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Antitumor component recognition from the Aconiti Lateralis Radix Praeparata and Glycyrrhizae Radix et Rhizoma herb pair extract by chemometrics and mean impact value†

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The purpose of this research is to recognize the active antitumor components from the mixed pair extract of Aconiti Lateralis Radix Praeparata (Fuzi in Chinese) and Glycyrrhizae Radix et Rhizoma (Gancao in Chinese) using chemometrics and mean impact value (MIV) methods. Firstly, 30 common components of 31 different samples were analyzed quantitatively and qualitatively by HPLC-UV and UPLC-Q-TOF tandem mass spectrometry, respectively. Meanwhile, MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assays were used to test the inhibition activities of the 31 different samples against HeLa cells. Then a back propagation (BP) neural network, support vector regression (SVR), and two optimization algorithms – genetic algorithm (GA) and particle swarm optimization (PSO) – were applied to construct composition–activity relationship (CAR) models for the Fuzi–Gancao extract. Based on the optimal CAR model, the MIV was introduced to evaluate the contribution of each individual component to the anticancer efficacy of the extract. Results indicated that the SVR-PSO model best depicted the complex relationship between the chemical composition and the inhibition effect of a Fuzi–Gancao extract. The 30 common components were ranked by their absolute MIVs, and the top 8, which corresponded to peaks 17, 25, 22, 13, 23, 28, 5, and 7 in the chromatogram, were tentatively deemed to be the main antitumor components. The integrated strategy shows a novel and efficient approach to understanding the potential contributions of components from complicated herbal medicines, and the identified results suggest certain directions for screening and research into new antitumor drugs.

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1. Introduction

Recently, herbal medicines (HMs) have appeared more and more attractive owing to their shining advantages of favorable cure effects, mild action, low levels of side effects and rich sources.¹ Especially for malignant and chronic diseases like cancer, HMs can tend to have remarkable therapeutic effects.² As we all know, HMs contain various compounds, which provide the material basis of the excellent curative effect of HMs. However, at present, most scientific research on HMs just focuses on one or a small number of ingredients that are usually present in high amounts, while some other low level components, which may also have potent bioactivity or can promote efficacy, are overlooked intentionally or unintentionally.

In order to find out as much as possible about the therapeutic constituents of HMs, it is necessary to study the relationship between chemical composition and the therapeutic

effect of the HMs, and chemometrics makes it possible to establish a composition–activity relationship (CAR) model. The CAR model can be used to predict the pharmaceutical activities of crude medicines according to chemical component information. Moreover, when combined with a variable relevance analysis method like mean impact value (MIV), the model can evaluate the bioactivities of the contained constituents, improving the efficiency of drug discovery. Some achievements have been made by this strategy, demonstrating its effectiveness.^{3–5} To date, various algorithms have been applied to CAR model construction. However, since HMs contain a wide variety of compounds and complicated mutual interactions exist among these components, the commonly used linear models like multiple linear regression analysis (MLR) usually do not fit the practical situation very well. By contrast, nonlinear algorithms like the back propagation (BP) neural network and support vector regression (SVR) are more suitable for explaining the CAR of HMs.^{6–8} BP is one of the most widely used neural networks, and it usually consists of three parts: an input layer, a hidden layer and an output layer. There are a certain number of neurons in each layer, which are involved in two processes, namely signal forward propagation and error back propagation,

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to adjust the weight and bias of each neuron to achieve functional approximation.⁹ SVR solves nonlinear problems by increasing the dimensions of the input data. In theory, a kernel function is utilized to map the input data to a high-dimensional characteristic space, converting the linearly non-separable data to linear separable data, and a linear regression function is established in the space to describe the relationship.^{10,11} BP and SVR have been successfully applied to quality prediction and evaluation for some herbal medicines.^{12–15}

Furthermore, the performance of CAR models can be markedly improved after parameter optimization.^{16,17} The genetic algorithm (GA) and particle swarm optimization (PSO) algorithm are two commonly used parameter optimization methods. GA is derived from the combination of natural selection and genetic inheritance, and it searches for the best optimal solution by selection, crossover and mutation like biological evolution.¹⁸ PSO is a kind of swarm intelligence algorithm, inspired by the foraging behavior of a biotic population, and it has many attractive traits such as easy operation, few adjustment parameters and fast convergence rates.¹⁹

Aconiti Lateralis Radix Praeparata (Fuzi in Chinese) and *Glycyrrhizae Radix et Rhizoma* (Gancao in Chinese) used to concurrently appear in classical Chinese medicine prescriptions for the treatment of rheumatoid arthritis and joint pain.^{20,21} Fuzi, the processed lateral roots of *Aconitum carmichaelii* Debeaux, has been in officinal use for a long time having various biological activities like analgesic, anti-inflammation, antidepressant, antihyperglycemic and cardiovascular protective effects.^{22–24} Gancao, the dried roots and stolons of *Glycyrrhiza uralensis* Fisch., is one of the most frequently used herbal medicines all over the world, and in addition to its pharmaceutical use, it often serves as an additive in herbal formulations to improve solubility, increase effects and alleviate toxicity.^{25,26} Recently, some researchers have reported that both Fuzi and Gancao possess favorable antitumor effects.^{27–29} However, the study of the antitumor constituents of the Fuzi–Gancao herb pair is extremely insufficient. As illustrated in Fig. 1, the present research makes use of a BP neural network and SVR to explore the CAR of the aqueous extract of the Fuzi–Gancao herb pair, and uses MIV to find out the main active antitumor components in this extract.

