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# A colorimetric and fluorescent chemosensor for Hg<sup>2+</sup> based on a photochromic diarylethene with a quinoline unit

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A new colorimetric and fluorescent 'on-off' chemosensor, **10**, based on a photochromic diarylethene with a quinoline unit was designed and synthesized. The chemosensor **10** demonstrated selective and sensitive detection of Hg<sup>2+</sup> ions in the presence of other competitive metal ions in acetonitrile. The stoichiometric ratio of the sensor **10** for Hg<sup>2+</sup> was determined to be 1 : 1, and the limit of detection of the probe **10** was calculated to be 56.3 nM for Hg<sup>2+</sup>. In addition, a molecular logic circuit with four inputs and one output was successfully constructed with UV/vis light and metal-responsive behavior. ESI-MS spectroscopy, Job's plot analysis, and <sup>1</sup>H NMR titration experiments confirm the binding behavior between **10** and Hg<sup>2+</sup>.

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## 1. Introduction

Nearly all heavy metals are toxic to human health and dangerous for the environment. In particular, Hg<sup>2+</sup> is one of the major and most deadly heavy metal ions,<sup>1–3</sup> and is widely distributed in the water, air, and soil of everyday living environments.<sup>4,5</sup> The accumulation of mercury even at low concentrations can cause many deleterious effects,<sup>6,7</sup> including DNA damage, movement disorders, mitosis impairment, and serious damage to the nervous and endocrine organ systems.<sup>8–15</sup> Many methods to measure Hg<sup>2+</sup> have been reported, including atomic absorption spectrometry (AAS), electrochemical sensing, inductively coupled plasma optical emission spectrometry (ICP-OES), voltammetry, and more.<sup>16–20</sup> However, these traditional methods have some common disadvantages, such as expensive specialized equipment, complicated sample preparation, and long assay times.<sup>21–23</sup> Thus, it is important to develop a facile, rapid, sensitive, and easy to use method for sensing mercury ions even at low concentrations.

Optical methods, colorimetric or fluorometric, are preferred methods owing to their operational simplicity, real time monitoring, high selectivity and sensitivity, and low detection limits.<sup>24–31</sup> In addition, optical probes for the determination of transition metal cations are popular because they can facilitate analyte detection in solutions by the naked eye<sup>32,33</sup> as well as in living cells by microscopy.<sup>34–36</sup> Actually, a lot of fluorescent sensors have been designed for the detection of Hg<sup>2+</sup>, such as rhodamine,<sup>37</sup> oligothiophene,<sup>38</sup> pyrazoline,<sup>39</sup> coumarin.<sup>40</sup> Unfortunately, most of these sensors for the detection of Hg<sup>2+</sup>

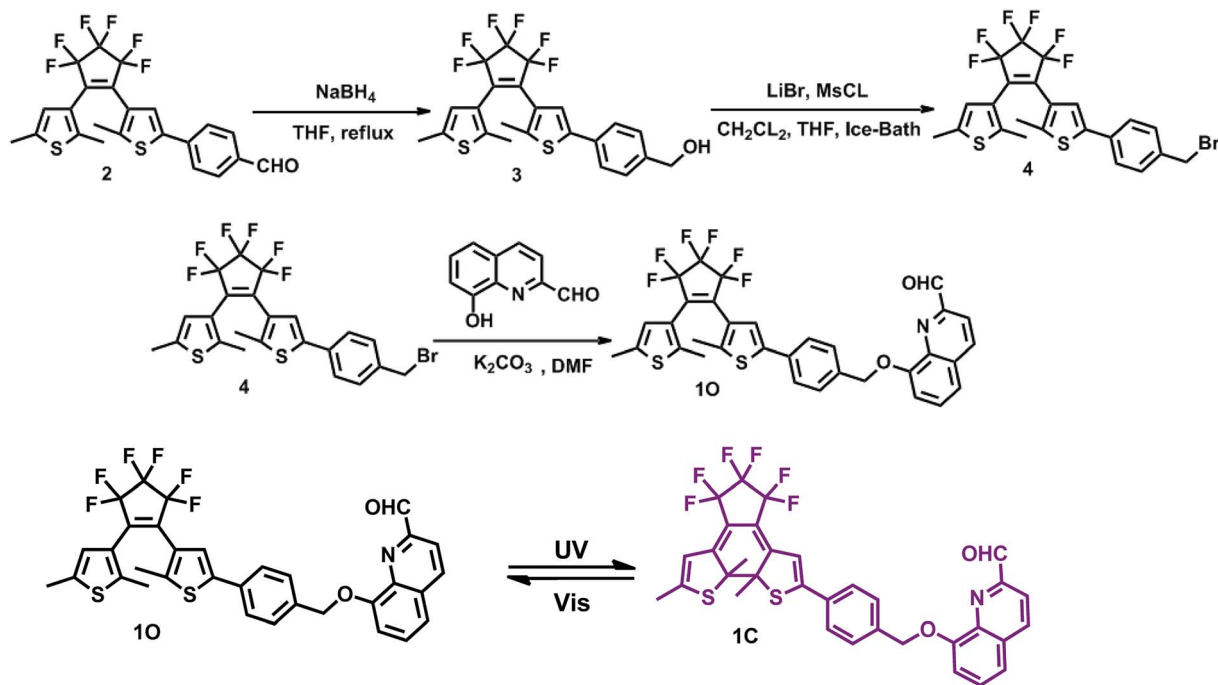
still have weak points, such as synthetic difficulty, complex molecular structure, poor selectivity and high detection limit. Therefore, it is necessary to design and develop efficient and convenient Hg<sup>2+</sup> sensors for their promising application potential in the environment.

Diarylethene derivatives, one of the most promising photo-switchable dyes have attracted considerable research interest due to their excellent photo reactivity, thermal stability, and fatigue resistance.<sup>41–44</sup> They can be used to construct photoswitches, logic circuits, optical memories, and chemosensors.<sup>45–49</sup> So far many diarylethene derivatives have been reported as optical sensors, such as diarylethene attach to 1-butyl-2-oxo-1,2-dihydroquinoline,<sup>50</sup> thieno-imidazole,<sup>51</sup> chromone,<sup>52</sup> 2-hydrazinopyridine,<sup>53</sup> 1,8-naphthyrindin,<sup>54</sup> rhodamine 6G.<sup>55</sup> However, most of these diarylethene derivatives are designed for detection of Zn<sup>2+</sup>, Al<sup>3+</sup>, Cd<sup>2+</sup> with Schiff base structure, and the reported sensing methods combining the colorimetric and fluorometric are relatively sparsely, especially with a quinoline group. Meanwhile, to our knowledge, Schiff base is unstable.<sup>56</sup> Thus, it is still highly desirable to design sensitive and stable sensors for Hg<sup>2+</sup> detection, which taking both advantages of colorimetric and fluorometric methods based on diarylethene with quinoline group.

Toward this goal, we describe herein a colorimetric and fluorescent chemosensor for Hg<sup>2+</sup> based on photochromic diarylethene with a quinoline unit. The selectivity and sensitivity for Hg<sup>2+</sup> were systematically investigated, and the results indicated that **10** exhibited changes in both absorbance and fluorescence upon exposure to Hg<sup>2+</sup> ions over other competing metal ions. In addition, **10** could detect Hg<sup>2+</sup> ions with a low detection limit (56.3 nM). Scheme 1 shows the synthetic route and photochromic process of **10**.

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Scheme 1 Scheme of the synthesis of diarylethene **10** and its photochromism.

## 2. Experiments

### 2.1. General methods

All solvents were of analytical grade and distilled before use. 8-Hydroxyquinoline was obtained from J & K Scientific Ltd and other chemical reagents were used as received. All metal ions except for  $\text{Ba}^{2+}$  and  $\text{K}^+$  that were prepared with their chloride salts.  $\text{Hg}^{2+}$  was prepared with its high chloride salts, and the remainings were obtained by dissolving the corresponding metal nitrates (0.1 mmol) in distilled water (10 mL). An EDTA solution was prepared with ethylenediaminetetraacetic acid disodium salt ( $\text{Na}_2\text{EDTA}$ ) (1.0 mmol) in distilled water (10 mL).

$^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra were recorded on a Bruker AV400 spectrometer (400 MHz) using  $\text{CD}_3\text{CL}$  and  $\text{CD}_3\text{CN}$  as solvents and tetramethylsilane (TMS) as the internal standard.

