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Semi-synthesis, structural modification and biological evaluation of 5-arylbenzofuran neolignans†

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5-Arylbenzofuran neolignans, a newfound class of natural products, were reported to possess several kinds of pharmacological activities. To solve the lack of natural sources and promote the research of 5-arylbenzofuran neolignans in all fields, an available semi-synthesis methodology of 5-arylbenzofuran neolignans was developed, and a detailed structural modification was conducted. In the meantime, a one-pot process of Waker-type cyclization and Wacker-type oxidation was developed. To explore the potential of 5-arylbenzofuran neolignans as bioactive substances, 5-arylbenzofuran neolignans and their derivatives were evaluated for their cytotoxicity. As a result, a preliminary structure–activity relationship was obtained. Most derivatives revealed low cytotoxic effects suggesting that they were relatively safer than the natural 5-arylbenzofuran neolignan. Several derivatives showed high cytotoxicities which were found to be closely associated with apoptosis-inducing. The selectivity assay for cytotoxicity showed tumor cells were more sensitive to the promising compounds than normal cells.

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Introduction

Neolignans are a large class of natural products, which consist of two propylbenzene (C₆C₃) units.¹ Benzofuran neolignans, a significant kind of neolignans, have drawn great interest due to their special skeleton and pharmacological activities. So far, the research of benzofuran neolignans has mainly focused on 2-arylbenzofuran neolignans (Fig. 1). Lots of 2-arylbenzofuran neolignans, such as ailanthoidol,² eupomatenoide 7,³ licarin A⁴ and acuminatin,⁵ have been proved to possess excellent biological activities.

In recent years, some 5-arylbenzofuran neolignans (Fig. 1) have come into sight with favorable anti-inflammatory,^{6,7} anti-microbial,⁸ and anti-influenza activities.⁹ Due to the difference in the bonding position of the two aromatic rings, 5-arylbenzofuran neolignans may have quite different biological effects compared to 2-arylbenzofuran neolignans. However, the low content in natural products and the difficulty of chemical synthesis have blocked the research and development of 5-arylbenzofuran neolignans.

In order to solve the problem of a lack of a natural source of 5-arylbenzofuran neolignans, we developed a semi-synthesis methodology utilizing honokiol, a high-yield natural product whose total synthesis methodologies are various, as the starting material (Fig. 1). To promote the study of 5-arylbenzofuran neolignans in all fields, we made structural modification and investigated their cytotoxicity. In addition, inspired by the achievements of 2-arylbenzofuran neolignans in tumor therapy,¹⁰ 5-arylbenzofuran neolignan derivatives with high cytotoxicities were further evaluated for their anti-proliferative effects and mechanisms.

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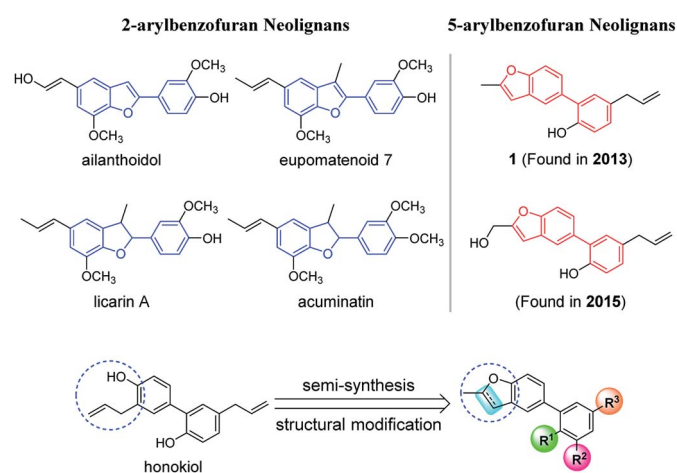


Fig. 1 The comparison of 5-arylbenzofuran neolignans with 2-arylbenzofuran neolignans, and the semi-synthesis methodology and structural modification of 5-arylbenzofuran neolignans.



Results and discussion

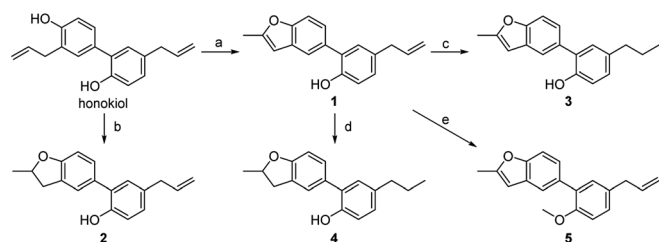
Chemistry

It is of great difficulty to totally synthesize 5-arylbenzofuran neolignans because the two propylbenzene units are directly connected by benzenes. Nonetheless we found there are a number of natural products containing the structure of 2-allyl-4-arylphenol which could be intramolecularly cyclized to afford 5-arylbenzofuran neolignans. In this paper, honokiol, a high-yield natural product with the structure of 2-allyl-4-arylphenol, was selected as the starting material (Scheme 1).

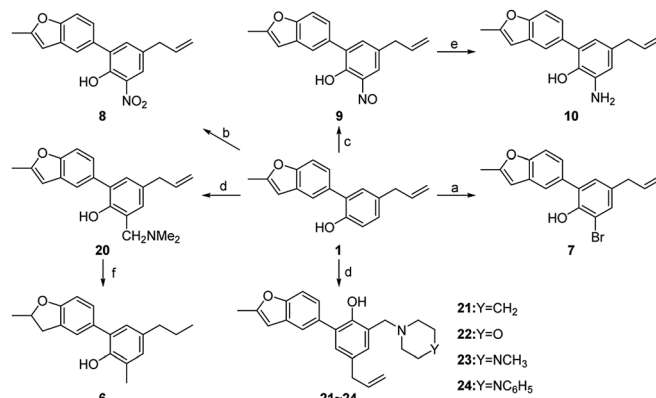
5-Arylbenzofuran neolignan **1**, which can only be extracted from natural product before,⁶ was synthesized starting from honokiol by Wacker-type intramolecular cyclization¹¹ in a high yield. For investigating the influence by furan ring, dihydrofuran derivative **2** was prepared through acid catalytic cyclization from honokiol in the presence of concentrated sulfuric acid.¹² To discuss the effects of phenolic hydroxyl and allyl group, we synthesized **3–5** by the method depicted in Scheme 1. Compound **3** and **4** were obtained by hydrogenation reduction of **1** using NaBH₄ (ref. 13) or H₂ (ref. 14) as hydrogen source respectively, while compound **5** was generated *via* the etherification of **1**.

In order to study the influences by different substituents on benzene ring, compound **7–26** with diverse substituents were designed and prepared. Among them, compounds **6–10** and **20–24** were synthesized from **1** according to the method described in Scheme 2. The bromination reaction of **1** provided compound **7** using NBS as the brominating reagent¹⁵ and the nitration of **1** prepared compound **8** by classic procedure. Compound **10** was synthesized *via* reduction reaction with zinc powder as a catalyst from **9**, which was prepared by nitrosation from **1**. Compounds **20–24** were obtained through Mannich reaction of **1** with formalin and appropriate secondary amines.¹⁶ And compound **6** was generated by hydrogenation reduction of **20** catalyzed by palladium-carbon.

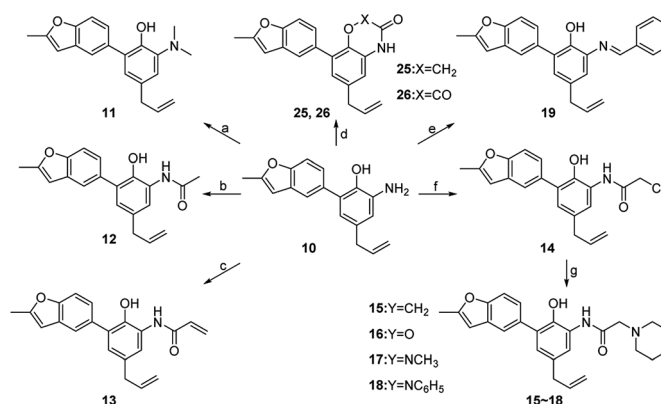
Compounds **11–19**, **25** and **26** were prepared from **10** according to the approach presented in Scheme 3. Compound **11** was produced *via* methylation of **10**. Acylamides **12–14** were obtained from **10** by acylation reaction with acetic anhydride, acryloyl chloride or chloroacetyl chloride, respectively. Aminoacetamides **15–18** were synthesized through reaction between compound **14** and appropriate secondary amines.¹⁷ Lactams **25** and **26** were prepared by treatment of **10** with chloroacetyl chloride¹⁸ or oxalyl chloride¹⁹ in the presence of 4-dimethylaminopyridine and triethylamine in THF, respectively.



Scheme 1 Reagents and conditions: (a) PdCl₂, NaOAc, O₂, DMA/H₂O, 60 °C; (b) H₂SO₄, DCE, 50 °C; (c) NaBH₄, NiCl₂·6H₂O, EtOH, r.t.; (d) Pd/C, H₂, MeOH, reflux; (e) CH₃I, KOH, THF, r.t.

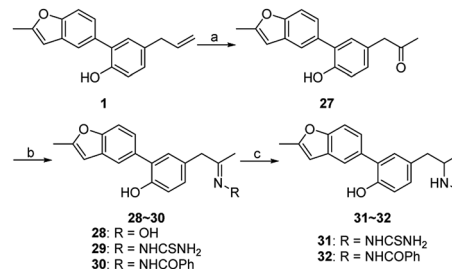


Scheme 2 Reagents and conditions: (a) NBS, MeCN, r.t.; (b) HNO₃, HOAc, CH₂Cl₂, 0 °C; (c) NaNO₂, concentrated hydrochloric acid (36–38%), MeCN/H₂O, r.t.; (d) secondary amines, HCHO (37%), MeOH, 60 °C; (e) Zn, NH₄Cl, HOAc, EtOH/H₂O, r.t.; (f) Pd/C, H₂, MeOH, reflux.



Scheme 3 Reagents and conditions: (a) CH₃I, NaOAc, DMF, r.t.; (b) acetic anhydride, K₂CO₃, ethyl acetate, r.t.; (c) acryloyl chloride, K₂CO₃, CH₂Cl₂, r.t.; (d) chloroacetyl chloride or oxalyl chloride, DMAP, NEt₃, THF, r.t. to reflux; (e) benzaldehyde, acetic acid, EtOH, r.t.; (f) chloroacetyl chloride, K₂CO₃, CH₂Cl₂, r.t.; (g) secondary amines, NEt₃, THF, r.t.

For further investigation on the influences of side chain, groups with large polarity or steric hindrance were introduced, and compounds **27–32** were synthesized *via* the route outlined in Scheme 4. Compound **27** was generated from **1** by Wacker oxidation adopting the condition reported previously.¹¹ The condensation reaction of **27** with appropriate primary amines



Scheme 4 Reagents and conditions: (a) PdCl₂, O₂, DMA/H₂O, 60 °C; (b) R-NH₂, EtOH, r.t.; (c) NaBH₄, EtOH, r.t.



prepared compounds **28–30**,^{20–22} and the reduction of **29** and **30** afforded compounds **31** and **32**, respectively.²³

In the meantime, a one-pot method of Wacker-type cyclization and Wacker-type oxidation was developed. The synthetic process was supplied in the Experimental section, and the discussion was provided in the ESI.†

Cytotoxicity and SAR analysis

The cytotoxicity of 5-arylbenzofuran neolignans (**1–32**) were evaluated *via* MTT assay against A549 (human lung carcinoma), K562 (human myelogenous leukemia) and HepG2 (human hepatocellular liver carcinoma) tumor cell lines described with IC₅₀ (concentration required for 50% inhibition). For a clear view, the IC₅₀ values of tested compounds were presented in Tables 1–3 according to their structural differences. It's found that most derivatives showed no cytotoxic effects against three tumor cell lines (IC₅₀ > 128 μM) which suggested they were relatively safer than the natural 5-arylbenzofuran neolignan. Several compounds, such as **23**, **26** and **30**, displayed obviously higher cytotoxicity than the natural 5-arylbenzofuran neolignan **1**. In addition, the A549 and K562 cells were more susceptible to 5-arylbenzofuran neolignans than HepG2 and the structural modifications caused appreciable impact on cytotoxicity.