2. Materials and methods

2.1. Chemicals and plant materials

HPLC-grade methanol and acetonitrile were both purchased from Concord Technology Co., Ltd. (Tianjin, China). Ammonium acetate (HPLC \geq 98%) and glacial acetic acid (HPLC \geq 99.8%) were both purchased from Tianjin Kemiou Chemical Reagent Co., Ltd. (Tianjin, China). Water was supplied by a Milli-Q water purification system (Millipore, Bedford, MA, USA). Lappaconitine hydrobromide (HPLC \geq 98%) was purchased from Nanjing SenBeiJia Biological Technology Co., Ltd. (Nanjing, China). Roswell Park Memorial Institute (RPMI) 1640 culture medium and trypsin were both purchased from Invitrogen Gibco Company (Gaithersburg, MD, USA). Fetal bovine serum (FBS) was purchased from Beijing Dingguo

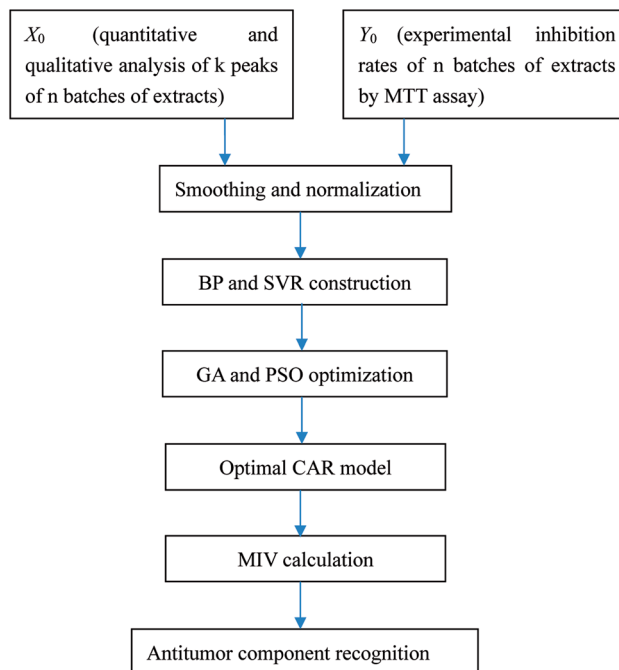


Fig. 1 A schematic representation of the framework of this research. CAR models for the Fuzi–Gancao herb pair were constructed by BP, SVR, GA and PSO, and used to fit experimental data. The main active antitumor components were recognized from MIVs based on the optimal CAR model.

Changsheng Biotechnology Co., Ltd. (Beijing, China). Sodium chloride injection (0.9%) was purchased from China Otsuka Pharmaceutical Co., Ltd. (Tianjin, China). MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) and dimethyl sulfoxide (DMSO) were purchased from Sigma-Aldrich Co. LLC. (Santa Clara, USA). HeLa cells were provided by Logistics University of People's Armed Police Force (Tianjin, China).

A total of 31 batches of raw Fuzi and Gancao from different places in China were collected and authenticated by Professor Tiejun Zhang from the Tianjin Institute of Pharmaceutical Research, and all voucher specimens are deposited at our laboratory.

2.2. Preparation of the extract

Dried medicinal slices of the two herbal medicines were accurately weighed, to give 20.00 g for Fuzi and 30.00 g for Gancao,

Table 1 Results of HPLC-UV method validation for precision, stability and reproducibility

Subject	RSD ^a of RRT ^b /%	RSD of RPA ^c /%
Precision	0.066–0.700	0.312–4.372
Stability	0.092–1.244	0.246–4.968
Reproducibility	0.124–0.430	0.508–4.857

^a RSD = (S.D./mean) \times 100%. ^b RRT: relative retention time. ^c RPA: relative peak area.



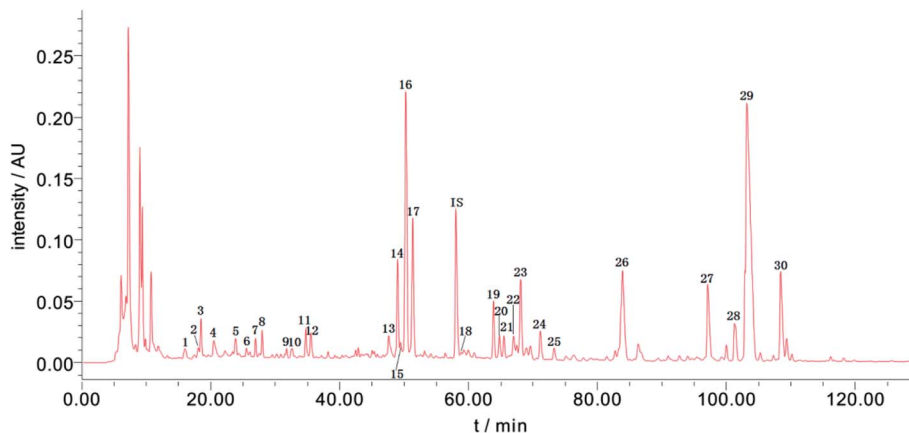


Fig. 2 A representative HPLC-UV fingerprint of Fuzi-Gancao extract. The 30 marked chromatographic peaks were determined as common peaks in 31 batches of Fuzi-Gancao extract.

and they were mixed in a 1000 mL round-bottom flask. The mixture was soaked in 600 mL water for 30 minutes and then extracted using a reflux device at 100 °C for 2 hours. The mixture was immediately filtered when the extraction was over. The filter liquor was concentrated under vacuum at 75 °C until the weight change in one minute was no more than 0.1 g, to obtain the extract sample, which was stored at 4 °C for next use.