Mass spectra were obtained using an Agilent 1100 Ion Trap LC/MS MSD system. Melting points were measured using a WRS-1B melting point apparatus. Infrared spectra (IR) were collected on a BrukerVertex-70 spectrometer. UV-vis absorption spectra were recorded on an Agilent 8453 UV/vis spectrophotometer equipped with an MUA-165 UV lamp and MVL-210 visible lamp for photoirradiation. Fluorescence spectra were measured with a Hitachi F-4600 fluorescence spectrophotometer. Fluorescence quantum yield was measured using a QY C11347-11 absolute PL quantum yield spectrometer. Photoirradiation was performed on a setup consisting of a SHG-200 UV lamp, Cx-21 ultraviolet fluorescence analysis cabinet, and BMH-250 visible lamp. The synthetic route to the target compound **10** is shown in Scheme 1. Intermediate compounds **2** were prepared by the reported method.<sup>57</sup>

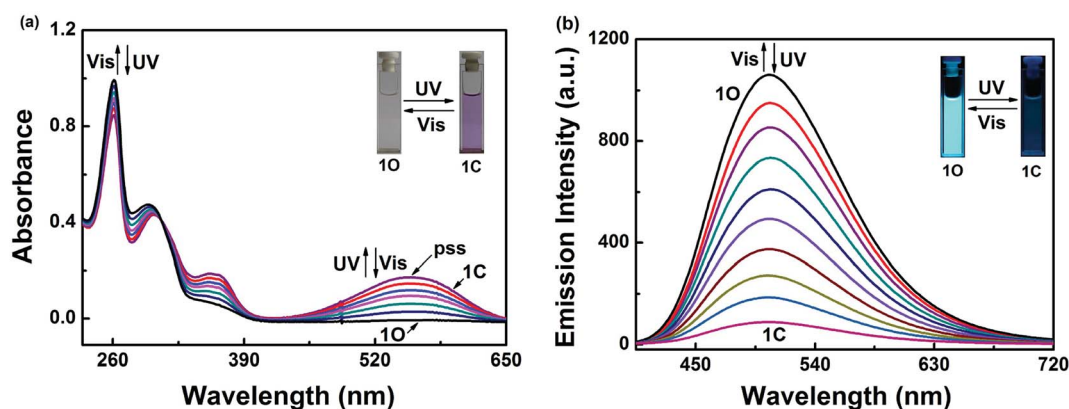


Fig. 1 Upon irradiation with UV-vis light: (a) absorption spectra changes of **10** in acetonitrile ( $2.0 \times 10^{-5} \text{ mol L}^{-1}$ ); (b) fluorescence spectra changes of **10** in acetonitrile ( $2.0 \times 10^{-5} \text{ mol L}^{-1}$ ). Excitation: 370 nm.



**2.1.1 Synthesis of 1-(2,5-dimethyl-3-thienyl)-2-[2-methyl-5-(4-hydroxymethylphenyl)-3-thienyl]perfluorocyclopentene (3).** 1.95 g, 4 mmol of compound 2 and 0.19 g, 4.8 mmol of NaBH<sub>4</sub> were taken in 50 mL of THF in a 100 mL round bottom flask. The mixture was stirred and refluxed for 2 h. After the reaction completed, the reaction product was cooled to RT and quenched with 20 mL water. The product was extracted with dichloromethane, the organic phase was washed with water, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered, and rotary evaporated. Finally, the compound was purified by column chromatography (petroleum ether : ethyl acetate = 1 : 1) to obtain a white solid 1.84 g

(3.76 mmol, 94%). Mp: 353–354 K; <sup>1</sup>H NMR (DMSO, 400 MHz),  $\delta$  (ppm): 1.843 (s, 3H, -CH<sub>3</sub>), 1.902 (s, 3H, -CH<sub>3</sub>), 2.293 (s, 3H, -CH<sub>3</sub>), 4.495 (t, 2H, -CH<sub>2</sub>-), 5.25 (s, 1H, -OH), 6.818 (s, 1H, Ar-H), 7.35 (d, 2H, Ar-H), 7.413 (s, 1H, Ar-H), 7.560 (d, 2H, Ar-H).

**2.1.2 Synthesis of 1-(2,5-dimethyl-3-thienyl)-2-[2-methyl-5-(4-bromomethylphenyl)-3-thienyl]perfluorocyclopentene (4).** To a stirred solution of 40 mL anhydrous CH<sub>2</sub>Cl<sub>2</sub> in ice bath, compound 3 (1.47 g, 3.00 mmol), MsCl (0.39 mL, 4.80 mmol) and TEA (0.75 mL, 5.40 mmol) were added. After rapid stirring for ten minutes at 273 K, added 10 mL of anhydrous THF containing LiBr (1.03 g, 12.00 mmol) to the reaction flask. Keep

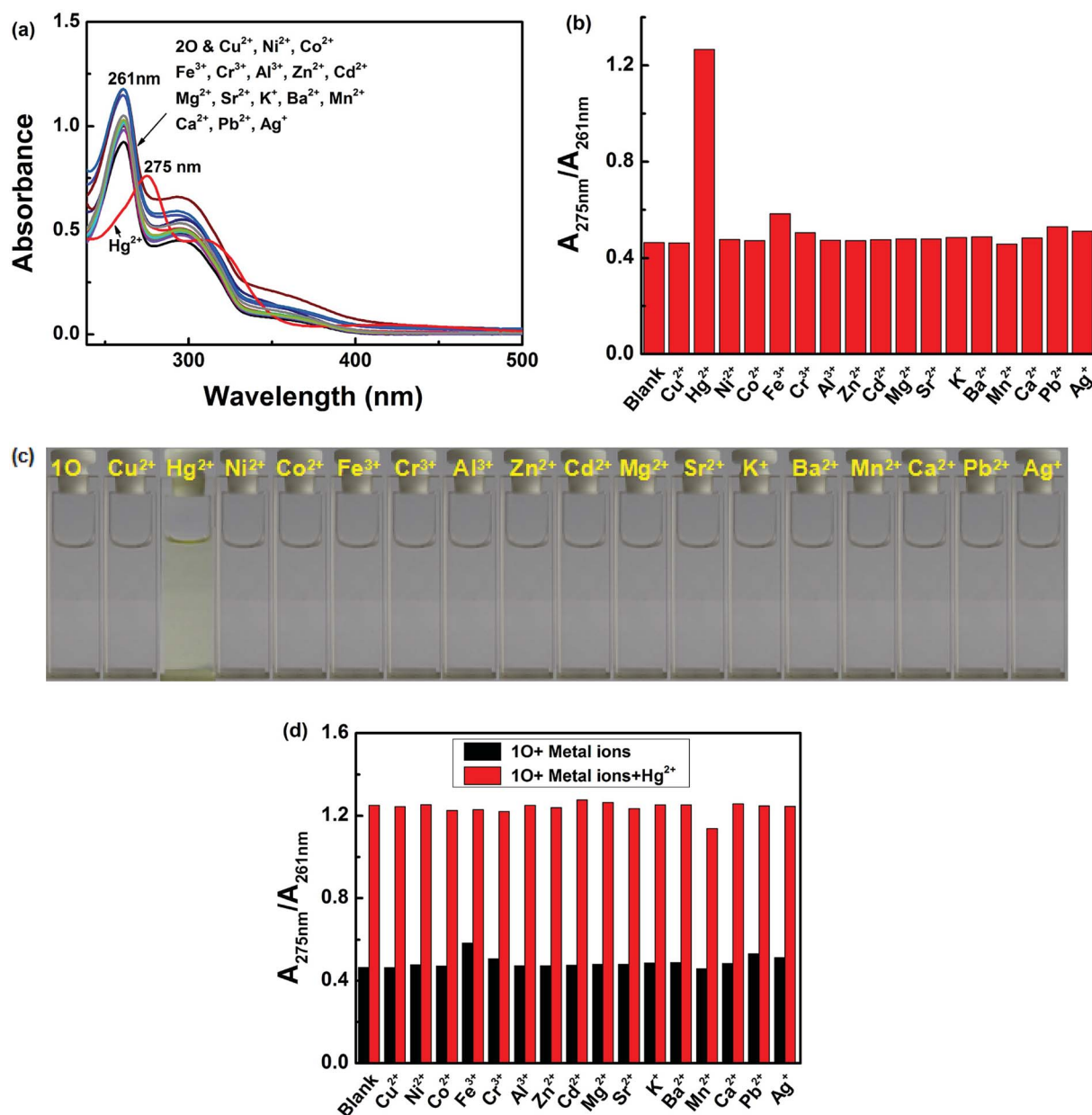


Fig. 2 Changes in the absorption of 1O induced by various metal ions (1.0 equiv.) in acetonitrile ( $2.0 \times 10^{-5}$  mol L<sup>-1</sup>). (a) UV/vis absorption spectra of 1O in the presence of different metal ions; (b) corresponding absorbance ratios ( $A_{275\text{ nm}}/A_{261\text{ nm}}$ ) of 1O in the presence of different metal ions; (c) photos demonstrating the color of 1O solutions containing different metal ions; (d) competitive tests for the absorption response of 1O in the presence of Hg<sup>2+</sup> and other competing metal ions in acetonitrile. Black bars represent the absorption of 1O solution in the presence of 1.1 equiv. metal ions and red bars represent the absorptions of 1O solutions containing the metal ions and Hg<sup>2+</sup> (1.0 equiv.).