As shown in Table 1, 5-arylbenzofuran **1** and **3** displayed higher cytotoxicity than 5-aryldihydrobenzofuran **2**, **4** and **6**. The reduction of allyl group made little difference to activity when comparing compound **1** and **3** with compound **2** and **4**. Phenolic hydroxyl was essential to the cytotoxicity as compound **5** possessing very low cytotoxicity against three tumor cells. In view of the above results, further research was carried out around the substructure of 5-(phenol-2-yl)benzofuran due to its comparatively good anti-proliferative activity.

In order to discuss the influences by substituents on benzene ring, compounds **7–26** were evaluated for the cytotoxicities and

the results were displayed in Table 2. Compounds with simple substituents such as bromo, nitro, nitroso, amino and dimethylamino (**7–11**) showed no cytotoxicity. Acylamides (**12–14**) seemed to have equipotent cytotoxicity compared to the natural product **1** while aminoacetamides (**15–18**) showed lower cytotoxicity except substituted by 4-methylpiperazine (**17**). Similar rule appeared in

Table 2 IC₅₀ values of compound **7–26** against HepG2, A549 and K562 cell lines

Compd.	R	IC ₅₀ (μM)		
		A549	K562	HepG2
7	Br	>128	>128	>128
8	NO ₂	>128	>128	>128
9	NO	>128	>128	>128
10	NH ₂	>128	>128	>128
11	N(CH ₃) ₂	>128	>128	>128
12	NHCOCH ₃	88.2 ± 3.4	80.0 ± 5.1	87.4 ± 6.1
13		53.0 ± 5.7	34.9 ± 1.2	63.8 ± 3.1
14		65.0 ± 2.0	58.5 ± 0.5	>128
15		>128	>128	>128
16		>128	>128	>128
17		75.4 ± 2.2	61.2 ± 5.6	98.5 ± 7.8
18		>128	>128	>128
19		>128	>128	>128
20		>128	87.78 ± 13.9	>128
21		>128	>128	>128
22		>128	>128	>128
23		25.1 ± 4.4	29.6 ± 2.5	36.8 ± 0.2
24		>128	>128	>128
25		53.4 ± 5.6	41.8 ± 5.2	>128
26		32.4 ± 1.7	18.0 ± 2.6	75.4 ± 0.3

Table 1 IC₅₀ values of compounds **1–6** against HepG2, A549 and K562 cell lines

Compd.	IC ₅₀ (μM)		
	A549	K562	HepG2
1	71.5 ± 1.94	67.8 ± 8.3	107.2 ± 12.7
2	115.5 ± 2.0	126.2 ± 1.5	125.6 ± 2.2
3	61.9 ± 2.7	73.9 ± 7.3	88.2 ± 8.0
4	96.1 ± 7.0	72.8 ± 8.9	121.6 ± 2.5
5	>128	>128	>128
6	84.9 ± 7.8	102.3 ± 15.5	>128



Table 3 IC₅₀ values of compound 27–32 against HepG2, A549 and K562 cell lines

Compd.	R ¹ or R ²	IC ₅₀ (μM)		
		A549	K562	HepG2
27	O	>128	>128	>128
28	N-OH	125.1 ± 12.2	>128	>128
29		64.7 ± 7.4	31.8 ± 4.4	92.1 ± 6.2
30		22.0 ± 2.1	29.0 ± 1.8	26.5 ± 3.5
31		103.8 ± 5.5	48.1 ± 3.4	80.9 ± 1.5
32		97.4 ± 8.0	66.8 ± 0.7	106.1 ± 13.6
5-FU ^a		34.3 ± 2.1	81.9 ± 7.4	>128
PTX ^b		21.0 ± 3.0	4.6 ± 0.7	16.5 ± 2.2

^a 5-FU: 5-fluorouracil. ^b PTX: paclitaxel.

mannich bases (20–24), that is the mannich base of 4-methylpiperazine (23) exhibited the highest cytotoxic effect which is much higher than the natural compound 1. Different from compound 5, lactams 25 and 26 showed relatively high cytotoxicity even though neither of their hydroxyl group was free. As a consequence, the structural modifications around phenolic hydroxyl group would have obvious impacts on cytotoxicity.

For further investigation on the modification of side chain, compounds 27–32 were synthesized and their cytotoxicities were exhibited in Table 3. As a result, compounds 29–32 presented higher cytotoxicity than 27 and 28, while 30 showed the highest cytotoxicity. For this position, the introduction of groups with non-polarity or large steric effect may enhance the cytotoxicity.

In addition, the selectivity for cytotoxicity was further studied by testing the cytotoxicity against human umbilical vein endothelial cell (HUVEC). As shown in Table 4, the selectivities of promising compounds 23 and 30 were 2.02 and 2.50 respectively, which were higher than the positive control paclitaxel. As already

Table 4 The selectivity of promising compounds

Compd.	IC ₅₀ (μM)		Selectivity ^a
	A549	HUVEC	
1	71.5 ± 1.94	134.4 ± 18.8	1.88
23	25.1 ± 4.4	50.8 ± 5.7	2.02
30	22.0 ± 2.1	55.1 ± 6.2	2.50
PTX ^b	21.0 ± 3.0	8.6 ± 1.6	0.41

^a Selectivity is defined as IC₅₀ (HUVEC)/IC₅₀ (A549). ^b PTX: paclitaxel.

reported, some human normal cells were more sensitive to paclitaxel than tumor cells,²⁴ and it's also a common problem for traditional anticancer drugs.²⁵ The promising compounds 23 and 30 seemed to possess certain advantage in the respect.

Morphological analysis

Microscopic observations were used to investigate the changes of cellular morphology and prove the anti-proliferative activities of compounds with high cytotoxicity. As displayed in Fig. 2A, no apparent changes of cell morphology were observed after HepG2 cells were treated with 20 μM of compound 1 for 24 h. However, HepG2 cells treated with compounds 23 and 30 displayed some morphological changes, such as cell shrinkage, rounding, loss of contact with neighbouring cells and detachment from plate. And the compound 30 had a greater effect on cell morphologies than that of compound 23, which was consistent with their cytotoxicities.

Apoptosis assessment by Hoechst 33258 staining

In order to observe the changes of the nuclear morphology, the HepG2 cells were stained by Hoechst 33258 and the results were exhibited in Fig. 2B. In the control group it's showed slightly blue and homogeneous fluorescence, and there was no obvious changes of fluorescence in the cells treated with compound 1 at the concentration of 20 μM. However, in the groups treated with 23 or 30, there was a lot of enhanced fluorescence which means karyopyknosis and chromatin condensation in apoptotic cells. It's suggested that compounds 23 and 30 could induce apoptosis of HepG2.

Apoptosis assessment by flow cytometry analysis

To clearly analyze the apoptosis-inducing effects of the modified compounds, the apoptosis ratios induced by compound 1, 23 and 30 in HepG2 cells were evaluated by flow cytometry analysis (Fig. 3). The result showed that compound 1 have little influence on the apoptosis of HepG2 compared with the control group. Compounds 23 and 30 caused more apoptosis than compound 1 at the same concentration, and it was consistent with the results of the Hoechst 33258 staining assay.

mRNA expression in HepG2 cells

Furthermore, for studying the mechanism of apoptosis-inducing caused by modified compounds, the expression levels of mRNAs related to the mitochondria-mediated apoptosis pathway were analyzed by qRT-PCR assay. The mRNA expression levels of *caspase-3*, *caspase-9*, *Bax*, *Bcl-2*, *p53* and *p21* in HepG2 cells were displayed in Fig. 4. Caspase-3 and caspase-9 contributes a major role in mitochondria-mediated apoptotic pathway.²⁶ And in the HepG2 cells treated with compound 23, the mRNA expression level of *caspase-3* and *caspase-9* was about twice the level of control group (Fig. 4A), while the mRNA expression level of *caspase-9* and *caspase-3* have no change treated with compound 1 and the *caspase-3* expression increased treated with compound 30.



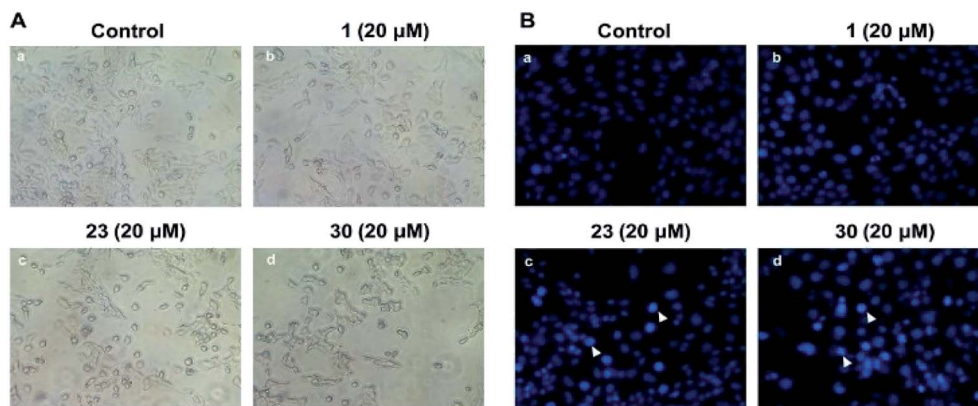


Fig. 2 (A) Cellular morphologies of HepG2 cells treated with different compounds (1, 23, 30) for 24 h. (B) Images of Hoechst 33258 stained HepG2 cells which were treated with 20 μM of different compounds (1, 23, 30) for 24 h. The arrows indicated the apoptotic cell. Original magnification is 400 \times .

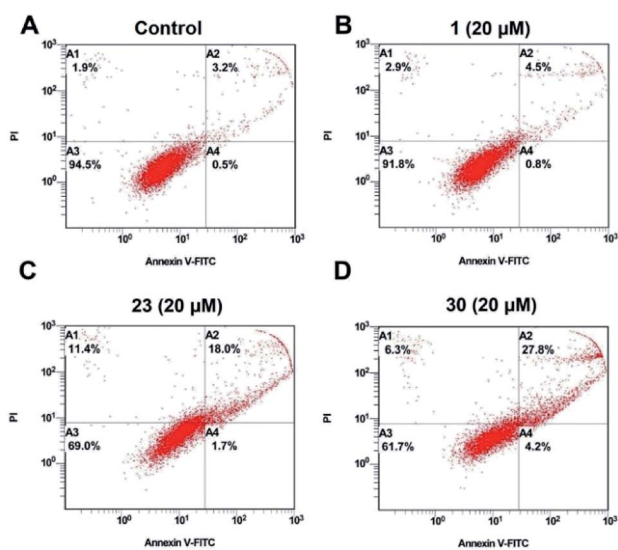


Fig. 3 Apoptosis effect of compounds 1, 23 and 30 at 20 μM in HepG2 cells for 24 h. The cells were stained with Annexin V-FITC/P and followed by flow cytometry analysis.

activation of caspases and led to apoptosis.²⁸ However, it made no difference for *Bax/Bcl-2* ratio after treated with compound 1 and 30.