2.3. Preparation of internal standard solution and sample solution

Lappaconitine hydrobromide served as the internal standard (IS). Lappaconitine hydrobromide (6.3 mg) was dissolved in water in a 10 mL volumetric flask using an ultra-sonicator, and diluted with water to obtain a concentration of 630 $\mu\text{g mL}^{-1}$ of IS stock solution. IS stock solution (1 mL) was diluted with water in a 5 mL volumetric flask to 126 $\mu\text{g mL}^{-1}$, to give the IS solution, which was stored at 4 °C for next use.

Precisely weighed extract (0.2 g) (see Section 2.2) was dissolved in water in a 5 mL volumetric flask using an ultra-sonicator, and then centrifuged at 7000 rpm for 10 minutes. Thereafter, an accurately measured amount of supernatant liquor (1.0 mL) was taken out and mixed thoroughly with an equal volume of IS solution. The mixture was filtered through a 0.22 μm nylon membrane, to give a sample solution for the following qualitative and quantitative analysis.

2.4. HPLC-UV instruments and conditions

A Waters e2695 high performance liquid chromatography (HPLC) Alliance system (Waters, Milford, MA, USA) consisting of a quaternary gradient pump, an online vacuum degasifier, an autosampler, and a column heating compartment, was used to separate the extract components and these separated components were then detected by a connected 2489 ultraviolet-visible (UV) detector. An Empower 3.0 workstation (Waters, Milford, MA, USA) was used for data acquisition and processing.

Chromatographic separation was fulfilled using a Waters Symmetry C₁₈ column (4.6 \times 250 mm, 5 μm) operated at 30 °C. The mobile phase was composed of 0.1% (v/v) glacial acetic acid

and 5 mmol L⁻¹ ammonium acetate aqueous solution (A) and acetonitrile (B). A linear gradient elution pattern was adopted keeping a constant flow rate of 0.4 mL min⁻¹, and the following elution program was used: 0–5 min, 5–5% B; 5–15 min, 5–10% B; 15–55 min, 10–26% B; 55–70 min, 26–27% B; 70–80 min, 27–30% B; 80–120 min, 30–46% B. The injection volume was 10 μL and the detection wavelength was set at 254 nm.

2.5. UPLC-Q-TOF instruments and conditions

An Agilent 1290 ultra-performance liquid chromatography (UPLC) Infinity system (Agilent Technologies, Palo, Alto, CA, USA), connected to a Bruker microTOF-Q II (Bruker Daltonics, Bremen, Germany) quadrupole time-of-flight mass spectrometer (Q-TOF-MS) with an electrospray ion source (ESI) was used for the qualitative identification of the components.

The separation conditions for UPLC were the same as those listed in Section 2.4. MS detection of the separated components was conducted in positive ion mode with a scan range from 100 *m/z* to 980 *m/z*. The other detailed conditions for MS analysis were as follows: the capillary voltage was set at 4.5 kV and the end plate offset voltage was -0.5 kV. The collision cell radio frequency was 100.0 Vpp and the nebulizer pressure was 0.12 MPa. The temperature of the dry heater was 190 °C and the flow rate of dry gas was 6.0 L min⁻¹. System control and data analysis were accomplished using a DataAnalysis 4.0 workstation (Bruker Daltonics, Bremen, Germany).

2.6. Antitumor efficacy test on the extract

MTT assays were performed to test the proliferation inhibition effects of the herb pair extracts of Fuzi-Gancao on the HeLa cervical cancer cell line. Three groups, namely a medicated group (all given the same amount of drug), a negative group not treated with drug solution, and a blank control group given only culture medium, were tested in parallel. Briefly, a certain amount of the prepared extract (see Section 2.2) was dissolved in sodium chloride (0.9%) by ultrasound and then centrifuged at 7000 rpm for 10 minutes. The supernatant liquor was filtered using a 0.22 μm nylon membrane, and the subsequent filtrate



was used as the mother solution (concentration 200 mg mL⁻¹ in terms of crude Fuzi). An accurately measured amount of mother solution (240 μL) was diluted with culture medium to 2 mL, to give a pharmaceutical medium of 24 mg mL⁻¹ in terms of crude Fuzi. Monolayer HeLa cells were digested with 0.25% trypsin and inoculated at a concentration of 5 × 10³ cells per well into 96-well plates containing RPMI 1640 complete medium and 10% FBS, and cultured at 37 °C, with saturated humidity, and 5% CO₂. After 24 hours, 100 μL of pharmaceutical medium was added into each well so that the final concentrations of crude Fuzi and Gancao in the cultivation system were 12 mg mL⁻¹ and 18 mg mL⁻¹, respectively, and then the cell culture was continued, keeping the conditions unchanged. After a further 48 hours, the supernatant culture solution was removed and 50 μL of MTT solution (1 mg mL⁻¹) was added into each well for generating formazan, and then the cell culture was continued as before. After 4 hours, the cultivation system was centrifuged and the supernatant liquor was discarded. Following this, 150 μL of DMSO solution was added into each well and the plate was slightly shaken until the generated formazan was dissolved thoroughly. The optical density (OD) of each well was measured using a Sunrise absorbance microplate reader (Tecan, Phoenix, CA, USA) at 490 nm, and the inhibition rate was calculated according to eqn (1). All experiments were repeated three times and the average value was calculated as the final inhibition rate of the sample.

