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Advances in targeting the folate receptor in the treatment/imaging of cancers

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The folate receptor (FR) is a recognised biomarker for tumour cells due to its overexpression on a large number of tumours. Consequently, the FR has been exploited by many diagnostic and therapeutic tools to allow targeted delivery to, and imaging of, cancer cells. Herein, we describe the many different approaches by which this has been achieved, including the attachment of folate to potent chemotherapeutic drugs to form FR-targeting small molecule–drug conjugates (SMDCs), FR-targeting antibodies (as antibody alone and as an antibody–drug conjugate), and in the form of complementary nanotechnology–folate platforms; as well as imaging variants thereof. The potential of exploiting the FR for targeted therapy/imaging has the potential to revolutionise the way several cancers are treated. These FR-targeted technologies can also pave the way for inspiring further sophisticated drug conjugates, especially as this receptor is being targeted by use of several complementary technologies: small molecule, nanoparticle and protein-based – thus providing broad and distinct knowledge in the area.

1 Introduction

1.1 Cancer treatment: chemotherapy and targeted therapy

Tumour-targeted drug delivery systems (TTDDSs) have emerged as a promising strategy in cancer treatment as they largely bypass the adverse side effects characteristic of conventional chemotherapy.¹ Targeting particular biomarkers that are overexpressed specifically on tumour cells enables the

selective delivery of cytotoxic cargo to cancerous tissue, thereby minimising toxic side effects in the patient and increasing the therapeutic index.^{2,3} In recent years, many receptors have been identified as being overexpressed on cancer cells, *e.g.* prostate-specific membrane antigen (PSMA), and the carbonic anhydrase IX and biotin receptors.^{2,4,5} In addition to these, the folate receptor (FR) has attracted considerable attention in the field. This review will highlight the recent progress in the use of the FR, particularly FR α as this is what the vast majority of FR-targeted drug delivery systems have focused on, as a promising candidate for tumour-targeted delivery due to its elevated expression on various cancer cell types.

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Faiza Javaid received a first-class honours MSci degree in Biochemistry from University College London (UCL), during which she was awarded the Plimmer Prize for Biochemistry (2014) and was twice a Dean's list commender. In 2016, she joined the Wellcome Trust 4 year Interdisciplinary Programme, and will be joining the lab of Dr Vijay Chudasama at UCL to study for her PhD, focusing on

the appraisal of the pyridazinedione platform for the development of a novel antibody drug conjugate.



1.2 The folate receptor

Several criteria must be considered when selecting an appropriate receptor for targeted drug delivery, the most important being considerably higher expression of the receptor of interest on cancerous tissue relative to normal cells. Additionally, in receptor-mediated internalisation pathways, the receptor would ideally rapidly recycle back to the cell surface, and/or be up-regulated, post interaction with the TTDDS to enable maximum delivery of further targeted therapeutics.⁶ Furthermore, to ensure specific delivery to the target cell, the receptor must be preferentially enriched at its surface and not released in circulation.⁷

Amongst the receptors that meet these requirements is a family of glycoproteins (35–40 kDa) known as the folate receptors (FRs),⁸ which can be divided into three different isoforms: FR α , FR β and FR γ . The α and β variants are attached to the cell membrane *via* glycosylphosphatidylinositol (GPI) anchors, whereas FR γ is found only in hematopoietic cells⁹ and lacks the GPI component, making it freely soluble.^{8,10}

The FR β isoform shares approximately 70% of its homology with FR α , and both possess a comparable affinity for folate.¹¹ FR β is upregulated on activated myeloid cells (primarily monocytes and macrophages) that participate in inflammatory and autoimmune diseases.^{12–15} The FR β isoform has also been detected in tumour-associated macrophages (TAMs) of many cancers, including those of the liver, kidney, skin, lung, blood and soft tissue.^{16–18} These macrophages can penetrate solid tumours and promote their metastasis and growth by suppression of CD8⁺ T cells and secretion of proangiogenic factors.¹⁹ The FR β isoform can consequently serve as a potential target for the selective delivery of cytotoxic agents in cancer treatment.

Notwithstanding FR β 's expression on some cancers, the FR α isoform has the most potential for targeted cancer therapy as it is the most widely expressed of all the FR isoforms²⁰ and is overexpressed in a large number of cancers of epithelial origin, including breast,²¹ lung, kidney and ovarian cancers,²² the

expression in these carcinomas being 100–300 times higher than on healthy cells and in the order of 1–10 million receptor copies per cell.^{23,24} Most healthy cells use the reduced folate carrier for folate uptake and thus the expression and distribution of FR α in non-cancerous tissue is largely confined to cells crucial for embryonic development, the choroid plexus in the brain and to the kidneys, where folates are filtered through the glomeruli and subsequently reabsorbed in the proximal tubule cells *via* FR-binding.^{8,25,26} Moreover, only the apical surface of these healthy cells expresses FR α , prohibiting the exposure of this receptor to folates found in circulation and cytotoxic agents administered parenterally, as intercellular junctions impede small molecules from crossing the epithelium.^{6,27} Upon tumorigenesis however, the entire architecture of a cell changes, with the vasculature becoming disorganised and chaotic and intercellular junctions being lost, giving rise to weak association between endothelial cells.²⁸ As a consequence, FR α loses its polarised cellular localisation and becomes randomly distributed over the entire cell surface.⁶ This phenomenon now renders FR α accessible to drug conjugates in blood circulation (Fig. 1).

This loss of receptor polarisation, combined with the high binding affinity ($K_D = 0.1$ – 1 nM)²⁹ of FR α for oxidised folates, such as folic acid, (FA), has led to the development of drug conjugates with FA as the targeting entity. The benefits of using folic acid in this way are manifold: it is non-immunogenic, cost efficient, has high stability and tissue permeability, possesses a low molecular weight and can be easily conjugated to diverse types of organic molecules, antibodies and nanoparticles.³⁰ There are only certain identified positions on the folic acid core scaffold where it can be attached to the rest of the conjugate without compromising the high binding strength to FR α , with one position proving to be the most favoured (Fig. 2).

Folates, once inside the cell, are needed for one carbon methylation reactions as well as the *de novo* synthesis of purines and thymidine, which are in turn required for DNA synthesis and repair. This makes FA integral to the survival of normal cells, but especially important for tumour tissues in order to sustain their rapid, uncontrolled and aggressive proliferation.^{10,24,31}

Typically, folates are taken up into cells by FR α and FR β *via* receptor-mediated endocytosis (RME).¹¹ The folate portion of the conjugate acts as the tumour-targeting ligand and will bind strongly to FR α and/or FR β receptors on a cancer cell, resulting in subsequent internalisation of the folate–drug conjugate. Once the construct is sequestered within the early endosome, resident proton pumps cause a slight decrease in pH, altering the FR's conformation and allowing the conjugate to detach from the receptor.^{24,32,33} The late endosome then fuses with the lysosome and intracellular thiols, such as glutathione (GluSH) which can degrade the conjugate by cleaving a self-immolative linker, enable release of the free toxic drug. This lethal payload can subsequently diffuse out of the endosome into the cytosol where it induces cell death; meanwhile, the folate receptors are recycled back to the surface of the cell to engage in further rounds of drug internalisation.²⁴ The availability of unoccupied receptors on the cell surface is dependent on the recycling rate of the empty receptors from the endosome, and



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on the research areas of aerobic C–H activation and various aspects of Chemical Biology. Vijay's research has recently been highlighted by Forbes, Scientific American, CNN, Nature Chemistry and the Royal Society of Chemistry.





Fig. 1 The transformation of healthy epithelial cells to tumour cells and the effect it has on receptor positioning. Upon tumorigenesis, intercellular junctions (tight junctions, adherens junction, desmosomes and gap junctions) are lost and receptors that were previously only found on the apical surface become randomly positioned on the tumour.⁶



Fig. 2 Structure of folic acid (FA). This vitamin's constituent units are pteric and glutamic acid.

due to its rapid recycling rate (8–12 h),²⁴ the FR has the potential to maximise drug capture and internalisation (Fig. 3).

This ability to attach chemical warheads to ligands that seek out FR α -expressing tumours, confers excellent selectivity to the construct while preserving drug potency, and this approach has led to the development of many small molecule–drug conjugates based on folic acid (FA-SMDCs).

2 Small molecule–drug conjugates (SMDCs)

2.1 Vintafolide

The most successful FA-SMDC is vintafolide, (formerly EC145): a water-soluble conjugate that selectively delivers the drug desacetyl vinblastine monohydrate (DAVLBH) to tumours that overexpress FR α .²⁹ Preclinical studies have shown vintafolide to bind to FR α with high affinity, and therefore has very specific and potent activity against FR α positive tumour xenografts as opposed to the untargeted DAVLBH.

The four constituent modules of vintafolide consist of: (1) a folic acid moiety to target FR α , (2) a hydrophilic peptide

spacer, (3) a self-immolative disulfide linker, and (4) a microtubule-destabilising drug DAVLBH (Fig. 4).²⁴ As folic acid is lipophilic, the spacer serves to ameliorate the overall water



Fig. 3 Receptor-mediated endocytosis of a folate–drug conjugate. The construct initially binds to FR α or FR β , forming an invagination and enclosing the conjugate in the early endosome. A mild drop in pH alters the receptor's conformation, resulting in FA–drug detachment. The late endosome's subsequent fusion with the lysosome leads to degradation of the conjugate and release of the free cytotoxic drug into the cell. The recycling endosome delivers the folate receptors back to the cell surface.⁶



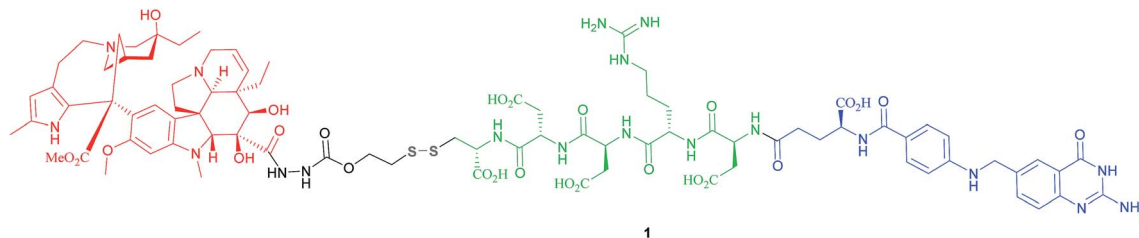


Fig. 4 Chemical structure of the folic acid-based SMDC vintafolide **1** is comprised of a folate targeting ligand (blue), a peptide spacer (green), a self-immolative disulfide linker (grey) and the potent cytotoxic drug DAVLBH (red).

solubility of the drug conjugate and in so doing, eliminates non-specific diffusion across cell membranes and ensures cell internalisation *via* RME. Typical examples of spacers commonly employed in FA-SMDCs include polysaccharides, peptides and polyethylene glycol (PEG) chains.^{6,24} An additional function provided by the spacer is to physically separate the drug cargo and targeting ligand, thereby minimising steric interference between the two and ensuring the retention of receptor binding affinity for the ligand.^{6,24} However, spacer length should not be too great as long, flexible spacers can allow the drug moiety to loop back and interact with the targeting ligand, jeopardising its affinity for the receptor.⁷

Small size (typically lower than 2000 Da) is critical for superior FA-SMDC tumour penetration and rapid systemic clearance.²⁴ Possessing a molecular weight of 1917 Da, vintafolide fulfils this criterion and displays a distribution time of 6 min.¹⁴ This short delivery time indicates rapid uptake of the drug conjugate by FR-positive tumour tissue, which is a desirable characteristic in minimising circulation time, and thus

precluding premature drug release. This FA-SMDC is also rapidly cleared from the body (elimination half-life of 26 min) *via* the kidneys and liver.³⁴