In the previous studies, *p53* and its downstream gene *p21* were also found able to cause apoptosis *via* the regulation of *Bax/Bcl-2* (ref. 29 and 30) and the activation of caspases.³¹ In addition, *p53* and *p21* played a key role in cell cycle regulation.³² As shown in Fig. 4C, the mRNA expression level of *p53* and *p21* in the HepG2 cells treated with compound 23 were about 2 times higher than the control.

In summary, the compound 23 induces apoptosis of HepG2 cells through the regulation of caspase-dependent *p53*-dependent and *Bax/Bcl-2* pathway. However, compound 30 also induce the expression of *caspase-3*, but had no effect on *Bax/Bcl-2* activation pathway. It is suggested that compound 30 induces apoptosis mainly by other signal pathways. Therefore, different structural modifications of 5-arylbenzofuran neolignan would induce apoptosis by regulation of different signal pathways.

Experimental section

General methods and materials

All the reagents were commercially available and used without further purification. Melting points were determined on an X-4 binocular microscope melting point apparatus. ¹H NMR and

The activation of caspase-dependent apoptosis pathway could be regulated by *Bcl-2* family members such as *Bax* and *Bcl-2*.²⁷ In Fig. 4B, the *Bax/Bcl-2* ratio of compound 23 treated HepG2 cells increased obviously, which always resulted in the

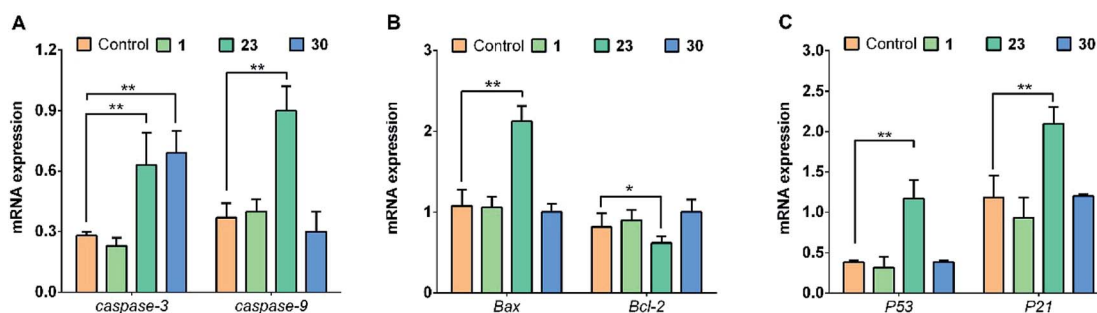


Fig. 4 mRNA expression levels of (A) *caspase-3*, *caspase-9*, (B) *Bax*, *Bcl-2*, (C) *p53* and *p21* in HepG2 cells treated with 20 μM of different compounds (1, 23, 30) for 24 h. * $P < 0.05$, ** $P < 0.01$.



^{13}C NMR spectra were recorded on a Bruker AV-400 spectrometer, using tetramethylsilane (TMS) as the internal standard and chemical shifts (δ) were expressed in ppm. Mass spectra were obtained by an Agilent 1100 series LC-MS. Elemental analyses were performed on a Vario EL III (Germany) instrument.

Synthesis of title compounds

4-Allyl-2-(2-methylbenzofuran-5-yl)phenol (1). A mixture of honokiol (10 mmol), NaOAc (1.0 mmol) and PdCl_2 (0.15 mmol) in the co-solvent of DMA and H_2O (35 mL, v/v = 6 : 1) was stirred under O_2 (0.8 Mpa) at 60 °C for 16 h. After cooling, the mixture was diluted with water and extracted with ethyl acetate. The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **1** as light yellow oil (86%). ^1H NMR (400 MHz, CDCl_3) δ 7.53 (d, J = 1.5 Hz, 1H, benzofuran 4 H), 7.50 (d, J = 8.4 Hz, 1H, benzofuran 7 H), 7.26 (dd, J = 8.4, 1.8 Hz, 1H, benzofuran 6 H), 7.10–7.07 (m, 2H, C_6H_3 , 3,5 H), 6.94 (d, 1H, J = 8.8 Hz, C_6H_3 , 6 H), 6.41 (s, 1H, benzofuran 3 H), 6.04–5.93 (m, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.21 (s, 1H, OH), 5.09 (dd, J = 23.1, 5.9 Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.36 (d, J = 6.7 Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.49 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 156.58, 154.44, 151.10, 138.08, 132.29, 131.79, 130.83, 130.21, 129.00, 128.75, 124.43, 120.87, 115.98, 115.75, 111.34, 102.93, 39.61, 14.19. LC-MS, m/z : 265.1 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{16}\text{O}_2$: C 81.79, H 6.10; found: C 81.85, H 6.08.

4-Allyl-2-(2-methyl-2,3-dihydrobenzofuran-5-yl)phenol (2). To a stirred solution of honokiol (10 mmol) in 1,2-dichloroethane (40 mL) was added concentrated sulfuric acid (98%, 0.2 mL) dropwise at room temperature. Then the mixture was heated to 50 °C and stirred for 24 h. After cooling, the solvent was evaporated under reduced pressure and the residue was then purified by silica gel chromatography to afford **2** as light yellow oil (12%). ^1H NMR (400 MHz, CDCl_3) δ 7.22 (s, 1H, Ar-H), 7.17 (d, J = 8.1 Hz, 1H, Ar-H), 7.03–7.01 (m, 2H, Ar-H), 6.87 (d, J = 8.1 Hz, 1H, Ar-H), 6.82 (d, J = 8.1 Hz, 1H, Ar-H), 5.96 (dt, J = 16.6, 6.9 Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.26 (s, 1H, OH), 5.11–5.01 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 4.99–4.91 (m, 1H, CH_3CHCH_2), 3.36–3.29 (m, 3H, $\text{CH}_2\text{CH}=\text{CH}_2$ and CH_3CHCH_2), 2.83 (dd, J = 15.5, 7.6 Hz, 1H, CH_3CHCH_2), 1.47 (d, J = 6.2 Hz, 3H, CH_3CHCH_2); ^{13}C NMR (101 MHz, CDCl_3) δ 159.36, 150.92, 137.88, 132.18, 130.38, 129.15, 128.96, 128.67, 128.33, 128.27, 125.95, 115.64, 115.60, 109.88, 80.11, 39.48, 37.10, 21.85. LC-MS, m/z : 267.2 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{18}\text{O}_2$: C 81.17, H 6.81; found: C 81.20, H 6.83.

2-(2-Methylbenzofuran-5-yl)-4-propylphenol (3). To a stirred solution of **1** (2.0 mmol) and $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$ (2.0 mmol) in EtOH (10 mL) was added NaBH_4 (10 mmol) in five portions over 15 min and kept stirring for 5 min at room temperature. Then the reaction was quenched with dilute hydrochloric acid and extracted with CH_2Cl_2 . The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **3** as light yellow oil (81%). ^1H NMR (400 MHz, CDCl_3) δ 7.53 (s, 1H, benzofuran 4 H), 7.48 (d, J = 8.4 Hz, 1H, benzofuran 7 H), 7.26 (d, J = 8.4 Hz, 1H, benzofuran 6 H), 7.08–7.04 (m, 2H, C_6H_3 , 4,6

H), 6.90 (d, J = 7.9 Hz, 1H, C_6H_3 , 3 H), 6.39 (s, 1H, benzofuran 3 H), 5.17 (s, 1H, OH), 2.55 (t, J = 7.6 Hz, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 2.47 (s, 3H, CH_3), 1.69–1.59 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 0.95 (t, J = 7.3 Hz, 3H, $\text{CH}_2\text{CH}_2\text{CH}_3$); ^{13}C NMR (101 MHz, CDCl_3) δ 156.60, 154.34, 150.53, 134.86, 131.60, 130.44, 130.17, 128.77, 128.22, 124.24, 120.67, 115.41, 111.36, 102.73, 37.27, 24.85, 14.17, 13.90. LC-MS, m/z : 267.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{18}\text{O}_2$: C 81.17, H 6.81; found: C 81.22, H 6.79.

2-(2-Methyl-2,3-dihydrobenzofuran-5-yl)-4-propylphenol (4). A mixture of **1** (5.0 mmol) and Pd/C (10%, 0.25 mmol) in MeOH (20 mL) was refluxed under H_2 (1.5 MPa) for 5 h. After cooling, the mixture was filtered and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **4** as light yellow oil (93%). ^1H NMR (400 MHz, CDCl_3) δ 7.24 (s, 1H, Ar-H), 7.19 (d, J = 8.1 Hz, 1H, Ar-H), 7.04–6.98 (m, 2H, Ar-H), 6.86 (d, J = 8.5 Hz, 1H, Ar-H), 6.83 (d, J = 8.1 Hz, 1H, Ar-H), 5.28 (s, 1H, OH), 5.01–4.92 (m, 1H, CH_3CHCH_2), 3.34 (dd, J = 15.5, 8.8 Hz, 1H, CH_3CHCH_2), 2.85 (dd, J = 15.6, 7.7 Hz, 1H, CH_3CHCH_2), 2.53 (t, J = 7.6 Hz, 2H, $\text{CH}_3\text{CH}_2\text{CH}_2$), 1.67–1.58 (m, 2H, $\text{CH}_3\text{CH}_2\text{CH}_2$), 1.49 (d, J = 6.2 Hz, 3H, CH_3CHCH_2), 0.94 (t, J = 7.3 Hz, 3H, $\text{CH}_3\text{CH}_2\text{CH}_2$); ^{13}C NMR (101 MHz, CDCl_3) δ 159.29, 150.56, 134.80, 130.22, 129.35, 128.94, 128.47, 128.25, 127.99, 125.93, 115.39, 109.81, 80.06, 37.25, 37.11, 24.82, 21.83, 13.88. LC-MS, m/z : 269.1 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{20}\text{O}_2$: C 80.56, H 7.51; found: C 80.52, H 7.50.

5-(5-Allyl-2-methoxyphenyl)-2-methylbenzofuran (5). To a stirred solution of **1** (1.0 mmol) and CH_3I (3.0 mmol) in DMF (5 mL) was added KOH (2 mmol). After 3 h at room temperature, the mixture was diluted with water and extracted with ethyl acetate. The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **5** as light yellow oil (71%). ^1H NMR (400 MHz, CDCl_3) δ 7.58 (s, 1H, benzofuran 4 H), 7.41 (d, J = 8.4 Hz, 1H, benzofuran 7 H), 7.35 (d, J = 8.5 Hz, 1H, benzofuran 6 H), 7.16 (d, J = 1.8 Hz, 1H, C_6H_3 , 6 H), 7.12 (d, J = 8.3 Hz, 1H, C_6H_3 , 4 H), 6.91 (d, J = 8.3 Hz, 1H, C_6H_3 , 3 H), 6.37 (s, 1H, benzofuran 3 H), 5.99 (ddt, J = 16.8, 10.0, 6.7 Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.13–5.03 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.78 (s, 3H, OCH_3), 3.38 (d, J = 6.7 Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.45 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 155.69, 154.97, 154.02, 137.82, 133.01, 132.26, 131.52, 131.21, 129.10, 128.06, 124.98, 121.04, 115.59, 111.37, 110.01, 102.78, 55.78, 39.46, 14.15. LC-MS, m/z : 279.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{19}\text{H}_{18}\text{O}_2$: C 81.99, H 6.52; found: C 81.96, H 6.50.

