

PAPER

 View Article Online
View Journal | View Issue
Cite this: *RSC Adv.*, 2018, 8, 41692

A new family of fullerene derivatives: fullerene-curcumin conjugates for biological and photovoltaic applications†

Edison Castro,^{id}*^a Maira R. Cerón,^{id}^{ab} Andrea Hernandez Garcia,^a Quentin Kim,^a Alvaro Etcheverry-Berrios,^{id}^c Mauricio J. Morel,^{id}^c Raúl Díaz-Torres,^d Wenjie Qian,^d Zachary Martinez,^e Lois Mendez,^a Frank Perez,^a Christy A. Santoyo,^a Raquel Gimeno-Muñoz,^d Ronda Esper,^a Denisse A. Gutierrez,^{id}^e Armando Varela-Ramirez,^{id}^e Renato J. Aguilera,^e Manuel Llano,^e Monica Soler,^{id}^c Núria Aliaga-Alcalde,^{id}^{df} and Luis Echegoyen^{id}*^a

The synthesis and characterization of a family of [60]fullerocurcuminoids obtained *via* Bingel reactions is reported. The new C₆₀ derivatives include curcumin and curcuminoids with a variety of end groups. Preliminary biological experiments show the potential activity of the compound containing a curcumin addend, which exhibits moderate anti-HIV-1 and radical scavenger properties, but no anti-cancer activity. In addition, the new fullerocurcuminoids exhibit HOMO/LUMO energy levels that are reasonably matched with those of perovskites and when they were tested in perovskite solar cells (PSCs) as the electron transporting material (ETM), photoconversion efficiencies ranging from 14.04–14.95% were obtained, whereas a value of 16.23% was obtained for [6,6]-phenyl-C₆₁-butyric acid methyl ester (PC₆₁BM) based devices.

 Received 8th October 2018
Accepted 30th November 2018

DOI: 10.1039/c8ra08334g

rsc.li/rsc-advances

Introduction

Since the discovery of C₆₀ in 1985 by Kroto *et al.*,¹ fullerenes have attracted considerable attention due to their potential applications in biology and materials science.^{2–4} In 1993, Wudl and co-workers reported that fullerenes can inhibit HIV-1 viral infectivity, and they proposed a fullerene-induced inhibition mechanism involving binding to the protease (PR) active site based on the effect of these compounds on the *in vitro* activity of HIV-1 protease combined with *in silico* predictions.^{5,6} However, this paradigm was recently challenged by some of us, who showed that fullerenes do not inhibit HIV-1 protease activity in

in vitro assays at doses that potentially block HIV-1 maturation and infection. These findings indicate an alternative mode of viral inhibition that is yet to be determined.^{7,8}

The potential use of fullerene C₆₀ and its derivatives as strong antioxidants to reduce reactive oxygen species (ROS) has also been explored.^{9–12} Hydroxyl radicals are the most harmful among all species within the ROS family,¹³ and previous studies have shown that C₆₀ and its derivatives effectively scavenge hydroxyl radicals *via* reactions with the double bonds.^{7–9,14–16}

Curcumin (CCM) is a polyphenolic natural product found in the food coloring turmeric (curry powder), a member of the ginger family.¹⁷ CCM possesses diverse biological properties, such as anti-tumor,¹⁸ anti-oxidant,¹⁹ anti-arthritis,²⁰ anti-amyloid,²¹ and anti-inflammatory,^{22,23} among others.^{24,25} These potential biomedical applications are hindered by its low bioavailability and poor water solubility, although it is readily soluble in ethanol, DMSO, and acetone.²⁶ On the other hand, straightforward synthetic methods for low cost and high yield preparation of curcumin have been reported.^{27,28} These facts have motivated the design and synthesis of curcumin derivatives, called curcuminoids (CCMoids), with the goal of overcoming some of the drawbacks, and many structural variations have been reported.²⁹

For many years researchers have been studying the interesting biological properties of both curcumin, fullerene C₆₀ and their derivatives as individual entities.^{3,29} However, to our

^aDepartment of Chemistry, The University of Texas at El Paso, 500W University Ave, El Paso, TX 79968, United States. E-mail: eacastroportillo@utep.edu; echegoyen@utep.edu

^bPhysical and Life Sciences, Lawrence Livermore National Laboratory, 7000 East Ave, Livermore, CA 94550, USA

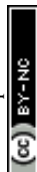
^cDepartamento de Ingeniería Química, Biotecnología y Materiales, Facultad de Ciencias Físicas y Matemáticas, Universidad de Chile, Beauchef 851, Santiago, Chile