Owing to these attractive and interesting properties, Leamon *et al.* carried out a study aiming to evaluate the impact of altering three out of the four of vintafolide's constituent elements. They demonstrated that varying the spacer composition, provided that it remained hydrophilic, had minimal effect on the potency of the conjugate. In contrast, bioreleasable linkers that can be cleaved by intracellular thiols such glutathione (GluSH) in the endosomal milieu are of critical importance for the conjugate's activity and have by far constituted the most successful approach for triggered drug release within the cell (Fig. 5).²⁴ For instance, it was shown that self-immolative disulfide and acyl hydrazone linkers exerted activity both *in vitro* and *in vivo*, whereas vintafolide analogues possessing more stable amide and thioether linkers did not.²⁹ Furthermore, it was demonstrated that upon substituting DAVLBH with other clinically approved vinca alkaloid drugs (vincristine, vindesine, vinorelbine and vinflunine)



Fig. 5 Mechanism of vintafolide drug release in the intracellular milieu. Glutathione cleaves the disulfide linker and the resulting thiolate undergoes a self-immolative, 1,2-elimination reaction to liberate the free drug DAVLBH.²⁴



while retaining the cleavable disulfide linker, that vintafolide was the only variant that exhibited biological activity *in vitro* and *in vivo*. This can be rationalised by considering vintafolide's high potency ($IC_{50} = 8.44 \pm 1.46$ nM) as compared to the over ten-fold greater IC_{50} values of all the other drugs ($IC_{50} > 100$ nM). It was further speculated that the absence of activity observed in the other vinca alkaloid forms could be due to a modification in the chemical structure following disulfide reduction and linker release.²⁹

Vintafolide has shown promise, both as a single agent, as well as in combination with doxorubicin in two phase II trials (ovarian and non-small cell lung cancers) and in a randomised open-label phase II study respectively (platinum-resistant ovarian cancer)²⁹ and entered phase III clinical trials in 2010 for advanced stage platinum-resistant ovarian cancer.^{10,35} However, this FA-SMDC failed to reach the pre-specified criteria for progression free survival and as a result, the trial was prematurely terminated. Despite the expectations to perform otherwise, the phase III trials were unable to solidify the superiority of the targeted FR therapy to conventional chemotherapeutic methods. Future trials and studies must therefore account for an appropriate selection of eligible patients that are likely to benefit sufficiently from anti-FR therapy.

2.2 Folate-taxoid conjugate

Seitz *et al.* have developed a highly potent next-generation folate-taxoid for use against drug-resistant and drug-sensitive cancer cell lines.¹ This folate-taxoid conjugate incorporates a folic acid targeting moiety and a highly potent taxoid SB-T-1214, which is a derivative of the chemotherapeutic drug Taxol. Similar to vintafolide, this SMDC possesses a self-immolative disulfide linker, and a hydrophilic PEGylated dipeptide spacer (Fig. 6).¹

In vitro analysis was carried out to compare the activity of the taxoid conjugate 2 and free SB-T-1214 in FR α -positive and FR α -negative cells. As expected, free SB-T-1214 was highly potent against all cell lines. Conversely, taxoid conjugate 2 exhibited appreciable cytotoxicity against the FR α -positive cell lines,

displaying IC_{50} values more than three times smaller than those observed for the FR α -negative cells. This notable potency has been ascribed to the uptake of the folate-taxoid 2 occurring *via* RME, an internalisation pathway unaffected by the folic acid naturally present in the cell culture medium, which suggests that folic acid required for cell growth is principally shuttled into cells through folate transport proteins *in lieu* of RME. Further, taxoid conjugate 2 also exhibited an over 1000-fold decrease in toxicity against healthy cells compared to the free drug. As with vintafolide, the cytotoxic activity of 2 stems from intracellular GluSH-triggered reduction of the disulfide linker to release the free toxic drug SB-T-1214.¹ Ideally for maximum biological activity, the drug should be released in its unmodified form, as with conjugate 2, giving further weight to the aforementioned speculation that the failure of vintafolide analogues may be due to the liberation of a chemically altered payload. Moreover, the efficient release of the chemical warheads is contingent on the GluSH levels present in the intracellular milieu, the concentration of which can vary in different cell lines. It is therefore important to consider this particular variation when selecting tumour cell lines to be targeted by SMDCs whose activity is dependent on the intracellular GluSH concentration. Partly in view of this potential complication/limitation with certain cancer cells and serum stability questionability, FA-SMDCs have been developed where degradation to release free drug is not mediated by intracellular GluSH.

The above examples comprise a small, but representative, selection of FA-SMDCs from a vast field of conjugates that employ a disulfide linker for cytotoxic drug release. It is of particular relevance to highlight that folate conjugates to many other drugs *via* a disulfide linker, such as maytansinoids,³⁶ mitomycins,³⁷ alkaloid/mitomycins,³⁸ tubulysins^{39,40} and camptothecins,⁴¹ have been prepared and appraised.

2.3 Dendritic β -galactosidase-responsive folate-monomethylauristatin E conjugate

There are a variety of free thiol-containing compounds present in the blood and as such, the disulfide bond in FA-SMDCs is



Fig. 6 Structure of the folate-taxoid conjugate 2 developed by Seitz *et al.*¹



susceptible to cleavage in circulation by these thiols, potentially giving rise to undesired premature drug release. Consequently, alternative approaches have been developed in which the FA-SMDCs do not possess disulfide linkers, a structural property which would ideally minimise off-target drug liberation in the bloodstream. One such example developed by Alsarraf *et al.* is the β -galactosidase-responsive drug conjugate **3** that delivers the potent antineoplastic drug monomethylauristatin E (MMAE) to cancer cells.⁴² This SMDC consists of a galactoside trigger, phenolic and aniline self-immolative linkers, a folic acid targeting ligand and two MMAE molecules centred around a chemical amplifier, enabling a release of two drug molecules

via a single internalisation and activation pathway. The warhead release mechanism was studied by incubating folate-conjugate **3** with β -galactosidase at pH 7.2 and at 37 °C. The cleavage mechanism begins with the enzyme-mediated hydrolysis of SMDC **3**'s glycosidic bond, generating a phenol intermediate **4** which undergoes 1,6-elimination and a successive decarboxylation to concomitantly yield quinone **5** and an aniline intermediate **6**. Ensuing 1,6- and 1,4- elimination processes result in the release of two MMAE molecules (Fig. 7).

The breakdown of conjugate **3** was monitored by HPLC and at $t = 35$ min, peaks corresponding to free MMAE, regioisomers of **5** and aniline intermediate **8** were detected. Trace amounts of

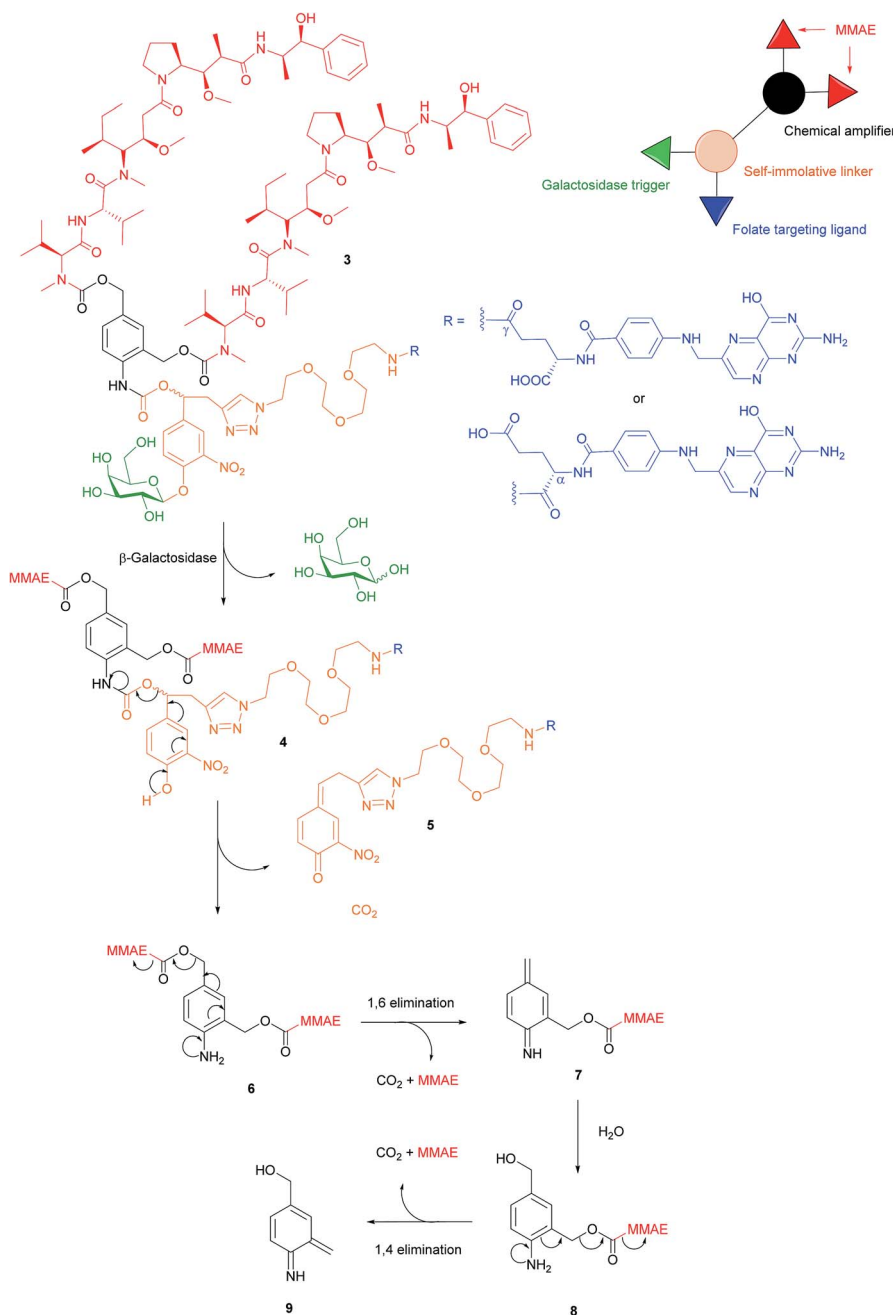


Fig. 7 Enzyme-catalysed double drug release mechanism of β -galactosidase-responsive folate-MMAE conjugate **3**.⁴²



phenol **4** and dimer **6** were also present in the mixture 35 minutes after the addition of β -galactosidase. Notwithstanding the complexity of the double drug liberation mechanism, MMAE release reached completion at $t = 2$ h, indicating that the process occurs reasonably quickly.⁴² The function of β -galactosidase in this release mechanism, which is found in the cell lysosome, was further confirmed by observing the absence of reaction when the same enzyme was incubated with non-cleavable glucuronide analogues of folate-conjugate **3**.

A 2,3-bis-(2-methoxy-4-nitro-5-sulfophenyl)-2H-tetrazolium-5-carboxanilide salt (XTT) cytotoxic assay was then performed using conjugate **3** and its monomeric analogue on FR α -positive cancer cell lines HeLa, SKOV-3 and A2780, but whose FR α expression is significantly lower levels than that of KB cells.⁴² Cell viability of all HeLa, SKOV-3 and A2780 was radically diminished by conjugate **3** (with IC₅₀ ranging from 9.62 to 64.51 nM), whereas a *ca.* 2- to 4-fold reduction in cytotoxicity was observed for the same cell lines when incubated with the monomer. This would appear to indicate that the higher levels of MMAE released from the dimer **3** result in increased tumour toxicity. The cytotoxic amplification of the dimer and monomer was then investigated by incubating A2780 cells with equal concentrations of both conjugates. If the β -galactosidase hydrolysed the same amounts of dimer **3** and the monomer, a 2-fold MMAE release would be expected from the former relative to the latter. However, analysis by HPLC/HRMS showed a 4-fold increase of MMAE drug liberation from **3** as compared to the monomer and a proposed hypothesis for this phenomenon is that β -galactosidase released from dead cancer cells can participate in the extracellular activation of non-internalised MMAE conjugates.⁴² This rationale is further supported by studies conducted by Antunes *et al.*, among others, showing that glucuronide prodrugs can be activated by β -glucuronidase liberated from dead tumour tissues.^{43,44}

A further example of an FA-SMDC that does not bear a disulfide linker and is cleaved by an enzyme is a folate-camptothecin conjugate degraded by the cathepsin B enzyme.⁴⁵ In addition to FA-SMDCs that are cleaved by enzymes already present in the tumour milieu, folate-enzyme conjugates have also been developed to deliver an enzyme to the folate receptor of the tumour cell prior to the administration of a prodrug that is converted to the active form by this enzyme. An example of this therapy utilises penicillin-V amidase and a doxorubicin prodrug.⁴⁶ Consequently, this work paves the way for the development of a new generation of enzyme-responsive FA-SMDCs that could broaden the scope of selectively targeting FR α -expressing tumours. To the best of our knowledge, no FA-SMDCs have been developed to specifically target the FR β isoform, but as both FR α and FR β have a similar affinity for folate and internalise *via* RME, there is scope for adapting the technologies applicable to FR α to the FR β isoform.