this temperature and continue to stir for 30 minutes. Then, the solution was warmed to room temperature and stirred for 12 h. After reaction, the product was quenched with 20 mL water and evaporated of organic solvents. Extracted with dichloromethane, dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered, and rotary evaporated. Finally, the product was purified by column chromatography (petroleum ether : ethyl acetate = 8 : 1) to obtain a white solid 1.48 g (2.68 mmol, 90%). Mp: 345–346 K;  $^1\text{H}$  NMR (DMSO, 400 MHz),  $\delta$  (ppm): 1.556 (s, 3H,  $-\text{CH}_3$ ), 1.927 (s, 3H,  $-\text{CH}_3$ ), 2.412 (s, 3H,  $-\text{CH}_3$ ), 4.763 (d, 2H,  $-\text{CH}_2-$ ), 6.837 (s, 1H, Ar-H), 7.497 (d, 3H, Ar-H), 7.631 (t, 2H, Ar-H).

**2.1.3 Synthesis of 1-(2,5-dimethyl-3-thienyl)-2-{2-methyl-5-phenyl-4-(2-formylquinoline-8-methoxyl)-3-thienyl}perfluorocyclopentene (10).** Compound 4 (0.55 g, 1 mmol) and 8-hydroxyquinoline-2-carboxaldehyde (0.19 g, 1.10 mmol) were added to anhydrous  $\text{CH}_3\text{CN}$  (30 mL) containing  $\text{K}_2\text{CO}_3$  (0.33 g, 2 mmol). The solution was stirred at 283 K for 48 h. The reaction mixture was filtered, and the solvent was removed under vacuum. The crude product was purified by column chromatography on silica gel using dichloromethane/ethyl acetate (v/v = 6/1) as the eluent to give the target compound **10** as a purple solid (0.42 g, 0.67 mmol) in 67% yield. Mp: 339–341 K.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm): 1.88 (s, 3H,  $-\text{CH}_3$ ), 1.94 (s, 3H,  $-\text{CH}_3$ ), 2.45 (s, 3H,  $-\text{CH}_3$ ), 5.53 (s, 2H,  $-\text{CH}_2-$ ), 6.76 (s, 1H, Ar-H), 7.18 (d, 1H, Ar-H), 7.53 (t, 2H, Ar-H), 5.9 (t, 5H, Ar-H), 8.12 (d, 1H, Ar-H), 8.32 (d, 1H, Ar-H), 10.36 (s, 1H,  $-\text{CHO}$ );  $^{13}\text{C}$

NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 14.34 ( $-\text{CH}_3$ ), 15.07 ( $-\text{CH}_3$ ), 29.69 ( $-\text{CH}_3$ ), 70.81 ( $-\text{CH}_2-$ ), 111.20, 117.92, 120.15, 122.76, 124.68, 125.86, 127.75, 129.56, 131.43, 133.21, 135.31, 136.26, 137.28, 137.78, 139.78, 140.38, 141.26, 141.54, 151.70, 155.13, 193.82; IR (KBr,  $\nu$ ,  $\text{cm}^{-1}$ ): 1272 ( $-\text{C}-\text{O}$ ), 1704 ( $-\text{CHO}$ ). Anal. calcd for  $\text{C}_{33}\text{H}_{23}\text{F}_6\text{NO}_2\text{S}_2$ : C, 61.58; H, 3.60; F, 17.71; N, 2.18; O, 4.97; S, 9.96; found: C, 61.53; H, 3.65; F, 17.74; N, 2.23; O, 4.91; S, 9.93. MS ( $m/z$ ): Calculated for  $\text{C}_{33}\text{H}_{23}\text{F}_6\text{NO}_2\text{S}_2$  [ $\text{M}$ ] $^+$ : 643.11, found: 666.0972 for [**10** + Na] $^+$  and 698.1233 for [**10** + 3 $\text{H}_2\text{O}$  + H] $^+$ .

### 3. Results and discussion

#### 3.1. Photochromic and fluorescent properties of diarylethene **10**

The photochromic behavior of diarylethene **10** induced by UV/vis light was monitored in acetonitrile ( $C = 2.0 \times 10^{-5} \text{ mol L}^{-1}$ ) at room temperature. As shown in Fig. 1a, diarylethene **10** displayed a sharp absorption band at 261 nm ( $\epsilon_{\text{max}} = 4.9 \times 10^4 \text{ mol}^{-1} \text{ L cm}^{-1}$ ), which is assigned to the  $\pi-\pi^*$  transition. Upon irradiation with 297 nm light, the absorption band at 279 nm was decreased significantly. A new absorption band centered at 557 nm ( $\epsilon_{\text{max}} = 0.9 \times 10^4 \text{ mol}^{-1} \text{ L cm}^{-1}$ ) appeared and the color of the solution changed from colorless to purple due to the formation of ring-closed isomer **1C**.<sup>58,59</sup>

This photocyclization reaction reached the photostationary state (PSS) after 3 min of irradiation with UV light, where a clear

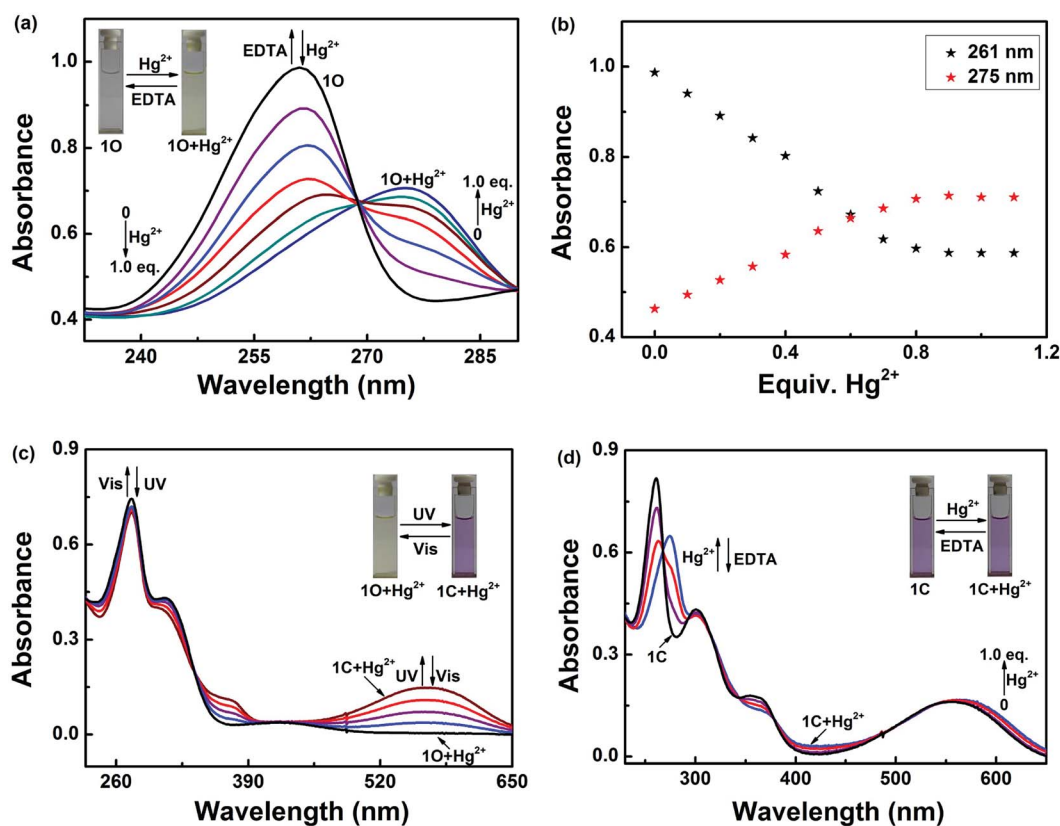


Fig. 3 Changes in the absorption spectrum and color of **10** ( $2.0 \times 10^{-5} \text{ mol L}^{-1}$ ) in acetonitrile induced by  $\text{Hg}^{2+}$ /EDTA and light stimuli: (a) UV/vis spectra of **10** induced by  $\text{Hg}^{2+}$ /EDTA; (b) absorbance at 261 and 275 nm versus the concentration of  $\text{Hg}^{2+}$ ; (c) UV/vis spectra of **10** +  $\text{Hg}^{2+}$  induced by UV/vis light; (d) UV/vis spectra of **1C** induced by  $\text{Hg}^{2+}$ /EDTA.