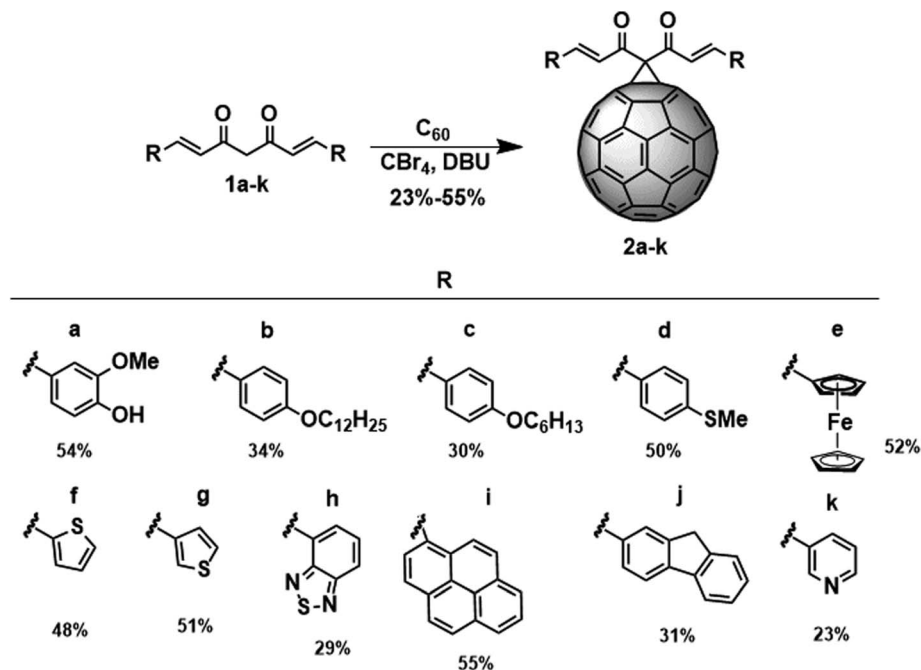
^dCSIC-ICMAB (Institut de Ciència dels Materials de Barcelona), Campus de la Universitat Autònoma de Barcelona, 08193 Bellaterra, Spain

^eDepartment of Biological Sciences, Border Biomedical Research Center, The University of Texas at El Paso, 500 West University Avenue, El Paso, TX 79968, USA

^fICREA (Institució Catalana de Recerca i Estudis Avançats), Passeig Lluís Companys 23, 08010 Barcelona, Spain

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c8ra08334g





Scheme 1 Synthesis of the fullerocurcuminoid derivatives 2a–k.

knowledge, no one has covalently combined these two structures into conjugated derivatives. In this work, we present a straightforward approach to synthesize several fullerocurcuminoid C_{60} derivatives using the methylene between the two carbonyl groups to form a methanoadduct through an addition/elimination reaction (Bingel reaction).³⁰ Preliminary biological and photophysical properties of these compounds were studied, testing the activity of C_{60} -CCM against HIV, as a radical scavenger and for anticancer action. Some of the new compounds were also tested in inverted PSCs as ETMs. The main focus of this work is to provide a methodology for the synthesis of C_{60} -CCMoids and to study some potential applications; thus detailed application studies are beyond the scope of this work.

Results and discussion

Synthesis of the fullerocurcuminoid C_{60} derivatives 2a–k

One of the most versatile reactions to functionalize fullerenes is the Bingel reaction, which involves an addition–elimination process to yield methanofullerenes.³⁰ The target compounds 2a–k were synthesized using this reaction, where the Br-CCMoid anions were generated *in situ* using carbon tetrabromide (CBr_4) and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) in *ortho*-dichlorobenzene (*o*-DCB) at room temperature. As represented in Scheme 1 eleven CCMoids 1a–k were reacted with C_{60} in the presence of CBr_4 and DBU to yield the fullerocurcuminoid derivatives 2a–k in 23–55% yields, using 3–30 min reaction times (see the ESI† for details). Longer reaction times led to the formation of *bis*- and *poly*-adduct products, which were not

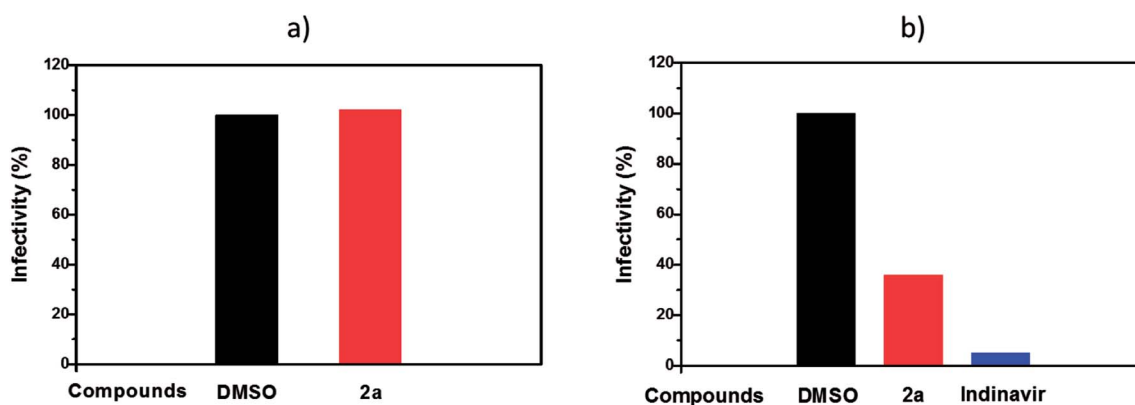


Fig. 1 Analysis of the viral life cycle step affected by compound 2a. (a) Effects on the early stages of the HIV-1 viral life cycle. (b) Effects of compound 2a and indinavir on the late phase of the HIV-1 infection. VSV-G pseudotyped, single-round HIV-1 expressing luciferase were produced in the presence of DMSO, compound 2a (3 μ M) and indinavir (0.1 μ M); the last one is a protease inhibitor used as a control.



isolated. Among all the new derivatives **2k** was the least soluble and the most unstable derivative; after one-week several dark spots were observed by thin layer chromatography (TLC). The chemical structures of compounds **1b–k** were confirmed by ^1H - and ^{13}C -NMR spectroscopy (Fig. S1–S10†) and **2a–k** were characterized by matrix-assisted-laser-desorption-ionization-mass-spectrometry (MALDI-TOF-MS), ^1H - and ^{13}C -NMR spectroscopy and UV-vis absorption (see ESI, Fig. S11–S32†).