2.4 Other linker platforms

2.4.1 Boron-nitrogen linker. In addition to the commonly employed disulfide and carbon-based linkers for drug release inside the cell, the covalent attachment of boronic acids to

Schiff base ligands to yield boronate complexes can also be utilised as a platform to selectively deliver cytotoxic drugs to cancer cells. Gois *et al.* designed such a complex (**10**), which comprises the cytotoxic drug bortezomib, PEG chains and folate targeting units (Fig. 8).⁴⁷

A bivalent folate targeting moiety was chosen to mimic the bivalent Fab regions present on immunoglobulin Gs (IgGs) that give rise to high affinity and specificity of antibodies for particular antigen epitopes.⁴⁷ Complex **10** exhibited an IC₅₀ value of 62 nM against MDA-MB-231 cancer cells, lower than that of free bortezomib, but superior selectivity for these FR α -overexpressing cells as compared to the free drug. As GluSH is present in millimolar concentrations in the cell, Gois *et al.* investigated the GluSH-mediated cleavage mechanism by synthesising complex **11**, a less sterically hindered analogue of complex **10**. The mechanism of drug release, as determined by HPLC, is thought to proceed *via* GluSH addition to the iminium carbon of the complex followed by opening of the five-membered ring and subsequent hydrolysis to promote release of drug **15** (Fig. 9).

2.4.2 Light-triggered drug release. Methods to induce cytotoxicity with light, such as photodynamic therapy (PDT) have also attracted considerable interest for applications in cancer therapy. This technology involves light-mediated activation of

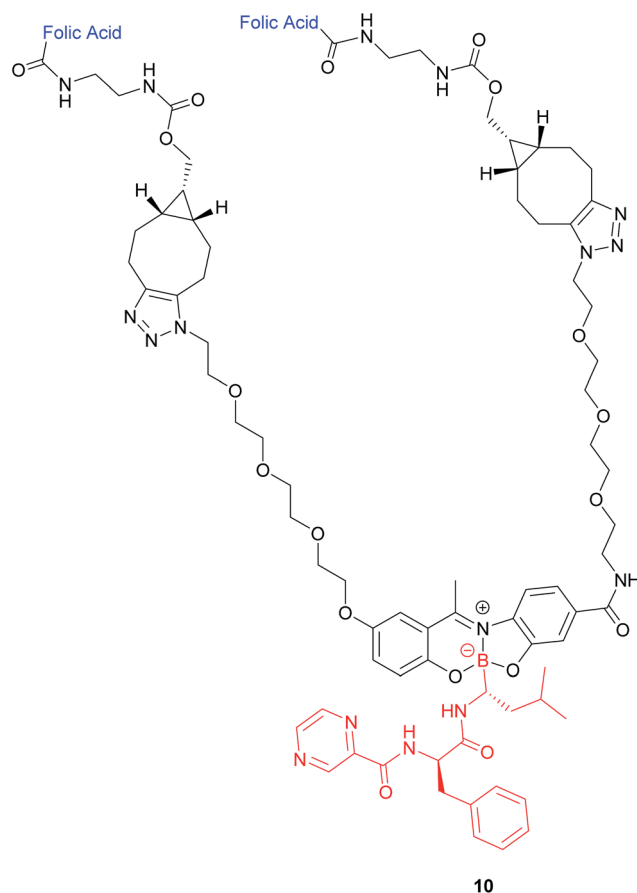


Fig. 8 Structure of the boron complex **10** developed by Gois *et al.* consisting of (i) a folic acid targeting moiety (blue), (ii) PEG chains and (iii) the cytotoxic agent bortezomib (red).⁴⁷





Fig. 9 Proposed mechanism for GluSH-mediated release of bortezomib (15) from complex 11.⁴⁷

a photosensitiser in the presence of oxygen and the subsequent generation of reactive oxygen species that neutralise the cells that have been exposed to the photosensitiser.⁴⁸ Moreover, the advantages of light-based techniques include non-invasive activation and added selectivity from the ease of this medium's spatial and temporal manipulation.⁴⁹ An example of a promising class of photosensitisers are boron dipyrromethene (BODIPY) derivatives that possess attractive optical and photophysical properties as well as displaying high stability in aqueous media.⁵⁰ Ke *et al.* have developed two diiododistyryl folate-conjugated BODIPY-based photosensitisers (**16a** and **16b**) with differing glycol linker lengths (Fig. 10).

The *in vitro* photosensitising ability of **16a** and **16b** was investigated by incubation both with KB human nasopharyngeal carcinoma cells, which have high expression of FR α and with

MCF-7 human breast adenocarcinoma cells, which have low expression of FR α .⁵⁰ No cytotoxic activity was detected for either in the absence of light, whereas activity was observed upon the illumination with IR light. Conjugate **16a**, with no triethylene glycol linker, displayed cytotoxic activity 3-fold higher (IC₅₀ of 60 nM) than that of **16b** (IC₅₀ of 180 nM).

The difference in cytotoxicity can be explained by the observation that **16b** aggregates more in RPMI culture medium than **16a**, probably due to the triethylene glycol linker of the former inducing dipole-dipole interactions in the neighbouring oligoethylene glycol chains.⁵⁰ Thus, conjugate **16a** with the shorter linker is an attractive candidate for use as a photosensitiser against cancer cells in PDT.

Other examples of FA-SMDCs whose cell-killing action is triggered by light include combretastatin A-4 prodrugs activated



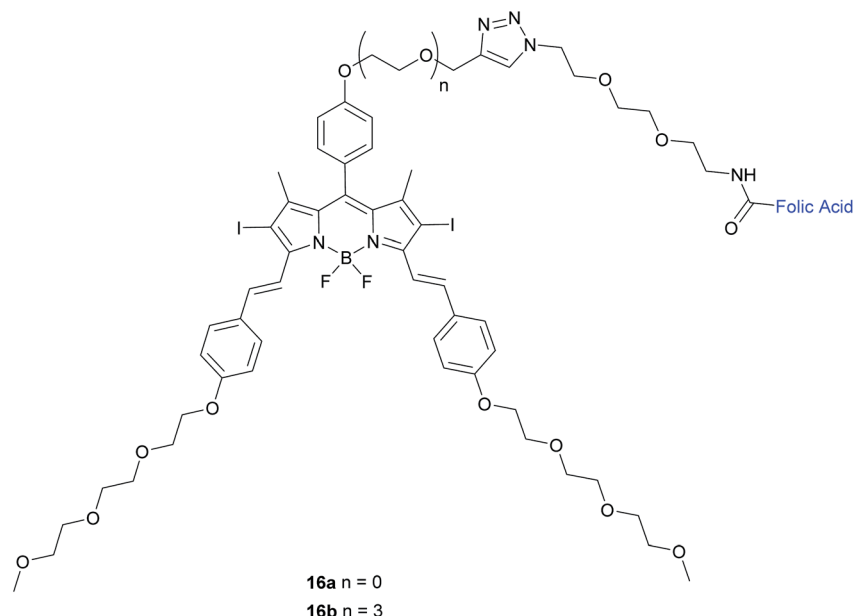


Fig. 10 Chemical structure of folate-BODIPY conjugates **16a** and **16b**.⁵⁰

by visible/near IR,⁵¹ a folate–doxorubicin conjugate that liberates doxorubicin when irradiated with UV-light⁴⁹ and a folate–chlorin conjugate where the photosensitising ability of the chlorin unit is activated upon irradiation of red light.⁴⁸

As described above, FA-SMDCs represent a varied class of conjugates for targeted drug delivery. Whilst a large number of these platforms have been targeted to FR α overexpression applications, these platforms can readily be applied to FR β overexpression scenarios (an emerging field) since folic acid binds to both these receptors. SMDCs are not the only group of treatments available for FR positive tumours, and the development of anti-folate antibodies that preferentially target FR α or FR β with specificity and selectivity (as they do not possess an indiscriminate folic acid targeting moiety) represents an alternative strategy.¹⁰

3 FR-targeted monoclonal antibodies

3.1 Farletuzumab (FR α targeted)

Farletuzumab (MORab003) is an example of a fully humanised anti-FR α mAb, weighing 145 kDa and produced in Chinese hamster ovary cells. This antibody has the added advantages of neither preventing the binding of folic acid to the FR α nor blocking FR α -mediated FA-transport into the cell.¹⁰ The therapeutic potential of farletuzumab has been shown in both *in vitro* and *in vivo* studies. Preclinical *in vitro* studies show that upon binding to FR α on tumour cells, farletuzumab promotes tumour cell lysis by various modes of action, including complement-dependent cytotoxicity (CDC) and antibody-dependent cell-mediated cytotoxicity (ADCC). Other mechanisms by which this anti-FR α mAb suppresses cancer cell proliferation activity include sustained autophagy and disruption of the FR α and lyn kinase interaction which curtails intracellular growth signalling.¹⁰

Farletuzumab in combination with carboplatin/taxane, followed by single-agent farletuzumab maintenance was employed in a phase II study performed in patients with platinum-sensitive recurrent ovarian cancer.¹⁰ Single-agent farletuzumab was well-tolerated by the patients, and no additional toxicity was observed when the antibody was administered in combination with chemotherapy. The study also demonstrated an overall improved response rate compared to historical, platinum-based combination chemotherapy regimens.^{52–55} Unfortunately, phase III trials in both platinum-resistant and platinum-sensitive ovarian cancer failed to replicate the promising results of the *in vitro* and *in vivo* studies. Results from a large phase III study in patients with platinum-sensitive recurrent ovarian cancer evaluated farletuzumab in combination with carboplatin and taxane, and was compared with carboplatin/taxane alone. Disappointingly, this trial did not meet the primary end point of improving progression-free survival (PFS).^{10,34} It is hoped that better patient selection will improve therapy for the future.

3.2 IMGN853 (FR α targeted)

In addition to stand-alone therapeutic antibodies such as the aforementioned farletuzumab, antibody–drug conjugates (ADCs), where a cytotoxic agent is covalently linked to an antibody, are now being employed as vehicles for the selective delivery of drugs to tumours. This technology combines the exquisite binding selectivity of antibodies and the potent toxicity of a chemical warhead, whose cell-killing potential is distinct from antibody-dependent cytotoxicity, whilst also minimising off-target toxicity.⁵⁶ This consequently enables the use of drugs that would otherwise be too toxic to be employed in conventional chemotherapeutic regimens. Moreover, the attachment of the cytotoxic agent magnifies the antibody's activity and has the potential to circumvent the rarely curative



action of unconjugated antibodies.⁵⁷ As opposed to the short circulation half-life typical of SMDCs, antibodies' large size confers a substantially longer half-life to the ADCs in the bloodstream, which in turn augments the proportion of the administered dose reaching and penetrating the tumour.