2-Methyl-6-(2-methyl-2,3-dihydrobenzofuran-5-yl)-4-propylphenol (6). A mixture of **20** (2.0 mmol) and Pd/C (10%, 0.1 mmol) in MeOH (10 mL) was refluxed under H_2 (1.5 MPa) for 5 h. After cooling, the mixture was filtered and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **6** as light yellow oil (88%). ^1H NMR (400 MHz, CDCl_3) δ 7.22 (s, 1H, dihydrobenzofuran 4 H), 7.17 (d, J = 8.1 Hz, 1H, dihydrobenzofuran 6 H), 6.91 (s, 1H, Ar-H), 6.86–6.81 (m, 2H, Ar-H), 5.12 (s, 1H, OH), 5.02–4.93 (m, 1H, CH_3CHCH_2), 3.35 (dd, J = 15.5, 8.8 Hz, 1H, CH_3CHCH_2), 2.86 (dd, J = 15.5, 7.6 Hz, 1H, CH_3CHCH_2), 2.50 (t, J = 7.6 Hz, 2H, $\text{CH}_3\text{CH}_2\text{CH}_2$), 2.27 (s, 3H, CH_3), 1.66–1.57 (m, 2H, $\text{CH}_3\text{CH}_2\text{CH}_2$),



1.50 (d, $J = 6.1$ Hz, 3H, CH_3CHCH_2), 0.94 (t, $J = 7.2$ Hz, 3H, $\text{CH}_3\text{CH}_2\text{CH}_2$); ^{13}C NMR (101 MHz, CDCl_3) δ 159.35, 148.66, 134.17, 130.08, 129.42, 128.95, 128.40, 127.64, 127.50, 125.97, 124.06, 109.90, 80.04, 37.27, 37.10, 24.86, 21.82, 16.25, 13.93. LC-MS, m/z : 283.1 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{19}\text{H}_{22}\text{O}_2$: C 80.82, H 7.85; found: C 80.87, H 7.84.

4-Allyl-2-bromo-6-(2-methylbenzofuran-5-yl)phenol (7). To a stirred solution of **1** (1.0 mmol) in MeCN (10 mL) was added NBS (1.0 mmol) in five portions over 1 h and kept stirring for 15 min at room temperature. Then the mixture was diluted with water and extracted with CH_2Cl_2 . The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **7** as light yellow oil (30%). ^1H NMR (400 MHz, CDCl_3) δ 7.58 (s, 1H, benzofuran 4 H), 7.46 (d, $J = 8.4$ Hz, 1H, benzofuran 7 H), 7.33–7.29 (m, 2H, Ar-H), 7.08 (s, 1H, Ar-H), 6.40 (s, 1H, benzofuran 3 H), 6.01–5.89 (m, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.59 (s, 1H, OH), 5.15–5.06 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.34 (d, $J = 6.5$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.48 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 156.37, 154.37, 147.64, 137.00, 133.38, 131.53, 131.07, 130.58, 129.90, 129.64, 124.33, 120.77, 116.26, 110.74, 110.57, 102.73, 39.07, 14.14. LC-MS, m/z : 342.9 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{15}\text{O}_2\text{Br}$: C 62.99, H 4.41; found: C 63.03, H 4.44.

4-Allyl-2-nitro-6-(2-methylbenzofuran-5-yl)phenol (8). To a stirred solution of **1** (1.0 mmol) in the co-solvent of CH_2Cl_2 and acetic acid (20 mL, $v/v = 1 : 1$) was added a solution of concentrated nitric acid (65–68%, 4 mmol) in 5 mL CH_2Cl_2 dropwise over 30 min in an ice bath and kept stirring for 30 min at room temperature. Then the reaction was quenched with water and extracted with CH_2Cl_2 . The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **8** as yellow oil (58%). ^1H NMR (400 MHz, CDCl_3) δ 7.92 (s, 1H, Ar-H), 7.62 (s, 1H, benzofuran 4 H), 7.49 (s, 1H, Ar-H), 7.46 (d, $J = 8.5$ Hz, 1H, benzofuran 7 H), 7.35 (d, $J = 8.4$ Hz, 1H, benzofuran 6 H), 6.41 (s, 1H, benzofuran 3 H), 5.95 (dt, $J = 16.6$, 7.0 Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.17–5.10 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.40 (d, $J = 6.6$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.48 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 156.34, 154.50, 151.41, 139.41, 136.04, 133.88, 133.54, 131.78, 130.30, 129.38, 124.59, 123.08, 121.08, 117.17, 110.43, 102.74, 38.96, 14.14. LC-MS, m/z : 310.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{15}\text{NO}_4$: C 69.89, H 4.89, N 4.53; found: C 69.94, H 4.90, N 4.51.

4-Allyl-2-nitroso-6-(2-methylbenzofuran-5-yl)phenol (9). To a stirred solution of **1** (10 mmol) and NaNO_2 (15 mmol) in the co-solvent of MeCN and H_2O (30 mL, $v/v = 5 : 1$) was added concentrated hydrochloric acid (36–38%, 3 mL) over 1 h in an ice bath and kept stirring for 30 min at room temperature. Then the reaction was quenched with water and extracted with CH_2Cl_2 . The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **9** as a yellow powder (77%). Mp 58–60 °C. ^1H NMR (400 MHz, CDCl_3) δ 11.04 (s, 1H, OH), 7.93 (d, $J = 2.2$ Hz, 1H, Ar-H), 7.63 (d, $J = 1.5$ Hz, 1H, benzofuran 4 H), 7.50 (d, $J = 2.2$ Hz, 1H, Ar-H), 7.47 (d, $J = 8.5$ Hz, 1H, benzofuran 7 H), 7.36 (dd, $J = 8.5$, 1.8 Hz, 1H,

benzofuran 6 H), 6.41 (s, 1H, benzofuran 3 H), 5.96 (ddt, $J = 17.2$, 10.7, 6.7 Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.18–5.11 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.41 (d, $J = 6.6$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.48 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 156.34, 154.51, 151.41, 139.42, 136.07, 133.88, 133.53, 131.78, 130.31, 129.38, 124.61, 123.07, 121.10, 117.17, 110.43, 102.75, 38.96, 14.15. LC-MS, m/z : 294.2 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{15}\text{NO}_3$: C 73.71, H 5.15, N 4.78; found: C 73.66, H 5.17, N 4.74.

4-Allyl-2-amino-6-(2-methylbenzofuran-5-yl)phenol (10). To a stirred solution of **9** (5.0 mmol) and NH_4Cl (1.0 mmol) in the co-solvent of EtOH, H_2O and acetic acid (62.5 mL, $v/v/v = 20 : 4 : 1$) was added zinc dust (25 mmol) in five portions over 30 min at room temperature. Then the mixture was filtered, diluted with water and extracted with ethyl acetate. The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **10** as a brown powder (73%). Mp 80–82 °C. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.56 (s, 1H, benzofuran 4 H), 7.46 (d, $J = 8.4$ Hz, 1H, benzofuran 7 H), 7.29 (d, $J = 8.5$ Hz, 1H, benzofuran 6 H), 6.58 (s, 1H, Ar-H), 6.46 (s, 1H, Ar-H), 6.31 (s, 1H, benzofuran 3 H), 5.98–5.87 (m, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.07 (d, $J = 17.0$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.00 (d, $J = 10.0$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 4.72 (br, 1H), 3.20 (d, $J = 6.7$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.45 (s, 3H, CH_3); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 155.97, 153.57, 138.97, 138.87, 138.82, 134.47, 131.91, 130.42, 129.11, 125.16, 121.06, 118.96, 115.56, 114.34, 110.29, 103.34, 14.26. LC-MS, m/z : 280.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{18}\text{H}_{17}\text{NO}_2$: C 77.40, H 6.13, N 5.01; found: C 77.46, H 6.17, N 4.99.

4-Allyl-2-(dimethylamino)-6-(2-methylbenzofuran-5-yl)phenol (11). To a solution of **10** (1.0 mmol) and CH_3I (3 mmol) in DMF (5 mL) was added NaOAc (1.2 mmol). After reacting for 3 h at room temperature, the mixture was diluted with water and extracted with ethyl acetate. The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **11** as red oil (71%). ^1H NMR (400 MHz, CDCl_3) δ 7.69 (s, 1H, benzofuran 4 H), 7.47–7.41 (m, 2H, benzofuran 6,7 H), 7.01 (s, 1H, Ar-H), 6.99 (s, 1H, Ar-H), 6.39 (s, 1H, benzofuran 3 H), 5.99 (dt, $J = 16.6$, 7.4 Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.14–5.04 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.36 (d, $J = 6.5$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.71 (s, 6H, $2 \times \text{CH}_3$), 2.46 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 155.74, 154.08, 146.85, 140.53, 137.84, 132.59, 131.16, 129.28, 127.61, 127.28, 124.51, 120.69, 119.38, 115.63, 110.23, 102.83, 45.25, 39.86, 14.14. LC-MS, m/z : 308.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{20}\text{H}_{21}\text{NO}_2$: C 78.15, H 6.89, N 4.56; found: C 78.11, H 6.87, N 4.58.

N-(5-Allyl-2-hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)acetamide (12). To a mixture of **10** (1.0 mmol) and K_2CO_3 (1.5 mmol) in ethyl acetate (5 mL) was added acetic anhydride (2 mmol) dropwise over 10 min at room temperature. Then the reaction mixture was washed with hot water and the aqueous layer was extracted with ethyl acetate. The combined organic layer was dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **12** as a brown powder (88%). Mp 70–72 °C. ^1H NMR (400 MHz, CDCl_3) δ 7.85 (br, 1H), 7.55 (s, 1H, benzofuran 4



H), 7.45 (d, $J = 8.4$ Hz, 1H, benzofuran 7 H), 7.40 (s, 1H, Ar-H), 7.29 (d, $J = 8.3$ Hz, 1H, benzofuran 6 H), 6.94 (s, 1H, Ar-H), 6.38 (s, 1H, benzofuran 3 H), 5.94 (dt, $J = 16.5, 6.8$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.11–5.02 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.32 (d, $J = 6.3$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.46 (s, 3H, CH_3), 2.20 (s, 3H, COCH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 169.77, 156.29, 154.26, 142.30, 137.50, 132.17, 131.91, 130.85, 129.72, 127.22, 126.25, 124.47, 120.87, 120.63, 115.84, 110.80, 102.74, 39.51, 24.19, 14.14. LC-MS, m/z : 322.1 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{20}\text{H}_{19}\text{NO}_3$: C 74.75, H 5.96, N 4.36; found: C 74.77, H 5.96, N 4.39.