The synthesized fullerocurcuminoids exhibited poor solubility in polar solvents such as DMSO or water, except for compound **2a**, due to the high solubility of the pristine CCM addend in DMSO. This low solubility in polar solvents and/or aggregation phenomena limited the biological studies exclusively to compound **2a**. Compound **2a** was tested as an anti-HIV-1 inhibitor of the viral replication in the early and late stages, as a radical scavenger, and as an anticancer agent against triple negative breast cancer (MDA-MB-231) and acute lymphoblastic leukemia (CEM) cells.

Effect of compound **2a** on the early stages of the HIV-1 life cycle

The effect of compound **2a** on the infectivity of VSV-G pseudotyped HIV-1 single-round infection viruses expressing LTR-driven luciferase (HIV-luc) was evaluated using the human $\text{CD4}^+ \text{T}$ cell line SupT1. These cells were infected with HIV-luc in the presence of DMSO (vehicle control) or fullerene **2a** (10 μM) and 24 h later the compound and the input virus were removed. After three days cellular luciferase and ATP levels were measured, and luciferase was normalized to ATP to adjust for cell viability and number. In these experiments, compound **2a**

only minimally affected HIV-1 infectivity (Fig. 1a). These results indicate that fullerene **2a** did not affect the early stages of the HIV-1 life cycle required for HIV-driven luciferase expression including viral entry and uncoating, reverse transcription, integration of viral cDNA, and HIV-1 Tat-mediated gene expression. These results also demonstrated that these compounds were not toxic to SupT1 cells at 10 μM concentration.

Effect of compound **2a** on the late stages of the HIV-1 life cycle

To evaluate the effects of **2a** in the late stages of viral development, we produced HIV-luc by plasmid transfection in HEK293T cells (obtained from Dr Eric Poeschla lab from Mayo Clinic) in the presence of DMSO, fullerene **2a** (3 μM), or indinavir³³ (0.1 μM). The latter is a PR inhibitor known to potently inhibit HIV-1 maturation. Viruses were concentrated by ultracentrifugation through a sucrose cushion, normalized for p24 levels, and their infectivity evaluated in untreated SupT1 cells (Fig. 1b). As expected, indinavir resulted in 96% inhibition at 0.1 μM , as compared to DMSO. On the other hand, compound **2a** inhibited by 64% the infectivity at 3 μM (Fig. 1b).

The plasmids used to generate retroviral vectors were described previously.³¹ HIV-1-derived vectors were produced using pHIV Luc and pMD.G. pHIV Luc was derived from pNL4-3.Luc.R-E³² by introducing a deletion in the env open reading frame (Bright-Glow™ Luciferase Assay System, Promega).

Compound **2a** exhibits cytoprotective activity in human neuroblastoma cells undergoing oxidative stress

A human SH-SY5Y cell line (ATCC) was selected to review the cytoprotective activity of compound **2a**; an established cell

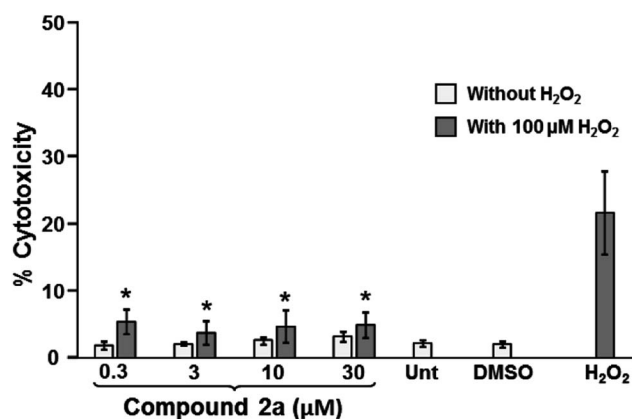


Fig. 2 Compound **2a** significantly mitigates H_2O_2 -induced oxidative stress in human neuroblastoma SH-SY5Y cells. Cells seeded in a 96 well plate were pretreated with a compound **2a** concentration gradient (μM) for 1 h, followed by addition of 100 μM of H_2O_2 and incubated for an additional 24 h (dark gray bars). A series of cell samples treated with just compound **2a** (light gray bars), without H_2O_2 , were analyzed concomitantly. Additional controls were untreated cells (Unt), cells treated with DMSO (solvent control) and cells treated with 100 μM H_2O_2 alone (positive control for cytotoxicity). Each bar represents the average of eight replicates and error bars their corresponding standard deviation. The P values were consistently < 0.00001 (*), when comparing the compound **2a**-pretreated cells with cells treated with H_2O_2 alone.