An example of such a FR α -targeting ADC is IMGN853 (17), and it comprises three elements: (1) an anti-FR α antibody that targets the FR α -expressing cancer cells, (2) DM4, an antimetabolic agent that inhibits tubulin polymerisation and microtubule assembly and (3) a disulfide-based linker that connects the drug to the antibody (Fig. 11).³⁴ As with the FA-SMDCs, IMGN853 binds to FR α , is internalised *via* RME, and ensuing enzymatic degradation of the antibody and linker releases the DM4 drug, which induces cell-cycle arrest and death by disrupting microtubule function. IMGN853 has demonstrated anti-tumour activity^{58,59} and is currently being assessed in phase II trials as a single agent and in combination regimens for patients with FR α -positive platinum-resistant ovarian cancer. This ADC represents a first generation construct of its type and there is plenty of scope to refine its chemistry should the clinical trials be unsuccessful.

3.3 Antibody m909 (FR β targeted)

In light of the observation that activated macrophages play an important role in the development and evolution of autoimmune diseases and certain cancers, Feng *et al.* developed an anti-human FR β -selective IgG1 antibody (m909) and demonstrated its ability to induce lysis in FR β -expressing cells.¹⁹

The ability of m909 to bind to cells expressing FR β was investigated by incubation of the antibody with CHO-hFR β (FR β positive) and CHO-K1 (FR β negative) cell lines. As expected, flow cytometry demonstrated binding of m909 to CHO-hFR β cells, but none to CHO-K1 cells. This selectivity was further assessed by incubating m909 with KB nasopharyngeal cells, known to express a large amount of FR α on their surface.⁶⁰ No binding of m909 was observed on the KB cells, confirming the selectivity of this antibody for FR β .

Subsequently, antibody m909's potential to induce ADCC was investigated by incubating m909, as well as a control isotype IgG, with three cell lines which themselves had been pre-incubated with NK cells: CHO-hFR β (FR β positive), preB L1.2 (FR β positive, but lower than CHO-hFR β) and CHO-K1 (FR β negative). Gratifyingly, cell lysis induced by m909 was observed

at higher levels in CHO-hFR β than in preB L1.2 and no cytotoxicity was detected in the FR β negative CHO-K1 cells. The control IgG exerted no lysis in any of the cell lines, even at a concentration of 200 nM.¹⁹ These results demonstrate that m909 is able to bind to FR β -expressing cells and mediate ADCC by recruiting NK cells and this antibody has been used to target FR β positive acute myeloid leukaemia (AML) blasts with chimeric antigen receptor T cells.

3.4 Chimeric antigen receptor T cell therapy for AML (FR β targeted)

Owing to the successful development of anti-FR β antibody m909, Low *et al.* incorporated it in the development of chimeric antigen receptor (CAR) T cell-therapy to treat AML, as approximately 70% of all AML tumours upregulate FR β .^{11,61,62} This treatment consists of attaching the single-chain variable fragment (scFv) of a monoclonal antibody (in this case m909) to T cell receptor signalling domains and in doing so, a patient's own T cells can bind to antigen-positive tumours with antibody-like affinity.¹¹

To demonstrate m909 CAR T cells' reactivity, they were incubated overnight with genetically engineered ovarian cancer C30-FR β cells. Subsequent analysis of the assays showed release of proinflammatory cytokines IFN- γ , IL-2, tumour necrosis factor α and inflammatory protein 1 α .¹¹ ELISA analysis was then used to demonstrate the same reactivity by overnight incubation of m909 CAR T cells and control CD19-28Z T cells with FR β positive AML lines, with the former incubation producing a greater secretion of IFN- γ than the latter. Low *et al.* also upregulated the expression of FR β on AML cell lines by treatment of all-*trans* retinoic acid (ATRA) and found that IFN- γ secreted by cells pre-treated with ATRA and then incubated with m909 CAR T cells was significantly greater than those without ATRA treatment.¹¹

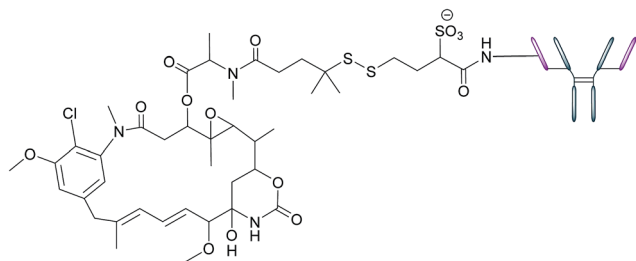
These *in vitro* results were then appraised *in vivo* using THPI (high FR β expression) AML cells inoculated in mice, with the administration of m909 CAR T cells leading to tumour regression. *In vivo* proliferation of these T cells was demonstrated by treating mice's peripheral blood with human CD3⁺ cells and subsequent administration of m909 CAR T cells. Analysis after 4 weeks of treatment showed significantly higher levels of peripheral blood T cells as compared to the controls, further demonstrating the selective binding of m909 CAR T cells to FR β .¹¹ Thus, this innovative development of anti-FR β m909 CAR T cells therapy presents a promising platform for the treatment of autoimmune diseases and AML.

Notwithstanding the possible future success of FR α and FR β -targeting antibodies, the field is still rather narrow. In contrast, the arena of FR-selective nanotechnology is more expansive and versatile.

4 Nanotechnology

4.1 Nanoemulsions (FR α targeted)

As highlighted above, conventional chemotherapy is limited by a lack of selectivity, and the unwanted side effects caused by the



17

Fig. 11 Structure of IMGN853 (17). The anti-FR α antibody is conjugated to the DM4 drug *via* a self-immolative disulfide linker.



non-specific cellular uptake of platinum-based regimens can be especially problematic. Nonetheless, due to its highly responsive nature, platinum-based therapy is still used as a leading chemotherapeutic agent in almost all stages of ovarian cancer. However, the case for further support of this choice of therapy is waning. For instance, the high frequency of Pt-based treatment cycles often result in acquired drug resistance which can occur *via* the decreased cellular uptake of Pt, which limits the formation of cytotoxic Pt–DNA adducts. Additionally, intracellular GluSH mediates the detoxification of Pt and leads to the inactivation of Pt by the formation of cisplatin–thiol conjugates, thereby preventing cell death occurring after the formation of the lethal Pt–DNA adducts.⁶³

In light of this, there is a critical need to modify the Pt therapeutic options currently available. To this effect, Patel *et al.* have reported the synthesis of NMI-350 Pt-theranostic nanoemulsions (NEs). The NMI-350 family is based on naturally occurring polyunsaturated fatty acid (PUFA) rich omega-3 and -6 fatty acid oils and gadolinium (Gd) labelled multi-compartmental NEs. Their oily core can encapsulate the cytotoxic and hydrophobic difattyacid platins and C₆-ceramide, and the NE surface can be employed for the attachment of imaging agents and folate ligands for targeting (Fig. 12).²¹

Through the aforementioned architecture, these NEs allow the controlled delivery of combined chemotherapy and additionally lengthen the blood circulation half-life of Pt to maximise uptake of nanodrug conjugates in malignant cells over a prolonged period of time. Moreover, the synthesis of the difattyacid platinum construct has been greatly improved: Patel *et al.* have developed a synthesis which takes 24 h, as opposed to previously reported procedures requiring 21 days.⁶⁴

Difattyacid platins of different chain lengths were synthesised using this more efficient method and folate was attached to the NE surface *via* a DSPE-PEG₃₄₀₀ spacer (Fig. 13). The fully functionalised NEs displayed a particle size in the range 120–150 nm.

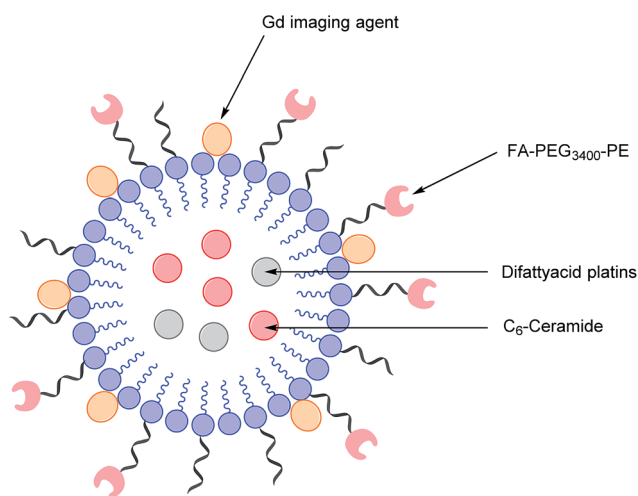


Fig. 12 Schematic representation of a NMI-350 nanoemulsion. Difattyacid platins and C₆-ceramide are encapsulated in the lipid core and lapidated gadolinium and folate are attached to the surface.²¹

FR α -binding efficiency of the NEs was then tested on two FR α -rich cell lines, KB-WT (Pt-sensitive) and KBCR-1000 (Pt-resistant) cell lines and analysed by flow cytometry. Both lines were treated with non-targeted rhodamine labelled NEs (NT-Rh-NE) and FA-targeted rhodamine labelled NEs (FA-Rh-NE), with the latter being functionalised with 100, 300, 1200 and 3600 FA molecules. As expected, cellular uptake in both the lines increased with higher levels of FA conjugation.²¹

The FA-Rh-NE labelled with 300 FA molecules was then selected for a cytotoxic assay due to being the most stable and cost effective relative to the other FA-Rh-NEs. This FA-Rh-NE was compared to cisplatin in a cytotoxic assay using the same Pt-sensitive and Pt-resistant cell lines, and this NE produced a *ca.* 30-fold increase in potency as compared to unconjugated cisplatin. This heightened cytotoxicity has the potential to reverse Pt-resistance and can be ascribed to the synergistic effect of the Pt and the exogenously added C₆-ceramide. After binding to FR α and ensuing internalisation *via* RME, dissociation of the NE is promoted by the acidic environment of the endosome, permitting the diffusion of the free Pt and C₆-ceramide across the endosome into the intracellular milieu, where they can exert their cytotoxic activity on chromosomal and mitochondrial DNA. Intracellular depletion of C₆-ceramide constitutes a resistance mechanism that shifts the equilibrium away from apoptosis in tumour cells.²¹ The addition of the ceramide to NEs serves to combat this resistance mechanism by shifting said equilibrium back towards apoptosis and encapsulation of the ceramide inside the NE shields it from metabolic degradation and inactivation.

The effect of the difattyacid cisplatin aliphatic linker length (C14, C16 and C18) was also evaluated and while the linkers had no effect on the stability of the NEs, the shortest chain **18a** produced the most potent cytotoxic activity. This observation can be rationalised by considering the shortest chain to be the best leaving group during Pt–O bond cleavage, resulting in quicker liberation of reactive Pt which can go then go on to form adducts with the tumour cell's DNA.²¹

4.2 Nanotubes (FR α targeted)

Another promising class of nanostructures that is generating increasing interest is that of coordination complex nano-assemblies.⁶⁵ These versatile structures possess desirable characteristics which can be difficult to achieve in other materials. For example, they have the potential to attain various geometries, and influencing tunable binding strength and directionality by choice of ligand.⁶⁵ Wang *et al.* have developed the first example of Ni–folate biomolecule-based coordination complex nanotubes (BMB-CCNTs) of an inner diameter of 5–8 nm and which incorporate FA as a targeting ligand, hydrazine as a linker, Ni as a connector and cisplatin as the cytotoxic agent.⁶⁵ These nanotubes' sufficiently large cavity permits a high drug loading which overcomes the small deliverable payload dose associated with other folate conjugates. Moreover, these nanotubes evade the undesirable accumulation in the kidneys typical of smaller folate–drug conjugates.⁶⁵



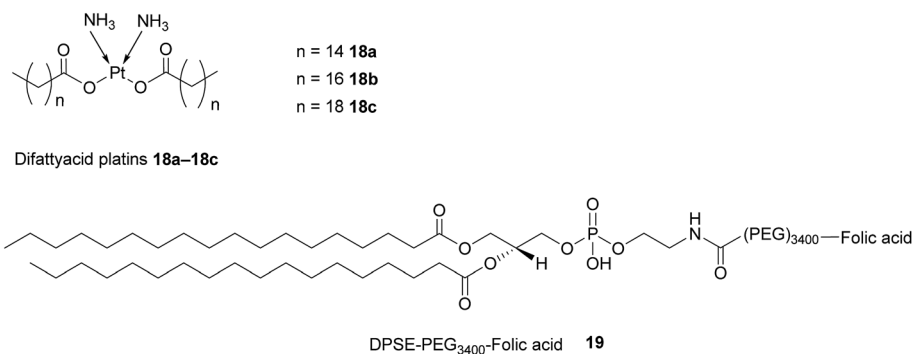


Fig. 13 Structure of the difattyacid cisplatins **18a–18c** and the DPSE-PEG₃₄₀₀-FA spacer **19**.