$$\text{Inhibition rate} = \frac{\text{OD}_{\text{negative}} - \text{OD}_{\text{medicated}}}{\text{OD}_{\text{negative}} - \text{OD}_{\text{blank}}} \times 100\% \quad (1)$$

2.7. Model construction

2.7.1. Data preprocessing. The credibility of a CAR model greatly depends on the data quality. The peak area data is derived from the acquisition and integration of the chromatographic peaks, and, during this process, the sample preparation, instrument operation, experimental environment and other factors may inevitably introduce systematic and random errors, reducing the data accuracy; these errors must be effectively eliminated by smoothing the processing. Moreover, the levels of the same constituent in different samples, as well as the levels of all characteristic constituents in the same sample, vary within a large range, which may make it difficult to create a unified model; it is essential to amalgamate the raw data into a small range in the same reference system by normalization, which can accelerate the speed of iteration and enhance the subsequent search efficiency for optimal parameters.³⁰ Hence, prior to CAR model construction, the acquired data was pre-processed for smoothing and normalization by eqn (2) and (3), respectively.

$$x_{\text{smoothing},i} = \frac{1}{2m+1} \sum_{j=i-m}^{j=i+m} x_j \quad (2)$$

where m is the number of variables on each side of x_i ($m = 9$ in this research).

$$x'_i = (x_i - x_{\min}) / (x_{\max} - x_{\min}) \quad (3)$$

where x'_i and x_i are normalized data and raw data, respectively; and x_{\max} and x_{\min} are the maximum and minimum of each raw variable, respectively.

2.7.2. Model construction and optimization. BP and SVR were employed to construct the CAR model in this research. For the BP method, the number of neurons in the input layer and the output layer correspond to the number of independent variables and dependent variables, respectively. The number of neurons in the hidden layer is of crucial importance for the fitting capacity of a BP neural network, and in this paper, it was set according to an empirical equation (eqn (4)). A hyperbolic tangent function (tansig) and linear function (purelin) served as the transfer functions for the hidden layer and the output layer, respectively. Besides, the weight and bias of each neuron in the network were constantly adjusted to achieve favorable fitting results.

In general, SVR includes two methods, ϵ -SVR and ν -SVR, and the only difference between them is that the insensitive loss function ϵ is artificially set for the former whereas it is automatically adapted for the latter.³¹ Although ϵ -SVR is simple and convenient, the manual selection of an appropriate value for ϵ is not always easy, and ν -SVR usually gives a better performance owing to its flexible changeability, so ν -SVR was adopted in this paper. The gauss radial basis function eqn (5) was used as the kernel function in this research due to its good learning and generalization ability.³² Moreover, the penalty parameter C and kernel function parameter g significantly impact the model reliability and should be optimized. In this research, GA was utilized to optimize the parameters of both the BP model and the SVR model, and PSO was used only to optimize the SVR model.

$$n = \log_2 m \quad (4)$$

where n is the number of neurons in the hidden layer and m is the number of independent variables.

$$K(x_i - x_j) = \exp(-\gamma \|x_i - x_j\|^2), \quad \gamma > 0 \quad (5)$$

In this study, 31 different samples were assigned to two groups at random; 20 samples were used as a training set to create and train the CAR model, while the other 11 samples were used as a test set to examine the predictive ability of the established CAR model for unknown data. The root mean square error (RMSE) and correlation coefficient (R) were used to evaluate the accuracy and reliability of the developed CAR model:

$$\text{RMSE} = \left[\frac{\sum_{i=1}^n (y_i^{\text{experimental}} - y_i^{\text{predicted}})^2}{n} \right]^{1/2} \quad (6)$$



$$R = \frac{\sum_{i=1}^n (x_i - \bar{x})(y_i - \bar{y})}{\left[\sum_{i=1}^n (x_i - \bar{x})^2 \sum_{i=1}^n (y_i - \bar{y})^2 \right]^{1/2}} \quad (7)$$

where n is the sample number.

2.8. Mean impact value

The mean impact value (MIV), originally put forward by Dombi in 1995 to characterize the variation of weight matrix of neurons in a neural network, is regarded as one of the most important indicators reflecting the effect of each independent variable on dependent variables.³³ In this research, MIVs were used to evaluate the bioactivities of the constituents in the samples based on the developed CAR model. The calculation process can briefly be described as follows: each independent variable (x_j) is assigned new values, plus and minus 10%, to obtain two new input data sets, $X_{(j)1}([x_1, x_2, \dots, 1.1x_j, \dots, x_k])$ and $X_{(j)2}([x_1, x_2, \dots, 0.9x_j, \dots, x_k])$, which are presently imported to the optimal CAR model to give the two corresponding predicted inhibition rates, $Y_{(j)1}$ and $Y_{(j)2}$. The difference between the $Y_{(j)1}$ and $Y_{(j)2}$ values is called the impact value (IV), and is caused by the variation of the independent variable, x_j . The MIV of x_j is worked out through dividing the difference value by the sample number. The absolute MIV for each independent variable stands for the contribution of that constituent to the inhibition activity of the Fuzi-Gancao extract on HeLa cells.

