


Cite this: *RSC Adv.*, 2018, 8, 24485

Synthesis, characterization, physical and thermodynamic properties of a novel anionic surfactant derived from *Sapindus laurifolius*

Neha Saxena,^a Nilanjan Pal,^a Keka Ojha,^a Swapan Dey^b and Ajay Mandal  ^{*a}

The present study deals with the synthesis, characterization, physical and thermodynamic properties of a novel anionic surfactant derived from *Sapindus laurifolius* for its potential application against conventional non-biodegradable surfactants. The synthesized surfactant was characterized by FTIR, GC-MS, EDX and FE-SEM analyses. The surfactant showed good thermal stability at different temperatures as obtained from TGA studies. Critical micelle concentration (CMC) values were obtained by surface tensiometry measurements. DLS studies revealed the micelle structures of the CMC aggregates at higher concentrations. Low interfacial tension values were obtained at the oil–aqueous interfaces for surfactant solutions. The effect of temperature on the interfacial behaviour was also investigated. Thermodynamic studies showed that adsorption was more favoured in comparison to micellization for all systems. Foam stability studies were performed as a function of time and concentration by the Bartsch method. The surfactant also formed stable emulsions at concentrations near the CMC value. A comprehensive assessment of the thermal, interfacial, foaming and emulsifying properties of the soap-nut-based surfactant provides grounds for potential application in a wide range of industries.

Received 7th May 2018
Accepted 23rd June 2018DOI: 10.1039/c8ra03888k
rsc.li/rsc-advances

Introduction

Surfactants are organic compounds possessing both hydrophilic and hydrophobic groups. Because of their behavioural characteristic of reducing surface tension at the water–air and oil–water interfaces, they are used widely in various industries as flocculating and wetting agents, adhesives, inks, emulsifiers and de-emulsifiers and in the oil and gas industries.^{1–4} However, most of the commercial synthetic surfactants have significant detrimental effects on the environment, which has driven researchers to synthesize natural surfactants from renewable resources.^{5–7} The focus on an eco-friendly environment concerns the use of surfactants and creates interest to develop a natural surfactant against commercial non-biodegradable surfactant.

Natural surfactants are directly obtained from natural resources or represent a modified form of natural resources that are biodegradable in nature. In the past years, various natural surfactants have been reported that have been directly or indirectly derived from natural raw material belonging to the genus *Sapindaceae* and that can serve as an alternative source to other commercially available surfactants.^{8,9} Surfactants obtained from fatty acid esters of sugars, amides and the esters of fatty

acids can be used as a substituent for synthetic surfactants.^{10,11} Natural surfactant can also be prepared from the esterification of vegetable oils^{12–14} or the epoxidation of various edible and non-edible oil. These naturally synthesized surfactants have an advantage over synthetic surfactants as they are cost effective and biodegradable. A new non-edible source, namely the *Sapindus mukorossi* fruit, has been identified and studies have been conducted to investigate the possibility to use it in the field of surfactant science. *S. mukorossi* is a large tree belonging to the family *Sapindaceae*. This species is widely spread in a wide range and flourishes well in loamy soil and in regions experiencing around 80–100 cm annual rainfall. The fruit is about 2–2.5 cm in diameter and consists of a pericarp and a hard seed. The soap-nut kernel has about 30% oil content by weight. The soap-nut fruits are reported to contain glycosides and fatty ester of tetracyclic triterpenoids. Soap-nut pericarp is traditionally used as a cleaning agent in laundries.

The increased use of commercial surfactants for industrial and domestic purposes imposes serious concerns for the environment as most of their constituents are dispersed in various compartments of the environment, like in soil, water, sediment, etc. The synthetic surfactants used nowadays can restrict the auto-filtering activity of marine animals.¹⁵ Such synthetic compounds thus cause serious pollution to the marine life and can also cause much damage to humans in the form of dermatitis, irritation and respiratory problems. Consequently, researchers around the world are actively seeking alternative

^aDepartment of Petroleum Engineering, Indian Institute of Technology (Indian School of Mines), Dhanbad-826 004, India. E-mail: ajay@iitism.ac.in

^bDepartment of Applied Chemistry, Indian Institute of Technology (Indian School of Mines), Dhanbad-826 004, India



sources of these synthetic surfactants that are economically viable and environmental friendly.¹⁶

The main objective of the present study is to synthesize a natural surfactant from soap-nut and to provide a substitute to commercially available surfactants. Characterization of the synthesized product was done using GC-MS, FTIR analysis, FE-SEM and EDX analysis. The interfacial properties at the air-water interface of the synthesized surfactant were investigated at different concentrations and as a function of temperature. The interfacial tension of the soap-nut surfactant at the oil-water interface was studied by varying the effective alkane carbon number (EACN). The thermodynamic and adsorption properties were also studied for the surfactant system using parameters like micellization, the Gibbs free energy of micellization and adsorption derived from a surface tension study. The foamability of the surfactant solution was studied as a function of time to predict the particle adsorption at the bubble surface and foam stability. The emulsion stability was also studied with heptane to determine the efficiency of the surfactant to be used as an emulsifier.

Experimental methods

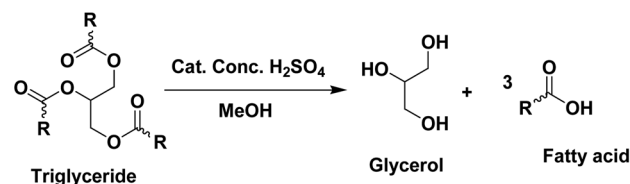
Materials required

Soap-nut fruits were procured from a local market. Ethanol, concentrated sulfuric acid, diethyl ether, pyridine, chloro-sulfonic acid, sodium carbonate, sodium hydrogen carbonate and normal butanol were obtained from Merck (India). Alkanes, like heptane, decane and dodecane, were obtained from Rankem, India. Table 1 represents the constituents of the soap-nut oil derived from the soap-nut fruit obtained from the central region of India.

Synthesis of an anionic surfactant from soap-nut pericarp

Step 1 Oil extraction and esterification. The fruit pericarp was separated from the seed by manual crushing. The pericarp and seeds were dried in an oven for 24 h to remove the moisture. The dried components were well crushed so they could be used for the oil extraction process. The oil was extracted from the seed in a Soxhlet apparatus using a solvent system. Here, 100 g of dried seed powder was placed in a round-bottomed flask with 600 mL of petroleum ether as a solvent, under the approximate temperature of 343 K overnight, protected from light for a period of 12 h. The oil was extracted by removing the solvent under reduced pressure by a rotatory evaporator at a temperature of 323 K. The oil obtained was weighted and the oil percentage was measured on the basis of dry weight. Soap-nut

Scheme 1a:



Scheme 1b:

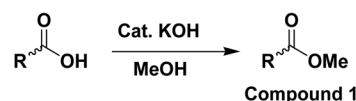


Fig. 1 Reaction scheme depicting the synthesis of the fatty acid methyl ester.

oil obtained from the soap-nut was esterified using an acid and base as a catalyst. The vegetable oil was mixed with methanol in the ratio (1 : 5 and 1 : 10) and a drop of KOH (base) was added to the reaction mixture to catalyze the reaction. The alcoholysis reaction in (Fig. 1) was reversible, therefore, an excess amount of alcohol was used to shift the equilibrium in the direction of product formation. The soap-nut oil used in the trans-esterified process was pre-treated to remove the free fatty acid content.

The percentage of esterification could be calculated by the formulae in eqn (1):

$$\text{Esterification\%} = \frac{\text{mass}_{\text{actual}}}{\text{mass}_{\text{theoretical}}} \times 100 \quad (1)$$

The degree of esterification was studied as a function of the oil methanol ratio (1 : 5, 1 : 10), and the percentage yield for the above two ratios was found to 87.6% and 88.9%. With the increasing concentration of methanol, the percentage yield was found to increase.¹⁷

Step 2 Sulfonation reaction. The intermediate product (compound 1) obtained in the trans-esterification reaction was used to synthesize the desired surfactant by sulfonation of the soap-nut oil methyl ester. In this the sulfonation process, chloro-sulfonic acid was added to pyridine in a round-bottomed reaction flask. The reaction was carried out under constant stirring for 30 min in an ice bath as the reaction was exothermic in nature and soap-nut methyl ester was added to the reaction mixture. The reaction mixture was heated at 333 K to obtain a clear solution, which was then quenched with a saturated solution of sodium carbonate and sodium hydrogen carbonate. Normal butanol was used to dissolve the unreacted components in the organic layer and to obtain the desired product in an aqueous layer, which was then rota-evaporated to obtain a solid mixture. The organic impurities from the final product were eliminated by washing it with petroleum ether carefully. The final slightly brown solid product (compound 2) was obtained by vacuum drying the product at 338 K overnight and was then characterized by FTIR and GC-MS. The soap-nut oil used in the synthesis chiefly contained oleic acid, palmitic acid, linoleic

Table 1 Properties and composition of the soap-nut oil

Properties	Value
Acid value (mg KOH per g)	13.14
Density (kg m ⁻³)@288 K	912
Kinematic viscosity (mm ² s ⁻¹)@313 K	40.08
Saponification value (mg KOH per g)	201.2