isosbestic point was observed at 308 nm. These observations suggest that the process is a two-component photochromic reaction. In addition, the purple solution of **1C** could be completely bleached upon irradiation with visible light ( $\lambda > 500$  nm) and the absorption spectrum returned back to the initial state **1O**. The cyclization and cycloreversion quantum yields were calculated to be 0.533 and 0.023, respectively, with 1,2-bis(2-methyl-5-phenyl-3-thienyl)-perfluorocyclopentene as the reference.

Fig. 1b shows the fluorescence emission spectra of **1O** in acetonitrile ( $2.0 \times 10^{-5}$  mol L<sup>-1</sup>) under photoirradiation at room temperature. A strong fluorescence peak of **1O** centered at

506 nm was observed under excitation at 370 nm. Upon irradiation with 297 nm UV light, the compound **1O** showed notable fluorescent switching properties, where the emission peak of **1O** decreased significantly and the solution color changed from bright cyan to dark. In the photostationary state, the fluorescence was 92% quenched due to the formation of the non-fluorescent ring closed isomer **1C**. The weak fluorescence may be due to incomplete cyclization. The fluorescence quantum yield of **1O** to **1C** was determined to be 0.245 to 0.021. Back irradiation with appropriate visible light ( $\lambda > 500$  nm) regenerated the open-ring isomer **1O** and restored the initial fluorescence.

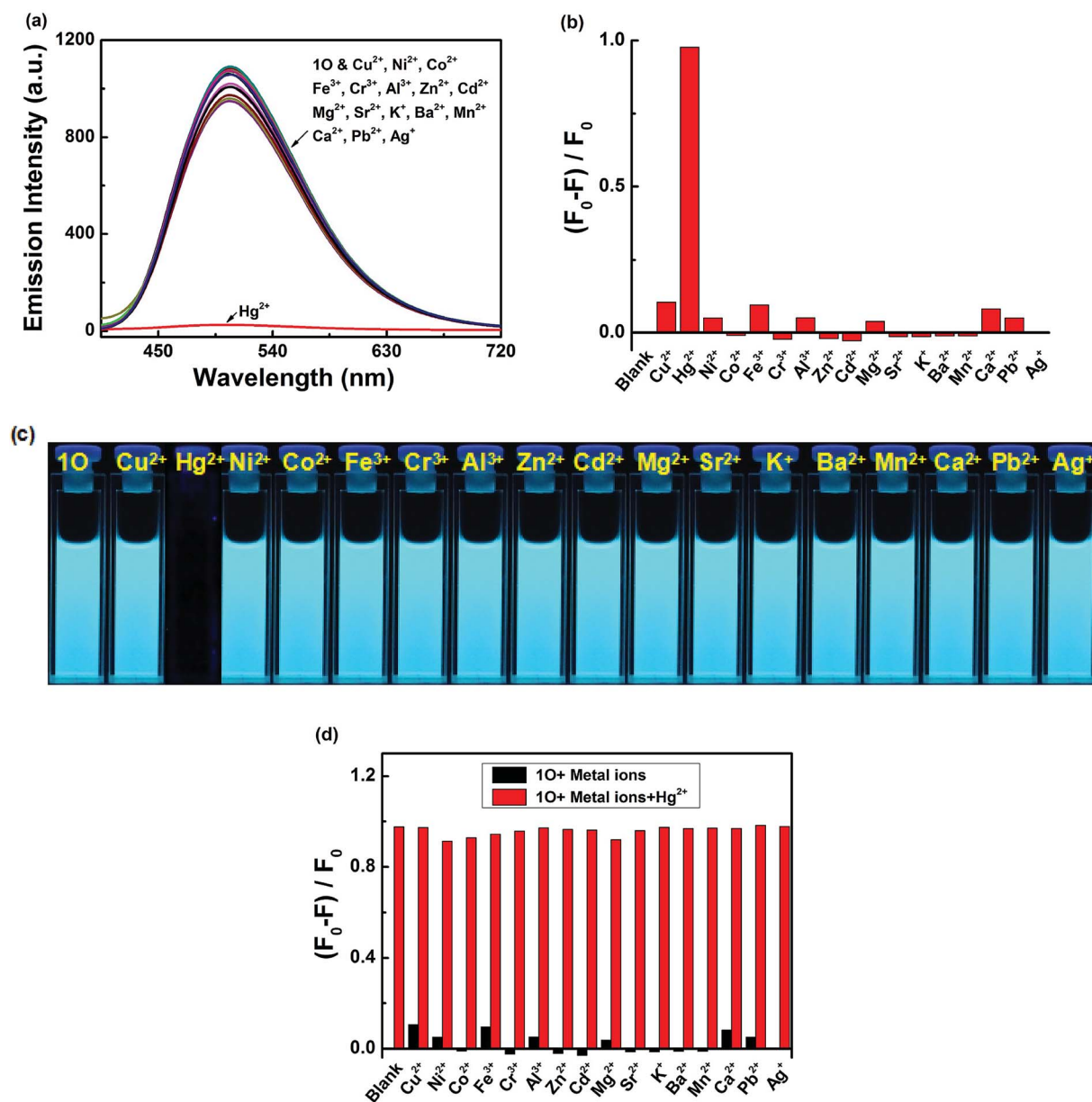


Fig. 4 Fluorescence responses of **1O** ( $2.0 \times 10^{-5}$  mol L<sup>-1</sup>) to various metal ions (2.1 equiv.) in acetonitrile. (a) Emission spectra of **1O** in the presence of different metal ions; (b) corresponding maximum emission intensities of **1O** in the presence of the metal ions; (c) photos demonstrating the colors of **1O** solutions containing different metal ions; (d) competitive tests for the fluorescent response of **1O** in the presence of Hg<sup>2+</sup> and other competing metal ions in acetonitrile. Black bars represent the fluorescence of **1O** solution in the presence of 2.1 equiv. metal ions and red bars represent the fluorescence of **1O** solutions containing the metal ions and Hg<sup>2+</sup> (2.1 equiv.).



### 3.2. Absorption response of **10** to $\text{Hg}^{2+}$

Colorimetric sensing behavior of **10** was examined by UV-vis in acetonitrile ( $2.0 \times 10^{-5} \text{ mol L}^{-1}$ ) at room temperature. As shown in Fig. 2, compound **10** was investigated in the presence of various metal ions, including  $\text{Cu}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Al}^{3+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Sr}^{2+}$ ,  $\text{K}^+$ ,  $\text{Ba}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Pb}^{2+}$  and  $\text{Ag}^+$ . Of all of these ions, only  $\text{Hg}^{2+}$  caused a modest color change from colorless to yellow (Fig. 2c). The UV-vis spectra showed that  $\text{Hg}^{2+}$  addition to the sensor **10** resulted in a 15 nm bathochromic shift from 261 nm to 275 nm, at the same time, a broad absorption band appeared in the visible region. These changes were ascribed to ligand-to-metal charge transfer (LMCT).<sup>60–62</sup> However, the other metal ions did not cause the absorption peak red shift and no new absorption band could be produced in the visible range (Fig. 2a). Fig. 2b shows the absorbance ratio ( $A_{275 \text{ nm}}/A_{261 \text{ nm}}$ ) of **10** upon addition of each ion, which also exhibited high selectivity of **10** for  $\text{Hg}^{2+}$  ions. To explore the practical applications of **10** in  $\text{Hg}^{2+}$  detection and to estimate the selectivity of **10** for  $\text{Hg}^{2+}$ , competitive experiments were conducted with the metal ions listed above. As depicted in Fig. 2d, other competitive metal ions did not interfere with the recognition of  $\text{Hg}^{2+}$  ions, confirming the selectivity of the sensor **10** for  $\text{Hg}^{2+}$  ions over other metal ions.