Fig. 14 Nanotube formation from nanosheets.⁶⁵

The initial stage of nanotube synthesis comprises the formation of a tape-like structure as the pteric acid unit of FA can form hydrogen bonds with the pteric acid moiety of other FA molecules. The glutamic acid portion of FA can then coordinate to Ni^{2+} without compromising the intermolecular hydrogen bonds and hydrazine serves as a bridging ligand between two Ni atoms, resulting in the formation of a nanosheet. The high temperature of this reaction aggravates the relative intermolecular movement of the nanosheets and thus stimulates curling in order to minimise the free surface energy. The high temperatures also promote nanotube formation by the breaking of partial initial bonds and the formation of new ones, with the hydrazine acting as a molecular string, tying the nanosheets into nanotubes (Fig. 14).⁶⁵

A cytotoxic assay was conducted to evaluate and compare the potency of cisplatin-loaded BMB-CCNTs (CDDP-CCNTs), blank BMB-CCNTs, CDDP and free folic acid in three different cell lines: HeLa (high FR α), and low FR α expressing A549 and HELF. As expected the CDDP was toxic to all three cell types. However, despite being less toxic than CDDP, the folate-nanotubes drastically reduced HeLa cell viability far more than HELF and A549. The nanotubes were labelled with fluorescein isothiocyanate (FITC) and their internalisation into HeLa (high FR α) and HELF (low FR α) cells was tracked by confocal laser scanning microscopy. After 24 h of incubation, a strong green fluorescence was observed in the HeLa cells, whereas that of the HELF cells was rather weak, indicating FR α -dependent uptake into the cells. The binding selectivity of the CCNTs to both cell lines was studied by flow cytometry, demonstrating a 34.1% uptake in HELF cells as opposed to 98.6% in HeLa cells. Together with the results of the fluorescence studies, Bio-TEM imaging showed that following uptake into cells *via* RME, the acidic endosomal environment triggers corrosion of the nanotubes and degradation into nanopieces, allowing drug release into the cytosol, giving rise to the inhomogeneous green fluorescence surrounding the nucleus.⁶⁵

To further corroborate the acid-dependent corrosion of the nanotube, the BMB-CCNTs were subjected to endosome-like conditions by soaking in PBS at pH 6.5 for 6 h. As anticipated, the open ends of the nanotubes began to disintegrate into nanopieces. Conversely, the CCNTs' original tubular architecture is preserved when soaked in PBS at pH 7.4, making it an effective targeted delivery system which is stable in blood and only disintegrates inside the cancerous tissue. This original nanotube complex provides insights for creating a novel multi-functional nanomedicine system, which may act as a target seeker and could concomitantly kill multiple malignant cells with a superior efficiency and fewer off-target side effects.⁶⁵

4.3 BAL-targeted liposomal doxorubicin delivery (FR β targeted)

Though the examples of targeted nanotherapy for FR α are more extensive, there are nevertheless nanotechnologies that utilise FR β as a delivery marker for targeted cytotoxics. One such example developed by Lu *et al.* is an FR β -targeted liposomal doxorubicin for treating biphenotypic acute leukaemia (BAL).⁶⁶

The group prepared several FR β -targeted liposomes (f-L-DOX) and varied the mole percentage of FR β -targeted distearoyl phosphatidylethanolamine (f-PEG-DSPE) in order to assess which lipid analogue constitutes the optimal formulation for therapeutic activity.⁶⁶

Cellular uptake of these liposomes was subsequently studied using fluorometry in order to evaluate whether the lipid mole percentage of f-PEG-DSPE would affect internalisation. As expected, binding and *in vitro* cytotoxicity assays demonstrated that the liposome formulation containing 0.5 mol% f-PEG-DSPE was more efficient in the uptake and more cytotoxic in BAL MV4-11 cells than those of 0.2 mol% and 0 mol%. The optimal mol% was determined to be 0.5 and further augmentation of f-PEG-DSPE (up to 2 mol%) displayed no additional improvement in cellular uptake cytotoxicity.⁶⁶

As previously mentioned, treatment with ATRA can upregulate FR β expression. In order to evaluate the impact of FR β upregulation on the cellular uptake and cytotoxicity of these formulations, cells were pre-treated with 1 μM ATRA. The results showed that pre-treating the cells for 5 days with ATRA caused a significant increase in the uptake rate of f-L-DOX by MV4-11 cells. Regarding the cytotoxicity studies, MTT assays showed that f-L-DOX exerted a 4.8-fold increase in cytotoxic activity than L-DOX to MV4-11 cells without pre-treatment with ATRA and 8.6 times greater lytic activity with ATRA pre-treatment. In contrast, the cytotoxic activity of free DOX and L-DOX in MV4-11 cells was not affected by ATRA pre-treatment, suggesting that the impact of ATRA pre-treatment on f-L-DOX-mediated cell death was directly due to FR β overexpression.⁶⁶ This nanotherapy enables efficient delivery of cytotoxic agents to tumours overexpressing FR β and could have potential future therapeutic applications in the clinic.

The FRs' upregulation on many different cancer types can be further exploited for diagnostic and imaging purposes.

5 Imaging: ^{99m}Tc-etarfolatide (FR α targeted)

Appraisal of FR α expression can be a useful diagnostic tool, allowing the FR α status to be monitored throughout the duration of treatment, with several avenues having been explored for FR α detection. However, despite the high specificity and sensitivity of these methods, their clinical use usually requires invasive tissue biopsies, which are typically taken from a single lesion.⁶⁷ Furthermore, the heterogeneous nature of FR α expression on tumours and the changing characteristics of tumours with time makes it difficult to construct an accurate representation of a patient's FR α status, thus generating an incomplete picture. Whole-body imaging that utilises folate radioconjugates can overcome this limitation by providing realtime and non-invasive FR α appraisal for multiple lesions at several time points.^{68,69}

A number of FR α -targeting imaging agents have been evaluated for tumour imaging.⁷⁰⁻⁷⁷ Etarfolatide (EC20) is one such example and is a folate-targeted radioimaging agent composed of ^{99m}technetium (Tc) complexed to folic acid *via* a short



non-cleavable peptide linker (Fig. 15).¹⁰ As opposed to the previously discussed cleavable linkers that are indispensable for drug release in the tumour milieu, EC20's linker is non-degradable as the release of the ^{99m}Tc is not a requirement for radiofolate imaging.

^{99m}Tc is a frequently employed radiographic tracer, possessing a half-life of 6 h and whose principle form of radioactive decay is gamma emission.¹⁰ Moreover, ^{99m}Tc-etarfolatide displays a strong binding affinity to FR α and tumours that overexpress FR α typically internalise a high proportion of the administered ^{99m}Tc-etarfolatide (~17% ID g⁻¹).⁷⁰ Added benefits of this probe conjugate include rapid accumulation at the tumour target site and subsequent swift clearance from the bloodstream *via* the kidneys. This in turn diminishes the non-specific tumour uptake of ^{99m}Tc-etarfolatide and permits the quick generation of images.¹⁰

^{99m}Tc-etarfolatide makes use of Tc's optimal single-photon emission computed tomography (SPECT) imaging characteristics, namely, a half-life of 6 h and a photon energy of 140 keV. Consequently, this probe conjugate has been subject to evaluation in numerous clinical trials, including those involving vintafolide, with ^{99m}Tc-etarfolatide as a companion imaging agent.^{69,71,78,79} Although no safety concerns have been established in this line of treatment, undesired adverse effects such as lower abdominal pain, nausea and vomiting, have all been identified as being ^{99m}Tc-etarfolatide-related, although these were only observed in <1% of patients.⁶⁷

While several phase II trials have demonstrated that ^{99m}Tc-etarfolatide imaging can be utilised to determine patients most likely to respond to vintafolide therapy,^{68,69} the imaging results and their interpretation can be influenced by physiological factors: principally the observation that ^{99m}Tc-etarfolatide is uptaken into the kidneys, bladder, spleen and somewhat into bone marrow. This may interfere with the interpretation of receptor expression in lesions close to these organs and for this reason, small quantities of folic acid are injected prior to ^{99m}Tc-etarfolatide administration in order to partially saturate the FR α s.⁶⁷ Another limitation of this probe conjugate stems from activated macrophages (that express FR β) also internalising ^{99m}Tc-etarfolatide, a phenomenon which can result in regions of inflammation or infection falsely appearing as FR α -positive tumour tissue.⁶⁷

Early studies on ^{99m}Tc-etarfolatide imaging were constrained by having to employ separate SPECT and computed tomography (CT) imaging, but contemporary SPECT/CT fusion imaging has greatly ameliorated spatial localisation and is able to

determine whether tumours are FR α -positive or FR α -negative. ^{99m}Tc-etarfolatide has proved to be valuable for the selection of patients likely to respond to treatments targeting the FR α . This probe conjugate has also shown promise for the staging and restaging of tumours, the assessment of disease prognosis and for the identification of patients who could benefit from intra-operative fluorescence FR α imaging to help reveal deep-seated tumours that can evade detection by intraoperative optical imaging due to limited signal penetration in human tissue.⁶⁷ ^{99m}Tc-etarfolatide may also have future applications for the prognosis of FR α -positive ovarian and lung cancer.^{80,81}

5.1 ⁶⁸Ga and ⁶⁴Cu radiofolates (FR α targeted)

Another approach for the imaging of FR α -positive tumours is the use of radiofolates for positron emission tomography (PET). This is a commonly used form of nuclear imaging and in a clinical context, is generally preferred to single-photon emission computed tomography (SPECT) owing to its superior resolution and sensitivity as well as its capability of quantifying the exact levels of accumulated radioactivity.^{82,83} Notwithstanding the widespread use of PET imaging, its application to small molecule-folate radioconjugates is limited and stems from their short circulation time in the bloodstream, resulting in high and undesirable renal accumulation of radioactivity. As human serum albumin is the major blood thiol, representing 80–90% of human plasma's thiol concentration,⁸⁴ Müller *et al.* sought to address the problem of renal radioactivity retention by functionalising folate conjugates with an albumin-binding moiety in the hope that this would confer a longer blood circulation time.⁸⁵ It was found that the novel albumin-binding folate radioconjugate (cm09) not only diminished renal radioactivity uptake, but demonstrated the added benefit of greater tumour accumulation.⁸⁵ Folate radioconjugate (cm09) and the next-generation analogue (cm10) have been employed for theranostic and therapeutic applications in preclinical studies, using radionuclides such as ¹⁷⁷Lu, ^{44/47}Sc and ^{149/161}Tb (Fig. 16).^{86–90}

Despite ⁶⁸Ga being the most frequently used radiometal in PET imaging, it possesses a half-life of only 68 min, rendering it non-ideal for chelation to the aforementioned albumin-binding radiofolates. In contrast, ⁶⁴Cu has a half-life of 12.7 h, permitting PET imaging at >24 h post-injection (p.i.) of the folate radioconjugate. Moreover, the positron energy of ⁶⁴Cu is almost as desirably low as that of ¹⁸F ($E_{\beta_{av}} = 250$ keV), which is currently the most commonly utilised clinical PET nuclide.⁸² As the established radiofolates cm09 and cm10 with a 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA) chelators, proved to be suboptimal for the coordination of ⁶⁸Ga and ⁶⁴Cu, Müller *et al.* synthesised a novel albumin-binding folate conjugate (rf42) with a 1,4,7-triazacyclononane, 1-glutaric acid-4,7-acetic acid (NODAGA)-chelator, permitting stable chelation to the radiometals ⁶⁸Ga and ⁶⁴Cu (Fig. 16).⁸²

The coordination of the radionuclides to rf42 proved to be very efficient, with both ⁶⁸Ga-rf42 and ⁶⁴Cu-rf42 displaying a radiochemical purity of >95% after 10 min at room temperature. To assess the protein binding of these conjugates, the

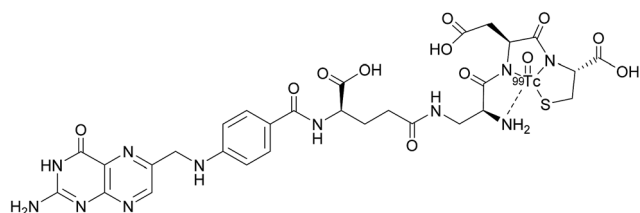


Fig. 15 Chemical structure of ^{99m}Tc-etarfolatide.