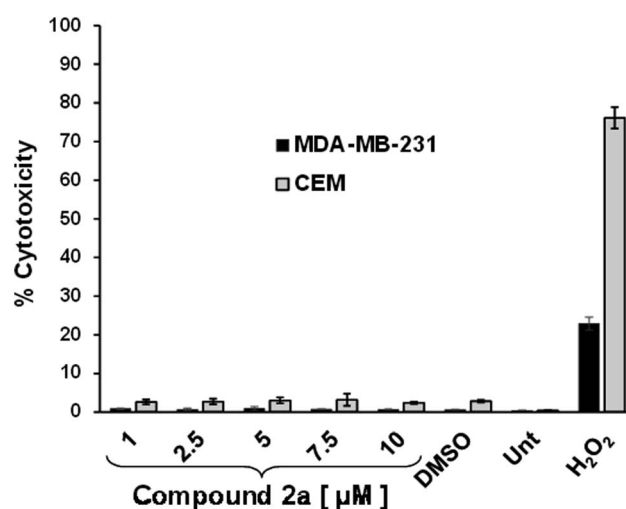


Fig. 3 Analysis of the cytotoxic properties of compound **2a** against MDA-MB-231 and CEM human cancer cell lines. To examine the percentage of cytotoxicity (y-axis) an IN Cell 2000 bioimager system (GE Healthcare) and differential nuclear staining assay were utilized. Cells were incubated for 48 h in the presence of various concentrations of compound **2a** (1 to 10 μM). The following controls were included: as a positive control for cytotoxicity H_2O_2 (1 mM); as a solvent control, 0.1% v/v DMSO; and untreated cells (Unt). Results shown are the average and standard deviation of four replicates.



model of dopaminergic neurons to study Parkinson's disease *in vitro*.³⁴ The cell death percentages were quantified by using the DNS assay and examined *via* high throughput screening bio-imager system (GE Healthcare).^{35,36} Cells pretreated (1 h) with 0.3, 3, 10 and 30 μM of compound **2a** were subsequently exposed (24 h) to 100 μM of H_2O_2 , to inflict the cellular oxidative stress and cell death. As expected, the H_2O_2 -treated cells, without any pretreatment, exhibited the maximum value of cytotoxicity ($21.71 \pm 6.22\%$; Fig. 2). All concentrations of compound **2a** tested displayed a consistently significant reduction in cytotoxicity after adding H_2O (100 μM ; 24 h) when compared with the cells treated with just H_2O_2 (Fig. 2). The cytotoxicity percentage values were 5.43 ± 1.89 , 3.76 ± 1.79 , 4.69 ± 2.41 and 4.92 ± 1.9 (average \pm SD), when 0.3, 3, 10 and 30 μM of compound **2a** were tested, respectively (Fig. 2); around 77% in reduction of cytotoxicity, which is considered highly significant ($P < 0.00001$). Moreover, compound **2a** itself showed no cytotoxic activity at all concentrations reviewed after 25 h of cell exposure (around 4.7%; light gray bars; Fig. 2). These cytotoxicity percentage values were very similar to DMSO and untreated negative control cells. These findings indicate that compound **2a** alone is not toxic to cells under the conditions tested in this study. Therefore, even at nanomolar concentrations (300 nM), compound **2a** possesses a strong antioxidant and

neuroprotective activity on human dopaminergic neurons SH-SY5Y cells, experiencing H_2O_2 -induced oxidative stress, probably acting as an effective free radical scavenger reagent.

Effect of compound **2a** as an anticancer agent

The potential anticancer activity of compound **2a** was examined in two human cancer cell lines; MDA-MB-231 triple negative breast cancer and CEM acute lymphoblastic leukemia cells (both from ATCC, Manassas, VA) by a differential nuclear staining assay^{34–36} (see ESI† for details). Both cell lines exhibited insignificant amounts of cytotoxicity when exposed to compound **2a** in a gradient of concentrations ranging from 1 to 10 μM for 48 h (Fig. 3). As shown in Fig. 3 the cytotoxicity values observed for compound **2a**-treated cells lines consistently resembled the values of the untreated (Unt) and DMSO vehicle controls, suggesting that compound **2a** has no cytotoxic activity against triple negative breast cancer and acute lymphoblastic leukemia cells. H_2O_2 (1 mM) was included as a positive control of death, and as expected, the highest cytotoxic values were observed; 22.9% and 76.1% for MDA-MB-231 and CEM cells respectively. These last results indicate that MDA-MB-231 cells were more resistant to the H_2O_2 (1 mM), as compared with CEM cells.

Photovoltaic properties of fullerocurcuminoids **2a–h**

Due to the extended conjugation of the CCM/CCMoid backbones, these compounds have been extensively studied as light-harvesting materials in bulk heterojunction (BHJ) and dye-sensitized (DS) solar cells,^{37–40} and PCE values up to 4.1%³⁷ and 5.8%⁴⁰ have been obtained, respectively. Since fullerocurcuminoids **2a–h** have not been synthesized before they have not been tested in PSCs as the electron transporting layers (ETLs) nor as the hole transporting layers (HTLs).