Matlab R2013b software (MathWork Inc., Natick, MA, USA) integrated with a ga toolbox and a libsvm-3.1 toolbox was used as a platform for all of the above computer aided operations, including data preprocessing, CAR model training, and MIV calculation.

3. Results and discussion

3.1. HPLC-UV method development and validation

In order to acquire as much chemical composition information about the Fuzi-Gancao extract as possible, and to achieve the best separation of the contained constituents, the chromatographic column, detection wavelength, composition and flow rate of the mobile phase, column temperature, and elution program were all optimized; the optimum separation was fulfilled by the HPLC-UV conditions described in Section 2.4.

To ensure the effectiveness and reliability of the developed HPLC-UV method, the precision, stability and reproducibility of the method were validated. The precision experiment was carried out by analyzing the same sample six times in succession. To validate intra-day stability, the same sample was analyzed at 0 h, 4 h, 8 h, 12 h and 24 h. For reproducibility, six different samples from the same extract were prepared and analyzed using the established method. For all of these experiments, the relative retention time (RRT) and relative peak area (RPA) of 30 common chromatographic peaks were measured using the IS as the reference peak, and then relative standard deviations (RSDs) of RRTs and RPAs of each of the 30 peaks were calculated. As shown in Table 1, the RSDs of the RRTs and

RPAs of all of the characteristic peaks were no more than 5% in the experiments for precision, stability and reproducibility. This means that the developed HPLC-UV method is feasible and dependable, and the samples are stable for 24 hours.

3.2. HPLC analysis

When developing the HPLC method, several detection wavelengths were tested; at 254 nm, the chromatographic peaks had relatively moderate responses and better peak resolutions than at other wavelengths. Thus, 254 nm was chosen as the detection wavelength for the Fuzi-Gancao extract. Using the developed HPLC method, a representative fingerprint of the Fuzi-Gancao extract was acquired and is shown in Fig. 2, which contains more than 70 chromatographic peaks. Among these peaks, some peaks existed in all or most of the 31 batches of Fuzi-Gancao extract; they were marked as common peaks and investigated, while some other peaks only existed in some of the batches of Fuzi-Gancao extract, and they were not investigated in this paper. By aligning the fingerprints of 31 batches of samples using the Similarity Evaluation System for Chromatographic Fingerprint of TCM by the Chinese Pharmacopoeia Commission, as marked in Fig. 2, 30 characteristic peaks were identified as common peaks.

RPAs of the 30 common peaks from all batches of the samples were measured and the data are listed in Table S1.† It can be seen that the contents of the chemical compounds in the different samples differed hugely. Taking peak 16 for instance, among the 31 batches of samples, the minimum RPA was 0.177 in sample 6 while the maximum RPA was 7.424 in sample 22. This is not an exception but a general case. These conspicuous differences usually result from various factors like the growing environment, harvesting time, post-processing and so on. These very different chemical compositions provide a solid material data basis for constructing a high-quality CAR model.

3.3. Peak identification by tandem mass spectrometry

A prepared sample that contained all the marked chromatographic peaks was subjected to UPLC-Q-TOF, and the 30 common peaks were tentatively characterized according to the m/z values of the parent ions and characteristic fragment ions in MS and MS/MS spectra, in comparison with those data in the published literature. The detailed identification information is listed in Table 2, and the structural formulas are shown in Fig. S1.†

For these chromatographic peaks of interest, there is only one component (peak 20) that has not been identified. According to the identification results, 18 components (peaks 1–13, 15, 18, 19, 22, 25) belong to Fuzi, which comprise three types of alkaloids, diester diterpenoid alkaloids (DDA), monoester diterpenoid alkaloids (MDA) and alkyolamine diterpenoid alkaloids (ADA); and the other 11 components originate from Gancao, containing two kinds of compounds, flavonoids and saponins.



Table 2 Identification of the chromatographic peaks in the fingerprint of the Fuzi–Gancao extract