Fig. 16 Chemical structures of albumin-binding folate conjugates cm09, cm10 and rf42. They are each composed of three modules: (i) folic acid for FR α -expressing tumour targeting (blue), (ii) an albumin-binding moiety (pink), and (iii) the chelator (green) (DOTA for cm09 and cm10 and NODAGA for rf42) for coordination of ^{68}Ga or ^{64}Cu . The three functionalities are connected via a lysine linker in radiofolates cm10 and rf42, and by a triazole linker in the case of conjugate cm09.^{22,82,85}



prepared ^{68}Ga -rf42 and ^{64}Cu -rf42 were then incubated with human plasma and the filtered fraction of the plasma samples was found to contain negligible levels of radioactivity, suggesting that the majority of both radiofolates was bound to plasma proteins, with no significant difference in bound fraction between ^{68}Ga -rf42 ($98.0 \pm 0.2\%$) and ^{64}Cu -rf42 ($96.3 \pm 1.3\%$).⁸² A control experiment, consisting of incubation of ^{68}Ga -rf42 and ^{64}Cu -rf42 in PBS (pH = 7.4), found that the subsequent filtrate contained >95% of the loaded radioactivity. This confirmed that the low detected levels of radioactivity in the plasma samples incubated with ^{68}Ga -rf42 and ^{64}Cu -rf42 are due to the binding of these radiofolates to plasma proteins. Moreover, the protein-bound fraction of both ^{68}Ga -rf42 and ^{64}Cu -rf42 was observed to be higher than that of ^{68}Ga -cm10 and ^{64}Cu -cm10.⁸²

Cellular internalisation of ^{68}Ga -rf42 and ^{64}Cu -rf42 was performed in KB cells using a γ -counter, and shown to be high, with an internalised fraction of 30% and 55% respectively relative to the totally bound ^{68}Ga -rf42 and ^{64}Cu -rf42. Samples coincubated with excess folic acid demonstrated a dramatic reduction in radioactivity (<0.2% of total added radioactivity), indicating that the radiofolates are internalised *via* RME.⁸² *In vivo* biodistribution studies in mice showed rapid tumour uptake of ^{64}Cu -rf42 after 4 h ($14.52 \pm 0.99\%$ IA g^{-1}) and 50% of the maximum radioactivity was still present in the tumours 72 h p.i. of the radiofolate. Elevated tumour uptake was also observed for ^{68}Ga -rf42 after 4 h ($11.92 \pm 1.68\%$ IA g^{-1}), though it was lower than that of ^{64}Cu -rf42. Off-target radioactivity was detected in the kidneys and salivary glands. The albumin-binding moiety of the conjugates also gave rise to relatively high initial blood radioactivity which cleared comparatively slowly.

PET/CT imaging was then performed on mice bearing KB tumours at 2 h p.i. of ^{64}Cu -rf42 and ^{64}Cu -cm10. Both radiofolates showed radioactivity in tumours and the kidneys, but the tumour uptake of ^{64}Cu -cm10 was lower than that of ^{64}Cu -rf42. No difference in tumour uptake was observed for ^{68}Ga -rf42 and ^{68}Ga -cm10. PET/CT imaging also showed that mice co-injected with excess folic acid displayed marked reduction in tumour radioactivity.⁸² Furthermore, ^{64}Cu -rf42's longer half-life allowed PET/CT imaging to be performed on mice up to 72 h p.i. and found that although maximum tumour uptake at 24 h p.i. was similar for ^{64}Cu -rf42 and previously reported ^{177}Lu -cm09, blood retention of the former was twofold higher at 24 h p.i. and more than four-fold higher at 72 h p.i. than the latter. These results indicate the superiority of the NODAGA chelator relative to the DOTA, and the promising features of ^{64}Cu -rf42 that allow PET/CT imaging at longer time points than is possible with ^{68}Ga -rf42.⁸²

5.2 Fluorescent off-on nanoprobe (FR α targeted)

In addition to the previously mentioned PET, CT and SPECT, fluorescence imaging based on nanomaterials has recently emerged as an attractive prospect owing to the relative ease with which it is possible to multi-functionalise a nanoprobe. Fluorescence spectroscopy itself is an imaging technique that possesses exceptional qualities in regards to molecular and

cellular imaging, namely, high spatiotemporal resolution and sensitivity.^{91–93} However, as the majority of nanomaterial-based fluorescent systems are in a permanently-on state, they are commonly afflicted with drawbacks such as a low signal/background ratio and false positives stemming from non-specific uptake of the nanoprobe on the surface of non-target cells.⁹⁴

In an effort to address these issues, Feng *et al.* have developed an off-on nanoprobe, which only fluoresces once bound and internalised in the target cell.⁹¹ The structure of this nanoprobe consists of three modules: (i) rhodamine B (RB), a fluorochrome that minimises false positive signals by virtue of being impervious to biological environments, (ii) folic acid (FA) as a cell-targeting moiety and (iii) graphene oxide (GO) which is both an efficient quencher for organic fluorochromes and biologically compatible. The GO surface was coated with amino disulfide bonds, with the amino portion of the linker allowing conjugation to FA and RB (Fig. 17).^{95–100}

Ideally, the intact nanoprobe should display negligible to weak fluorescence due to the quenching action of the GO. Once the probe is internalised by the cell, intracellular GluSH should cleave the disulfide bond, thereby liberating the RB from the GO and generating fluorescence.⁹¹

The sensitivity of the nanoprobe was investigated by incubation in PBS (pH = 7.4) at 37 °C with two different concentrations of GluSH, 1 mM and 10 μM , the lowest intracellular and the highest extracellular concentration respectively. Incubation with 1 mM GluSH displayed a large increase in fluorescence, suggesting efficient cleavage of the nanoprobe's disulfide bonds by GluSH. On the contrary, incubation with 10 μM GluSH produced a rather weak fluorescent response with a low signal/background ratio. This significant discrepancy in fluorescence is an attractive feature for sensitive intracellular imaging.⁹¹ The cleavage action of thiols was confirmed through separate incubations of the nanoprobe with 1 mM cysteine and homocysteine, both experiments showing increases in fluorescence. The essential incorporation of the disulfide bond for fluorescence was also investigated by observing a very small increase in fluorescence when incubating GluSH with a control nanoprobe containing an alkane linker *in lieu* of a disulfide connector.

The hypothesis that this nanoprobe is internalised *via* RME was confirmed by incubating the nanoconstruct with FR α -overexpressing HeLa cells. Confocal laser scanning microscopy showed a steady increase of fluorescence which levels out after 3 h, suggesting that total degradation of the nanoprobe is completed after this time. Separate experiments where HeLa cells were pre-treated with (i) excess FA and with (ii) the nanoprobe lacking the FA moiety, showed weak fluorescence after subsequent incubation with the unmodified nanoprobe, corroborating internalisation *via* RME. This internalisation pathway and thus the selectivity of the nanoprobe were further validated by incubating the nanoprobe with HeLa (FR α -positive) and NIH-3T3 and MCF-7 (FR α -negative) cell lines. As expected, only HeLa displayed fluorescence ~ 16 and 7 times higher relative to NIH-3T3 and MCF-7 cells respectively.⁹¹

An additional challenge in the cancer diagnosis field is the distinction of tumours that are morphologically similar. As





Fig. 17 Fluorescence off-on response of nanoprobe in FR α -positive cell.⁹¹

HeLa and MCF-7 cells are akin in their morphologies, both lines were co-incubated for 12 h with the HeLa cells being subjected to an initial 30 min staining with a fluorescent dye in order to efficiently identify both cell types. Once the co-incubation was complete, the nanoprobe was added to the cell mixture and incubated for 3 h. Confocal laser scanning microscopy revealed a strong fluorescent signal in HeLa cells, whereas that detected from the MCF-7 cells was negligible, confirming this nanoprobe's ability to differentiate between FR α -positive and FR α -negative cell lines as well as those with similar morphologies, properties which are promising for cancer diagnostics.⁹¹ Future work on this nanoprobe could include incubation with human plasma to appraise the nature of its stability in blood-mimicking conditions.

The majority of these imaging technologies have been targeted/focused on FR α -expressing tumours. Although FR β imaging has been studied in the context of many inflammatory diseases,^{13,19,101–104} there does not appear to be as wide a range of imaging techniques specifically geared towards FR β -expressing cancers. Sun *et al.* have investigated the use of folate-FITC as a fluorescent imaging agent for the visualisation of FR β -expressing TAMs in head and neck squamous cell carcinoma.¹⁷ However, this technique appears to possess limited clinical value: TAMs penetrate tumours in a non-uniform manner and

therefore targeting of folate-FITC to cells in the tumour microenvironment leads to heterogeneous fluorescence. Although not detrimental for the detection of the tumour, this heterogeneity has limitations in the intraoperative imaging of the tumours. Additionally, due to the poor tissue penetration (a few millimetres)¹⁰⁵ of folate-FITC, applications for the use of this fluorophore are limited, but could be addressed by those that emit in the near infrared (NIR) region instead.

6 Conclusions

A variety of folate receptor targeting constructs have been developed against FR α -expressing tumours, each with their own advantages and limitations (Table 1). SMDCs, where the cytotoxic drug is linked to a folic acid tumour-targeting moiety, have perhaps attracted the most interest. Within this area, a number of technologies have been explored for drug release, including boron-thiol based, light-triggered and enzyme-cleavable strategies, as well as the classical disulfide linker model. In particular, the disulfide linker-bearing SMDC vintafolide has been investigated in clinical trials, making it as far as phase III. These strategies and platforms are also amenable to FR β -expressing tumours (as folic acid binds both FR α and FR β efficiently) and represent an exciting emerging field. Complementary and





Table 1 A table summarising the different types of FR α - and FR β -targeting platforms with focus on structure, applications, and their various advantages and limitations

Platforms	Structure	Applications	Advantages	Limitations
Small molecule conjugates (SMCs)	Imaging (applicable to FR α and FR β expressing tumours) SMDs (only exists for FR α expressing tumours currently)	Disease diagnosis, prognosis, monitoring Tumour-targeted therapy	Facile tumour penetration due to small size, convenient chemical synthesis, short circulation time, rapid systemic clearance	Undesirable accumulation of drug (SMDs) and often radioactivity (imaging) in liver and kidneys, small deliverable dose
Antibody-based	mAbs (tunable to FR α and FR β expressing tumours) ADCs (only exists for FR α targeting currently)		Exquisite selectivity, used alone or in combination therapy, vehicles for drug delivery to tumour site Allows the use of very potent cytotoxic drugs, minimal off-target toxicity	Expensive, potentially immunogenic, poor tumour penetration due to large size, long systemic circulation time of ADC may lead to potential off-site toxicity
Nano-based (tunable to FR α and FR β expressing tumours as folic acid is used for targeting)	Cytotoxic drug encapsulated within the nanoparticle core, external FA targeting moiety		High drug loading capacity, minimal off-target toxicity (in theory), encapsulation allows use of very potent cytotoxic drugs, biocompatible	Convolved synthesis, difficult purification and storage

orthogonal targeting constructs such as the FR α -specific mAb farletuzumab have also been investigated. Whilst this mAb experienced limited success as a single agent, the drug-conjugated antibody (IMGN853) is showing far more promise and is currently being investigated in ongoing phase II studies. Nanoparticle-based constructs such as nanoemulsions and nanotubes are also able to exert cytotoxic activity as well as enabling higher drug loadings than SMDs. In addition to cell death mediated by therapy-based FR α -targeted modules, the overexpression of FR α has also been exploited as a biomarker in fluorescence, PET and SPECT imaging; typically employing similar constructs to therapy approaches but with a non-cleavable linker. The folate-conjugated imaging agent that has progressed the furthest, ^{99m}Tc -etarfolatide, has been utilised as a companion agent in vintafolide's clinical trials and provides an accurate assessment of response to FR α -targeted therapies. These folic acid-based targeting methodologies can also be applied to FR β -expressing tumours. In the context of antibody-based technologies, the m909 CAR T cell therapy has led to *in vivo* AML tumour regression, while in the field of nanotherapeutics, the FR β -targeted liposome f-L-DOX has demonstrated the ability to induce cytotoxicity in FR β -positive BAL cancer cells. In terms of imaging, folate-FITC has been used to optically visualise cancers, but it is associated with various disadvantages, with NIR technology having the potential to supersede it in the future.