***N*-(5-Allyl-2-hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)acrylamide (13)**. To a stirred mixture of **10** (1.0 mmol) and K_2CO_3 (1.5 mmol) in CH_2Cl_2 (10 mL) was added acryloyl chloride (1.0 mmol) in CH_2Cl_2 (1.0 mL) dropwise over 10 min at room temperature. Then the reaction was quenched with water and extracted with CH_2Cl_2 . The combined organic layer was dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **13** as a yellow powder (46%). Mp 64–66 °C. ^1H NMR (400 MHz, CDCl_3) δ 7.94 (s, 1H), 7.58 (s, 1H, Ar-H), 7.55 (d, $J = 1.4$ Hz, 1H, benzofuran 4 H), 7.46 (d, $J = 8.4$ Hz, 1H, benzofuran 7 H), 7.29 (dd, $J = 8.4, 1.7$ Hz, 1H, benzofuran 6 H), 7.15 (s, 1H), 6.94 (s, 1H, Ar-H), 6.44 (d, $J = 16.2$ Hz, 1H, $\text{COCH}=\text{CH}_2$), 6.39 (s, 1H, benzofuran 3 H), 6.31 (dd, $J = 16.8, 10.1$ Hz, 1H, $\text{COCH}=\text{CH}_2$), 5.95 (ddt, $J = 16.8, 10.0, 6.7$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.77 (d, $J = 9.9$ Hz, 1H, $\text{COCH}=\text{CH}_2$), 5.12–5.02 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.33 (d, $J = 6.7$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.47 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 164.33, 156.35, 154.29, 142.17, 137.45, 132.29, 131.71, 130.51, 129.80, 128.45, 127.21, 126.03, 124.39, 120.83, 120.64, 115.87, 110.90, 102.74, 39.56, 14.14. LC-MS, m/z : 334.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{21}\text{H}_{19}\text{NO}_3$: C 75.66, H 5.74, N 4.20; found: C 75.60, H 5.76, N 4.19.

***N*-(5-Allyl-2-hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)-2-chloroacetamide (14)**. The title compound was prepared from **10** and chloroacetyl chloride in a similar procedure described for **13**. Purification by silica gel chromatography afforded **14** as a white powder (81%). Mp 118–120 °C. ^1H NMR (400 MHz, CDCl_3) δ 8.81 (brs, 1H), 7.84 (s, 1H, Ar-H), 7.53 (s, 1H, benzofuran 4 H), 7.49 (d, $J = 8.3$ Hz, 1H, benzofuran 7 H), 7.27 (d, $J = 6.3$ Hz, 1H, benzofuran 6 H), 6.94 (s, 1H, Ar-H), 6.41 (s, 1H, benzofuran 3 H), 6.15 (br, 1H), 5.98 (dt, $J = 16.6, 6.7$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.14–5.05 (m, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 4.22 (s, 2H, COCH_2), 3.37 (d, $J = 6.3$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 2.49 (s, 3H, CH_3); ^{13}C NMR (101 MHz, CDCl_3) δ 164.31, 156.72, 154.42, 141.21, 137.44, 132.57, 131.00, 130.13, 129.48, 126.82, 125.19, 124.13, 120.66, 119.98, 115.94, 111.34, 102.70, 43.01, 39.67, 14.19. LC-MS, m/z : 356.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{20}\text{H}_{18}\text{NO}_3\text{Cl}$: C 67.51, H 5.10, N 3.94; found: C 67.55, H 5.11, N 3.91.

***N*-(5-Allyl-2-hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)-2-(piperidin-1-yl)acetamide (15)**. A solution of **14** (0.5 mmol), piperidine (1.5 mmol) and triethylamine (1.0 mmol) in THF (5 mL) was stirred at room temperature for 8 h. Then the mixture was concentrated under reduced pressure and purified by silica gel chromatography to afford **15** as a brown powder (48%). Mp 125–127 °C. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 9.95 (s, 1H), 8.80 (br, 1H), 7.92 (s, 1H, Ar-H), 7.61 (s, 1H, benzofuran 4 H), 7.52 (d,

$J = 8.4$ Hz, 1H, benzofuran 7 H), 7.32 (d, $J = 8.4$ Hz, 1H, benzofuran 6 H), 6.83 (s, 1H, Ar-H), 6.61 (s, 1H, benzofuran 3 H), 5.96 (dt, $J = 16.7, 6.8$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.10 (d, $J = 17.0$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.04 (d, $J = 9.9$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.32 (d, $J = 6.6$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.09 (s, 2H, COCH_2), 2.51 (s, 3H, CH_3), 2.47 (br, 4H, piperidine 2,6 H), 1.58 (br, 4H, piperidine 3,5 H), 1.41 (s, 2H, piperidine 4 H); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 169.15, 156.27, 153.86, 141.70, 138.41, 133.24, 132.13, 131.27, 129.36, 129.23, 125.81, 125.10, 121.23, 119.03, 116.07, 110.66, 103.36, 62.81, 54.63, 39.64, 26.19, 23.81, 14.25. LC-MS, m/z : 405.1 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{25}\text{H}_{28}\text{N}_2\text{O}_3$: C 74.23, H 6.98, N 6.93; found: C 74.27, H 7.00, N 6.96.

***N*-(5-Allyl-2-hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)-2-morpholinoacetamide (16)**. The title compound was prepared from **14** and morpholine in a similar procedure described for **15**. Purification by silica gel chromatography afforded **16** as a light purple powder (85%). Mp 147–149 °C. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 9.91 (s, 1H), 8.82 (s, 1H), 7.85 (s, 1H, Ar-H), 7.62 (s, 1H, benzofuran 4 H), 7.52 (d, $J = 8.4$ Hz, 1H, benzofuran 7 H), 7.33 (d, $J = 8.4$ Hz, 1H, benzofuran 6 H), 6.86 (s, 1H, Ar-H), 6.61 (s, 1H, benzofuran 3 H), 5.96 (dt, $J = 16.6, 6.8$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.11 (d, $J = 17.0$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.05 (d, $J = 10.0$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.66 (br, 4H, morpholine 2,6 H), 3.33 (d, $J = 6.6$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.18 (s, 2H, COCH_2), 2.56 (br, 4H, morpholine 3,5 H), 2.47 (s, 3H, CH_3); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 168.66, 156.29, 153.86, 142.04, 138.38, 133.23, 132.13, 131.37, 129.35, 128.99, 126.11, 125.11, 121.24, 119.54, 116.10, 110.65, 103.36, 66.77, 62.18, 53.63, 39.59, 14.24. LC-MS, m/z : 407.1 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{24}\text{H}_{26}\text{N}_2\text{O}_4$: C 70.92, H 6.45, N 6.89; found: C 70.90, H 6.49, N 6.86.

***N*-(5-Allyl-2-hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)-2-(4-methylpiperazin-1-yl)acetamide (17)**. The title compound was prepared from **14** and 4-methylpiperazine in a similar procedure described for **15**. Purification by silica gel chromatography afforded **17** as a brown powder (76%). Mp 101–103 °C. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 9.86 (s, 1H), 8.82 (brs, 1H), 7.89 (s, 1H, Ar-H), 7.60 (s, 1H, benzofuran 4 H), 7.51 (d, $J = 8.4$ Hz, 1H, benzofuran 7 H), 7.31 (d, $J = 8.4$ Hz, 1H, benzofuran 6 H), 6.82 (s, 1H, C_6H_2 , Ar-H), 6.61 (s, 1H, benzofuran 3 H), 5.95 (dt, $J = 16.7, 6.9$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.09 (d, $J = 17.0$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 5.03 (d, $J = 10.0$ Hz, 1H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.31 (d, $J = 6.6$ Hz, 2H, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.14 (s, 2H, COCH_2), 2.54 (br, 4H, piperazine-H), 2.47 (s, 3H, CH_3), 2.40 (brs, 4H, piperazine-H), 2.17 (s, 3H, NCH_3); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 168.79, 156.27, 153.86, 141.81, 138.42, 133.26, 132.07, 131.29, 129.35, 129.16, 125.89, 125.12, 121.25, 119.15, 116.07, 110.65, 103.37, 61.93, 55.19, 53.16, 46.09, 39.62, 14.26. LC-MS, m/z : 420.2 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{25}\text{H}_{29}\text{N}_3\text{O}_3$: C 71.57, H 6.97, N 10.02; found: C 71.55, H 6.99, N 10.05.

***N*-(5-Allyl-2-hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)-2-(4-phenylpiperazin-1-yl)acetamide (18)**. The title compound was prepared from **14** and 4-phenylpiperazine in a similar procedure described for **15**. Purification by silica gel chromatography afforded **18** as a light purple powder (79%). Mp 136–138 °C. ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 9.98 (s, 1H), 8.81 (s, 1H), 7.92 (s, 1H, Ar-H), 7.60 (s, 1H, benzofuran 4 H), 7.50 (d, $J = 8.4$ Hz, 1H,



benzofuran 7 H), 7.31 (d, $J = 8.4$ Hz, 1H, benzofuran 6 H), 7.24–7.17 (m, 2H, C₆H₅, 3,5 H), 6.94 (d, $J = 7.9$ Hz, 2H, C₆H₅, 2,6 H), 6.84 (s, 1H, Ar-H), 6.81–6.75 (m, 1H, C₆H₅, 4 H), 6.58 (s, 1H, benzofuran 3 H), 5.96 (dt, $J = 16.7, 6.6$ Hz, 1H, CH₂CH=CH₂), 5.10 (d, $J = 16.7$ Hz, 1H, CH₂CH=CH₂), 5.04 (d, $J = 9.9$ Hz, 1H, CH₂CH=CH₂), 3.32 (d, $J = 6.6$ Hz, 2H, CH₂CH=CH₂), 3.23 (s, 2H, COCH₂), 3.20 (br, 4H, piperazine-H), 2.70 (br, 4H, piperazine-H), 2.45 (s, 3H, CH₃); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 168.72, 156.26, 153.87, 151.43, 141.86, 138.42, 133.22, 132.11, 131.23, 129.39, 129.13, 125.94, 125.10, 121.24, 119.43, 119.19, 116.09, 115.94, 110.66, 103.37, 61.84, 53.19, 48.92, 39.65, 14.26. LC-MS, m/z : 482.1 [M + H]⁺. Elemental anal (%) calcd for C₃₀H₃₁N₃O₃: C 74.82, H 6.49, N 8.73; found: C 74.88, H 6.50, N 8.75.

(E)-4-Allyl-2-(benzylideneamino)-6-(2-methylbenzofuran-5-yl)phenol (19). A mixture of **10** (1.0 mmol), benzaldehyde (1.0 mmol) and acetic acid (0.1 mL) in EtOH (10 mL) was stirred at room temperature for 30 min. The reaction was left overnight until precipitation. The precipitate was separated by filtration, washed with EtOH and dried to afford **19** as a yellow powder (68%). Mp 91–93 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.86 (s, 1H, N=CH), 8.51 (br, 1H, OH), 8.19–8.09 (m, 2H, Ar-H), 7.71 (s, 1H, benzofuran 4 H), 7.55–7.49 (m, 4H, Ar-H), 7.44 (d, $J = 8.6$ Hz, 1H, benzofuran 6 H), 7.20 (s, 1H, Ar-H), 7.07 (s, 1H, Ar-H), 6.61 (s, 1H, benzofuran 3 H), 6.03 (dt, $J = 16.8, 6.8$ Hz, 1H, CH₂CH=CH₂), 5.14 (d, $J = 17.0$ Hz, 1H, CH₂CH=CH₂), 5.06 (d, $J = 9.8$ Hz, 1H, CH₂CH=CH₂), 3.38 (d, $J = 6.6$ Hz, 2H, CH₂CH=CH₂), 2.46 (s, 3H, CH₃); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 158.93, 156.21, 153.71, 147.78, 138.53, 137.44, 136.70, 133.26, 131.87, 131.15, 129.70, 129.16, 129.12, 128.95, 125.12, 121.13, 116.46, 116.10, 110.38, 103.35, 39.47, 14.27. LC-MS, m/z : 368.0 [M + H]⁺. Elemental anal (%) calcd for C₂₅H₂₁NO₂: C 81.72, H 5.76, N 3.81; found: C 81.67, H 5.78, N 3.84.