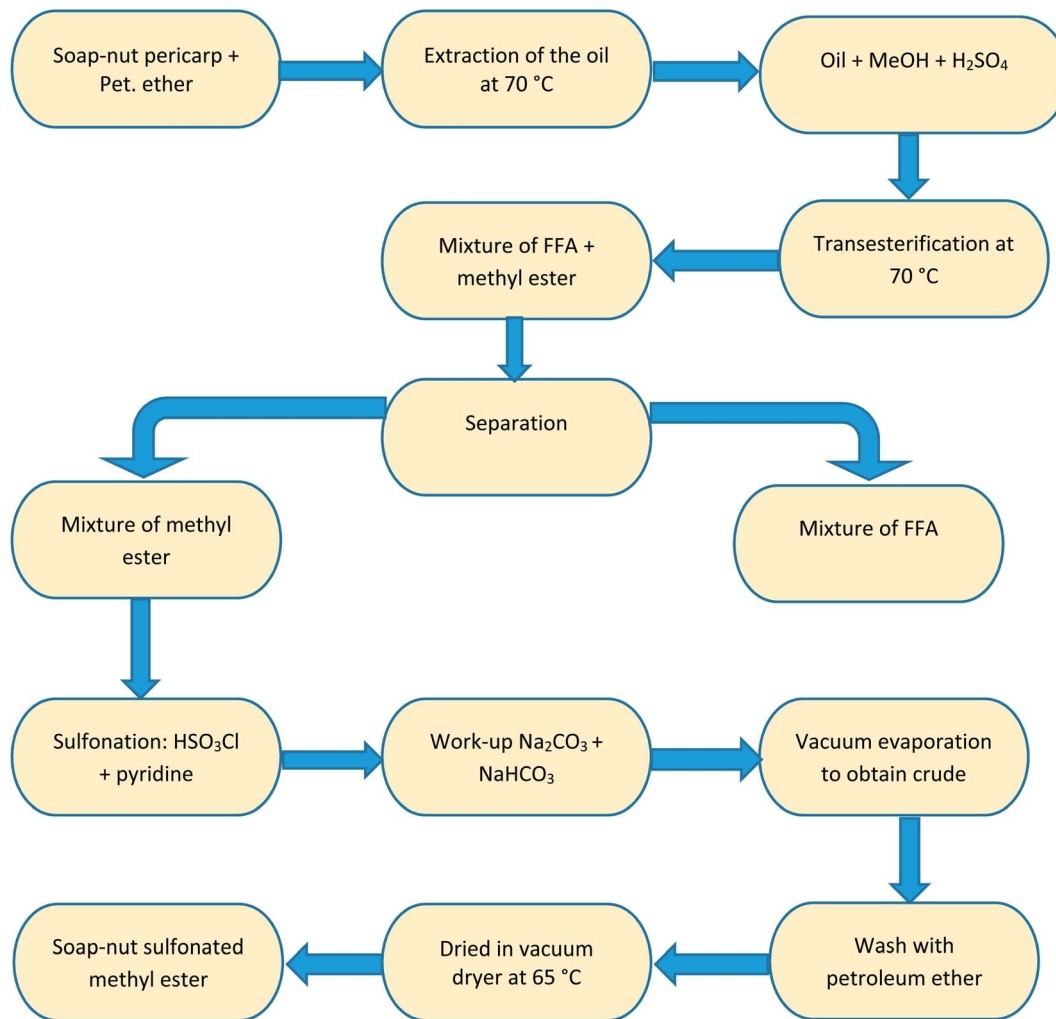


Fig. 2 Flowchart showing the stepwise synthesis of the soap-nut surfactant.

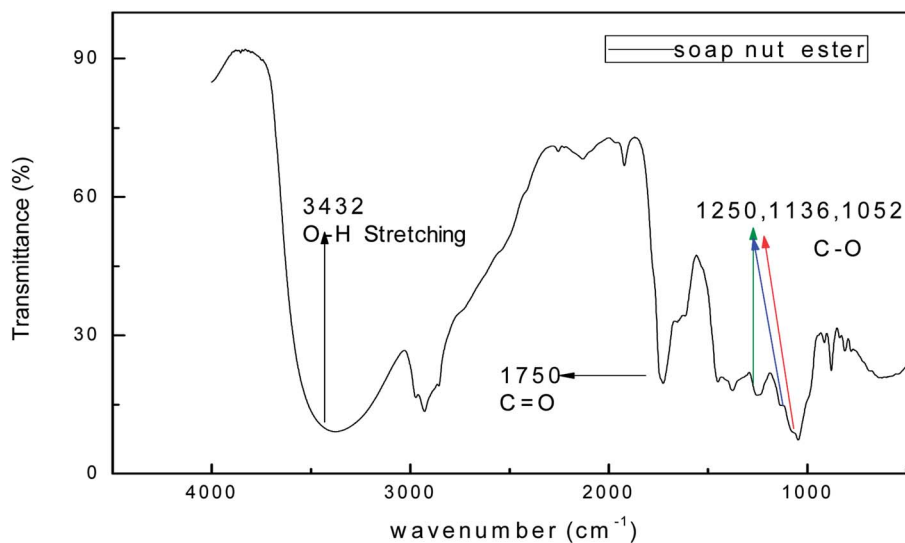


Fig. 3 FTIR spectrum of the soap-nut ester formed as an intermediate product.

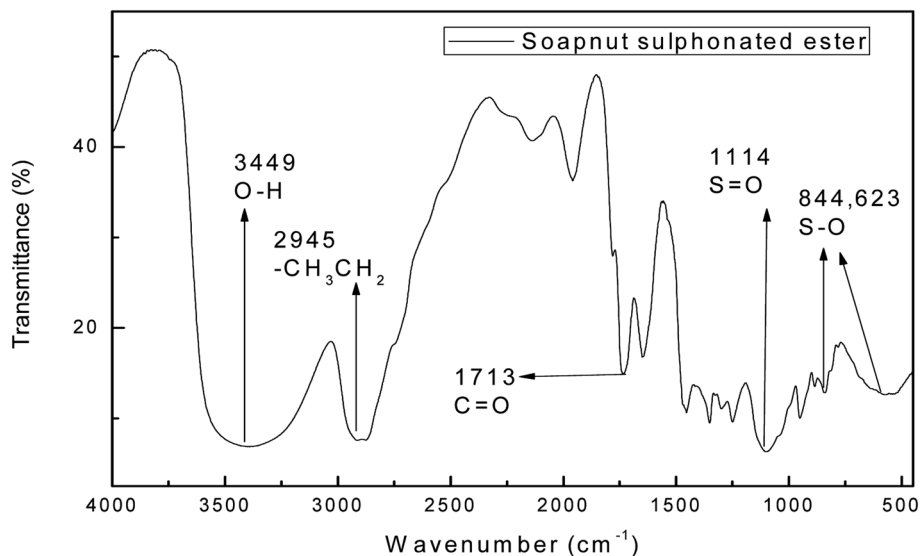


Fig. 4 FTIR study of the soap-nut sulphonated ester surfactant.

acid and arachidic acid along with traces of unidentified saturated and unsaturated fatty acids. Hence, the final solid surfactant formed in the sulfonation process was a mixture of the sulfonated ester. A flowchart representing the preparation step of the soap-nut surfactant is shown in Fig. 2.

Characterization by FTIR, GC-MS, TGA and FE-SEM analyses

The FTIR analysis of the synthesized surfactant was done using a Perkin Elmer-Spectrum 2 spectrophotometer to detect the functional group constituents of the newly synthesized sulfonated ester surfactant derived from the soap-nut oil. A pellet of synthesized soap-nut surfactant was prepared using KBr for FTIR analysis. The surface morphology of the

synthesized surfactant was analyzed by FE-SEM analysis using a SUPRA-55 instrument (ZEISS, Germany). The soap-nut surfactant was made moisture free by keeping it in a desiccator for 24 h. Gas chromatography coupled with mass spectrometry (Jeol GC-Mate II GC-MS) was used to determine the constituents of the methyl ester synthesized from the soap-nut oil. TGA analysis of the surfactant synthesized from the soap-nut oil was done to calculate its thermal stability using a thermogravimetric analyzer (Netzsch-STA 449 Jupiter).

Conductivity measurements

The electrical conductivities of the synthesized soap-nut surfactant solutions with varying concentrations were

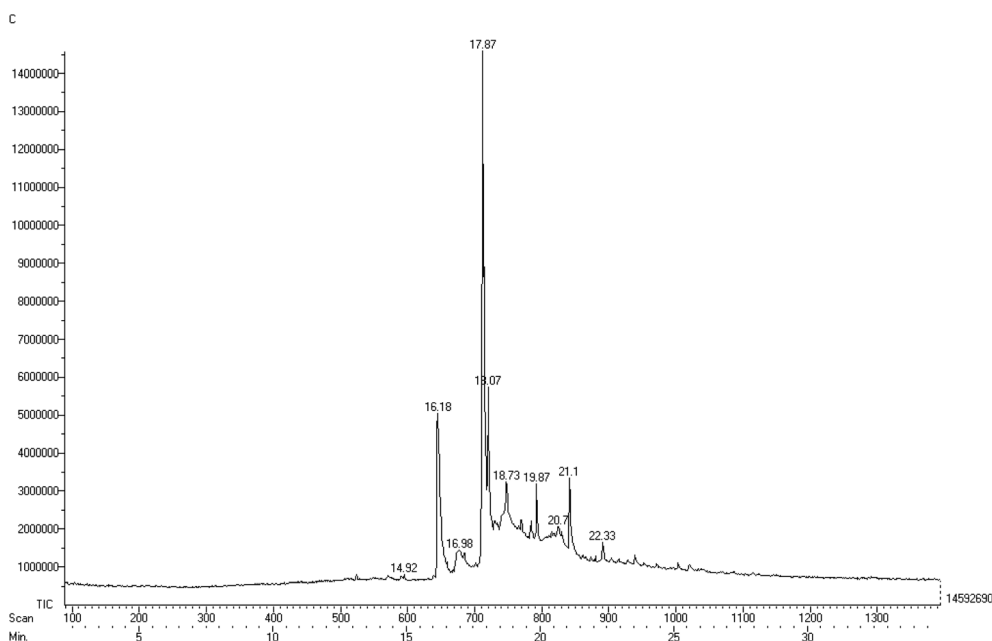


Fig. 5 Gas chromatogram of the different components of the soap-nut oil methyl ester.