In summary, at present there exists a plethora of linker technologies available for efficient drug release and tumour-imaging for disease diagnosis, prognosis and monitoring in the field of FR targeting. Albeit with some exceptions along the way, considerable recent progress has been made. With all the lessons learned from both the successes and failures, and with several new sophisticated linkers at the fore, there is great promise in how the next-generation of FR-targeted constructs is shaping up.

Conflicts of interest

There are no conflicts to declare.

References

- J. D. Seitz, J. G. Vineberg, E. Herlihy, B. Park, E. Melief and I. Ojima, *Bioorg. Med. Chem.*, 2015, **23**, 2187–2194.
- N. Krall, F. Pretto, W. Decurtins, G. J. L. Bernardes, C. T. Supuran and D. Neri, *Angew. Chem., Int. Ed.*, 2014, **53**, 4231–4235.
- E. Valeur, L. Knerr, M. Ölwegård-Halvarsson and M. Lemurell, *Drug Discovery Today*, 2017, **22**, 841–847.
- S. A. Kularatne, K. Wang, H. K. Santhapuram and P. S. Low, *Mol. Pharm.*, 2009, **6**, 780–789.
- S. Bhuniya, S. Maiti, E. J. Kim, H. Lee, J. L. Sessler, K. S. Hong and J. S. Kim, *Angew. Chem., Int. Ed.*, 2014, **53**, 4469–4474.
- M. Srinivasarao, C. V. Galliford and P. S. Low, *Nat. Rev. Drug Discovery*, 2015, **14**, 203–219.

- 7 C. V. Galliford and P. S. Low, in *Drug Delivery: Principles and Applications*, ed. B. Wang, L. Hu and T. J. Siahaan, John Wiley & Sons, 2nd edn, 2016, pp. 451–474.
- 8 S. Quici, A. Casoni, F. Foschi, L. Armelao, G. Bottaro, R. Seraglia, C. Bolzati, N. Salvatorese, D. Carpanese and A. Rosato, *J. Med. Chem.*, 2015, **58**, 2003–2014.
- 9 T. Mironava, M. Simon, M. H. Rafailovich and B. Rigas, *Toxicol. In Vitro*, 2013, **27**, 882–889.
- 10 J. A. Ledermann, S. Canevari and T. Thigpen, *Ann. Oncol.*, 2015, **26**, 2034–2043.
- 11 R. C. Lynn, M. Poussin, A. Kalota, Y. Feng, P. S. Low, D. S. Dimitrov and D. J. Powell Jr, *Blood*, 2015, **125**, 3466–3477.
- 12 W. Xia, A. R. Hilgenbrink, E. L. Matteson, M. B. Lockwood, J.-X. Cheng and P. S. Low, *Blood*, 2009, **113**, 438–446.
- 13 C. M. Paulos, M. J. Turk, G. J. Breur and P. S. Low, *Adv. Drug Delivery Rev.*, 2004, **56**, 1205–1217.
- 14 C. Bailly, *Phytochem. Rev.*, 2014, **13**, 171–181.
- 15 A. Puig-Kröger, E. Sierra-Filardi, A. Dominguez-Soto, R. Samaniego, M. T. Corcuera, F. Gomez-Aguado, M. Ratnam, P. Sanchez-Mateos and A. L. Corbi, *Cancer Res.*, 2009, **69**, 9395–9403.
- 16 H. Kurahara, S. Takao, T. Kuwahata, T. Nagai, Q. Ding, K. Maeda, H. Shinchi, Y. Mataka, K. Maemura, T. Matsuyama and S. Natsugoe, *Ann. Surg. Oncol.*, 2012, **19**, 2264–2271.
- 17 J. Y. Sun, J. Shen, J. Thibodeaux, G. Huang, Y. Wang, J. Gao, P. S. Low, D. S. Dimitrov and B. D. Sumer, *Laryngoscope*, 2014, **124**, E312–E319.
- 18 J. Shen, K. S. Putt, D. W. Visscher, L. Murphy, C. Cohen, S. Singhal, G. Sandusky, Y. Feng, D. S. Dimitrov and P. S. Low, *Oncotarget*, 2015, **6**, 14700–14709.
- 19 Y. Feng, J. Shen, E. D. Streaker, M. Lockwood, Z. Zhu, P. S. Low and D. S. Dimitrov, *Arthritis Res. Ther.*, 2011, **13**, R59.
- 20 C. Chen, J. Ke, X. Edward Zhou, W. Yi, J. S. Brunzelle, J. Li, E. L. Yong, H. E. Xu and K. Melcher, *Nature*, 2013, **500**, 486–489.
- 21 N. R. Patel, A. Piroyan, A. H. Nack, C. A. Galati, M. McHugh, S. Orosz, A. W. Keeler, S. O'Neal, W. C. Zamboni, B. Davis and T. P. Coleman, *Mol. Pharm.*, 2016, **13**, 1996–2009.
- 22 K. Siwowska, S. Haller, F. Bortoli, M. Benešová, V. Groehn, P. Bernhardt, R. Schibli and C. Müller, *Mol. Pharm.*, 2017, **14**, 523–532.
- 23 L. Sun, Q. Wu, F. Peng, L. Liu and C. Gong, *Colloids Surf., B*, 2015, **135**, 56–72.
- 24 I. R. Vlahov and C. P. Leamon, *Bioconjugate Chem.*, 2012, **23**, 1357–1369.
- 25 T. Betzel, C. Müller, V. Groehn, A. Müller, J. Reber, C. R. Fischer, S. D. Krämer, R. Schibli and S. M. Ametamey, *Bioconjugate Chem.*, 2013, **24**, 205–214.
- 26 H. Birn, O. Spiegelstein, E. I. Christensen and R. H. Finnell, *J. Am. Soc. Nephrol.*, 2005, **16**, 608–615.
- 27 K. Simons and S. D. Fuller, *Annu. Rev. Cell Biol.*, 1985, **1**, 243–288.
- 28 S. Azzi, J. K. Hebda and J. Gavard, *Front Oncol.*, 2013, **3**, 1–14.
- 29 C. P. Leamon, I. R. Vlahov, J. A. Reddy, M. Vetzal, H. K. Santhapuram, F. You, A. Bloomfield, R. Dorton, M. Nelson, P. Kleindl, J. F. Vaughn and E. Westrick, *Bioconjugate Chem.*, 2014, **25**, 560–568.
- 30 C.-Y. Cheng, K.-L. Ou, W.-T. Huang, J.-K. Chen, J.-Y. Chang and C.-H. Yang, *ACS Appl. Mater. Interfaces*, 2013, **5**, 4389–4400.
- 31 J. G. Vineberg, E. S. Zuniga, A. Kamath, Y. J. Chen, J. D. Seitz and I. Ojima, *J. Med. Chem.*, 2014, **57**, 5777–5791.
- 32 R. J. Lee, S. Wang and P. S. Low, *Biochim. Biophys. Acta*, 1996, **1312**, 237–242.
- 33 I. R. Vlahov, H. K. Santhapuram, P. J. Kleindl, S. J. Howard, K. M. Stanford and C. P. Leamon, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 5093–5096.
- 34 I. Vergote and C. P. Leamon, *Ther. Adv. Med. Oncol.*, 2015, **7**, 206–218.
- 35 M. Bartouskova, B. Melichar and B. Mohelnikova-Duchonova, *Pteridines*, 2015, **26**, 1–12.
- 36 C. A. Ladino, R. V. Chari, L. A. Bourret, N. L. Kedersha and V. S. Goldmacher, *Int. J. Cancer*, 1997, **73**, 859–864.
- 37 J. A. Reddy, E. Westrick, I. R. Vlahov, S. J. Howard, H. K. Santhapuram and C. P. Leamon, *Cancer Chemother. Pharmacol.*, 2006, **58**, 229–236.
- 38 C. P. Leamon, J. A. Reddy, I. R. Vlahov, E. Westrick, A. Dawson, R. Dorton, M. Vetzal, H. K. Santhapuram and Y. Wang, *Mol. Pharm.*, 2007, **4**, 659–667.
- 39 C. P. Leamon, J. A. Reddy, M. Vetzal, R. Dorton, E. Westrick, N. Parker, Y. Wang and I. R. Vlahov, *Cancer Res.*, 2008, **68**, 9839–9844.
- 40 J. A. Reddy, R. Dorton, A. Dawson, M. Vetzal, N. Parker, J. S. Nicoson, E. Westrick, P. J. Klein, Y. Wang, I. R. Vlahov and C. P. Leamon, *Mol. Pharm.*, 2009, **6**, 1518–1525.
- 41 W. A. Henne, S. A. Kularatne, J. Hakenjos, J. D. Carron and K. L. Henne, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 5810–5813.
- 42 J. Alsarraf, E. Péraudeau, P. Poinot, I. Tranoy-Opalinski, J. Clarhaut, B. Renoux and S. Papot, *Chem. Commun.*, 2015, **51**, 15792–15795.
- 43 I. F. Antunes, H. J. Haisma, P. H. Elsinga, V. Di Galleonardo, A. Van Waard, A. T. M. Willemsen, R. A. Dierckx and E. F. J. de Vries, *Mol. Pharm.*, 2012, **9**, 3277–3285.
- 44 I. Tranoy-Opalinski, T. Legigan, R. Barat, J. Clarhaut, M. Thomas, B. Renoux and S. Papot, *Eur. J. Med. Chem.*, 2014, **74**, 302–313.
- 45 P. V. Paranjpe, S. Stein and P. J. Sinko, *Anticancer Drugs*, 2005, **16**, 763–775.
- 46 J. Y. Lu, D. A. Lowe, M. D. Kennedy and P. S. Low, *J. Drug Targeting*, 1999, **7**, 43–53.
- 47 F. M. F. Santos, A. I. Matos, A. E. Ventura, J. Gonçalves, L. F. Veiros, H. F. Florindo and P. M. P. Gois, *Angew. Chem., Int. Ed.*, 2017, **56**, 9346–9350.
- 48 P. X. Li, J. H. Mu, H. L. Xiao and D. H. Li, *Oncol. Rep.*, 2015, **33**, 125–132.
- 49 M. M. Dcona, J. E. Sheldon, D. Mitra and M. C. T. Hartman, *Bioorg. Med. Chem. Lett.*, 2017, **27**, 466–469.