4-Allyl-2-((dimethylamino)methyl)-6-(2-methylbenzofuran-5-yl)phenol (20). A solution of **1** (2.0 mmol), dimethylamine (aqueous solution, 40%, 6.0 mmol) and formalin (37%, 6.0 mmol) in MeOH (5.0 mL) was stirred at 60 °C for 6 h. Then the mixture was concentrated under reduced pressure and purified by silica gel chromatography to afford **20** as light yellow oil (53%). ¹H NMR (400 MHz, CDCl₃) δ 7.67 (s, 1H, benzofuran 4 H), 7.47–7.39 (m, 2H, benzofuran 6,7 H), 7.11 (s, 1H, Ar-H), 6.78 (s, 1H, Ar-H), 6.37 (s, 1H, benzofuran 3 H), 6.05–5.92 (m, 1H, CH₂CH=CH₂), 5.14–5.02 (m, 2H, CH₂CH=CH₂), 3.68 (s, 2H, ArCH₂N), 3.33 (d, $J = 6.5$ Hz, 2H, CH₂CH=CH₂), 2.45 (s, 3H, CH₃), 2.33 (s, 6H, N(CH₃)₂); ¹³C NMR (101 MHz, CDCl₃) δ 155.52, 153.97, 153.39, 138.04, 133.03, 130.20, 130.18, 129.19, 129.14, 127.40, 124.83, 122.06, 120.92, 115.44, 110.01, 102.84, 62.97, 44.35, 39.53, 14.15. LC-MS, m/z : 322.1 [M + H]⁺. Elemental anal (%) calcd for C₂₁H₂₃NO₂: C 78.47, H 7.21, N 4.36; found: C 78.42, H 7.19, N 4.38.

4-Allyl-2-(2-methylbenzofuran-5-yl)-6-(piperidin-1-ylmethyl)phenol (21). The title compound was prepared from **1** and piperidine in a similar procedure described for **20**. Purification by silica gel chromatography afforded **21** as light yellow oil (81%). ¹H NMR (400 MHz, CDCl₃) δ 7.66 (s, 1H, benzofuran 4 H), 7.44–7.41 (m, 2H, benzofuran 6,7 H), 7.09 (s, 1H, Ar-H), 6.78 (s, 1H, Ar-H), 6.38 (s, 1H, benzofuran 3 H), 5.98 (dt, $J = 16.3, 8.0$ Hz, 1H, CH₂CH=CH₂), 5.12–5.02 (m, 2H, CH₂CH=CH₂),

3.70 (s, 2H, ArCH₂N), 3.33 (d, $J = 6.5$ Hz, 2H, CH₂CH=CH₂), 2.52 (br, 4H, piperidine 2,6 H), 2.46 (s, 3H, CH₃), 1.61 (br, 4H, piperidine 3,5 H), 1.47 (br, 2H, piperidine 4 H); ¹³C NMR (101 MHz, CDCl₃) δ 155.51, 153.95, 153.40, 138.05, 133.18, 130.13, 129.21, 129.15, 127.61, 127.55, 124.80, 121.87, 120.87, 115.40, 110.05, 102.85, 62.43, 53.87, 39.51, 25.66, 23.97, 14.14. LC-MS, m/z : 362.1 [M + H]⁺. Elemental anal (%) calcd for C₂₄H₂₇NO₂: C 79.74, H 7.53, N 3.87; found: C 79.79, H 7.54, N 3.88.

4-Allyl-2-(2-methylbenzofuran-5-yl)-6-(morpholinomethyl)phenol (22). The title compound was prepared from **1** and morpholin in a similar procedure described for **20**. Purification by silica gel chromatography afforded **22** as colourless oil (58%). ¹H NMR (400 MHz, CDCl₃) δ 7.65 (s, 1H, benzofuran 4 H), 7.45–7.40 (m, 2H, benzofuran 6,7 H), 7.12 (s, 1H, Ar-H), 6.81 (s, 1H, Ar-H), 6.38 (s, 1H, benzofuran 3 H), 5.97 (dt, $J = 16.7, 6.8$ Hz, 1H, CH₂CH=CH₂), 5.13–5.03 (m, 2H, CH₂CH=CH₂), 3.74 (s, 2H, ArCH₂N), 3.72 (br, 4H, morpholine 2,6 H), 3.33 (d, $J = 6.6$ Hz, 2H, CH₂CH=CH₂), 2.59 (br, 4H, morpholine 3,5 H), 2.45 (s, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃) δ 155.65, 154.01, 152.83, 137.90, 132.92, 130.63, 130.57, 129.41, 129.20, 127.93, 124.77, 120.91, 120.88, 115.56, 110.11, 102.83, 66.69, 62.17, 52.90, 39.48, 14.15. LC-MS, m/z : 364.1 [M + H]⁺. Elemental anal (%) calcd for C₂₃H₂₅NO₃: C 76.01, H 6.93, N 3.85; found: C 75.97, H 6.94, N 3.82.

4-Allyl-2-(2-methylbenzofuran-5-yl)-6-((4-methylpiperazin-1-yl)methyl)phenol (23). The title compound was prepared from **1** and 4-methylpiperazine in a similar procedure described for **20**. Purification by silica gel chromatography afforded **23** as light yellow oil (68%). ¹H NMR (400 MHz, CDCl₃) δ 7.65 (s, 1H, benzofuran 4 H), 7.45–7.39 (m, 2H, benzofuran 6,7 H), 7.10 (s, 1H, Ar-H), 6.80 (s, 1H, Ar-H), 6.38 (s, 1H, benzofuran 3 H), 5.97 (dt, $J = 16.6, 7.7$ Hz, 1H, CH₂CH=CH₂), 5.29 (br, 1H, OH), 5.12–5.02 (m, 2H, CH₂CH=CH₂), 3.75 (s, 2H, ArCH₂N), 3.33 (d, $J = 6.6$ Hz, 2H, CH₂CH=CH₂), 2.62 (br, 8H, piperazine-H), 2.46 (s, 3H, NCH₃), 2.29 (s, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃) δ 155.59, 153.97, 152.98, 137.91, 133.02, 130.50, 130.41, 129.31, 129.16, 127.78, 124.77, 121.32, 120.86, 115.52, 110.06, 102.82, 61.61, 54.51, 52.10, 45.52, 39.48, 14.14. LC-MS, m/z : 377.1 [M + H]⁺. Elemental anal (%) calcd for C₂₄H₂₈N₂O₂: C 76.56, H 7.50, N 7.44; found: C 76.55, H 7.47, N 7.48.

4-Allyl-2-(2-methylbenzofuran-5-yl)-6-((4-phenylpiperazin-1-yl)methyl)phenol (24). The title compound was prepared from **1** and 4-phenylpiperazine in a similar procedure described for **20**. Purification by silica gel chromatography afforded **24** as a white powder (55%). Mp 50–52 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.67 (s, 1H, benzofuran 4 H), 7.45–7.40 (m, 2H, Ar-H), 7.27–7.21 (m, 2H, Ar-H), 7.23 (s, 1H, Ar-H), 6.92–6.81 (m, 4H, Ar-H), 6.37 (s, 1H, benzofuran 3 H), 5.98 (dt, $J = 16.4, 6.7$ Hz, 1H, CH₂CH=CH₂), 5.14–5.03 (m, 2H, CH₂CH=CH₂), 3.79 (s, 2H, ArCH₂N), 3.34 (d, $J = 6.1$ Hz, 2H, CH₂CH=CH₂), 3.20 (br, 4H, piperazine-H), 2.74 (br, 4H, piperazine-H), 2.44 (s, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃) δ 155.64, 154.00, 152.99, 150.97, 137.95, 132.98, 130.58, 130.52, 129.41, 129.21, 127.87, 124.81, 121.30, 120.92, 120.26, 116.47, 115.60, 110.13, 102.87, 61.79, 52.55, 49.18, 39.54, 14.20. LC-MS, m/z : 439.1 [M + H]⁺. Elemental anal (%) calcd for C₂₉H₃₀N₂O₂: C 79.42, H 6.90, N 6.39; found: C 79.48, H 6.91, N 6.42.



6-Allyl-8-(2-methylbenzofuran-5-yl)-2H-benzo[*b*][1,4]-oxazin-3(4*H*)-one (25). To a stirred solution of **10** (1.0 mmol), triethylamine (2.5 mmol) and 4-dimethylaminopyridine (0.2 mmol) in THF (5 mL) was added chloroacetyl chloride (1.0 mmol) in CH₂Cl₂ (1.0 mL) dropwise over 10 min at room temperature. Then the reaction was heated to reflux and stirred for 12 h. After cooling, the mixture was concentrated under reduced pressure and purified by silica gel chromatography to afford **25** as a white powder (44%). Mp 169–171 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.72 (s, 1H, NH), 7.62 (s, 1H, benzofuran 4 H), 7.51 (d, *J* = 8.5 Hz, 1H, benzofuran 7 H), 7.33 (d, *J* = 8.5 Hz, 1H, benzofuran 6 H), 6.84 (s, 1H, Ar-H), 6.73 (s, 1H, Ar-H), 6.60 (s, 1H, benzofuran 3 H), 5.95 (ddt, *J* = 16.5, 9.5, 6.7 Hz, 1H, CH₂CH=CH₂), 5.12 (d, *J* = 17.2 Hz, 1H, CH₂CH=CH₂), 5.06 (d, *J* = 9.9 Hz, 1H, CH₂CH=CH₂), 4.54 (s, 2H, OCH₂), 3.33 (d, *J* = 6.7 Hz, 1H, CH₂CH=CH₂), 2.46 (s, 3H, CH₃); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 165.89, 156.43, 153.93, 139.24, 138.07, 134.40, 131.93, 130.32, 129.25, 128.52, 125.01, 124.86, 121.14, 116.42, 115.21, 110.55, 103.32, 67.32, 39.31, 14.25. LC-MS, *m/z*: 320.0 [M + H]⁺. Elemental anal (%) calcd for C₂₀H₁₇NO₃: C 75.22, H 5.37, N 4.39; found: C 75.18, H 5.34, N 4.42.

6-Allyl-8-(2-methylbenzofuran-5-yl)-2H-benzo[*b*][1,4]-oxazine-2,3(4*H*)-dione (26). To a stirred solution of **10** (1.0 mmol) and triethylamine (2.5 mmol) in CH₂Cl₂ (10 mL) was added oxalyl chloride (1.5 mmol) in CH₂Cl₂ (1.0 mL) dropwise over 10 min at room temperature. Then the reaction was quenched with water and extracted with CH₂Cl₂. The combined organic layer was dried over Na₂SO₄ and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **26** as a brown powder (42%). Mp 160–162 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.57 (br, 1H, NH), 7.87 (s, 1H, benzofuran 4 H), 7.58 (d, *J* = 8.5 Hz, 1H, benzofuran 7 H), 7.48 (d, *J* = 8.5 Hz, 1H, benzofuran 6 H), 7.16 (s, 1H, Ar-H), 6.86 (s, 1H, Ar-H), 6.43 (s, 1H, benzofuran 3 H), 6.04–5.93 (m, 1H, CH₂CH=CH₂), 5.17–5.10 (m, 2H, CH₂CH=CH₂), 3.45 (d, *J* = 6.6 Hz, 2H, CH₂CH=CH₂), 2.49 (s, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃) δ 156.34, 154.63, 139.49, 137.06, 136.76, 130.12, 129.70, 129.18, 124.66, 123.56, 122.68, 120.16, 116.35, 110.81, 108.82, 102.90, 40.16, 14.15. LC-MS, *m/z*: 334.1 [M + H]⁺. Elemental anal (%) calcd for C₂₀H₁₅NO₄: C 72.06, H 4.54, N 4.20; found: C 72.11, H 4.56, N 4.15.