The optical properties of the fullerocurcuminoid derivatives **2a–k** were investigated by means of UV/Vis spectroscopy. As seen in Fig. S33,† **2a–k** exhibit absorption bands below 400 nm arising from both functionalities, C_{60} and the aromatic endings of the CCMoids. Between 400–450 nm the bands observed for the majority of the compounds correspond to the skeleton of the CCMoid moieties and in the case of **2e**, the absorption band at the highest wavelength is linked to the ferrocene groups.^{41–43}

Table 1 Electrochemical and photophysical data of compounds **2a–h**. PC_{61}BM values were taken from the literature^{46a}

Compound	λ_{abs} (nm)	E_g (eV)	$E_{\text{onset red}}$ (V)	LUMO (eV)	HOMO (eV)
PC_{61}BM	718	1.73	−0.90	−3.90	−5.63
2a	716	1.73	−1.09	−3.93	−5.66
2b	718	1.73	−1.08	−3.94	−5.67
2c	718	1.73	−1.09	−3.93	−5.66
2d	714	1.74	−1.13	−3.89	−5.63
2e	720	1.72	−1.07	−3.95	−5.67
2f	716	1.73	−1.02	−4.00	−5.73
2g	718	1.73	−1.10	−3.92	−5.65
2h	718	1.73	−1.09	−3.93	−5.66

^a Values obtained using the following formula
 $E_{\text{LUMO}} = -(E_{\text{onset red}}^{\text{vs Fc}^+/\text{Fc}} + 4.8) \text{ eV}.$ ⁴⁵

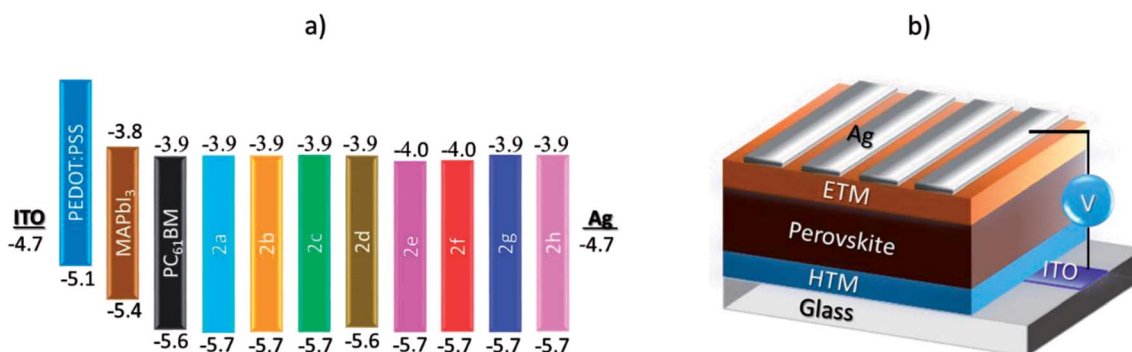


Fig. 4 (a) Schematic illustration of the estimated HOMO and LUMO energy levels, calculated from CV and UV-vis, (b) PSCs' cell architectures.