Fig. 3 shows the absorption spectral and color changes of **10** in acetonitrile ( $2.0 \times 10^{-5} \text{ mol L}^{-1}$ ) induced by the stimulation of  $\text{Hg}^{2+}$ /EDTA and UV/vis. As depicted in Fig. 3a, the first absorption band around 341 nm gradually reduced and a new absorption band at 275 nm obviously increased with the sequential addition of  $\text{Hg}^{2+}$  amount. It reached the maximum at 1.0 equiv. of  $\text{Hg}^{2+}$ , and a notable color change from colorless to yellow was apparent from bare eye. Further addition of  $\text{Hg}^{2+}$  did not change the absorption spectrum, suggesting that the **10** +  $\text{Hg}^{2+}$  complex was formed. The absorbance intensity at 261 and at 275 nm exhibited a good linear relationship with the  $\text{Hg}^{2+}$  in the range of 0–1.0 equiv. (Fig. 3b).

Compound **10** +  $\text{Hg}^{2+}$  could also undergo photoisomerization upon alternating irradiation with UV and visible light. Upon irradiation with 297 nm UV light, the yellow

solution of **10** +  $\text{Hg}^{2+}$  turned purple and a new absorption band centered at 532 nm ( $\epsilon_{\text{max}} = 7.4 \times 10^3 \text{ mol}^{-1} \text{ L cm}^{-1}$ ) emerged due to the formation of the closed-ring isomer **1C** +  $\text{Hg}^{2+}$  (Fig. 3c). In addition, **1C** +  $\text{Hg}^{2+}$  also could be formed by reacting **1C** with  $\text{Hg}^{2+}$  (Fig. 3d). It was noticed that the absorption properties of **10** and **1C** could be restored by adding excess EDTA to the solutions containing **10** +  $\text{Hg}^{2+}$  and **1C** +  $\text{Hg}^{2+}$ , respectively.

### 3.3. Fluorescence response of **10** to $\text{Hg}^{2+}$

As shown in Fig. 4, after the addition of 2.1 equiv. of different metal ions  $\text{Cu}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Al}^{3+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Sr}^{2+}$ ,  $\text{K}^+$ ,  $\text{Ba}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Pb}^{2+}$  and  $\text{Ag}^+$ , no significant changes in fluorescence intensity were observed except for in case of  $\text{Hg}^{2+}$ , where an emission band peak at 506 nm exhibited quenching (Fig. 4a). Compared with that of **10**, the fluorescence intensity of the complex **10** +  $\text{Hg}^{2+}$  was reduced to 97% at 506 nm (Fig. 4b). Simultaneously,  $\text{Hg}^{2+}$  could make the color of the **10** solution “switched-off” from a fluorescent bright cyan to a non-fluorescent color, no obvious color changes could be observed in the presence of other metal ions (Fig. 4c). These results suggest that **10** is highly selective towards  $\text{Hg}^{2+}$ . To explore practical applicability of **10** in  $\text{Hg}^{2+}$  detection, competitive experiments were performed in the presence of the above-mentioned metal ions (Fig. 4d). It was observed that the fluorescence quenching of **10** +  $\text{Hg}^{2+}$  was not affected by the presence of other metal ions (2.1 equiv.) in acetonitrile, indicating the high binding affinity of **10** for  $\text{Hg}^{2+}$ .

Meanwhile, changes in fluorescence of **10** induced by  $\text{Hg}^{2+}$ /EDTA were performed in acetonitrile ( $2.0 \times 10^{-5} \text{ mol L}^{-1}$ ) at room temperature, and the results are shown in Fig. 5. A decline in emission intensity at 506 nm was noted for **10** when the  $\text{Hg}^{2+}$  concentration increased from 0 to 2.1 equiv., accompanied by a notable fluorescent color change from bright cyan to dark and followed by a plateau upon further titration. Notably, the fluorescence of **10** was successfully restored with the addition of 2.1 equiv. of EDTA. In addition, the fluorescence quantum yield of **10** +  $\text{Hg}^{2+}$  was determined to be 0.008, 29.6 folds lower than that

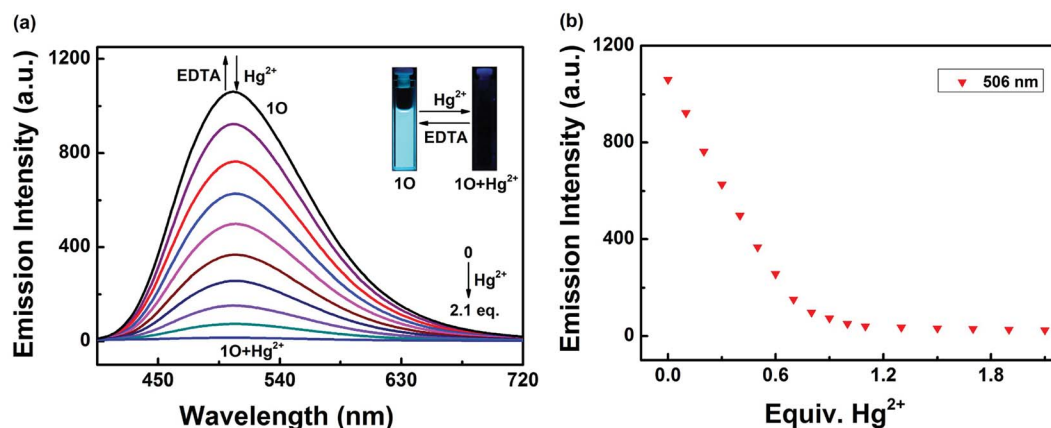


Fig. 5 Changes in the fluorescence of **10** in acetonitrile ( $2.0 \times 10^{-5} \text{ mol L}^{-1}$ ) induced by  $\text{Hg}^{2+}$ /EDTA, excited at 370 nm: (a) fluorescence emission spectra of **10** titrated with different amounts of  $\text{Hg}^{2+}$ ; (b) the fluorescence intensities of **10** at 506 nm in the presence of different equiv.  $\text{Hg}^{2+}$ .