No.	t_R /min	MS	MS/MS	Error (ppm)	Formula	Identification
1	16.8	394.5073[M + H] ⁺	376.2489, 358.2396, 340.2323, 328.2262, 322.2198	4	C ₂₂ H ₃₅ NO ₅	Chuanfumine
2	21.2	440.2661[M + H] ⁺	422.2511, 408.4881	−4	C ₂₃ H ₃₇ NO ₇	9-Hydroxysenbusine A
3	21.7	378.4667[M + H] ⁺	360.2534, 342.2443, 332.2296, 300.7898	−0.9	C ₂₂ H ₃₅ NO ₄	KaraKoline
4	22.7	486.4962[M + H] ⁺	468.2523, 454.2445, 436.2346, 422.2274, 404.2056	−2.4	C ₂₄ H ₃₉ NO ₉	Mesaconine
5	24.0	424.4683[M + H] ⁺	406.2590, 388.2472, 374.2306	−0.8	C ₂₃ H ₃₇ NO ₆	Senbusine A/B
6	25.3	408.5207[M + H] ⁺	390.2637, 372.2516, 358.2383, 340.2322	−4.9	C ₂₃ H ₃₇ NO ₅	Isotalatizidine
7	26.6	500.5257[M + H] ⁺	482.2795, 468.2628, 450.2505, 436.2210, 418.2287	0.9	C ₂₅ H ₄₁ NO ₉	Aconine
8	27.7	358.4685[M + H] ⁺	340.2277, 322.2143	5.8	C ₂₂ H ₃₁ NO ₃	Songorine
9	30.7	470.4375[M + H] ⁺	438.2501, 420.2327, 406.2375, 388.2110	−1.2	C ₂₄ H ₃₉ NO ₈	Hypaconine
10	31.3	454.5523[M + H] ⁺	436.2706, 418.2640, 404.2458, 386.2331	−4.7	C ₂₄ H ₃₉ NO ₇	Fuziline
11	33.4	438.5300[M + H] ⁺	420.2749, 402.2658, 388.2500, 370.2386, 356.2235	−1.3	C ₂₄ H ₃₉ NO ₆	Neoline
12	34.9	450.4916[M + H] ⁺	432.2742, 422.2562, 414.2669, 404.2507, 390.2517	−2.7	C ₂₃ H ₃₉ NO ₆	Kondelfin
13	47.2	464.5773[M + H] ⁺	432.2741, 414.2627, 400.2456, 372.2519, 358.2419, 340.2204, 322.2145	3.9	C ₂₆ H ₄₁ NO ₆	14-Acetyltalatizamine
14	48.5	257.2084[M + H] ⁺	137.0216, 119.0490	1.2	C ₁₅ H ₁₂ O ₄	Liquiritigenin
15	49.2	606.5307[M + H] ⁺	588.2766, 574.2666, 556.2552, 542.2361, 524.2215, 506.2155	0.3	C ₃₁ H ₄₃ NO ₁₁	14-Benzoyl-10-hydroxymesaconine
16	49.8	551.1760[M + H] ⁺	419.1353, 257.2213, 137.0227, 119.0501	1.8	C ₂₆ H ₃₀ O ₁₃	Liquiritin apioside
17	50.9	419.1329[M + H] ⁺	257.1971, 137.0237, 119.0496	1.9	C ₂₁ H ₂₂ O ₉	Liquiritin
18	58.8	590.6342[M + H] ⁺	572.2874, 558.2707, 540.2597, 526.2448, 508.2342, 494.2155	0.8	C ₃₁ H ₄₃ NO ₁₀	Benzoylmesaconine
19	63.4	604.6657[M + H] ⁺	586.3044, 572.2835, 554.2715, 540.2547, 522.2469	0.6	C ₃₂ H ₄₅ NO ₁₀	Benzoylaconine
20	64.5	543.5424	—	—	—	Unknown
21	65.5	419.1369[M + H] ⁺	389.9859, 239.0720, 137.0219	5.5	C ₂₁ H ₂₂ O ₉	Isoliquiritin
22	67.3	574.7080[M + H] ⁺	542.2746, 510.2440, 492.2435, 478.2282	−1.8	C ₃₁ H ₄₃ NO ₉	Benzoylhypaconine
23	68.2	269.2086[M + H] ⁺	237.0521, 169.0647	1.7	C ₁₆ H ₁₂ O ₄	Formononetin
24	71.6	855.6927[M + K] ⁺	679.3701, 503.3346, 485.3246, 467.3159, 449.3103	0.1	C ₄₂ H ₆₄ O ₁₆	Licorice saponin J2
25	73.1	558.5207[M + H] ⁺	540.2869, 526.2824, 508.2663	2.2	C ₃₁ H ₄₃ NO ₈	14-Benzoyl-deoxyaconine
26	85.1	469.5422[M + H] ⁺	379.2757	−0.4	C ₃₀ H ₄₄ O ₄	Glabrolide
27	97.3	839.4065[M + H] ⁺	663.3755, 487.3440, 469.3316, 451.3204	−0.7	C ₄₂ H ₆₂ O ₁₇	Licorice saponin G2
28	101.2	487.3388[M + H] ⁺	469.3329, 451.3196, 439.3216, 423.3244, 405.3087	−1.9	C ₃₀ H ₄₆ O ₅	24-Hydroxy glycyrrhetic acid
29	103.4	823.4108[M + H] ⁺	647.3800, 471.3416, 453.3373, 435.3268	2.7	C ₄₂ H ₆₂ O ₁₆	Glycyrrhizic acid
30	108.1	453.5640[M + Na] ⁺	257.1911, 217.1702, 204.1916	0.4	C ₂₂ H ₂₂ O ₉	Ononin

3.4. Antitumor efficacy of Fuzi–Gancao extract

In our previous study, an extract of 1 : 1.5 (weight ratio) of Fuzi and Gancao showed a noticeable antitumor effect. In this paper,

the inhibition rates on HeLa cells of 31 batches of samples were measured by MTT assays, as listed in Table S2.† As is shown in Fig. 3, most samples could strongly prevent the growth of HeLa