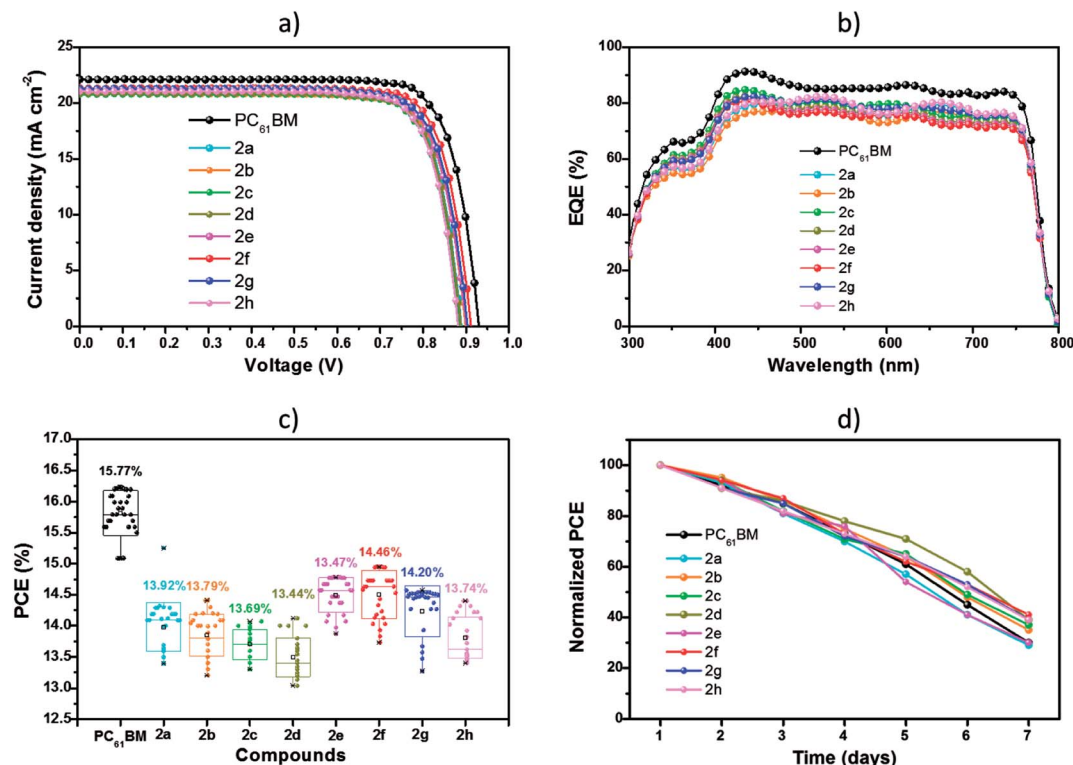


Fig. 5 (a) *J*–*V* curves under 1 sun illumination (100 mW cm^{-2}) in forward voltage scans. (b) EQE spectra for the PSCs fabricated using compounds **2a–h** and **PC₆₁BM** as the ETMs. (c) The PCE histograms measured for 32 independent cells. (d) Normalized PCEs of PSCs measured as a function of time in $\sim 20\%$ humidity at room temperature for seven days.

The electrochemical properties of **2a–h** and **PC₆₁BM** were determined by cyclic voltammetry (CV) in *o*-DCB as described in the EIS. As shown in Fig. S34,[†] compounds **2a–h** exhibit reversible and quasi-reversible cathodic electrochemical behavior between -1.0 and -1.3 V, whereas more negative potentials lead to irreversible retro-cycloadditions.⁴⁴ In particular compounds **2d**, **2e** and **2f** showed chemical reversibility for the first reduction step at a scan rate of 100 mV s^{-1} , followed by chemical and electrochemically irreversible waves. The highest occupied molecular orbital/lowest unoccupied molecular orbital (HOMO/LUMO) values were estimated from the UV and CV measurements.⁴⁵ The optical properties of compounds **2a–h** are summarized in Table 1.

Compounds **2a–h** exhibit similar HOMO/LUMO values to those of **PC₆₁BM**, thus they are a good match for the conduction and valence bands of the perovskite, respectively,⁴⁷ and should provide efficient electron collection and hole blocking ability. The schematic representation of the work function of the PSCs is illustrated in Fig. 4a.

We studied the electron transporting ability of compounds **2a–h** in PSCs with the configuration of ITO/PEDOT:PSS/perovskite/ETM/Ag (Fig. 4b), and compared them to **PC₆₁BM**. The cells were fully characterized using SEM, XRD, and photovoltaic measurements (Fig. S35 and S36,[†] respectively). As shown in Fig. 5a, devices based on **2a–h** show PCEs ranging from 14.04 – 14.95% . The main photovoltaic characteristics are

Table 2 Summary of the main photovoltaic performance of PSCs using **2a–h** as the ETMs^a

Compound	Calculated J_{sc} (mA cm^{-2})	J_{sc} (mA cm^{-2})	V_{oc} (V)	FF (%)	PCE (%)
PC₆₁BM	21.87	22.10	0.93	0.79	15.73 ± 0.46 (16.23)
2a	21.02	21.14	0.89	0.76	13.92 ± 0.38 (14.30)
2b	20.54	20.79	0.90	0.77	13.79 ± 0.62 (14.41)
2c	20.35	20.77	0.89	0.76	13.69 ± 0.35 (14.04)
2d	20.63	20.87	0.89	0.76	13.44 ± 0.68 (14.12)
2e	21.27	21.33	0.90	0.77	13.47 ± 1.31 (14.78)
2f	21.09	21.33	0.91	0.77	14.46 ± 0.49 (14.95)
2g	21.16	21.25	0.90	0.76	14.20 ± 0.34 (14.54)
2h	20.86	21.02	0.89	0.77	13.74 ± 0.66 (14.40)

^a Calculated J_{sc} from EQE measurements, values in parentheses are the highest PCEs.



summarized in Table 2. These PCEs, although lower than for the control **PC₆₁BM** (16.23%), are significant and promising. The integrated photocurrent densities based on EQE measurements (Fig. 5b) are consistent with those obtained from *J*-*V* measurements (Table 2). Devices prepared with all the fullerene derivatives including **PC₆₁BM** exhibit a high photo response within the entire visible range. Device performance reproducibility was calculated from the PCE distributions measured for 32 independent cells (Fig. 5c). The long term-device stabilities for PSCs were conducted under ~20% humidity at room temperature without encapsulation for seven days. Fig. 5d, shows the normalized PCEs vs. time. After seven days of periodic measurements, devices based on **2a-h** and **PC₆₁BM** lost 60% of their initial PCE. No significant differences were observed for devices prepared either with compounds **2a-h** or **PC₆₁BM**. These encouraging results will be studied in more detail in the future to fully understand the roles of aggregation as well as that of interface interactions between the C₆₀-CCMoids and the perovskite films.