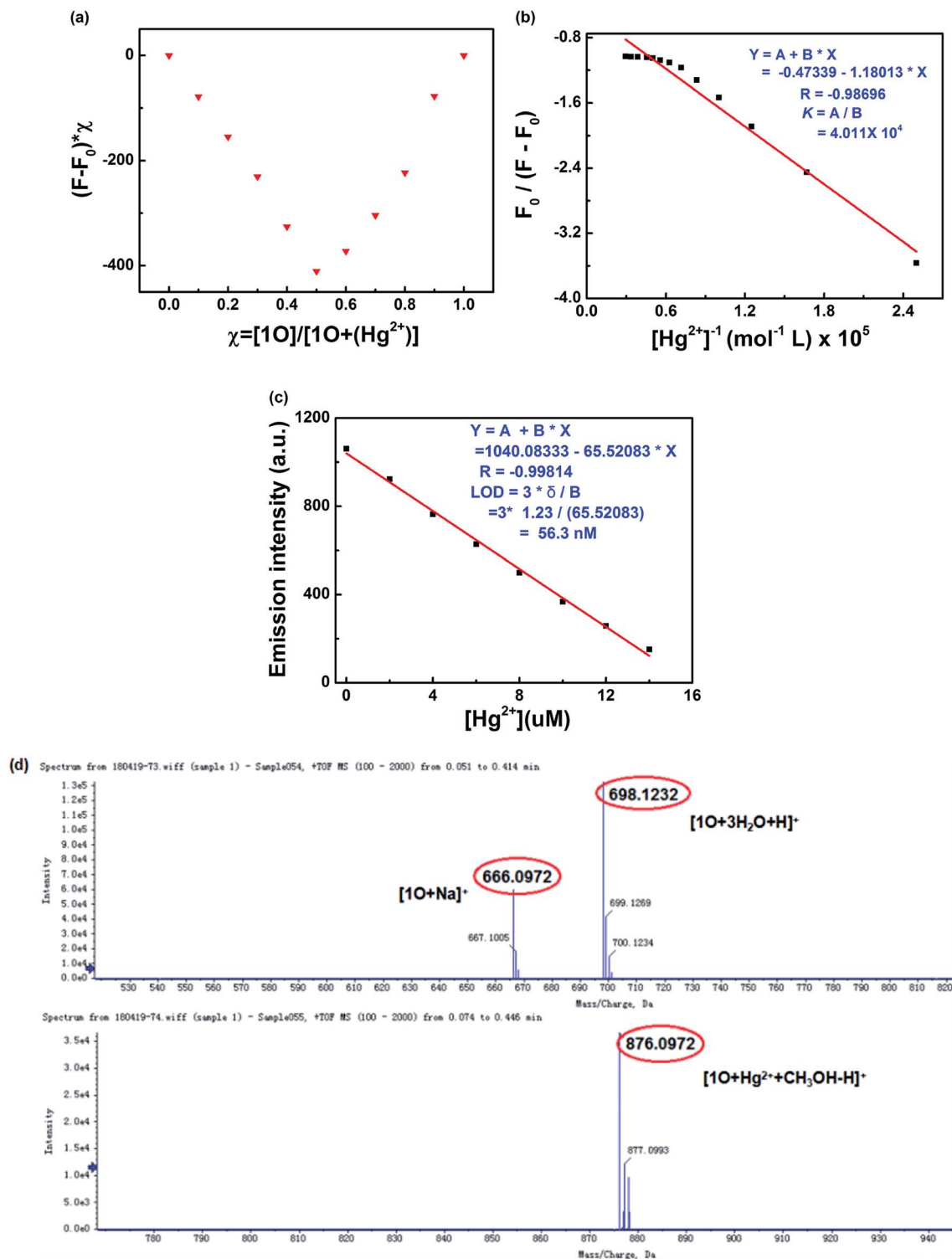


Fig. 6 (a) Job's plot showing the 1 : 1 complex of **10** with  $\text{Hg}^{2+}$ ; (b) Hildebrand–Benesi plot based on the 1 : 1 binding stoichiometry. The binding constant of **10** with  $\text{Hg}^{2+}$  was calculated to be  $4.011 \times 10^4 \text{ L mol}^{-1}$ ; (c) the limit of detection (LOD) was estimated to be 18.5 nM; (d) ESI-MS spectrum of **10** and **10** +  $\text{Hg}^{2+}$ .

of **10** ( $\Phi_f$ , **10** = 0.245), suggesting that the diarylethene **10** could also be potentially used as a fluorescent probe of  $\text{Hg}^{2+}$ .

The binding stoichiometry of **10** and  $\text{Hg}^{2+}$  was calculated from the modified Hildebrand–Benesi equation. As shown in

Fig. 6a, the lowest concentration of **10** +  $\text{Hg}^{2+}$  was achieved at  $[\text{Hg}^{2+}]/([\text{Hg}^{2+}] + [\mathbf{10}]) = 0.5$ , indicating that **10** was bound to  $\text{Hg}^{2+}$  in a binding stoichiometry of 1 : 1. The association constant ( $K_a$ ) of **10** with  $\text{Hg}^{2+}$  was calculated to be  $4.011 \times 10^4 \text{ L}$



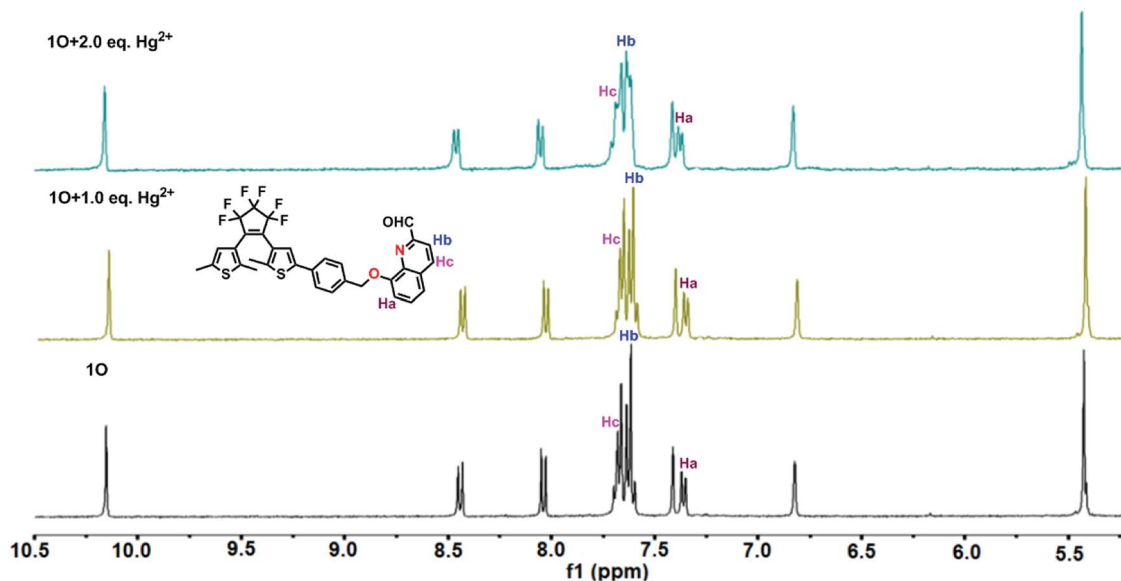
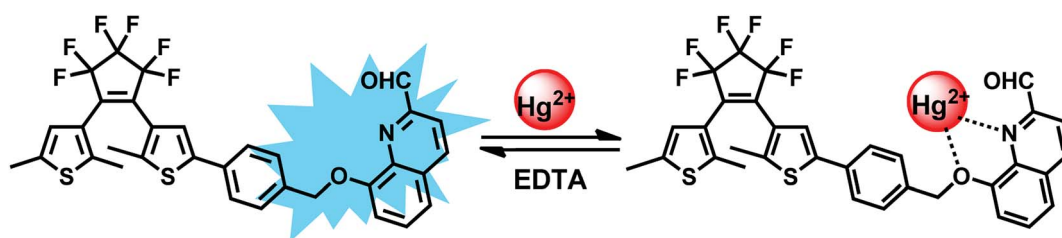


Fig. 7  $^1\text{H}$  NMR spectral changes in deuterated acetonitrile with the increasingly addition of  $\text{Hg}^{2+}$ .



Scheme 2 The binding mode between **10** and  $\text{Hg}^{2+}$ .

$\text{mol}^{-1}$  ( $R = -0.98696$ ) (Fig. 6b). The limit of detection of **10** for  $\text{Hg}^{2+}$  was measured to be 56.3 nM (Fig. 6c), lower than those of other  $\text{Hg}^{2+}$  sensors reported in literature, suggesting that **10** could be used as a highly sensitive fluorescence sensor of  $\text{Hg}^{2+}$ . The 1 : 1 coordination stoichiometry of **10** with  $\text{Hg}^{2+}$  was further confirmed by the ESI mass spectrometry (ESI-MS) analysis. **10** exhibited a characteristic peak at 666.0972  $m/z$  for  $[\mathbf{10} + \text{Na}]^+$  (calcd 666.11) and 698.1233  $m/z$  for  $[\mathbf{10} + 3\text{H}_2\text{O} + \text{H}]^+$  (calcd 698.11), which disappeared as 2.1 equiv. of  $\text{Hg}^{2+}$  were added, accompanied by the appearance of a new peak at 876.0972  $m/z$  for  $[\mathbf{10} + \text{Hg}^{2+} + \text{CH}_3\text{OH}-\text{H}]^+$  (calcd 876.10) (Fig. 6d).

Furthermore,  $^1\text{H}$  NMR titrations verified the binding mode of **10** with  $\text{Hg}^{2+}$ . As shown in Fig. 7, the double peaks at 7.353, 7.371 ppm belonging to Ha gradually shifted to 7.367 and 7.385 ppm, respectively. In addition, the proton signal of Hb at 7.617 ppm and Hc at 7.680 ppm reduced with the addition of  $\text{Hg}^{2+}$ . Therefore, we deduced that one oxygen atom close to methylene, and the nitrogen atom of quinolone together participated in binding with a  $\text{Hg}^{2+}$  ion, supporting that **10** was an “O–N” type ligand. Similar phenomena also observed in other reported probes.<sup>63</sup> Based on the UV-vis and fluorescence titration experiments, Job's plot, ESI-MS,  $^1\text{H}$  NMR titrations analysis, the proposed structure of **10** +  $\text{Hg}^{2+}$  is shown in Scheme 2.