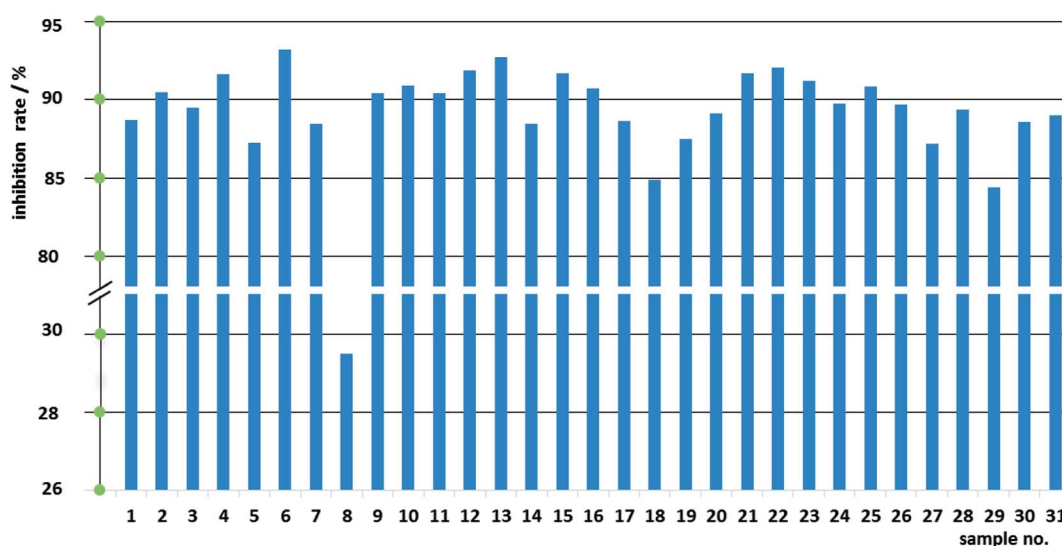


Fig. 3 Inhibition rates of 31 batches of Fuzi–Gancao extract on HeLa cell proliferation.



Table 3 The results of RMSE and *R* of the three CAR models

Model	Training set		Test set	
	RMSE ^a	<i>R</i> ^b	RMSE	<i>R</i>
BP-GA	0.001	1.000	0.0052	0.9413
SVR-GA	0.1902	1.000	0.1969	0.9822
SVR-PSO	0.0006	1.000	0.0082	0.9879

^a RMSE: root mean square error. ^b *R*: correlation coefficient.

cells. At the same weight ratio (2 : 3) of Fuzi and Gancao and the same dose of 15 mg mL⁻¹ (with respect to crude Fuzi), different samples exerted obviously different inhibition effects against HeLa cells: the lowest and the highest inhibition rates were 29.50% (sample 8) and 93.21% (sample 6), respectively. The significant differences in anticancer efficacy among the 31 different samples work in concert with the diverse chemical compositions, and imply that these data are suited to the construction of a CAR model.

3.5. Model comparison and selection

The data in Tables S1 and S2† were used to create the independent variable matrix $X_{31 \times 30}$ and the dependent variable matrix $Y_{31 \times 1}$, and three models were constructed by different algorithms to fit the relationship between these two groups of data. The RPAs of the 30 concerned chromatographic peaks were imported to Matlab and the corresponding inhibition rates of the extracts were the theoretical output values of the CAR model. The RMSE and *R* of the training set and test set of the three investigative models are listed in Table 3. For the training set, the three models have the same *R* values, and BP-GA and SVR-PSO have similar RMSEs that are both much lower than the RMSE of SVR-GA, suggesting that the former two models have better fitting precision than SVR-GA. For the test set, the RMSEs of BP-GA and SVR-PSO are still nearly the same and lower than that of SVR-GA, however the *R* of SVR-PSO is far higher than that

of BP-GA, indicating that SVR-PSO has the stronger prediction and generalization capability. Comparatively speaking, SVR-PSO performs best in describing the complicated composition–activity relationship for the Fuzi–Gancao extract. The predictive regression curves of the training set and test set by the SVR-PSO model with best $C = 14.3265$ and best $g = 0.1$ are shown in Fig. 4. Clearly, the predicted values correspond with the original experimental inhibition rates very well in both the training set and the test set, indicating that the established SVR-PSO model combines high fitting accuracy with good generalization capability and is capable of describing the relationship between chemical composition and pharmacological activity of Fuzi–Gancao extract.

3.6. Antitumor component recognition by MIV

Based on the superior SVR-PSO model, the MIV method was employed to identify the active antitumor components from the Fuzi–Gancao extract. MIVs of the 30 common peaks from the 31 different samples were acquired as described in Section 2.8 and the 30 peaks were then ranked by their absolute MIVs, as shown in Table 4. A higher absolute MIV means that the corresponding component contributes more to the inhibition effect of the Fuzi–Gancao extract on HeLa cells, and therefore, the top 8 components (peaks 17, 25, 22, 13, 23, 28, 5, 7) are temporarily deemed to possess good antitumor activity.