Table 2 Results of the GC-MS analysis of the soap-nut oil methyl ester

Acid methyl ester	Molecular ion M^+ (m/z)	Retention time in the chromatogram (min)	Concentration (% \pm SD of total area in chromatogram)		
			Present study	Deka <i>et al.</i> ²¹	Jia <i>et al.</i> ²²
Palmitic	270	16.18	(20.83 \pm 1.26)	4.78	3.89
Linoleic	294	16.98	(0.91 \pm 0.10)	—	8.24
Oleic	296	17.87	(52.68 \pm 0.99)	58.8	54.19
Stearic	298	18.07	(6.58 \pm 0.09)	1.78	1.17
11-Eicosenoic	324	19.87	(4.11 \pm 0.23)	25.93	22.31
Arachidic	326	20.7	(13.09 \pm 0.09)	8.61	5.31
Behenic	354	21.1	(1.02 \pm 0.03)	—	0.97

determined using a Hanna HI-2003 conductivity meter at 298 K, 313 K, 323 K, 333 K and 348 K. The electrical conductivity of a solution depends on the mobility and number of ions or charged particles present in the solution. In the case of ionic surfactants, the electrical conductivity increases upon increasing the surfactant concentration. The formation of micelles affect the conductivity of an ionic solution. The critical micelle concentration (CMC) was obtained from the break or inflection point (intersection of linear fits) in the conductivity *versus* surfactant concentration curve. The value of the conductivity was taken after allowing the readings to stabilize and reach a constant value. After each measurement, the probe was immersed in a standard solution for calibration.

Interfacial tension measurements at the air–water interface

The interfacial tension at the air–water interface and the CMC of the synthesized surfactant were measured using the Du Nouy ring method (Easy Dyne, Kruss Germany) in a wide range of temperatures (298 K, 313 K, 323 K, 333 K, 348 K). The surface tension was calculated for the aqueous solution of the synthesized surfactant at different surfactant concentrations. Surface tension values for each individual sample were measured three times to maintain the consistency of the results, and the final recorded values were taken as the average of the measured values. After measuring each set of different concentrations, the platinum ring used in the experiment was cleaned using acetone and was dried with a flame for the next set of experiments. Electrical conductivity measurements were performed with a Hanna HI-2003 conductivity meter to determine the Krafft temperature.

Dynamic light scattering measurements

The size distribution of surfactant molecules in aqueous solution was determined by dynamic light scattering experiments at 298 K using Zetasizer Nano S90 (Malvern Instruments, UK). It measures the particle size by measuring rate of diffusion of particles in solution using Stokes–Einstein equation. Measurements were performed at two different concentrations of surfactant solution, one at the CMC (9.3 mM) and the other at a higher concentration (11.3 mM). In this technique, the diffusion of particles moving under Brownian motion is measured and converted to the hydrodynamic diameter (d_h) of

droplets using Stokes–Einstein eqn (2), which is expressed below as

$$d_h = \frac{k_b T}{3\pi\eta D} \quad (2)$$

where k_b is the Boltzmann constant, D is the diffusion coefficient and η is the viscosity of solvents at temperature (T). Three measurements were taken for each sample to check the consistency of the results.

Interfacial tension measurements at the oil–water interface

The interfacial tension at the oil–water interface and the CMC of the synthesized soap-nut surfactant was measured by using a spinning drop tensiometer (Data Physics) at 303 K. Interfacial tension was measured for an aqueous solution of the synthesized surfactant at different concentrations. Interfacial tension values for each individual sample were measured twice to check the accuracy of the results. The measured value of the interfacial tension of distilled water at the oil–water interface using heptane was found to be 48 mN m^{−1} at 303 K.

Foamability and foam stability studies

Foams were prepared by the Bartsch method,¹⁸ whereby the foam was created by 20 vigorous shakings by hand of a closed cylinder containing 15 mL of surfactant solution and the remaining air (60 mL height). The mixture was shaken 10 times, with the same intensity for every set of experiments, and the foam height was observed for a period of 60 minutes.¹⁹ This method was used to evaluate the foam stability. At any instant, the foam height was measured as the difference between the total height of the foam–liquid system and the liquid level in the column. In this study, another important parameter known as the half-decay time was determined for each surfactant concentration. The half-decay time is defined as the time taken by the foam to reach one-half of its initial foam height value. It is also a parameter of the foam stability property, whereby the longer the half-decay time value, the greater is the foam stability.

Emulsion stability

The emulsification properties of the soap-nut surfactant were studied.²⁰ Oil and the aqueous surfactant solution at different concentrations were mixed thoroughly by vigorous shaking in



Scheme 2

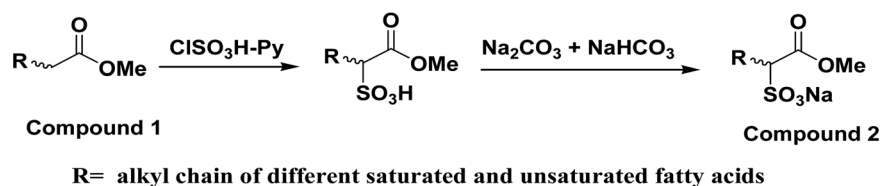


Fig. 6 Reaction scheme for the synthesis of the soap-nut surfactant.

conical graduated tubes. The stability of the emulsion was tested by measuring the time required to separate the surfactant solution from the emulsion.

Results and discussion

FTIR spectrum analysis

The FTIR spectrum of the synthesized surfactant was recorded in the range of 4000–400 cm^{-1} . The IR spectra of the soap-nut ester and soap-nut sulfonated ester are shown in (Fig. 3 and 4), respectively. The absorption band at 3452 cm^{-1} displays a weak stretching vibration of an hydroxyl group. The peak at 2939 cm^{-1} indicated the symmetrical vibration of a cis olefinic $=\text{C}-\text{H}$ double bond. The sharp peak at 1750 cm^{-1} is the characteristic stretching vibration of a carbonyl carbon $\text{C}=\text{O}$ bond constituted by the ester group. Bending vibrations of aliphatic groups $-\text{CH}_2$ and $-\text{CH}_3$ are depicted at 1448 cm^{-1} . The absorption bands at 1250 cm^{-1} , 1136 cm^{-1} and 1052 cm^{-1} indicate the asymmetrical axial stretching vibration of the $\text{C}-\text{O}$ ester group.

Fig. 4 shows the FTIR spectrum of the surfactant derived from the soap-nut oil methyl ester. The characteristic peak for the ester group of the intermediate ester is also present in the final synthesized product. The ester group peak is slightly shifted from the previous point, which may be due to the introduction of a polar group, which leads to the shift in peak to 1713 cm^{-1} . The absorbance bands for different functional groups and bonds are depicted in Fig. 4. The newly introduced absorption peaks for the stretching vibration of sulfonate groups $\text{S}=\text{O}$ and $\text{S}-\text{O}$ appear at 1114 cm^{-1} , 844 cm^{-1} and 623 cm^{-1} respectively. Thus, the FTIR results confirm the

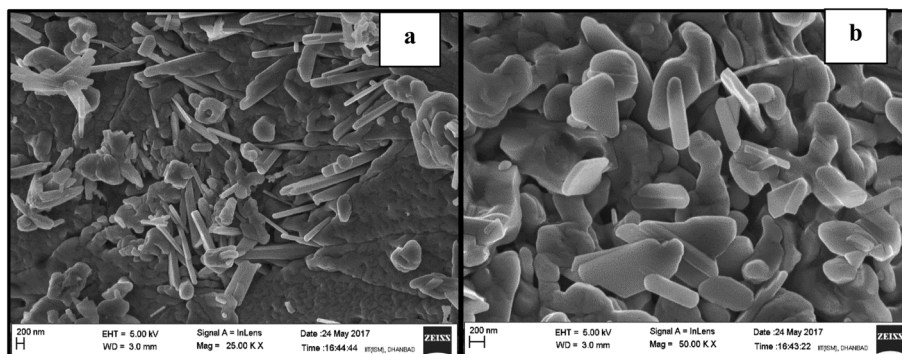
sulfonation of the soap-nut methyl ester to form the resultant desired product of a methyl ester sulfonate.

GC-MS analysis of the soap-nut surfactant

Seven fatty acid methyl esters were analyzed and identified using gas chromatography combined with mass spectrometry (GC-MS) in the soap-nut oil methyl ester formed after transesterification of the extracted soap-nut oil. The fatty acid methyl ester detected was a mixture of unsaturated and saturated forms, mainly palmitic (16 : 0), linoleic (18 : 2), oleic (18 : 1), stearic (18 : 0), eicosenoic (20 : 1), arachidic (20 : 0) and behenic (22 : 0), as depicted in Fig. 5. The fatty acid methyl ester composition was identified using mass spectrometry and compared to the library, which permitted us to attribute the molecular formulae of different derivatives forming the constituents of the fatty acid. Table 2 also provides the retention time of the elutions along with the relative area of each peak identified in the gas chromatogram. Among the all identified esters, it was observed that oleic acid was the major component in the soap-nut oil extracted from the soap-nut fruit. The composition of the soap-nut oil methyl ester was found to be similar to the composition reported by other research groups.^{21,22}

Reaction scheme of the soap-nut surfactant

Fig. 6 shows the mechanism of reaction for the synthesis of the soap-nut surfactant. The fatty acid obtained after extraction was first esterified in an acid-catalyzed reaction in the presence of methanol to obtain a methyl ester moiety in the fatty acid chain. In the sulfonation step, pyridine is added as a base to abstract

Fig. 7 FE-SEM images of the soap-nut surfactant at magnifications of: (a) 25k \times (b) 50k \times .