1-(4-Hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)propan-2-one (27). A mixture of **1** (10 mmol) and PdCl₂ (0.15 mmol) in the co-solvent of DMA and H₂O (40 mL, v/v = 4 : 1) was stirred under O₂ (0.8 Mpa) at 60 °C for 16 h. After cooling, the mixture was diluted with water and extracted with ethyl acetate. The combined extracts were dried over Na₂SO₄ and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **27** as a white powder (64%). Mp 118–120 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.53–7.48 (m, 2H, benzofuran 4,7 H), 7.26–7.24 (m, 1H, benzofuran 6 H), 7.10–7.07 (m, 2H, Ar-H), 6.97 (d, 1H, *J* = 7.8 Hz, Ar-H), 6.41 (s, 1H, benzofuran 3 H), 5.35 (br, 1H, OH), 3.66 (s, 2H, CH₂), 2.49 (s, 3H, CH₃), 2.18 (s, 3H, COCH₃); ¹³C NMR (101 MHz, CDCl₃) δ 190.98, 158.27, 157.11, 154.69, 132.77, 131.21, 130.51, 130.05, 129.53, 129.28, 123.93, 120.69, 116.34, 111.78, 102.69, 50.17, 29.23, 14.17. LC-MS, *m/z*: 280.9 [M + H]⁺. Elemental anal (%) calcd for C₁₈H₁₆O₃: C 77.12, H 5.75; found: C 77.07, H 5.79.

(*E/Z*)-1-(4-Hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)propan-2-one oxime (28). A mixture of **27** (2.0 mmol), hydroxylamine hydrochloride (3.0 mmol) and NaOAc (3.0 mmol) in EtOH (20 mL) was stirred and refluxed for 30 min. After cooling, the mixture was diluted with water, shocked and placed until precipitation. Then the precipitate was separated by filtration, washed with water and dried to afford **28** as a white powder (95%). Mp 38–40 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.52 (s, 1H, benzofuran 4 H), 7.49 (d, *J* = 7.8 Hz, 1H, benzofuran 7 H), 7.27–7.24 (m, 1H, benzofuran 6 H), 7.13–7.09 (m, 2H, Ar-H), 6.98–6.90 (m, 1H, Ar-H), 6.41 (s, 1H, benzofuran 3 H), 5.07 (br, 1H, OH), 3.71 (s, 1H, N-OH), 3.52–3.44 (m, 2H, CH₂), 2.49 (s, 3H, CH₃), 1.86 (s, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃) δ 158.53, 156.47, 154.30, 151.67, 131.50, 131.22, 129.99, 129.23, 128.97, 128.56, 124.34, 120.78, 116.13, 111.13, 102.80, 41.21, 34.09, 19.64, 14.13, 13.45. LC-MS, *m/z*: 296.1 [M + H]⁺. Elemental anal (%) calcd for C₁₈H₁₇NO₃: C 73.20, H 5.80, N 4.74; found: C 73.17, H 5.79, N 4.70.

(*E/Z*)-2-(1-(4-Hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)propan-2-ylidene)hydrazine-1-carbothioamide (29). A solution of **27** (1.0 mmol), thiosemicarbazide (4.0 mmol) and acetic acid (0.2 mL) in EtOH (20 mL) was stirred and refluxed for 1 h. After cooling, the mixture was diluted with water, shocked and placed until precipitation. Then the precipitate was separated by filtration, washed with water and dried to afford **29** as a light yellow powder (57%). Mp 97–99 °C. ¹H NMR (400 MHz, Acetone-*d*₆) δ 8.96 (brs, 1H), 8.01 (brs, 1H), 7.58–6.75 (m, 8H), 6.37 (s, 1H, benzofuran 3 H), 3.72–3.34 (m, 2H, CH₂), 2.31 (s, 3H, CH₃), 1.94–1.77 (m, 3H, CH₃); ¹³C NMR (101 MHz, acetone-*d*₆) δ 180.10, 155.88, 153.92, 152.99, 152.21, 133.13, 131.51, 129.23, 129.04, 128.68, 128.35, 124.86, 120.95, 116.27, 109.86, 102.75, 43.98, 14.51, 13.11. LC-MS, *m/z*: 354.0 [M + H]⁺. Elemental anal (%) calcd for C₁₉H₁₉N₃O₃S: C 64.57, H 5.42, N 11.89; found: C 64.61, H 5.44, N 11.85.

(*E/Z*)-*N'*-(1-(4-Hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)propan-2-ylidene)benzohydrazide (30). A solution of **27** (0.5 mmol), benzoylhydrazide (1.0 mmol) and acetic acid (0.2 mL) in EtOH (20 mL) was stirred and refluxed for 1 h. After cooling, the mixture was diluted with water, shocked and placed until precipitation. Then the precipitate was separated by filtration, washed with water and dried to afford **30** as a light yellow powder (90%). Mp 99–101 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.87–7.73 (m, 2H, Ar-H), 7.53–7.44 (m, 5H, Ar-H), 7.26–6.93 (m, 4H, Ar-H), 6.40 (s, 1H, benzofuran 3 H), 5.56 (br, 1H, OH), 3.78–3.62 (m, 2H, CH₂), 2.48 (s, 3H, CH₃), 2.18 (s, 1H, CH₃), 1.88 (s, 2H, CH₃); ¹³C NMR (101 MHz, acetone-*d*₆) δ 164.42, 156.78, 155.00, 154.08, 149.27, 135.21, 134.23, 133.62, 132.46, 131.36, 130.25, 129.95, 129.71, 129.41, 128.56, 127.66, 125.89, 121.97, 117.63, 117.36, 110.80, 103.76, 45.19, 15.91, 14.10. LC-MS, *m/z*: 399.2 [M + H]⁺. Elemental anal (%) calcd for C₂₅H₂₂N₂O₃: C 75.36, H 5.57, N 7.03; found: C 75.30, H 5.54, N 6.99.

2-(1-(4-Hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)propan-2-yl)hydrazine-1-carbothioamide (31). To a stirred solution of **29** (0.5 mmol) in EtOH (5.0 mL) was added NaBH₄ (5.0 mmol) in five portions over 1 h and kept stirring for 1 h at room temperature. Then the reaction was quenched with dilute



hydrochloric acid and extracted with CH_2Cl_2 . The combined extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was then purified by silica gel chromatography to afford **31** as a white powder (31%). Mp 81–83 °C ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 9.30 (brs, 1H), 8.76 (brs, 1H), 7.68 (brs, 1H), 7.65 (d, $J = 1.0$ Hz, 1H, benzofuran 4 H), 7.47 (d, $J = 8.5$ Hz, 1H, benzofuran 7 H), 7.38 (dd, $J = 8.5, 1.3$ Hz, 1H, benzofuran 6 H), 7.17 (br, 1H), 7.08 (d, $J = 1.5$ Hz, 1H, C_6H_3 2 H), 6.95 (dd, $J = 8.2, 1.6$ Hz, 1H, C_6H_3 6 H), 6.88 (d, $J = 8.1$ Hz, 1H, C_6H_3 5 H), 6.58 (s, 1H, benzofuran 3 H), 4.88 (br, 1H), 3.08–2.96 (m, 1H, CH), 2.72 (dd, $J = 13.2, 4.7$ Hz, 1H, CH_2), 2.44 (s, 3H, CH_3), 2.32 (dd, $J = 13.1, 8.7$ Hz, 1H, CH_2), 0.89 (d, $J = 6.1$ Hz, 3H, CH_3); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 181.88, 156.04, 153.55, 152.87, 133.71, 131.81, 130.12, 129.16, 129.06, 128.36, 125.17, 121.10, 116.32, 110.23, 103.30, 57.09, 17.93, 14.24. LC-MS, m/z : 356.0 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{19}\text{H}_{21}\text{N}_3\text{O}_2\text{S}$: C 64.20, H 5.96, N 11.82; found: C 64.23, H 6.00, N 11.78.

***N'*-(1-(4-Hydroxy-3-(2-methylbenzofuran-5-yl)phenyl)propan-2-yl)benzohydrazide (32)**. The title compound was prepared from **30** in a similar procedure described for **31**. Purification by silica gel chromatography afforded **32** as a white powder (47%). Mp 188–190 °C. ^1H NMR (400 MHz, CDCl_3) δ 7.62 (d, $J = 7.5$ Hz, C_6H_5 2,6 H), 7.49–7.45 (m, 3H, Ar-H), 7.43–7.38 (m, 2H, benzofuran 4,7 H), 7.22 (d, $J = 8.5$ Hz, 1H, benzofuran 6 H), 7.15–7.11 (m, 2H, Ar-H), 6.95 (d, $J = 8.1$ Hz, 1H, Ar-H), 6.39 (s, 1H, benzofuran 3 H), 5.26 (br, 2H), 3.45–3.35 (m, 1H, CH), 2.81 (dd, $J = 12.7, 6.7$ Hz, 1H, CH_2), 2.67 (dd, $J = 13.0, 6.6$ Hz, 1H, CH_2), 2.48 (s, 3H, CH_3), 1.16 (d, $J = 6.0$ Hz, 3H, CH_3); ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 166.06, 156.07, 153.55, 152.74, 133.72, 133.58, 131.92, 131.76, 130.19, 129.28, 129.07, 128.80, 128.24, 127.59, 125.19, 121.12, 116.21, 110.24, 103.31, 56.95, 40.80, 18.68, 14.22. LC-MS, m/z : 401.2 $[\text{M} + \text{H}]^+$. Elemental anal (%) calcd for $\text{C}_{25}\text{H}_{24}\text{N}_2\text{O}_3$: C 74.98, H 6.04, N 7.00; found: C 75.03, H 6.00, N 7.02.

Biological assays

Cell culture. The A549, K562, HepG2 and HUVEC cells were obtained from Institute of Cell Biology, Chinese Academy of Science (Shanghai China) and cultured in RPMI-1640 medium (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum (Gibco; Thermo Fisher Scientific, Inc.) and 100 U mL^{-1} penicillin–streptomycin at 37 °C in incubator with 5% CO_2 /95% air atmosphere.

Cytotoxicity assay. The cytotoxicity of all synthesized compounds were evaluated by typical MTT assay or MTS assay according to our previous method.³³ The IC_{50} (the concentration required for 50% inhibition) was calculated by the Logit method.

Morphological analysis. The changes of cellular morphology were observed under an inverted fluorescence microscope. The HepG2 cells of experimental groups were treated with 20 μM of different compounds (**1**, **23** and **30**) for 24 h, and the control group was treated with DMSO.

