellent agreement between the measured and modeled force quadratures is also found in liquid, as shown in Fig. 6(e) and (f). Here the curves show a very different shape, but also observe hysteresis. Fig. 6(g) and (h) show that in liquid the cantilever is oscillating much closer to the surface, such that its equilibrium position of the tip $h = -5.6$ nm is actually below the equilibrium position of the surface. At low amplitude, the tip does not separate from the surface, with intermittent contact starting when the amplitude exceeds some 4 nm. We also see that, as expected, the adhesion force $F_{\text{ad}} = 0$ nN in liquid. Hence the absence of surface lifting in liquid.

A comparison of the moving surface model parameters in air and liquid shows that LDPE is much stiffer in liquid, both interfacial and bulk contributions. We also see that the relaxation time of the free surface τ_s in liquid is much longer, but the volumetric relaxation time τ_v becomes shorter. These observations can be explained by the increased Laplace pressure in liquid: in air the topography of the LDPE domain has a convex curvature, forming a dome-like protrusion above the flat PS surface. When submerged in the liquid, an increase in interfacial energy results in an increase in Laplace pressure across this curved interface, which causes the dome to deform and crumple (see the height profiles in Fig. 2(a) and (b)). This deformation is associated with the stiffening of LDPE, causing the parameter k_v to increase by two orders of magnitude. The viscous force constant $\eta = \tau_k$ also increases, but the relaxation time becomes shorter.

IV. Conclusion

We have demonstrated that ImAFM works extremely well in liquid environments where the cantilever's fundamental flexural resonance has the quality factor of order $Q \approx 10$. We performed experiments capturing the full nonlinear response including Fourier components up to frequencies many times the resonant frequency. Numerical simulation of a realistic model of the cantilever in liquid, including two eigenmodes, showed that the ImAFM force reconstruction methodology works very well using only the response in a narrow frequency band near resonance. This reconstruction method applied to experimental data shows hysteresis in the force quadrature curves, typical of soft viscoelastic materials. In both air and liquid, these experimental results were well explained by a moving surface model, which takes into account both the viscous and elastic nature of the sample.

Other fields such as biology and bioengineering can possibly take the advantage of measuring interfacial properties with ImAFM in environments relevant for cells, *i.e.* in liquid. Biological materials have widely varying mechanical properties. Bulk Young's moduli between $E \approx 10^3$ Pa and $E \approx 10^9$ Pa have been reported.⁴² In the case of extremely soft materials ($E \leq 10^6$ Pa), typical AFM measurements are dominated by surface forces. AFM work has largely ignored viscosity but, in our experience with dynamic AFM on soft polymers, it plays a dominant role. It will be necessary to

adapt the measurement conditions, *i.e.* cantilever resonance frequency, stiffness and drive amplitude, on a case-to-case basis. However for ImAFM, under damped dynamics, $Q \gtrsim 10$ is desirable. If Q becomes too small, force reconstruction using harmonics may be more appropriate.²⁰

Conflicts of interest

Two of the authors, DBH and DF, are part owners in the company Intermodulation Products AB.

Acknowledgements

PAT, DF, RB and DBH gratefully acknowledge financial support by the Swedish Vetenskapsrådet (VR), Stiftelsen Olle Engkvist Byggmästare and Knut och Alice Wallenbergs Stiftelse. FC, BRR and AF gratefully acknowledge financial support by the Swiss National Center of Competence in Research Bio-Inspired Materials and the Swiss National Science Foundation.