Conclusions

In this work, we describe for the first time the conjugation of curcumin-like compounds to fullerene C₆₀ through a Bingel type reaction to obtain new fullerocurcuminoid derivatives. Fullerocurcuminoid **2a** was soluble in DMSO and was tested as an anti-HIV-1 agent, as a radical scavenger, and as an anticancer agent. When studying the late stages of the HIV viral cycle, compound **2a** inhibits 64% of the HIV-1 viral infection at 3 μM and it also prevents radical damage at this concentration. No anticancer activity was detected in our assays. These results are consistent with those for curcuminoid, which exhibits low toxicity and no anticancer activity toward several cell lines. In addition, fullerocurcuminoids **2a-h** were soluble in chlorobenzene, and exhibit HOMO/LUMO energy levels that match those of perovskites, therefore, when they were tested in PSCs as the ETMs, PCEs in the range of 14.04–14.95%, were obtained. A value of 16.23% was obtained for **PC₆₁BM**-based devices. Further studies are needed to understand the nature of the photovoltaic activity as well as to improve the observed performances.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors thank the US National Science Foundation (NSF) for the generous support of this work under the NSF-PREM program DMR 1205302 and CHE-1801317 (to L. E.). The Robert A. Welch Foundation is also gratefully acknowledged for an endowed chair to L. E. (Grant AH-0033). MS acknowledges support through the FONDECYT Grant Number 1161775, 3170509. AEB thanks Conicyt PFCHA 21140734 for a PhD scholarship. The work of N. A.-A. has been supported by the Spanish Government by the project MAT2016-77852-C2-1-R and

acknowledge the “Severo Ochoa” Program for Centres of Excellence in R&D (SEV-2015-0496). N. A.-A. and W. Q. thanks to the Generalitat de Catalunya, FI-DGR grant 2014. R. D.-T. thanks to the Fundació Montcelimar. A. E. B., M. S and N. A.-A. thank to the CSIC (project iCOOP20162). ML thanks the National Institute of General Medical Sciences (NIGMS) Grant Number 5 SC1 AI098238-02 and the National Institutes of Health (NIH). The Cytometry, Screening and Imaging Core Facility-BBRC-UTEP that was used in this work was supported by the National Institute on Minority Health and Health Disparities (NIMHD) through a Research Centers for Minority Institutions (RCMI) grant [5G12MD007592] a component of NIH. We also thank Dr Narayan for helping us with the anti-radical experiments.

References

- 1 H. W. Kroto, J. R. Heath, S. C. O'Brien, R. F. Curl and R. E. Smalley, *Nature*, 1985, **318**, 162–163.
- 2 A. Montellano Lopez, A. Mateo-Alonso and M. Prato, in *Fullerenes: Principles and Applications (2)*, The Royal Society of Chemistry, 2012, pp. 389–413.
- 3 E. Castro, A. H. Garcia, G. Zavala and L. Echegoyen, *J. Mater. Chem. B*, 2017, **5**, 6523–6535.
- 4 E. Castro, J. Murillo, O. Fernandez-Delgado and L. Echegoyen, *J. Mater. Chem. C*, 2018, **6**, 2635–2651.
- 5 S. H. Friedman, D. L. DeCamp, R. P. Sijbesma, G. Srdanov, F. Wudl and G. L. Kenyon, *J. Am. Chem. Soc.*, 1993, **115**, 6506–6509.
- 6 R. Sijbesma, G. Srdanov, F. Wudl, J. A. Castoro, C. Wilkins, S. H. Friedman, D. L. DeCamp and G. L. Kenyon, *J. Am. Chem. Soc.*, 1993, **115**, 6510–6512.
- 7 E. Castro, Z. S. Martinez, C. S. Seong, A. Cabrera-Espinoza, M. Ruiz, G. Hernandez, A. F. Valdez, M. Llano and L. A. Echegoyen, *J. Med. Chem.*, 2016, **59**, 10963–10973.
- 8 Z. S. Martinez, E. Castro, C. S. Seong, M. R. Ceron, L. Echegoyen and M. Llano, *Antimicrob. Agents Chemother.*, 2016, **60**, 5731–5741.
- 9 N. Gharbi, M. Pressac, M. Hadchouel, H. Szwarc, S. R. Wilson and F. Moussa, *Nano Lett.*, 2005, **5**, 2578–2585.
- 10 T. Kop, M. Bjelaković and D. Milić, *Tetrahedron*, 2015, **71**, 4801–4809.
- 11 T. Baati, F. Bourasset, N. Gharbi, L. Njim, M. Abderrabba, A. Kerkeni, H. Szwarc and F. Moussa, *Biomaterials*, 2012, **33**, 4936–4946.
- 12 N. Higashi, T. Shosu, T. Koga, M. Niwa and T. Tanigawa, *J. Colloid Interface Sci.*, 2006, **298**, 118–123.
- 13 G. V. Andrievsky, V. I. Bruskov, A. A. Tykhomyrov and S. V. Gudkov, *Free Radical Biol. Med.*, 2009, **47**, 786–793.
- 14 R. Angelique, Y. W. Yan, F. J. Marcel and H. Barbara, *Exp. Dermatol.*, 2017, **26**, 220–224.
- 15 N. S. Allen, E. B. Zeynalov, K. Taylor and P. Birkett, *Polym. Degrad. Stab.*, 2009, **94**, 1932–1940.
- 16 R. Czochara, J. Kusio and G. Litwinienko, *RSC Adv.*, 2017, **7**, 44021–44025.
- 17 S. Hewlings and D. Kalman, *Foods*, 2017, **6**, 92.