Hoechst 33258 staining. After treated with 20 μM of different compounds (**1**, **23** and **30**) for 24 h, the cells were fixed with stationary liquid (24% glacial acetic, 72% methyl alcohol, 4%

paraformaldehyde). Then the HepG2 cells were rinsed three times with PBS, each time for 5 min, and then stained with a solution of Hoechst 33258 ($2 \mu\text{g mL}^{-1}$) for 15 min at room temperature, and then washed with PBS again for three time, each time for 5 min. Subsequently, the stained nuclei were observed under an inverted fluorescence microscope. The control group was treated with DMSO.

Flow cytometry analysis. The extent of apoptosis was quantitatively measured by an Annexin V-FITC/PI dual staining assay with annexin V-FITC/PI apoptosis detection Kit (Solarbio, CA1020). Briefly, HepG2 Cells were treated with 20 μM of different compounds (**1**, **23** and **30**) for 24 h. Then detached and adherent cells were centrifuged at $1500 \times g$ for 5 min, and then rinsed with ice-cold PBS for two times. Adjust the density of HepG2 cells to 5×10^5 cells per mL. And the cells were subsequently stained with binding buffer for 10 min at 4 °C, and labeled with Annexin V-fluorescein isothiocyanate (FITC) and propidium iodide. Sample were analyzed using a flow cytometer (BD Biosciences).

qRT-PCR assay. Total RNA was isolated from the HepG2 cells ($6 \times 6 \text{ cm}$, 2×10^6 /well) by using Trizol according to manufacturer's instructions and digested using DNase I (Takara) to remove the genomic DNA. The first-strand cDNA was synthesized using a cDNA synthesis kit (Fermentas, K1622). The mRNA expression levels of *Bax*, *Bcl-2*, *caspase-3*, *caspase-9*, *p53* and *p21* were measured by qRT-PCR in the 96-well plates with the real-time detection PCR system (Bio-Rad, Shanghai, China) with SYBR Premix *ExTaq*II (Takara) and the gene β -actin was used as an endogenous control. The primers used for the qPCR analysis were as follows: *Bcl-2* forward, 5'-CAAAGGTGGATCAGATTCAAG-3'; *Bcl-2* reverse, 5'-GGTGAGCATTATCACCCAGAA-3'; *Bax* forward, 5'-TGGCAGCAGTGACAGCAGCG-3'; *Bax* reverse, 5'-TACGGAGGTGGAGTGGGTGT-3'; *caspase-3* forward, 5'-AGAACACTGAAAACCTCAGTGGATTTC-3'; *caspase-3* reverse, 5'-TCCATTTTATAACTG-TTGTCCAGGG-3'; *caspase-9* forward, 5'-AGTCTATTTTATTATGGGCTCG-3'; *caspase-9* reverse, 5'-TGGATGTTTATGTCACCTTTTC-3'; *p21* forward, 5'-ATGGAGAACTGAAAACCTC-3'; *p21* reverse, 5'-TGTGAGCATGGAAACAA-TAC-3'; *p53* forward, 5'-ACTCCCATCTTCCACCTTTG-3'; *p53* reverse, 5'-CCCTGTTGCTGTAGCCATATT-3'; β -actin forward, 5'-CCATAAACGATGCCGGA-3'; β -actin reverse, 5'-CACCACCCATA-GAATCAAGA-3'.

Conclusions

In conclusion, a semi-synthesis methodology of 5-arylbenzofuran neolignans was developed. A series of 5-arylbenzofuran neolignan derivatives were synthesized and evaluated for their cytotoxicity *in vitro* against three tumor cell lines (HepG2, A549 and K562). Most derivatives revealed low cytotoxic effects and several compounds such as **23**, **26** and **30** displayed obviously better cytotoxicity than the natural 5-arylbenzofuran neolignan **1**. A preliminary structure–activity relationship was obtained, which showed that the modification of benzofuran, phenolic hydroxyl, allyl and the substituent on benzene ring made different effects on the cytotoxicity. For further understanding the anti-proliferative activities and mechanism of 5-



arylbenzofuran neolignans, a series of biological assays were done around these compounds. As a result, compounds **23** and **30** could induce the apoptosis of HepG2 cells through regulation of different signal pathways. The selectivity assay for cytotoxicity showed tumor cells were more sensitive to the promising compounds than normal cells. In addition, a one-pot method of Wacker-type cyclization and Wacker-type oxidation was developed, which may promote the synthesis of benzofuran ketones and related structures.

It's the first time for detailed research of 5-arylbenzofuran neolignans including semi-synthesis methodology, structural modification, cytotoxicity, structure–activity relationship and anti-proliferation mechanism after they were isolated from plants. The methods and conclusions in this paper would promote the research on 5-arylbenzofuran neolignan and its derivatives in the cancer therapy and other fields.

Conflicts of interest

The authors declare no competing interests.

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Notes and references

- 1 S. Apers, A. Vlietinck and L. Pieters, *Phytochem. Rev.*, 2003, **2**, 201–217.
- 2 J. K. Kim and J. G. Jun, *J. Cell. Biochem.*, 2011, **112**, 3816–3823.
- 3 M. S. Setzer, K. G. Byler, I. V. Ogungbe and W. N. Setzer, *Sci. Pharm.*, 2017, **85**, 5–32.
- 4 T. Morikawa, I. Hachiman, K. Matsuo, E. Nishida, K. Ninomiya, T. Hayakawa, O. Yoshie, O. Muraoka and T. Nakayama, *J. Nat. Prod.*, 2016, **79**, 2005–2013.
- 5 L. F. L. Barros, A. Barison, M. J. Salvador, R. D. Mello-Silva, E. C. Cabral, M. N. Eberlin and M. E. A. Stefanello, *J. Nat. Prod.*, 2009, **72**, 1529–1532.
- 6 W. L. Kuo, C. Y. Chung, T. L. Hwang and J. J. Chen, *Phytochemistry*, 2013, **85**, 153–160.
- 7 C. Y. Chung, W. L. Kuo, T. L. Hwang, M. I. Chung and J. J. Chen, *Chem. Biodiversity*, 2015, **12**, 1263–1270.
- 8 J. Ping; J. Qi, H. M. Feng, and K. Misun, PCT Pat., WO2017083363, 2017.
- 9 A. X. Hu, D. Lin, K. M. Li, J. Ye, A. L. Liu and W. W. Lian, *Chin. Pat.*, CN105622558B, 2016.
- 10 Z. B. Zhou, J. G. Luo, K. Pan, S. M. Shan, W. Zhang and L. Y. Kong, *Planta Med.*, 2013, **79**, 1730–1735.
- 11 T. Mitsudome, T. Umetani, N. Nosaka, K. Mori, T. Mizugaki, K. Ebitani and K. Kaneda, *Angew. Chem., Int. Ed.*, 2006, **45**, 481–485.
- 12 Q. Shen, Y. Dai, G. Wang, F. Yao, Y. Duan, H. Chen, W. Zhang, X. Zhang and X. Yao, *Bioorg. Med. Chem.*, 2014, **22**, 2671–2677.
- 13 Z. L. Kong, S. C. Tzeng and Y. C. Liu, *Bioorg. Med. Chem. Lett.*, 2005, **15**, 163–166.
- 14 N. C. Ulrich, J. G. Kodet, N. R. Mente, C. H. Kuder, J. A. Beutler, R. J. Hohl and D. F. Wiemer, *Bioorg. Med. Chem.*, 2010, **18**, 1676–1683.
- 15 T. Yamasaki, H. Hirose, T. Yamashita, N. Takakura, S. Morimoto, T. Nakahata, A. Kina, Y. Nakano, Y. O. Tamura, J. Sugama, T. Odani, Y. Shimizu, S. Iwasaki, M. Watanabe, T. Maekawa and S. Kasai, *Bioorg. Med. Chem.*, 2017, **25**, 4153–4162.
- 16 Y. Li, X. Qiang, L. Luo, X. Yang, G. Xiao, Y. Zheng, Z. Cao, Z. Sang, F. Su and Y. Deng, *Bioorg. Med. Chem.*, 2017, **25**, 714–726.
- 17 S. D. Tala, T. H. Ou, Y. W. Lin, K. S. Tala, S. H. Chao, M. H. Wu, T. H. Tsai, R. Kakadiya, S. Suman, C. H. Chen, T. C. Lee and T. L. Su, *Eur. J. Med. Chem.*, 2014, **76**, 155–169.
- 18 R. Bollu, J. D. Palem, R. Bantu, V. Guguloth, L. Nagarapu, S. Polepalli and N. Jain, *Eur. J. Med. Chem.*, 2015, **89**, 138–146.
- 19 D. S. Xie, Y. C. Wang, J. Xie, J. Lu, J. J. Cui, M. Zhang, L. Fu and Y. X. Wang, *Med. Chem. Res.*, 2014, **23**, 4977–4989.
- 20 S. M. Cloonan, J. J. Keating, S. G. Butler, A. J. S. Knox, A. M. Jørgensen, G. H. Peters, D. Rai, D. Corrigan, D. G. Lloyd, D. C. Williams and M. J. Meegan, *Eur. J. Med. Chem.*, 2009, **44**, 4862–4888.
- 21 J. B. Liu, R. H. Cao, W. Yi, C. M. Ma, Y. Q. Wan, B. H. Zhou, L. Ma and H. C. Song, *Eur. J. Med. Chem.*, 2009, **44**, 1773–1778.
- 22 G. T. Notte and J. L. Leighton, *J. Am. Chem. Soc.*, 2008, **130**, 6676–6677.
- 23 P. Thanigaimalai, K. C. Lee, V. K. Sharma, E. Roh, Y. Kim and S. H. Jung, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 3527–3530.
- 24 D. S. Grant, T. L. Williams, M. Zahaczewsky and A. P. Dicker, *Int. J. Cancer*, 2003, **104**, 121–129.
- 25 O. Meurette, A. Fontaine, A. Pebillard, G. L. Moigne, T. Lamy, D. Lagadic-Gossman and M. T. Dimanche-Boitrel, *Ann. N. Y. Acad. Sci.*, 2006, **1090**, 209–216.
- 26 X. Zhao, S. Y. Kim and K. Y. Park, *J. Med. Food*, 2013, **16**, 9–19.
- 27 G. Rodríguez-Berriguete, L. Galvis, B. Fraile, F. R. de Bethencourt, P. Martínez-Onsurbe, G. Olmedilla, R. Paniagua and M. Royuela, *Hum. Pathol.*, 2012, **43**, 229–237.
- 28 Z. Sun, Q. Han, L. Duan, Q. Yuan and H. Wang, *Oncol. Lett.*, 2018, **15**, 1999–2005.
- 29 S. M. Woo, Y. K. Choi, A. J. Kim, S. G. Cho and S. G. Ko, *Mol. Med. Rep.*, 2016, **13**, 1091–1096.
- 30 G. Hollmann, R. Linden, A. Giangrande and S. Allodi, *Aquat. Toxicol.*, 2016, **173**, 1–8.
- 31 M. Moulin, S. Carpentier, T. Levade and A. P. Arrigo, *Apoptosis*, 2007, **12**, 1703–1720.
- 32 A. Karimian, Y. Ahmadi and B. Yousefi, *DNA Repair*, 2016, **42**, 63–71.
- 33 L. Wang, Q. Lin, T. Yang, Y. Liang, Y. Luo, Y. Nie, J. Shen, X. Fu, Y. Tang and F. Luo, *J. Agric. Food Chem.*, 2017, **65**, 8374–8385.