### 3.4. pH effects

In order to study practical applicability, the pH effect on the fluorescence intensity changes of the **10** solution in the absence and presence of 2.1 equiv. of  $\text{Hg}^{2+}$  was evaluated. As shown in Fig. 8, when the pH value was lower than 4, it showed the fluorescence emission was significantly reduced, owing to the protonation of quinoline unit.<sup>64</sup> When the pH value range from 4 to 10, no apparent changes of the fluorescence intensity were

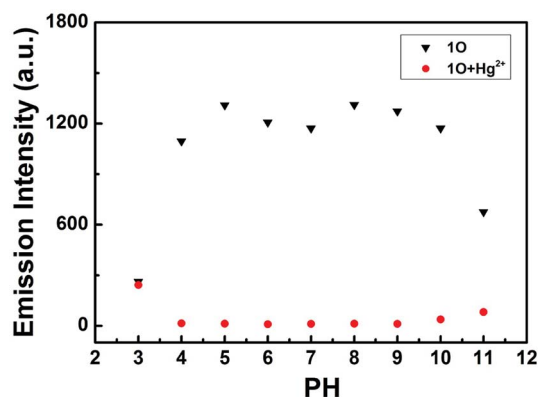


Fig. 8 Changes in the fluorescence intensity of sensor **10** at 506 nm with and without  $\text{Hg}^{2+}$  at different pH values ( $\lambda_{\text{ex}} = 370$  nm).



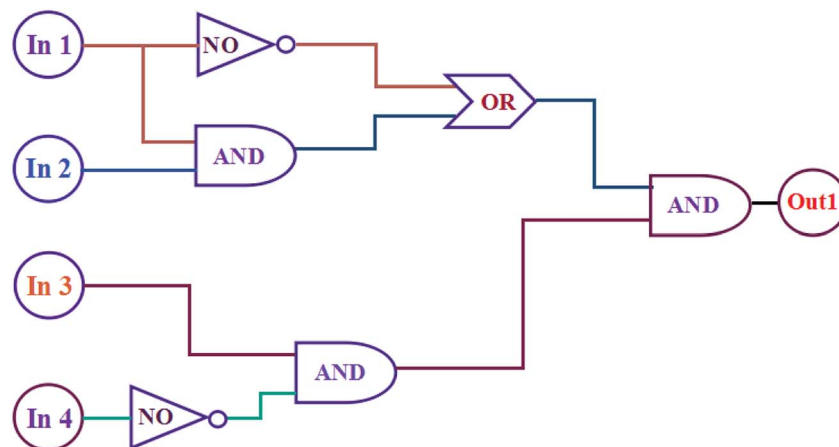


Fig. 9 The combinatorial logic circuits to the truth table given in Table 1: In1 (297 nm light), In2 ( $\lambda > 500$  nm visible light), In3 ( $\text{Hg}^{2+}$ ), In4 (EDTA) and output1 (fluorescence at 506 nm).

observed for **10**. Coincidentally, the **10** +  $\text{Hg}^{2+}$  complex was also not sensitive to pH in the range of 4–10, and at the same time, we can see the big gaps between the fluorescence intensity of **10** and **10** +  $\text{Hg}^{2+}$  within this pH range. Therefore, suggesting that sensor **10** was stable at biological conditions and could detect  $\text{Hg}^{2+}$  in a relatively wide pH range (4 to 10).

### 3.5. Application in logic circuits

As mentioned above, the fluorescence of **10** can be independently modulated by exposure to UV/vis light and to  $\text{Hg}^{2+}$ /EDTA. Based on these properties, a combinatorial logic circuit can be constructed using the stimulation of light irradiation (In1: 297 nm light and In2:  $\lambda > 500$  nm visible light) and chemical compounds (In3:  $\text{Hg}^{2+}$  and In4: EDTA) as four input signals, and the fluorescence intensity at 506 nm as the output signal (Fig. 9). The output signal could serve as ‘on’ with a Boolean

value of ‘1’ when fluorescence intensity was quenched to *ca.* 97%, otherwise defined as ‘off’ with a Boolean value of ‘0’. For example, inputs of ‘0, 0, 1 and 0’, corresponding to In1, In2, In3 and In4, result in an output of ‘off, off, on and off’. Under these conditions, **10** was converted to **10** +  $\text{Hg}^{2+}$  by stimulation with  $\text{Hg}^{2+}$ , and its emission intensity was quenched dramatically. As a result, the output signal is ‘on’ and the output digit is ‘1’. All possible logical combinations of the four inputs and their corresponding outputs are listed in Table 1.

## 4. Conclusion

In conclusion, we have developed a novel diarylethene-based colorimetric and on-off type fluorescent chemosensor **10** for the selective and sensitive sensing of  $\text{Hg}^{2+}$  in acetonitrile. The complexation of **10** with various metal ions was studied using UV-vis absorption and fluorescent emission spectra. Importantly, competition experiments suggested that the selective colorimetric and fluorescence quenching of  $\text{Hg}^{2+}$  is not affected by other metal ions. The detection limit of **10** towards  $\text{Hg}^{2+}$  was 56.3 nM. These results showed that probe **10** is an efficient sensor for detecting  $\text{Hg}^{2+}$ . In addition, the absorption and fluorescence emission of **10** were efficiently adjusted by stimuli with light and chemical species. Based on these characteristics, an integrated logic circuit with multiple control switches was successfully designed and constructed.

## Conflicts of interest

There are no conflicts of interest to declare.

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Table 1 Truth table for all possible strings of four binary-input data and the corresponding output<sup>a</sup>

Input				Output
In1 (UV)	In2 (Vis)	In3 ( $\text{Hg}^{2+}$ )	In4 (EDTA)	$\lambda_{\text{em}} = 506$ nm
0	0	0	0	0
1	0	0	0	0
0	1	0	0	0
0	0	1	0	1
0	0	0	1	0
1	1	0	0	0
1	0	1	0	0
1	0	0	1	0
0	1	1	0	1
0	1	0	1	0
0	0	1	1	0
1	1	1	0	1
1	1	0	1	0
1	0	1	1	0
0	1	1	1	0
1	1	1	1	0

<sup>a</sup> At 506 nm, when fluorescence is quenched to *ca.* 97%, is defined as 1. Otherwise, it is 0.