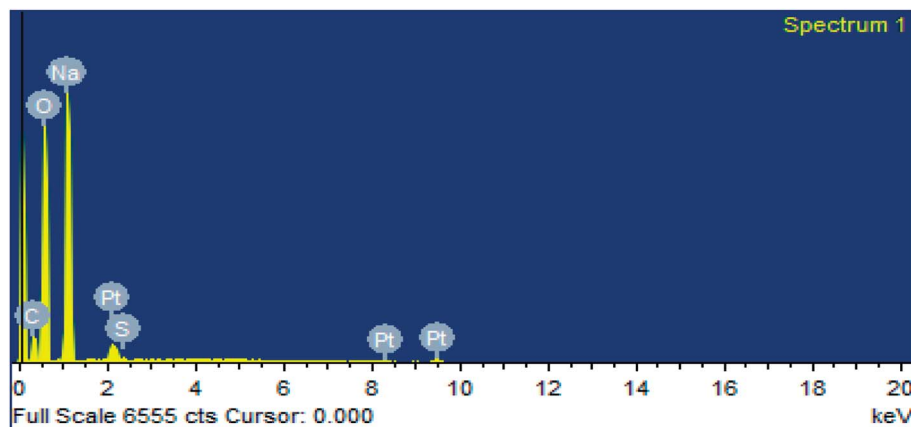


Fig. 8 EDX analysis of the synthesized surfactant.

the α -hydrogen from the ester group, being the most reactive site for the reaction to take place easily compared to the other available reactive sites, like the unsaturation sites present in the fatty acid. The sulfonate group was attached to the α -position of the ester group.^{23–25}

FE-SEM analysis of the soap-nut surfactant

FE-SEM analysis of the surfactant derived from soap-nut oil was performed to study the surface morphology. Fig. 7a at 25k \times zoom shows that the surfactant molecules appear to be in cluster form and are compact and bound tightly with the vicinal molecules. The image shown in Fig. 7b at 50k \times zoom illustrates that the particles are rod-like and irregular in shape, and are well dispersed within the area under analysis.

EDX spectrum of the soap-nut oil surfactant

In EDX analysis of the soap-nut surfactant, five elements were observed: carbon (C), oxygen (O), sodium (Na), sulfur (S) and platinum (Pt), as shown in (Fig. 8). The platinum (Pt) peak

characterized in the spectra was due to the presence of coating materials used along with the surfactant during the analysis. All the remaining elements present correspond to different components of the synthesized surfactant. The EDX data clearly show the presence of sulfur as a chloro-sulfonic acid used in the sulfonation reaction, which indicates the formation of the desired product. The presence of sodium shows that the anionic surfactant exists in the form of a sodium salt. No extra peak of any other element is depicted in the EDX spectra, which indicates that the synthesized surfactant is pure and free from any other elemental impurities.

Thermal gravimetric analysis of the synthesized surfactant

The thermal stability of the soap-nut surfactant was studied by thermogravimetric analysis (TGA). The weight loss percentages are depicted with the increase in temperature in Fig. 9. The TGA plot shows that the initial thermal loss occurs from 373 K to 533 K, wherein about a 6% loss in weight is observed. The initial decomposition up to 373 K marks the removal of moisture from

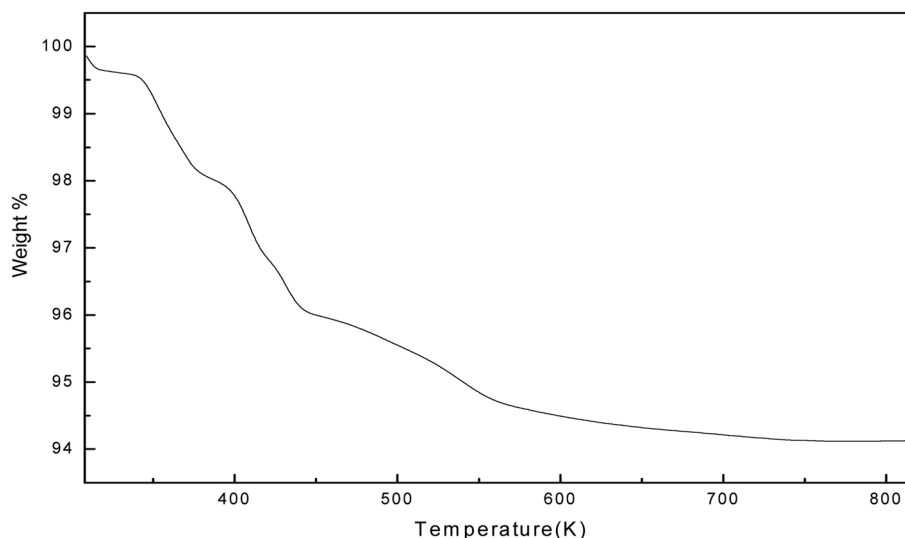


Fig. 9 Thermal stability curve for the synthesized surfactant.



the surfactant. The second decomposition peak marks the decomposition of any oxides present as impurities in the surfactant and the final peaks marks the degradation of surfactant at a temperature of around 523 K. No further loss in weight was observed above this temperature, which shows that the soap-nut surfactant was thermally stable even at higher temperatures. The presence of both saturated and unsaturated fatty acids in the soap-nut oil supports the good thermal stability of the synthesized surfactant as these fatty acids require a high temperature for their degradation. The anti-oxidants present in soap-nut oil aid have good thermal stability.

The thermogravimetric analysis of the soap-nut surfactant was compared with the other commercial surfactants available on the market, like sodium dodecyl sulfate (SDS) and a natural surfactant derived from castor oil. Upon comparing the results of TGA, it was found that the soap-nut surfactant was stable at up to 473 K for its first weight loss but SDS and castor oil surfactant showed their initial degradation below 400 K and even a loss of 30–40% weight up to 450 K. This clearly indicates that the surfactant synthesized from soap-nut oil can be applied for application in diverse fields where there is a requirement for high temperature stability.^{26,27}

Determination of the critical micelle concentration

Conductivity studies were conducted to determine the CMC of the surfactant system. The effects of concentration and temperature on the CMC are shown in Fig. 10. Using Williams' method, the CMC value was obtained at the breakpoint of the linear points of the conductivity *versus* concentration plots.^{28,29} The sudden change in slope was attributed to the binding of counterions to the micelles to flocculate and shows the lower mobility of micelles in comparison to surfactant molecules that do not flocculate at a lower surfactant concentration. It was observed that the CMC increased with temperature due to a delay in the micellization process in the aqueous phase. The

increase in temperature distorts the structured water contiguous to the hydrophobic group, which leads to a shifting of the monomers–micelle equilibrium in favour of the monomer and promotes micellization at a higher concentration of surfactant.³⁰ The CMC values of the soap-nut surfactant were found to be 9.3 mM at 298 K, 9.8 mM at 313 K, 10.3 mM at 323 K, 10.8 mM at 333 K and 11.3 mM at 348 K.

Surface tension, Krafft temperature and critical micelle concentration at the air–water interface

The surface tension of water at the air–water interface decreases to a minimum value and becomes almost constant or it may show a slight increase in this value with the increase in concentration of the surfactant solution, causing an improved adsorption ability at the interface with the increase in concentration. The sharp decrease in the value of surface tension indicates that the surfactant molecules have greater affinity towards the air–water interface. The specific concentration value at which the surfactant molecules are completely adsorbed at the air–water interface, thus favouring the formation of micelles, is called the critical micelle concentration (CMC). To determine the Krafft temperature, the surfactant solution (0.9 wt%) was prepared by adding 50 mL of hot water and then keeping under refrigeration at around 277–278 K for 24 h. The temperature of the cooled surfactant solution was raised gradually under constant stirring, and the conductance (κ) was measured with every 2 K rise in temperature. A sudden change in the value of κ *versus* temperature was not seen as the Krafft temperature of the surfactant solution was observed to be less than 277 K. The surfactant solution showed clear visibility at room temperature of 303 K, which showed its good water solubility due to the polar sulfonate group present in the chemical compound.

Natural surfactants prepared from vegetables oil have higher CMC values and show a greater tendency of adsorption at air–water interfaces. These types of surfactant require more a number of surfactant molecules to be adsorbed at the interface. The surface activity of the surfactant molecules at the air–water interface varies with the concentration of the surfactant solution in the bulk phase. The surface tension *versus* concentration curves of surfactant solutions at different temperatures are depicted in Fig. 11. As the concentration of the synthesized surfactant solution increases, the surface tension values decrease as there is an enhancement in adsorption of the surfactant molecules at the interface of air and water,³¹ and this trend was observed until the concentration reached the CMC value at approx. (as also observed by conductivity studies) 9.3 mM, 9.8 mM, 10.3 mM, 10.8 mM and 11.3 mM at 298 K, 313 K, 323 K, 333 K and 348 K having respective surface tension values of 37.1, 33.4, 32.4, 29.6 and 28.6 mN m^{−1}. It was observed that as the temperature increases, the CMC of the surfactant increases, due to the lower probability of hydrogen bond formation at higher temperatures. A similar trend of increasing CMC and decreasing surface tension values was observed by other researchers.^{32,33} After the CMC, the surface tension curve attains a constant value as the surface becomes saturated with

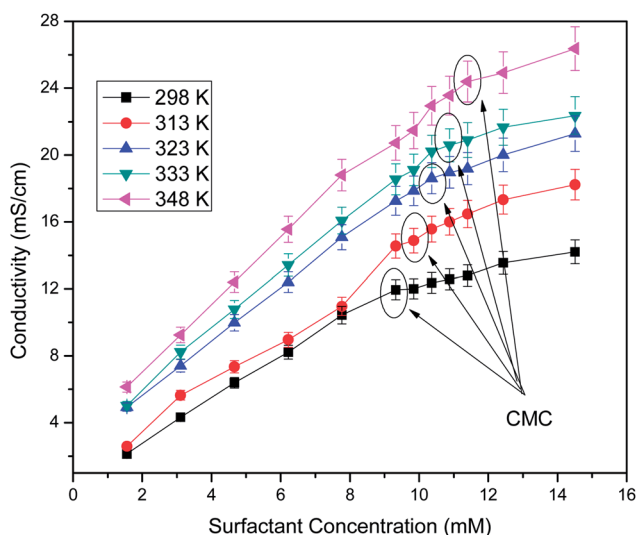


Fig. 10 Electrical conductivity *versus* surfactant concentration at different temperatures.