References

- 1 S. E. Cross, Y.-S. Jin, J. Tondre, R. Wong, J. Rao and J. K. Gimzewski, AFM-based analysis of human metastatic cancer cells, *Nanotechnology*, 2008, **19**(38), 384003.
- 2 M. Li, L. Liu, X. Xiao, N. Xi and Y. Wang, Viscoelastic Properties Measurement of Human Lymphocytes by Atomic Force Microscopy Based on Magnetic Beads Cell Isolation, *IEEE Trans. Nanobioscience*, 2016, **15**(5), 398–411.
- 3 A. B. Mathur, A. M. Collinsworth, W. M. Reichert, W. E. Kraus and G. A. Truskey, Endothelial, cardiac muscle and skeletal muscle exhibit different viscous and elastic properties as determined by atomic force microscopy, *J. Biomech.*, 2001, **34**(12), 1545–1553.
- 4 S. Tripathy and E. J. Berger, Measuring Viscoelasticity of Soft Samples Using Atomic Force Microscopy, *J. Biomech. Eng.*, 2009, **131**(9), 094507.
- 5 T. R. Matzelle, G. Geuskens and N. Kruse, Elastic properties of poly(N-isopropylacrylamide) and poly(acrylamide) hydrogels studied by scanning force microscopy, *Macromolecules*, 2003, **36**(8), 2926–2931.
- 6 A. J. Engler, S. Sen, H. L. Sweeney and D. E. Discher, Matrix Elasticity Directs Stem Cell Lineage Specification, *Cell*, 2006, **126**(4), 677–689.
- 7 D. E. Discher, Tissue Cells Feel and Respond to the Stiffness of Their Substrate, *Science*, 2005, **310**(5751), 1139–1143.
- 8 E. Martinez, A. Lagunas, C. A. Mills, S. Rodriguez-Segui, M. Estevez, S. Oberhansl, J. Comelles and J. Samitier, Stem cell differentiation by functionalized micro- and nanostructured surfaces, *Nanomedicine*, 2009, **4**(1), 65–82.
- 9 G. C. Binnig, C. F. Quate and Ch. Gerber, Atomic Force Microscope, *Phys. Rev. Lett.*, 1986, **56**(9), 930.
- 10 A. Cartagena and A. Raman, Local viscoelastic properties of live cells investigated using dynamic and quasi-static atomic force microscopy methods, *Biophys. J.*, 2014, **106**(5), 1033–1043.