- 18 P. Neerati, Y. A. Sudhakar and J. R. Kanwar, *J. Cancer Sci. Ther.*, 2013, **5**, 313–319.
- 19 M. Asouri, R. Ataee, A. A. Ahmadi, A. Amini and M. R. Moshaei, *Asian J. Chem.*, 2013, **25**, 7593–7595.
- 20 J. K. Jackson, T. Higo, W. L. Hunter and H. M. Burt, *Inflammation Res.*, 2006, **55**, 168–175.
- 21 G. M. Cole, F. Yang, G. P. Lim, J. L. Cummings, D. L. Masterman and S. A. Frautschy, *Curr. Med. Chem.: Immunol., Endocr. Metab. Agents*, 2003, **3**, 15–25.
- 22 H. Nishikawa, J. Tsutsumi and S. Kitani, *J. Funct. Foods*, 2013, **5**, 763–772.
- 23 B. B. Aggarwal and K. B. Harikumar, *Int. J. Biochem. Cell Biol.*, 2009, **41**, 40–59.
- 24 A. Goel, A. B. Kunnumakkara and B. B. Aggarwal, *Biochem. Pharmacol.*, 2008, **75**, 787–809.
- 25 K. M. Nelson, J. L. Dahlin, J. Bisson, J. Graham, G. F. Pauli and M. A. Walters, *J. Med. Chem.*, 2017, **60**, 1620–1637.
- 26 M. Bernabé-Pineda, M. a. T. Ramírez-Silva, M. Romero-Romo, E. González-Vergara and A. Rojas-Hernández, *Spectrochim. Acta, Part A*, 2004, **60**, 1091–1097.
- 27 H. Itokawa, Q. Shi, T. Akiyama, S. L. Morris-Natschke and K.-H. Lee, *Chin. Med.*, 2008, **3**, 11.
- 28 E. V. Rao and P. Sudheer, *Indian J. Pharm. Sci.*, 2011, **73**, 262–270.
- 29 A. Amalraj, A. Pius, S. Gopi and S. Gopi, *J. Tradit. Complement. Med.*, 2017, **7**, 205–233.
- 30 C. Bingel, *Chem. Ber.*, 1993, **126**, 1957–1959.
- 31 M. Llano, D. T. Saenz, A. Meehan, P. Wongthida, M. Peretz, W. H. Walker, W. Teo and E. M. Poeschla, *Science*, 2006, **314**, 461–464.
- 32 J. He, S. Choe, R. Walker, P. Di Marzio, D. O. Morgan and N. R. Landau, *J. Virol.*, 1995, **69**, 6705–6711.
- 33 D. Leung, G. Abbenante and D. P. Fairlie, *J. Med. Chem.*, 2000, **43**, 305–341.
- 34 P. Kabiraj, J. E. Marin, A. Varela-Ramirez and M. Narayan, *ACS Chem. Neurosci.*, 2016, **7**, 1519–1530.
- 35 C. Lema, A. Varela-Ramirez and R. J. Aguilera, *Curr. Cell. Biochem.*, 2011, **1**, 1–14.
- 36 E. Robles-Escajeda, U. Das, N. M. Ortega, K. Parra, G. Francia, J. R. Dimmock, A. Varela-Ramirez and R. J. Aguilera, *Cell. Oncol.*, 2016, **39**, 265–277.
- 37 F. Archet, D. Yao, S. Chambon, M. Abbas, A. D'Aléo, G. Canard, M. Ponce-Vargas, E. Zaborova, B. Le Guennic, G. Wantz and F. Fages, *ACS Energy Lett.*, 2017, **2**, 1303–1307.
- 38 D. Sinha, D. De and A. Ayaz, *Spectrochim. Acta, Part A*, 2018, **193**, 467–474.
- 39 M. Tsuchikawa, A. Takao, T. Funaki, H. Sugihara and K. Ono, *RSC Adv.*, 2017, **7**, 36612–36616.
- 40 H. Ozawa, H. Kawaguchi, Y. Okuyama and H. Arakawa, *Ambio*, 2012, **41**, 149–150.
- 41 F. Langa and J. F. Nierengarten, *Fullerenes: Principles and Applications*, RSC, Cambridge, United Kingdom, 2011.
- 42 J. G. Domínguez-Chávez, E. Cruz-Chávez, I. Moggio, E. Arias-Marín, T. Klimova, I. Lijanová and M. Martínez-García, *Fullerenes, Nanotubes, Carbon Nanostruct.*, 2012, **20**, 249–265.
- 43 M. H. Ucisik, S. Küpcü, B. Schuster and U. B. Sleytr, *J. Nanobiotechnol.*, 2013, **11**, 37.
- 44 G. L. Closs, P. Gautam, D. Zhang, P. J. Krusic, S. A. Hill and E. Wasserman, *J. Phys. Chem.*, 1992, **96**, 5228–5231.
- 45 Q. Sun, H. Wang, C. Yang and Y. Li, *J. Mater. Chem.*, 2003, **13**, 800–806.
- 46 C. Tian, E. Castro, G. Betancourt-Solis, Z.-A. Nan, O. Fernandez-Delgado, S. Jankuru and L. Echegoyen, *New J. Chem.*, 2018, **42**, 2896–2902.
- 47 P. Schulz, E. Edri, S. Kirmayer, G. Hodes, D. Cahen and A. Kahn, *Energy Environ. Sci.*, 2014, **7**, 1377–1381.