- 50 M. R. Ke, S. L. Yeung, D. K. P. Ng, W. P. Fong and P. C. Lo, *J. Med. Chem.*, 2013, **56**, 8475–8483.
- 51 G. Nkepanang, M. Bio, P. Rajaputra, S. G. Awuah and Y. You, *Bioconjugate Chem.*, 2014, **25**, 2175–2188.
- 52 D. K. Armstrong, A. J. White, S. C. Weil, M. Phillips and R. Coleman, *Gynecol. Oncol.*, 2013, **129**, 452–458.
- 53 J. Pfisterer, M. Plante, I. Vergote, A. du Bois, H. Hirte, A. J. Lacave, U. Wagner, A. Stähle, G. Stuart, R. Kimmig, S. Olbricht, T. Le, J. Emerich, W. Kuhn, J. Bentley, C. Jackisch, H. J. Lück, J. Rochon, A. H. Zimmermann and E. Eisenhauer, *J. Clin. Oncol.*, 2006, **24**, 4699–4707.
- 54 G. Bolis, G. Scarfone, G. Giardina, A. Villa, G. Mangili, M. Melpignano, M. Presti, S. Tateo, M. Franchi and F. Parazzini, *Gynecol. Oncol.*, 2001, **81**, 3–9.
- 55 A. J. González-Martín, E. Calvo, I. Bover, M. J. Rubio, A. Arcusa, A. Casado, B. Ojeda, C. Balañá, E. Martínez, A. Herrero, B. Pardo, E. Adrover, J. Rifá, M. J. Godes, A. Moyano and A. Cervantes, *Ann. Oncol.*, 2005, **16**, 749–755.
- 56 V. Chudasama, A. Maruani and S. Caddick, *Nat. Chem.*, 2016, **8**, 114–119.
- 57 P. D. Senter, *Curr. Opin. Chem. Biol.*, 2009, **13**, 235–244.
- 58 C. Kurkjian, P. LoRusso, K. Sankhala, M. Birrer, M. Kirby, S. Ladd, S. Hawes, K. Running, J. O'Leary and K. Moore, *J. Clin. Oncol.*, 2013, **31**, 2573.
- 59 K. Moore, J. Ponte, P. LoRusso, M. Birrer, T. Bauer, H. Borghaei, D. O'Malley, R. Ruiz-Soto, R. Lutz and L. Malik, *J. Clin. Oncol.*, 2014, **32**, 5571.
- 60 R. M. Schultz, S. L. Andis, K. A. Shackelford, S. B. Gates, M. Ratnam, L. G. Mendelsohn, C. Shih and G. B. Grindey, *Oncol. Res.*, 1995, **7**, 97–102.
- 61 J. F. Ross, H. Wang, F. G. Behm, P. Mathew, M. Wu, R. Booth and M. Ratnam, *Cancer*, 1999, **85**, 348–357.
- 62 X. Q. Pan, X. Zheng, G. Shi, H. Wang, M. Ratnam and R. J. Lee, *Blood*, 2002, **100**, 594–602.
- 63 G. Tapia and I. Díaz-Padilla, in *Ovarian Cancer - A Clinical and Translational Update*, ed. I. Díaz-Padilla, InTech, Rijeka, 2013, pp. 205–214.
- 64 M. Maeda, N. A. Uchida and T. Sasaki, *Jpn. J. Cancer Res.*, 1986, **77**, 523–525.
- 65 Y. Wang, C. Zhang, H. Li, G. Zhu, S. Bao, S. Wei, L. Zheng, M. Ren and Z. Xu, *J. Mater. Chem. B*, 2015, **3**, 296–305.
- 66 Y. Lu, J. Wu, J. Wu, M. Gonit, X. Yang, A. Lee, G. Xiang, H. Li, S. Liu, G. Marcucci, M. Ratnam and R. J. Lee, *Mol. Pharm.*, 2007, **4**, 707–712.
- 67 A. H. Maurer, P. Elsinga, S. Fanti, B. Nguyen, W. J. G. Oyen and W. A. Weber, *J. Nucl. Med.*, 2014, **55**, 701–704.
- 68 R. W. Naumann, R. L. Coleman, R. A. Burger, E. A. Sausville, E. Kutarska, S. A. Ghamande, N. Y. Gabrail, S. E. Depasquale, E. Nowara, L. Gilbert, R. H. Gersh, M. G. Teneriello, W. A. Harb, P. A. Konstantinopoulos, R. T. Penson, J. T. Symanowski, C. D. Lovejoy, C. P. Leamon, D. E. Morgenstern and R. A. Messmann, *J. Clin. Oncol.*, 2013, **31**, 4400–4406.
- 69 R. T. Morris, R. N. Joyrich, R. W. Naumann, N. P. Shah, A. H. Maurer, H. W. Strauss, J. M. Uszler, J. T. Symanowski, P. R. Ellis and W. A. Harb, *Ann. Oncol.*, 2014, **25**, 852–858.
- 70 C. P. Leamon, M. A. Parker, I. R. Vlahov, L. C. Xu, J. A. Reddy, M. Vetzal and N. Douglas, *Bioconjugate Chem.*, 2002, **13**, 1200–1210.
- 71 R. E. Fisher, B. A. Siegel, S. L. Edell, N. M. Oyesiku, D. E. Morgenstern, R. A. Messmann and R. J. Amato, *J. Nucl. Med.*, 2008, **49**, 899–906.
- 72 C. J. Mathias, S. Wang, P. S. Low, D. J. Waters and M. A. Green, *Nucl. Med. Biol.*, 1999, **26**, 23–25.
- 73 S. Wang, J. Luo, D. A. Lantrip, D. J. Waters, C. J. Mathias, M. A. Green, P. L. Fuchs and P. S. Low, *Bioconjugate Chem.*, 1997, **8**, 673–679.
- 74 W. Guo, G. H. Hinkle and R. J. Lee, *J. Nucl. Med.*, 1999, **40**, 1563–1569.
- 75 B. A. Siegel, F. Dehdashti, D. G. Mutch, D. A. Podoloff, R. Wendt, G. P. Sutton, R. W. Burt, P. R. Ellis, C. J. Mathias, M. A. Green and D. M. Gershenson, *J. Nucl. Med.*, 2003, **44**, 700–707.
- 76 G. M. van Dam, G. Themelis, L. M. Crane, N. J. Harlaar, R. G. Pleijhuis, W. Kelder, A. Sarantopoulos, J. S. de Jong, H. J. Arts, A. G. van der Zee, J. Bart, P. S. Low and V. Ntziachristos, *Nat. Med.*, 2011, **17**, 1315–1319.
- 77 I. Aljammaz, B. Al-Otaibi, F. Al-Rumayan, S. Al-Yanbawi, S. Amer and S. M. Okarvi, *Nucl. Med. Biol.*, 2014, **41**, 457–463.
- 78 L. Teng, J. Xie, L. Teng and R. J. Lee, *Expert Opin. Drug Delivery*, 2012, **9**, 901–908.
- 79 S. Banerjee, M. R. Pillai and N. Ramamoorthy, *Semin. Nucl. Med.*, 2001, **31**, 260–277.
- 80 D. J. O'Shannessy, G. Yu, R. Smale, Y. S. Fu, S. Singhal, R. P. Thiel, E. B. Somers and A. Vachani, *Oncotarget*, 2012, **3**, 414–425.
- 81 Y. L. Chen, M. C. Chang, C. Y. Huang, Y. C. Chiang, H. W. Lin, C. A. Chen, C. Y. Hsieh and W. F. Cheng, *Mol. Oncol.*, 2012, **6**, 360–369.
- 82 R. Farkas, K. Siwowska, S. M. Ametamey, R. Schibli, N. P. van der Meulen and C. Müller, *Mol. Pharm.*, 2016, **13**, 1979–1987.
- 83 A. Rahmim and H. Zaidi, *Nucl. Med. Commun.*, 2008, **29**, 193–207.
- 84 F. Kratz, A. Warnecke, K. Scheuermann, C. Stockmar, J. Schwab, P. Lazar, P. Drückes, N. Esser, J. Drevs, D. Rognan, C. Bissantz, C. Hinderling, G. Folkers, I. Fichtner and C. Unger, *J. Med. Chem.*, 2002, **45**, 5523–5533.
- 85 C. Müller, H. Struthers, C. Winiger, K. Zhernosekov and R. Schibli, *J. Nucl. Med.*, 2013, **54**, 124–131.
- 86 C. Müller, K. Zhernosekov, U. Köster, K. Johnston, H. Dorrer, A. Hohn, N. T. van der Walt, A. Türlér and R. Schibli, *J. Nucl. Med.*, 2012, **53**, 1951–1959.
- 87 C. Müller, J. Reber, S. Haller, H. Dorrer, P. Bernhardt, K. Zhernosekov, A. Türlér and R. Schibli, *Eur. J. Nucl. Med. Mol. Imaging*, 2014, **41**, 476–485.
- 88 C. Müller, J. Reber, S. Haller, H. Dorrer, U. Köster, K. Johnston, K. Zhernosekov, A. Türlér and R. Schibli, *Pharmaceuticals*, 2014, **7**, 353–365.
- 89 C. Müller, M. Bunka, J. Reber, C. Fischer, K. Zhernosekov, A. Türlér and R. Schibli, *J. Nucl. Med.*, 2013, **54**, 2168–2174.



- 90 C. Müller, M. Bunka, S. Haller, U. Köster, V. Groehn, P. Bernhardt, N. P. van der Meulen, A. Türlér and R. Schibli, *J. Nucl. Med.*, 2014, **55**, 1658–1664.
- 91 D. Feng, Y. Song, W. Shi, X. Li and H. Ma, *Anal. Chem.*, 2013, **85**, 6530–6535.
- 92 M. Z. Zhang, R. N. Yu, J. Chen, Z. Y. Ma and Y. D. Zhao, *Nanotechnology*, 2012, **23**, 1–11.
- 93 X. Q. Chen, M. Sun and H. M. Ma, *Curr. Org. Chem.*, 2006, **10**, 477–489.
- 94 W. Shi and H. M. Ma, *Chem. Commun.*, 2012, **48**, 8732–8744.
- 95 W. Shi, X. Li and H. Ma, *Angew. Chem., Int. Ed.*, 2012, **51**, 6432–6435.
- 96 J. Jia, K. Wang, W. Shi, S. M. Chen, X. H. Li and H. M. Ma, *Chem.–Eur. J.*, 2010, **16**, 6638–6643.
- 97 Y. K. Kim, M. H. Kim and D. H. Min, *Chem. Commun.*, 2011, **47**, 3195–3197.
- 98 C. L. Zhang, Y. X. Yuan, S. M. Zhang, Y. H. Wang and Z. H. Liu, *Angew. Chem., Int. Ed.*, 2011, **50**, 6851–6854.
- 99 J. Kim, L. J. Cote, F. Kim and J. X. Huang, *J. Am. Chem. Soc.*, 2010, **132**, 260–267.
- 100 C. H. Lu, H. H. Yang, C. L. Zhu, X. Chen and G. N. Chen, *Angew. Chem., Int. Ed.*, 2009, **48**, 4785–4787.
- 101 W. Han, R. Zaynagetdinov, F. E. Yull, V. V. Polosukhin, L. A. Gleaves, H. Tanjore, L. R. Young, T. E. Peterson, H. C. Manning, L. S. Prince and T. S. Blackwell, *Am. J. Respir. Cell Mol. Biol.*, 2015, **53**, 50–59.
- 102 T. Nagai, M. Tanaka, K. Hasui, H. Shirahama, S. Kitajima, S. Yonezawa, B. Xu and T. Matsuyama, *Clin. Exp. Immunol.*, 2010, **161**, 348–356.
- 103 B. Varghese, N. Haase and P. S. Low, *Mol. Pharm.*, 2007, **4**, 679–685.
- 104 C. Müller, F. Forrer, R. Schibli, E. P. Krenning and M. de Jong, *J. Nucl. Med.*, 2008, **49**, 310–317.
- 105 N. A. Jager, J. Westra, R. Golestani, G. M. van Dam, P. S. Low, R. A. Tio, R. H. J. A. Slart, H. H. Boersma, M. Bijl and C. J. Zeebregts, *J. Nucl. Med.*, 2014, **55**, 1945–1951.