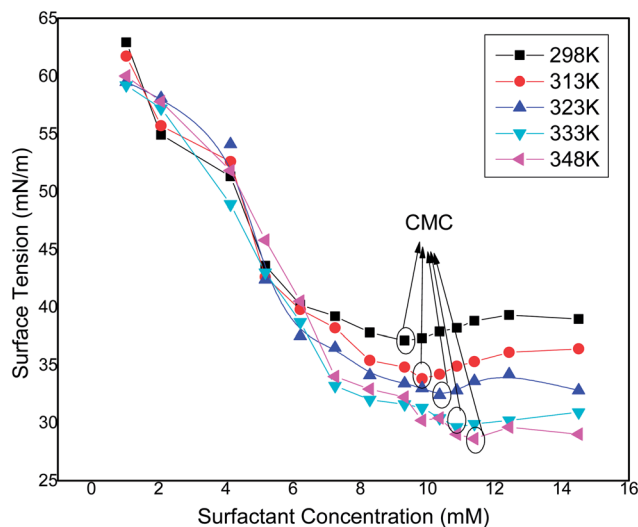


Fig. 11 Variation of surface tension (standard deviation $\pm 0.05 \text{ mN m}^{-1}$) with the concentration of the synthesized surfactant at different temperatures.

the surfactant molecules. The experiment was repeated three times to check the consistency of the results and the average of three readings was taken into consideration.

Dynamic light scattering measurements

Fig. 12 shows the intensity-weighted distribution profile of different surfactants at 9.3 mM (CMC), 10.3 mM and 11.3 mM. At the CMC, the micellar structures begin to form and a single peak in the size ranges from 5–25 nm of the micelles is observed. At higher concentrations, additional surfactant molecules aggregate towards the micellar structures. This is because of the non-availability of vacant sites for surfactant adsorption at the interface. The intensity of particle distribution

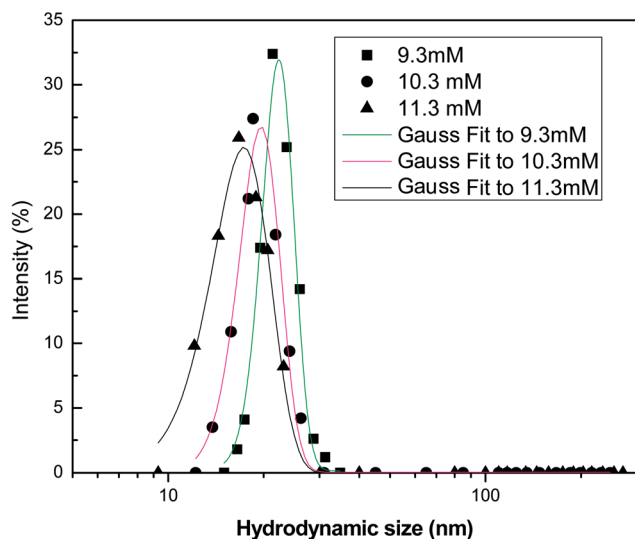


Fig. 12 Intensity-weighted particle size distribution profile of the soap-nut surfactant at different surfactant concentrations.

Table 3 Analysis of the scattering intensity versus size plots of the surfactant system using a Gaussian probability distribution function

Surfactant concentration	Fitting parameters	Distribution function (Gaussian)
9.3 mM	Std. deviation (w)	2.598
	Area (A)	207.093
	Offset (y_0)	0.145
	Centre (x_c)	22.321
	Adj R^2	0.967
10.3 mM	Std. deviation (w)	2.922
	Area (A)	195.942
	Offset (y_0)	0.038
	Centre (x_c)	19.62
	Adj R^2	0.984
11.3 mM	Std. deviation (w)	3.679
	Area (A)	232.901
	Offset (y_0)	−0.07
	Centre (x_c)	17.313
	Adj R^2	0.991

profile increases with concentration, due to the greater tendency of the surfactant monomers to form aggregates. Thus, micellization is more favoured when the surfactant concentration exceeds the (CMC value). At the CMC, the adsorption of surfactant molecules at the interface ceases and additional monomers interact with the micelles present at the bulk aqueous phase, resulting in the micellar size to increase.

In a surfactant system, continuous flocculation and deflocculation of the surfactant molecules occur, which is due to the variation in particle size and their distribution. Therefore, modelling of the DLS data is important to analyse the size and behaviour of molecules in the aqueous system. The probability distribution analysis showed good fitting with the experimental data as the adjusted coefficients of determination for the Gaussian model were found to be 0.967, 0.984 and 0.991 for the 9.3 mM, 10.3 mM and 11.3 mM solutions, respectively. This shows that the size distribution profile obtained from analysis of the surfactant solutions follows a normal or Gaussian statistical behaviour. This was also supported by the low values of offset obtained for the different analyzed systems. The adjusted coefficient values of distribution parameters are represented in Table 3.

Interfacial tension measurements at the oil–water interface

Studies of the interfacial tension between oil (heptane, decane and dodecane) and the soap-nut surfactant solutions at different concentrations at 303 K are depicted in (Fig. 13). The interfacial tension initially decreases to a minimum value, due to the dynamic equilibrium existing between adsorption and desorption of the surfactant molecules at the oil–water interface. As the aqueous phase interacts with the oil phase, surfactant molecules show a diffusion phenomenon from the bulk to the water interface, while at the interface of oil and water, phase adsorption occurs. Here, at first, the adsorption rate of molecules was higher than the desorption rate, leading to a lowering of the interfacial tension. Finally, when an equilibrium state was achieved and the adsorption and desorption



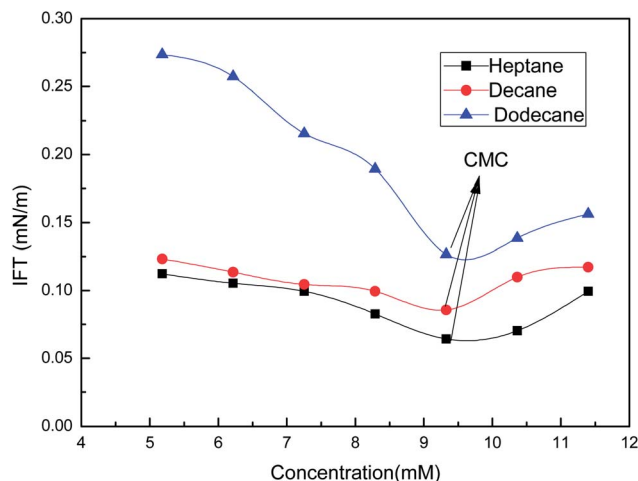


Fig. 13 Variation of the interfacial tension with the concentration of the soap-nut surfactant at the oil-water interface.

forces were balanced, the IFT approached towards a constant value.³⁴ At 9.3 mM (the CMC), the value of IFT was found to be lowest in all the oils tested. As the surfactant concentration was further increased, micelle formation took place, resulting in a slight increase in IFT. It was found that with the increasing EACN number of the alkanes, the tendency to reduce the IFT decreases.³⁵

Foamability and foam stability of the synthesized surfactant

The foam height can be calculated by the net foam generated in the foaming process. At a particular instance of time, the foam height can be evaluated from eqn (3);

$$h_{\text{foam}} = h_{\text{total}} - h_{\text{liquid}} \quad (3)$$

where h_{foam} , h_{total} and h_{liquid} as a function of time are depicted in Fig. 14a. The foam stability increases with increasing the concentration³⁶ of the surfactant solution and becomes constant after it has reached a certain value. At low surfactant concentration, the free concentration of the surfactant is depleted because of its adsorption onto the particles. The surfactant molecules cannot stabilize the foam, but with increasing the concentration, the drainage from the foam slows down, which favours the foam stability.³⁷ Amphiphilic particles promote foaming by attachment to the bubble surface as individual particles but at higher concentration, say above the CMC, the particles concentration foamability is inhibited due to flocculation of the bubbles.³⁸ In addition, it can be observed from Fig. 14b that the half-life time increases with the surfactant concentration. The half-decay time values for the surfactant solutions were 48, 49, 52, 54, 55, 58 and 59 min for 7, 9, 11, 13 mM solutions, respectively. However, it was also evident that the foam stability does not increase drastically, but gradually with the surfactant concentration, showing that a quite longer foam life could be observed as a result of the optimization of the surface and interfacial properties of the surfactant.³⁹

Emulsion stability

Surfactants possess an important property of emulsification, which leads to their use as emulsifying agents in various industrial applications. The amphiphilic property of a surfactant helps in solubilizing the water in non-polar liquids, like oils. Surfactants get adsorbed on the oil-water interface to reduce interfacial tension. This subsequently leads to a decrease in the energy required to form the oil-water interface for the emulsion. The oil-in-water emulsion so formed shows the emulsification ability of the surfactant. Here, emulsions formed by a surfactant near the CMC were found to be stable and did not separate for more than 180 min, when compared to the other lower concentrations, which were less stable for the time lapse the experiment was under observation for, as shown in (Fig. 15). At the CMC, it was observed that the stability time was the highest, showing the greatest packing density within the cross-sectional area per adsorbed surfactant molecule.^{40–44} With the further increase in concentration, the packing density of surfactant molecules at the oil-water interface remain unaltered, due to the unavailability of vacant sites for adsorption.