The top 8 components are liquiritin, 14-benzoyl-deoxyaconine, benzoylhypaconine, 14-acetylaltatizamine, formononetin, 24-hydroxy glycyrrhetic acid, senbusine A/B and aconine. Of these, five components (peaks 25, 22, 13, 5, 7) are from Fuzi and three components (peaks 17, 23, 28) are from Gancao. Among the 8 components, there are not only high level components like formononetin (peak 23), but also lower level components like 14-benzoyl-deoxyaconine (peak 25). Indeed, the ranking of 14-benzoyl-deoxyaconine is even higher than that of formononetin, meaning that the recognition results do not rely on the component content, which is beneficial for finding

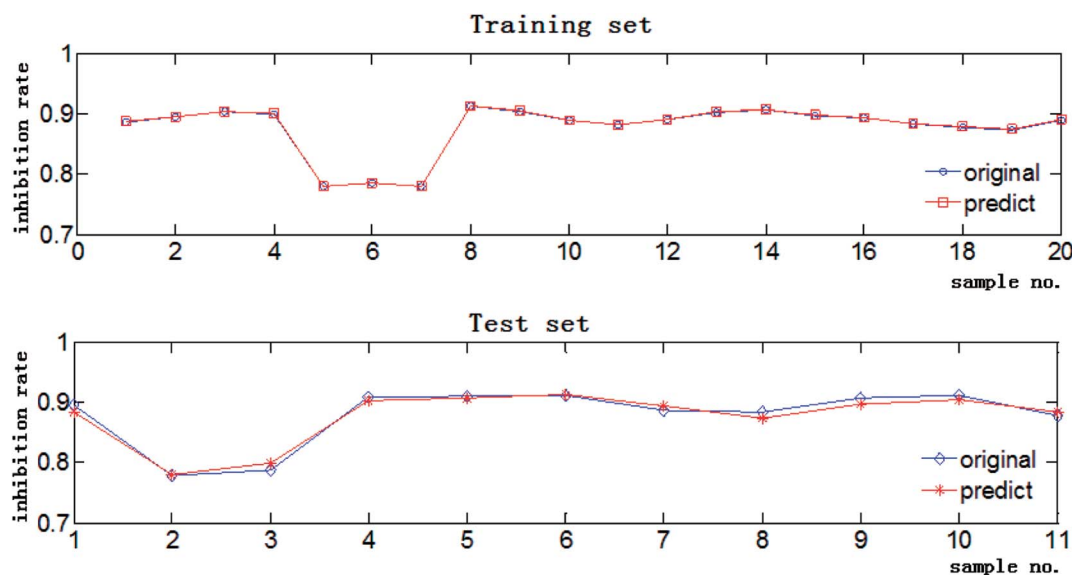


Fig. 4 The predictive regression curves of the SVR-PSO model.



Table 4 Absolute MIV and ranking of the 30 chromatographic peaks of the Fuzi–Gancao extract^a

Peak	Absolute MIV	Ranking	Peak	Absolute MIV	Ranking
1	0.0008	28	16	0.0015	25
2	0.0102	15	17	0.0424	1
3	0.0042	19	18	0.0109	14
4	0.0016	24	19	0.0012	26
5	0.0184	7	20	0.0160	9
6	0.0008	29	21	0.0009	27
7	0.0169	8	22	0.0293	3
8	0.0137	10	23	0.0247	5
9	0.0002	30	24	0.0042	20
10	0.0022	23	25	0.0315	2
11	0.0113	13	26	0.0028	22
12	0.0121	11	27	0.0072	16
13	0.0284	4	28	0.0238	6
14	0.0060	17	29	0.0029	21
15	0.0120	12	30	0.0051	18

^a MIV: mean impact value.

those low level compounds with high activity. Most of the recognized components have been confirmed to have prominent anti-tumor efficacy in the published literature. For example, liquiritin (peak 17) is a major flavonoid constituent in *Glycyrrhizae Radix* and it was found that liquiritin could effectively suppress the proliferation and migration of human gastric cancer cells which are resistant to cisplatin, and, furthermore, the combined therapy of liquiritin and cisplatin exhibited potent inhibition activity against cisplatin-resistant gastric cancer cells through multiple pathways.³⁴ Moreover, the component corresponding to peak 22 was identified as benzoylhypaconine, which has been reported to possess selective cytotoxicity against the MGC80 gastric cancer cell line with a low IC₅₀ value of 24.7 μg mL⁻¹.³⁵

4. Conclusions

An integrated strategy was efficiently applied to recognize antitumor components from a mixed extract of Fuzi–Gancao. Firstly, desirable separation was fulfilled by HPLC for the constituent ingredients in the Fuzi–Gancao extract, and 30 common chromatographic peaks from 31 different samples were analyzed quantitatively and qualitatively. Secondly, three CAR models were established using different algorithms to interpret the relationship between the chemical composition and the antitumor effect of the Fuzi–Gancao extracts, and the SVR-PSO model stood out after consideration of the fitting precision and prediction ability. Furthermore, the antitumor activities of the 30 identified compounds were evaluated by MIV, and 8 components with high absolute MIVs were identified as promising anticancer compounds. This proposed strategy provides an attractive approach to the identification of potent antitumor compounds. It will benefit drug-discovery from herbal medicines, and the identified compounds provide a reference for early antitumor pharmaceutical screening.

Conflicts of interest

The authors declare no competing financial interest.

Non-standard abbreviations

BP	Back propagation
CAR	Composition–activity relationship
GA	Genetic algorithm
MIV	Mean impact value
MLR	Multiple linear regression
PSO	Particle swarm optimization
RPA	Relative peak area
RRT	Relative retention time
SVR	Support vector regression

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