- 11 H. Schillers, M. Wälte, K. Urbanova and H. Oberleithner, Real-time monitoring of cell elasticity reveals oscillating myosin activity, *Biophys. J.*, 2010, **99**(11), 3639–3646.
- 12 H. Babahosseini, B. Carmichael, J. S. Strobl, S. N. Mahmoodi and M. Agah, Sub-cellular force microscopy in single normal and cancer cells, *Biochem. Biophys. Res. Commun.*, 2015, **463**(4), 587–592.
- 13 B. Fallqvist, M. L. Fielden, T. Pettersson, N. Nordgren, M. Kroon and A. K. B. Gad, Experimental and computational assessment of F-actin influence in regulating cellular stiffness and relaxation behaviour of fibroblasts, *J. Mech. Behav. Biomed. Mater.*, 2016, **59**, 168–184.
- 14 S. Iyer, R. M. Gaikwad, V. Subba-Rao, C. D. Woodworth and I. Sokolov, AFM Detects Differences in the surface Brush of normal and cancerous cervical cells, *Nat. Nanotechnol.*, 2011, **4**(6), 389–393.
- 15 L. Hamon, D. Pastré, P. Dupaigne, C. L. Breton, E. L. Cam and O. Piétrement, High-resolution AFM imaging of single-stranded DNA-binding (SSB) protein–DNA complexes, *Nucleic Acids Res.*, 2007, **35**(8), 1–7.
- 16 M. B. Viani, T. E. Schäffer, A. Chand, M. Rief, H. E. Gaub and P. K. Hansma, Small cantilevers for force spectroscopy of single molecules, *J. Appl. Phys.*, 1999, **86**(4), 2258–2262.
- 17 R. Borgani, D. Forchheimer, J. Bergqvist, P.-A. Thorén, O. Inganäs and D. B. Haviland, Intermodulation electrostatic force microscopy for imaging surface photo-voltage, *Appl. Phys. Lett.*, 2014, **105**(14), 143113.
- 18 W. Melitz, J. Shen, A. C. Kummel and S. Lee, Kelvin probe force microscopy and its application, *Surf. Sci. Rep.*, 2011, **66**(1), 1–27.
- 19 X. Zhao, J. Schwenk, A. O. Mandru, M. G. Penedo, M. Bacani, M. A. Marioni and H. J. Hug, Magnetic force microscopy with frequency-modulated capacitive tip-sample distance control, *New J. Phys.*, 2018, 013018.
- 20 J. Legleiter, M. Park, B. Cusick and T. Kowalewski, Scanning probe acceleration microscopy (SPAM) in fluids: mapping mechanical properties of surfaces at the nanoscale, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**(13), 4813–4818.
- 21 X. Xu, J. Melcher and A. Raman, Accurate force spectroscopy in tapping mode atomic force microscopy in liquids, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2010, **81**(3), 1–7.
- 22 T. Fukuma, M. J. Higgins and S. P. Jarvis, Direct imaging of individual intrinsic hydration layers on lipid bilayers at Ångstrom resolution, *Biophys. J.*, 2007, **92**(10), 3603–3609.
- 23 J. I. Kilpatrick, S. H. Loh and S. P. Jarvis, Directly probing the effects of ions on hydration forces at interfaces, *J. Am. Chem. Soc.*, 2013, **135**(7), 2628–2634.
- 24 J. P. K. Instruments AG. QI™ mode – Quantitative Imaging with the Nanowizard® 3 AFM. Technical report.
- 25 D. Platz, E. A. Tholén, D. Pesen and D. B. Haviland, Intermodulation atomic force microscopy, *Appl. Phys. Lett.*, 2008, **92**(15), 153106.
- 26 S. Jesse, S. V. Kalinin, R. Proksch, A. P. Baddorf and B. J. Rodriguez, The band excitation method in scanning probe microscopy for rapid mapping of energy dissipation on the nanoscale, *Nanotechnology*, 2007, **18**(43), 435503.
- 27 J. P. Killgore, D. G. Yablon, A. H. Tsou, A. Gannepalli, P. A. Yuya, J. A. Turner, R. Proksch and D. C. Hurley, Viscoelastic property mapping with contact resonance force microscopy, *Langmuir*, 2011, **27**(23), 13983–13987.
- 28 D. B. Haviland, Quantitative force microscopy from a dynamic point of view, *Curr. Opin. Colloid Interface Sci.*, 2017, **27**, 74–81.
- 29 D. Platz, D. Forchheimer, E. A. Tholén and D. B. Haviland, Interaction imaging with amplitude-dependence force spectroscopy, *Nat. Commun.*, 2013, **4**, 1360.
- 30 D. Forchheimer, D. Platz, E. A. Tholén and D. B. Haviland, Model-based extraction of material properties in multi-frequency atomic force microscopy, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2012, **85**(19), 195449.
- 31 P.-A. Thorén, R. Borgani, D. Forchheimer, I. Dobryden, P. M. Claesson, H. G. Kassa, P. Leclère, Y. W. Heinrich, M. Jaeger and D. B. Haviland, On modeling and measuring viscoelasticity with dynamic Atomic Force Microscopy, 2017, arXiv:1711.09024.
- 32 <http://www.intermodulation-products.com/>.
- 33 www.jpk.com.
- 34 J. E. Sader, Frequency response of cantilever beams immersed in viscous fluids with applications to the atomic force microscope, *J. Appl. Phys.*, 1998, **84**(1), 64–76.
- 35 M. J. Higgins, R. Proksch, J. E. Sader, M. Polcik, S. M. Endoo, J. P. Cleveland and S. P. Jarvis, Noninvasive determination of optical lever sensitivity in atomic force microscopy, *Rev. Sci. Instrum.*, 2006, **77**(1), 13701.
- 36 R. Borgani, P.-A. Thorén, D. Forchheimer, I. Dobryden, S. M. Sah, P. M. Claesson and D. B. Haviland, Background-force compensation in dynamic atomic force microscopy, *Phys. Rev. Appl.*, 2017, **7**, 064018.
- 37 D. Forchheimer, S. S. Borysov, D. Platz and D. B. Haviland, Determining surface properties with bimodal and multimodal AFM, *Nanotechnology*, 2014, **25**(48), 485708.
- 38 S. S. Borysov, D. Platz, A. S. de Wijn, D. Forchheimer, E. A. Tholén, A. V. Balatsky and D. B. Haviland, Reconstruction of tip–surface interactions with multimodal intermodulation atomic force microscopy, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2013, **88**(11), 115405.
- 39 R. Garcia and E. T. Herruzo, The emergence of multifrequency force microscopy, *Nat. Nanotechnol.*, 2012, **7**(4), 217–226.
- 40 E. Jones, T. Oliphant and P. Peterson, *et al.*, SciPy: Open source scientific tools for Python, 2001, [SciPy version 0.19.1].
- 41 D. B. Haviland, C. A. van Eysden, D. Forchheimer, D. Platz, H. G. Kassa and P. Leclère, Probing viscoelastic response of soft material surfaces at the nanoscale, *Soft Matter*, 2016, **12**(2), 619–624.
- 42 E. Moeendarbary and A. R. Harris, Cell mechanics: principles, practices, and prospects, *Wiley Interdiscip. Rev.: Syst. Biol. Med.*, 2014, **6**, 371.
- 43 In the interest of full disclosure, we hereby declare that two of the authors, DBH and DF, are part owners in the company Intermodulation Products AB.