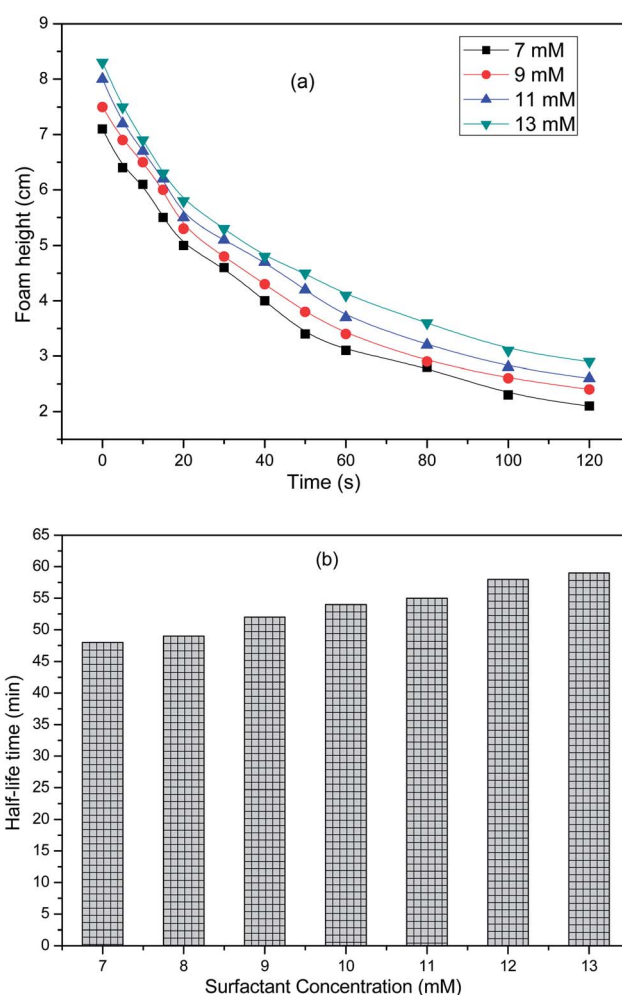


Fig. 14 Foaming properties of the surfactant solutions: (a) foam height as a function of time at different surfactant concentrations, (b) half-life time as a function of the surfactant concentration.



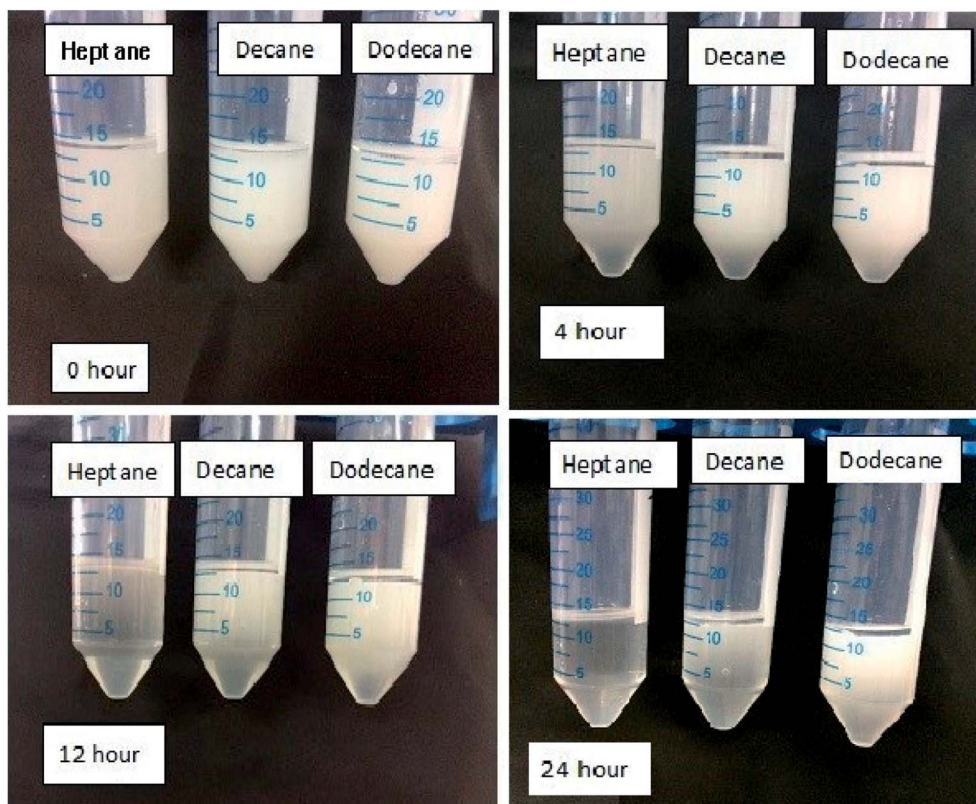


Fig. 15 Images depicting the stability of emulsions at the CMC with the lapse of time at 303 K.

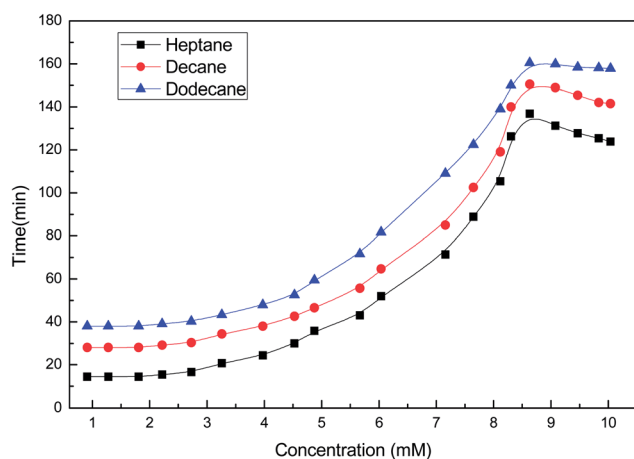


Fig. 16 Emulsion stability tests of the soap-nut surfactant at different concentrations.

The stabilizing effect of oil-in-water emulsions at concentrations near the CMC could be attributed to the ability of long-chain alkanes in the oil to associate and form mixed monolayers with the adsorbed ionic surfactant at the oil-water interface.^{45–47} Therefore, the adsorption tendency is favoured till the CMC concentration is reached, following which the micellization tendency in the bulk oil phase is favoured. The emulsions formed in the presence of 5% oil (heptane, decane and dodecane) were found to remain stable even after 24 h visual observation, as evident from Fig. 15 and 16. Emulsions formed

with decane and dodecane were found to be more pronounced as compared to the heptane system with the lapse of time.

Adsorption behaviour at the air–aqueous interface

The surface excess concentration, (Γ_{\max}) is an important parameter for calculating the degree of adsorption of surfactant molecules at the air interface⁴⁸ as obtained by using Gibb's isotherm eqn (4):

$$\Gamma_{\max} = -1/RT(\partial\gamma/(\partial \ln c))_T \quad (4)$$

where Γ_{\max} corresponds to the surface excess concentration of the surfactant solution calculated in mol m^{-2} , R represents the gas constant ($R = 8.314 \text{ J mol}^{-1} \text{ K}^{-1}$), T represents the temperature measured in K, γ represents the surface tension measured in mN m^{-1} , c is the surfactant concentration in the aqueous phase and $(d\gamma/d \ln c)$ corresponds to the surface tension isotherm slope up to the CMC (the concentration at which the surface tension isotherm curve shows a sharp break). To determine the surface activity of a surfactant solution, it is essential to consider the packing density of the surfactant solution at the air interface. The orientation and degree of packing of the adsorbed surfactant molecules can be calculated using the area per molecule at the interface. From the surface excess concentration, the minimum area (A_{\min}) occupied per surfactant molecule (in \AA^2 per molecule) at the air interface can be obtained using eqn (5):



Table 4 Adsorption parameters calculated for the soap-nut surfactant at different temperatures

Temp (K)	CMC (M)	γ_{CMC} (mN m ⁻¹)	Γ_{max} ($\mu\text{mol m}^{-2}$) $\times 10^{-3}$	A_{min} (\AA^2 per molecule)	pC_{20}	Π_{CMC} (mN m ⁻¹)
298	9.3×10^{-3}	37.1	5.67	29.24	2.69	34.8
313	9.8×10^{-3}	33.8	5.79	28.64	2.70	35.8
323	10.3×10^{-3}	32.4	5.86	28.31	2.74	35.6
333	10.8×10^{-3}	29.6	5.93	27.65	2.75	36.3
348	11.3×10^{-3}	28.6	5.95	27.86	2.77	36.0

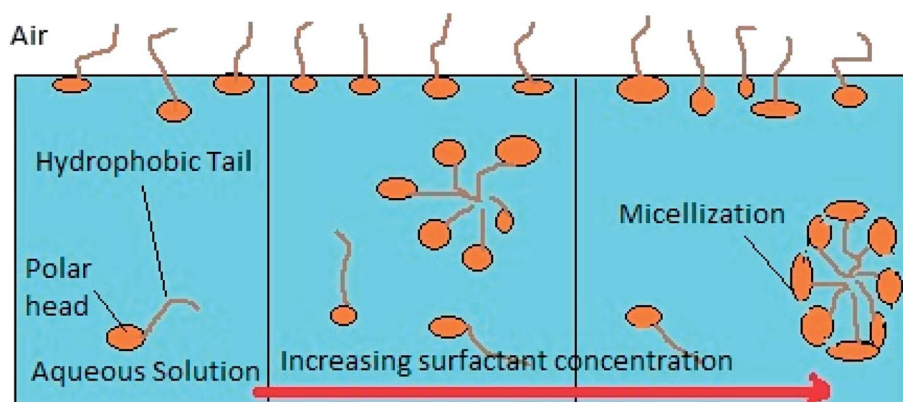


Fig. 17 Micellization process with increasing surfactant concentration.

$$A_{\text{min}} = \frac{10^{20}}{N_A \Gamma_{\text{max}}} \quad (5)$$

where N_A is Avogadro's number and Γ_{max} represents the surface excess concentration measured in mol m^{-2} . The values of Γ_{max} and A_{min} calculated from Gibbs adsorption isotherm predict the molecular arrangement of surfactant molecules at the air interface.⁴⁹ The greater the adsorption, the smaller is the effective area (A_{min}) of surfactant molecules at the surface.