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## References

- 1 E. M. Nolan and S. J. Lippard, *Chem. Rev.*, 2008, **108**, 3443–3480.
- 2 T. Balaji, S. A. El-Safty, H. Matsunaga, T. Hanaoka and F. Mizukami, *Angew. Chem., Int. Ed.*, 2006, **118**, 7360–7366.
- 3 G. Sener, L. Uzun and A. Denizli, *Anal. Chem.*, 2014, **86**, 514–520.
- 4 C. J. Wu, J. B. Wang, J. J. Shen, C. Bi and H. W. Zhou, *Sens. Actuators, B*, 2017, **243**, 678–683.
- 5 Y. Zhang, J. J. Jiang, M. Li, P. F. Gao, L. H. Shi, G. M. Zhang, C. Dong and S. M. Shuang, *Sens. Actuators, B*, 2017, **238**, 683–692.
- 6 G. Q. Chen, Z. Guo, G. M. Zeng and L. Tang, *Analyst*, 2015, **140**, 5400–5443.
- 7 E. M. Nolan and S. J. Lippard, *Chem. Rev.*, 2008, **108**, 3443–3480.
- 8 Y. Wang, F. Yang and X. R. Yang, *Biosens. Bioelectron.*, 2010, **25**, 1994–1998.
- 9 Y. L. Wang, Y. Y. Cui, R. Liu, F. P. Gao, L. Gao and X. Y. Gao, *Sci. China: Chem.*, 2015, **58**, 819–824.
- 10 J. B. S. Fretham, S. Caito, J. E. Martinez-Finley and M. Aschner, *Toxicol. Res.*, 2012, **1**, 132–138.
- 11 P. S. Zhang, J. Li, B. W. Li, J. S. Xu, F. Zeng, J. Lv and S. Z. Wu, *Chem. Commun.*, 2015, **51**, 4414–4416.
- 12 J. S. Lee, M. S. Han and C. A. Mirkin, *Angew. Chem., Int. Ed.*, 2007, **46**, 4096.
- 13 T. Sun, Q. F. Niu, Y. Li, T. D. Li and H. X. Liu, *Sens. Actuators, B*, 2017, **248**, 24–34.
- 14 H. J. Huang, Y. Xu and X. Qian, *J. Org. Chem.*, 2009, **74**, 2167–2170.
- 15 R. Singh and G. Das, *Sens. Actuators, B*, 2018, **258**, 478–483.
- 16 H. Erxleben and J. Ruzicka, *Anal. Chem.*, 2005, **77**, 5124–5128.
- 17 Z. Q. Zhu, Y. Y. Su, J. Li, D. Li, J. Zhang, S. P. Song, Y. Zhao, G. X. Li and C. H. Fan, *Anal. Chem.*, 2009, **81**, 7660–7666.
- 18 M. Ozdemir, *Sens. Actuators, B*, 2017, **249**, 217–228.
- 19 J. M. Gong, T. Zhou, D. D. Song, L. Z. Zhang and X. L. Hu, *Anal. Chem.*, 2010, **82**, 567–573.
- 20 M. P. N. Bui, J. Brockgreitens, S. Ahmed and a. Abbas, *Biosens. Bioelectron.*, 2016, **85**, 280–286.
- 21 S. C. Lopes Pinheiro, I. M. Raimundo Jr, M. C. Moreno-Bondi and G. Orellana, *Anal. Bioanal. Chem.*, 2010, **398**, 3127–3138.
- 22 M. Ozdemir, *J. Photochem. Photobiol., A*, 2016, **31**, 7–13.
- 23 M. G. K. Salmani, G. H. Rounaghi and M. Chamsaz, *Sens. Actuators, B*, 2018, **256**, 968–975.
- 24 G. Q. Chen, Z. Guo, G. M. Zeng and L. Tang, *Analyst*, 2015, **140**, 5400–5443.
- 25 M. H. Lee, J. S. Kim and J. L. Sessler, *Chem. Soc. Rev.*, 2015, **44**, 4185–4191.
- 26 D. T. Quang and J. S. Kim, *Chem. Rev.*, 2010, **110**, 6280–6301.
- 27 K. P. Carter, A. M. Young and A. E. Palmer, *Chem. Rev.*, 2014, **114**, 4564–4601.
- 28 S. Y. Qin, B. Chen, J. Huang and Y. F. Han, *New J. Chem.*, 2018, **42**, 12766–12772.
- 29 J. F. Zhang, Y. Zhou, J. Yoon and J. S. Kim, *Chem. Soc. Rev.*, 2011, **40**, 3416–3429.
- 30 K. Ponnuvel, V. Padmini and R. Sribalan, *Sens. Actuators, B*, 2016, **222**, 605–611.
- 31 K. Ponnuvel, M. Kumar and V. Padmini, *Sens. Actuators, B*, 2016, **227**, 242–247.
- 32 J. M. Jung, C. Kim and R. G. Harrison, *Sens. Actuators, B*, 2018, **255**, 2756–2763.
- 33 Y. Zhao, L. L. Cui and Z. B. Chen, *Sens. Actuators, B*, 2017, **241**, 262–267.
- 34 H. Wang, P. S. Zhang, J. Chen, Y. Li, M. L. Yu, Y. F. Long and P. G. Yi, *Sens. Actuators, B*, 2017, **242**, 818–824.
- 35 S. M. Xu, Y. Liu, H. Yang, K. Zhao, J. G. Li and A. P. Deng, *Anal. Chim. Acta*, 2017, **964**, 150–160.
- 36 L. Yang, Y. A. Su, Y. N. Geng, H. Q. Xiong, J. L. Han, Q. Fang and X. Z. Song, *Org. Biomol. Chem.*, 2018, **16**, 5036–5042.
- 37 K. S. Min, R. Manivannan and Y. A. Son, *Sens. Actuators, B*, 2018, **261**, 545–552.
- 38 T. Sun, Q. F. Niu, Y. Li, T. D. Li and H. X. Liu, *Sens. Actuators, B*, 2017, **248**, 24–34.
- 39 E. Bozkurt and H. I. Gul, *Sens. Actuators, B*, 2018, **255**(1), 814–825.
- 40 C. J. Wu, J. B. Wang, J. J. Shen, C. Bi and H. W. Zhou, *Sens. Actuators, B*, 2017, **243**, 678–683.
- 41 M. Irie, T. Fukaminato and K. Matsuda, *Chem. Rev.*, 2014, **114**, 12174–12277.
- 42 W. L. Li, X. Li, Y. S. Xie, Y. Wu, M. Q. Li and X. Y. Wu, *Sci. Rep.*, 2015, **5**, 9186.
- 43 G. T. Xu, B. Li, J. Y. Wang, D. B. Zhang and Z. N. Chen, *Chem.–Eur. J.*, 2015, **21**, 3318–3326.
- 44 B. Li, H. M. Wen, J. Y. Wang, L. X. Shi and Z. N. Chen, *Inorg. Chem.*, 2015, **54**, 11511–11519.
- 45 J. J. He, J. X. He, T. T. Wang and H. P. Zeng, *J. Mater. Chem. C*, 2014, **2**, 7531–7540.
- 46 Y. L. Fu, C. B. Fan, G. Liu and S. Z. Pu, *Sens. Actuators, B*, 2017, **239**, 295–303.
- 47 S. J. Xia, G. Liu and S. Z. Pu, *J. Mater. Chem. C*, 2015, **3**, 4023–4029.
- 48 Y. Yamazaki, A. Sekine and H. Uekusa, *Cryst. Growth Des.*, 2017, **17**, 19–27.
- 49 R. J. Mart and R. K. Allemann, *Chem. Commun.*, 2016, **52**, 12262–12277.
- 50 E. T. Feng, Y. Y. Tu, C. B. Fan, G. Liu and S. Z. Pu, *RSC Adv.*, 2017, **7**, 50188.
- 51 Y. M. Xue, R. J. Wang, C. H. Zheng, G. Liu and S. Z. Pu, *Tetrahedron Lett.*, 2016, **57**, 1877–1881.
- 52 E. T. Feng, C. B. Fan, N. S. Wang, G. Liu and S. Z. Pu, *Dyes Pigm.*, 2018, **151**, 22–27.
- 53 S. Y. Li, D. B. Zhang, J. Y. Wang, R. M. Lu and C. H. Zheng, *Sens. Actuators, B*, 2017, **245**, 263–272.
- 54 X. X. Zhang, R. J. Wang, C. B. Fan, G. Liu and S. Z. Pu, *Dyes Pigm.*, 2017, **139**, 208–217.
- 55 H. T. Xu, H. C. Ding, G. Li, C. B. Fan, G. Liu and S. Z. Pu, *RSC Adv.*, 2017, **7**, 29827.



- 56 Z. L. Shi, Y. Y. Tu, R. J. Wang, G. Liu and S. Z. Pu, *Dyes Pigm.*, 2018, **149**, 764–773.
- 57 R. J. Wang, S. Z. Pu, G. Liu and S. Q. Cui, *Beilstein J. Org. Chem.*, 2012, **8**, 1018–1026.
- 58 C. B. Fan, L. L. Gong, L. Huang, F. Luo, R. Krashina, X. F. Yi, *et al.*, *Angew. Chem., Int. Ed.*, 2017, **56**, 7900–7906.
- 59 S. L. Guo, G. Liu, C. B. Fan and S. Z. Pu, *Sens. Actuators, B*, 2018, **266**, 603–613.
- 60 Y. Y. Fu, H. X. Li and W. P. Hu, *Eur. J. Org. Chem.*, 2007, **2007**, 2459–2463.
- 61 K. L. Zhong, X. Zhou, R. B. Hou, P. Zhou, S. H. Hou, Y. J. Bian, G. Zhang, L. J. Tang and X. H. Shang, *RSC Adv.*, 2014, **4**, 16612–16617.
- 62 Q. Li, Y. Guo and S. J. Shao, *Sens. Actuators, B*, 2012, **171–172**, 872–877.
- 63 L. L. Chang, Q. Gao, S. Liu, C. C. Hu, W. J. Zhou and M. M. Zheng, *Dyes Pigm.*, 2018, **153**, 117–124.
- 64 M. Amelia, M. Baroncini and A. Credi, *Angew. Chem., Int. Ed.*, 2008, **47**, 6240–6243.