The adsorption efficiency, pC_{20} , and surface tension reduction effectiveness, Π_{CMC} are supplementary parameters that show the surface activity of the surfactant molecules in solution.⁵⁰ The adsorption efficiency can be calculated using eqn (6):

$$pC_{20} = -\log C_{20} \quad (6)$$

where pC_{20} is the surfactant concentration which lowers the pure solvent surface tension by 20 mN m^{-2} and is the minimum concentration required for saturation of the surface adsorption.

The other parameter Π_{CMC} indicates the minimum value of the surface tension caused by dissolution of the surfactant molecules and hence represents the surfactant efficiency to

lower the surface tension of the solvent, as calculated using eqn (7):

$$\Pi_{\text{CMC}} = \gamma_o - \gamma_{\text{CMC}} \quad (7)$$

where γ_{CMC} and γ_o are the surface tensions of the solution at the CMC and of pure solvent, respectively. A greater value of Π_{CMC} or smaller value of γ_{CMC} predicts a better ability to lower the surface tension of a solution. This clearly shows that with an increase in temperature, the ability to lower the surface tension of aqueous solutions is significantly enhanced. The different adsorption parameters are depicted in Table 4.

Thermodynamics of micellization and adsorption

Micellization is the process of an association–dissociation phenomenon in which both micelles and surfactant molecules simultaneously exist in a surfactant solution. Fig. 17 shows a schematic explaining the micellization process with increasing surfactant concentration. A coulombic force of repulsion exists between the polar head group acting against the attractive force between the hydrophobic hydrocarbon tail

Table 5 Micellization parameters for the soap-nut surfactant at different temperatures

Temperature (K)	ΔG_{mic} (kJ mol ⁻¹)	ΔH_{mic} (kJ mol ⁻¹)	ΔS_{mic} (kJ mol ⁻¹ K)	$T\Delta S_{\text{mic}}$ (kJ mol ⁻¹)
298	−21.53	−2.96	0.062	18.47
313	−22.47	−3.26	0.061	19.18
323	−23.06	−3.48	0.060	19.38
333	−23.67	−3.70	0.059	19.64
348	−24.57	−4.04	0.058	20.18



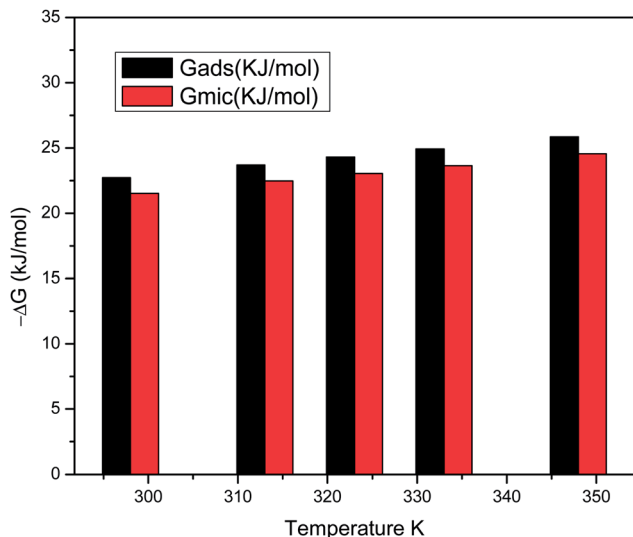


Fig. 18 Comparison of ΔG_{mic} and ΔG_{ads} values for the surfactant solutions at different temperatures.

chains of the surfactant in order to form agglomerates, which separate them continuously.⁴⁹ The Gibbs free energy change of micellization (ΔG_{mic}) can be calculated by using eqn (8):

$$\Delta G_{mic} = RT \ln X_{CMC} \quad (8)$$

where R is the universal gas constant ($8.314 \text{ J K}^{-1} \text{ mol}^{-1}$), T is the temperature (K) and X_{CMC} is a ratio of the molar concentration of the surfactant in the liquid phase at the CMC and that of the solvent in the liquid phase. The values of ΔG_{mic} were found to be negative for the soap-nut oil-based surfactant system at different temperatures, showing that the micellization process was feasible.

The enthalpy of micellization for the soap-nut oil derived surfactant system was determined by using the values of ΔG_{mic} in the Gibbs-Helmholtz relation, as shown in eqn (9):

$$\Delta H_{mic} = -RT^2 \left(\frac{\partial \ln(X_{CMC})}{\partial T} \right) \quad (9)$$

The value of the ΔH_{mic} is negative, which proves that micellization was favourable for its application. The higher the negative value of ΔH_{mic} , the greater is the tendency of micellization.

The entropy change can be calculated easily with the already calculated values of Gibbs free energy change and the enthalpy change. Taking entropy as a function of the temperature and volume of surfactant solution, the $T\Delta S_{mic}$ equation can be expressed according to the second law of thermodynamics in eqn (10):

$$T\Delta S_{mic} = \Delta H_{mic} - \Delta G_{mic} \quad (10)$$

The values of ΔS_{mic} were found to be positive. As we know, the entropy of a system is the degree of randomness, here it was observed that the entropy increased with the increase in temperature as the randomness of the system increased by the destruction of micellar structures. This result further validates the more negative values of ΔG_{mic} with the increase in temperature. Table 5 shows different parameters calculated during the micellization study.

A complete analysis of the adsorption parameters at the air interface was performed from the surface tension data for air-aqueous systems at different temperatures. The amount of surfactant solution adsorbed at the air interface per unit area was calculated using Gibbs adsorption equation. With the addition of the surfactant in the aqueous phase, the surfactant molecules initially adsorb at the interface between air and aqueous solution. The Gibbs free energy of adsorption (ΔG_{ads}) in kJ mol^{-1} was determined from the calculated value of ΔG_{mic} (kJ mol^{-1}) in eqn (11):

$$\Delta G_{ads} = [\Delta G_{ads} - 6.023\pi_{CMC}A_{min}] \times 10^{-3} \quad (11)$$

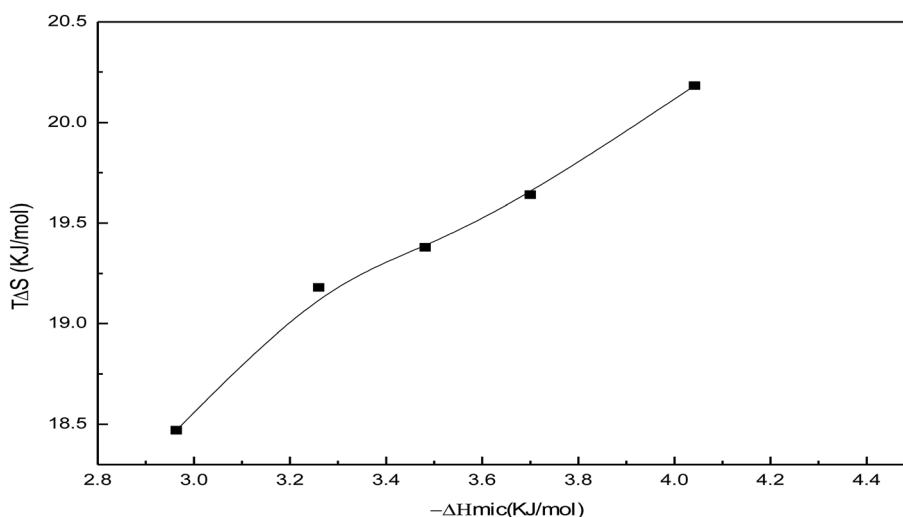


Fig. 19 Enthalpy-entropy compensation showing $-\Delta H_{mic}$ versus $T\Delta S_{mic}$ plots for the soap-nut oil surfactant.



here, the ΔG_{ads} values are more negative than ΔG_{mic} for the surfactants at all temperatures, confirming that the adsorption process is generally favoured over micellization, as depicted in (Fig. 18).

Fig. 19 shows the variation of $T\Delta S_{\text{mic}}$ with $-\Delta H_{\text{mic}}$ for the soap-nut oil surfactant solutions. Initially, ΔH_{mic} shows a linear relationship with $T\Delta S_{\text{mic}}$. After a threshold value of ΔH_{mic} is reached, the slope of the curve slightly decreases due to the reduction in micellization. Studies involving enthalpy–entropy compensations show that $(-\Delta H_{\text{mic}}) < (T\Delta S_{\text{mic}})$. Therefore, it is evident that the micellization behaviour of the surfactant is entropy driven, not enthalpy driven.

Conclusion

The effectiveness of an anionic surfactant synthesized from a natural vegetable resource, soap-nut pericarp was evaluated with an objective to find an alternative to the commercial surfactants available in the market. The surfactant synthesized was suitably characterized by FTIR, GC-MS, EDX and FE-SEM techniques for the determination of the functional groups, various composition of esters, elemental composition and molecular surface morphology, respectively. Thermogravimetric analysis was performed and showed good thermal stability of the synthesized surfactant, suggesting it could be used for industrial applications at varying temperatures. The Krafft temperature of the synthesized surfactant solution was observed at below 277 K. The CMC of the soap-nut surfactant was calculated using the conductivity method and was found to be 9.3 mM at 298 K. Low interfacial tension values at the air–water and oil–water interfaces predicted its efficacy to be employed as a surface active agent in different industries. DLS studies revealed that a fraction of the micellar structures at the CMC aggregate to form clusters as the additional surfactant is added to an aqueous solution. The effect of temperature on the surface tension values was also studied and it was found that the surface tension values decreased with the increase in temperature. Further, it was noticed that the CMC of the surfactant increased with increases in the temperature because of distortion of the structured water molecules adjoining to the hydrophobic group of the surfactant. The aggregation process of surfactant molecules was entropy driven, not enthalpy driven. Thermodynamic studies revealed that the adsorption tendency was more favoured than micellization in air–aqueous systems at a varied temperature range. This may be attributed to the greater tendency of surfactant molecules to migrate to the interface rather than aggregating to micellar structures. The adsorption and micellization behaviours were observed to critically depend on temperature. The foam stability of the surfactant solution is an important property as it facilitates the adsorption of surfactant molecules at the air–water interface. In the emulsion stability at the CMC with oil, a low value of surface tension at the air–water interface and oil–water interfaces provided ground to suggest its potential application in various fields of application as an eco-friendly and biodegradable surfactant.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

The authors gratefully acknowledge the financial assistance provided by Oil India Limited [Contract No. 6206917], Duliajan, Assam, India to the Department of Petroleum Engineering, Indian Institute of Technology (ISM), Dhanbad, India.

References

- 1 D. Mańko, A. Zdziennicka and B. Jańczuk, *Appl. Surf. Sci.*, 2017, **392**, 17–125.
- 2 A. N. Bondarchuk, L. M. Peter, G. P. Kissling, E. Madrid, J. A. Aguilar-Martínez, Z. Rymansaib, P. Irvani, M. Gromboni, L. H. Mascaro, A. Walsh and F. Marken, *Appl. Catal., B*, 2017, **211**, 289–295.
- 3 R. Zhang, J. Huo, Z. Peng, Q. Feng, J. Zhang and J. Wang, *Colloids Surf., A*, 2017, **520**, 855–863.
- 4 L. Torres, A. Moctezuma, J. R. Avendaño, A. Muñoz and J. Gracida, *J. Pet. Sci. Eng.*, 2011, **76**(1), 6–11.
- 5 N. Pal, K. Babu and A. Mandal, *J. Pet. Sci. Eng.*, 2016, **146**, 591–600.
- 6 I. Zhang, B. Cai, W. Xu, H. Gang, J. Liu, S. Yang and B. Mu, *Colloids Surf., A*, 2015, **483**, 87–95.
- 7 A. M. Rabiou, S. Elias and O. Oyekola, *Energy Procedia*, 2016, **100**, 88–192.
- 8 D. Roy, R. R. Kommalapati, S. S. Mandava, K. T. Valsaraj and W. D. Constant, *Environ. Sci. Technol.*, 1997, **31**, 670–675.
- 9 N. Singh, A. Kaur and K. Yadav, *Nat. Sci.*, 2010, **8**, 41–47.
- 10 K. Holmberg, *Curr. Opin. Colloid Interface Sci.*, 2001, **6**, 148–159.
- 11 S. Salati, G. Papa and F. Adani, *Biotechnol. Adv.*, 2011, **29**, 913–922.
- 12 K. Babu, N. Pal, V. K. Saxena and A. Mandal, *Korean J. Chem. Eng.*, 2016, **33**, 711–719.
- 13 N. A. Negm, A. S. El-Tabl, I. A. Aiad, K. Zakareya and A. H. Moustafa, *J. Surfactants Deterg.*, 2013, **16**(6), 857–863.
- 14 S. Kumar, A. Kumar and A. Mandal, *AIChE J.*, 2017, **63**, 2731–2741.
- 15 S. A. Ostroumov, *Hydrobiologia*, 2003, **500**, 341–344.
- 16 S. Song, L. Zhu and W. Zhou, *Environ. Pollut.*, 2008, **156**, 1368–1370.
- 17 S. S. Vieira, Z. M. Magriotis, N. A. V. Santos, A. A. Saczk, C. E. Hori and P. A. Arroyo, *Bioresour. Technol.*, 2013, **133**, 248–255.
- 18 O. Bartsch and U. Schaumssysteme, *Fortschrittsber. Kolloide Polym.*, 1924, **20**, 1–49.
- 19 K. G. Marinova, E. S. Basheva, B. Nenova, M. Temelska, A. Y. Mirarefi, B. Campbell and I. B. Ivanov, *Food Hydrocolloids*, 2009, **23**, 1864–1876.
- 20 S. C. Kothekar, A. M. Ware, J. T. Waghmare and S. A. Momin, *J. Dispersion Sci. Technol.*, 2007, **28**, 477–484.
- 21 M. A. H. Shah, K. Dutta and D. C. Deka, *Adv. Appl. Sci. Res.*, 2015, **5**(4), 43–50.



- 22 C. Sun, L. Jia, B. Xi, L. Wang and X. Wang, *Ind. Crops Prod.*, 2017, **102**, 97–104.
- 23 A. Mariod, H. Hassan and K. M. Saeed, *Eurasian Journal of Forest Science*, 2018, **6**(1), 52–67.
- 24 K. A. Elaries, I. M. Tan, M. Awang and I. Saaïd, *Pet. Sci. Technol.*, 2010, **28**, 1799–1806.
- 25 J. A. Torres, L. A. Diaz and J. F. Sanchez, *Ing. Invest.*, 2009, **29**, 48–53.
- 26 B. Meschi, A. J. Trinidad, G. Rivers, S. Sy, P. Russo, A. Yu, N. Y. Zhou and B. Zhao, *Carbon*, 2015, **91**, 188–199.
- 27 K. Babu, N. K. Maurya, A. Mandal and V. K. Saxena, *Braz. J. Chem. Eng.*, 2015, **32**, 795–803.
- 28 R. J. Williams, J. N. Phillips and K. J. Mysels, *Trans. Faraday Soc.*, 1954, **51**, 728–737.
- 29 N. Pal, N. Saxena and A. Mandal, *Colloid Polym. Sci.*, 2017, **295**, 2261–2277.
- 30 S. Kumar and A. Mandal, *Tenside, Surfactants, Deterg.*, 2017, **54**(5), 427–436.
- 31 B. Gao and M. M. Sharma, *J. Colloid Interface Sci.*, 2013, **407**, 375–381.
- 32 E. Mohajeri and G. D. Noudeh, *J. Chem.*, 2012, **9**(4), 2268–2274.
- 33 P. Pillai, N. Pal and A. Mandal, *J. Surfactants Deterg.*, 2017, **20**, 1321–1335.
- 34 H. Zhao, S. Xia and P. Ma, *J. Chem. Technol. Biotechnol.*, 2005, **80**(10), 1089–1096.
- 35 J. Lu, P. J. Liyanage, S. Solairaj, S. Adkins, G. P. Arachchilage, D. H. Kim, C. Britton, U. Weerasooriya and G. A. Pope, *J. Pet. Sci. Eng.*, 2014, **120**, 94–101.
- 36 A. Pradhan and A. Bhattacharyya, *J. Surf. Sci. Technol.*, 2014, **30**, 59–76.
- 37 A. Verma, G. Chauhan and K. Ojha, *Asia-Pac. J. Chem. Eng.*, 2017, **12**(6), 872–883.
- 38 H. Azira, A. Tazerouti and J. P. Canselie, *J. Surfactants Deterg.*, 2008, **11**(4), 279–286.
- 39 S. Kumar and A. Mandal, *Appl. Surf. Sci.*, 2017, **420**, 9–20.
- 40 S. J. Rehfeld, *J. Phys. Chem.*, 1962, **66**(10), 1966–1968.
- 41 H. Katepalli and A. Bose, *Langmuir*, 2014, **30**(43), 12736–12742.
- 42 Z. G. Cui, L. L. Yang, Y. Z. Cui and B. P. Binks, *Langmuir*, 2010, **26**(7), 4717–4724.
- 43 D. Kurukji, R. Pichot, F. Spyropoulos and I. T. Norton, *J. Colloid Interface Sci.*, 2013, **409**, 88–97.
- 44 N. Pal, N. Saxena and A. Mandal, *J. Chem. Eng. Data*, 2017, **62**, 1278–1291.
- 45 J. E. Carless and G. W. Hallworth, *J. Colloid Interface Sci.*, 1968, **26**(1), 75–88.
- 46 J. Sjöblom, P. Stenius, S. Simon and B. A. Grimes, *Encyclopedia of Colloid and Interface Science*, 2013, pp. 415–454.
- 47 W. Wu, H. Fang, F. Yang, S. Chen, X. Zhu, Q. Yuan and W. Gan, *J. Phys. Chem. C*, 2016, **120**(12), 6515–6523.
- 48 Z. Zhao, Z. Li, W. Qiao and L. Cheng, *Colloids Surf., A*, 2005, **259**(1–3), 71–80.
- 49 N. Pal, N. Saxena and A. Mandal, *Colloids Surf., A*, 2017, **533**, 20–32.
- 50 M. J. Rosen, *Surfactants and Interfacial Phenomena*, John Wiley and Sons, Inc., New York, 3rd edn, 2004.

