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Effects of arbuscular mycorrhizal fungi on *Leymus chinensis* seedlings under salt–alkali stress and nitrogen deposition conditions: from osmotic adjustment and ion balance

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Nitrogen deposition and soil salinization–alkalization have become major environmental problems throughout the world. *Leymus chinensis* is the dominant, and considered the most valuable, species for grassland restoration in the Northeast of China. However, little information exists concerning the role of arbuscular mycorrhizal fungi (AMF) in the adaptation of seedlings to the interactive effects of nitrogen and salt–alkali stress, especially from the perspective of osmotic adjustment and ion balance. Experiments were conducted in a greenhouse and *Leymus chinensis* seedlings were cultivated with NaCl/NaHCO₃ under two nitrogen treatments (different concentrations of NH₄⁺/NO₃[−]). Root colonization, seedling growth, ion content, and solute accumulation were measured. The results showed that the colonization rate and the dry weights of the seedlings were both decreased with the increasing salt–alkali concentration, and were much lower under alkali stress. Both of the nitrogen treatments decreased the colonization rate and dry weights compared with those of the AM seedlings, especially under the N2 (more NH₄⁺–N content) treatment. The Na⁺ content increased but the K⁺ content decreased under salt–alkali stress, and more markedly under alkali stress. AMF colonization decreased the Na⁺ content and increased the K⁺ content to some extent. In addition, the nitrogen treatments had a negative effect on the two ions in the AM seedlings. Under salt stress, the seedlings accumulated abundant Cl[−] to maintain osmotic and ionic balance, but alkali stress inhibited the absorption of anions and the seedlings accumulated organic acids in order to resist the imbalance of both osmosis and ions, whether under the AM or nitrogen treatments. In addition, proline accumulation is thought to be a typical adaptive feature in both AM and non-AM plants under nitrogen and salt–alkali stress. Our results suggest that the salt–alkali tolerance of *Leymus chinensis* seedlings is enhanced by association with arbuscular mycorrhizal fungi, and the seedlings can adapt to the nitrogen and salt–alkali conditions by adjusting their osmotic adjustment and ion balance. Excessive nitrogen partly decreased the salt–alkali tolerance of the *Leymus chinensis* seedlings.

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Introduction

Natural processes and human activities, such as fossil fuel combustion and fertilizer use, have strongly enhanced the rate of atmospheric nitrogen deposition and have received much attention.¹ According to the fifth IPCC report, nitrogen input into the ecosystem due to human causes each year has

increased tenfold over the past 150 years and it has been predicted to be at least twice the current level by the 2050s.² Excess input of nitrogen can have adverse ecological effects, such as changes in soil function and eutrophication.^{3,4} Nitrogen is also a limiting resource in the grassland ecosystem of northern China, which exerts a profound effect on plant and soil micro-organisms.⁵ In addition, not only is the total N content increasing, but the ratio of NH₄⁺–N to NO₃[−]–N is also changing as a result of N deposition. However, to the best of our knowledge, the physiological effect of NH₄⁺/NO₃[−] has been always overlooked.

Salinity is considered one of the most important environmental factors limiting plant growth and yield throughout the world.⁶ The stress caused by soil salinity generally involves osmotic stress and ion-induced injury.⁷ These effects can

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inhibit nutrient absorption and prevent them from being transported within plants. In order to resist environmental stress, most plants have developed a variety of adaptive mechanisms, such as the synthesis of compatible solutes, the accumulation or exclusion of selected ions, and the control of ion uptake by the roots.⁸ In addition, soil alkalization is also a marked feature in Northeastern China. The existence of alkali stress has been demonstrated to be more severe than salt stress due to the high pH, which may inhibit ion uptake and disrupt ionic balance.^{9–11} Thus, they are actually two totally different types of stress. Yet, in spite of this, people pay little attention to the effect of alkali stress compared with that of salt stress.

Arbuscular mycorrhizal fungi (AMF), a kind of ancient soil microorganism, widely occurs in saline soil, and can form mutualistic relationships with over 80% of terrestrial plants.¹² It is known that arbuscular mycorrhiza can enhance plant growth and development, and can also alleviate the adverse effects of salt stress.^{13,14} The potential mechanisms for mycorrhizal plants responding to salt stress may include improving the nutrient uptake ability of the plant (especially the uptake of phosphorus), maintaining higher antioxidant enzymatic activities, elevating the K^+ content, and changing the root functions.^{15–17} However, the ecological role of arbuscular mycorrhizal fungi on the osmotic adjustment and ion transport of plants under alkali stress (high pH), especially the interactive effects of alkali stress and nitrogen deposition (different concentrations of NH_4^+ / NO_3^-) conditions, has still rarely been reported.

Leymus chinensis, also called alkali grass, is a perennial rhizomatous species of the family Poaceae. This grass not only has a high tolerance to salt–alkali soil, but also contains quite a lot of nutrients, such as carbohydrates, minerals, and proteins.¹⁸ Thus, some reports have indicated that *Leymus chinensis* is considered to be one of the most valuable species for grassland restoration in the Northeast of China.^{19,20} Here, we evaluated the contribution of arbuscular mycorrhizal fungi to the growth, ion content, and solute accumulation of *Leymus chinensis* seedlings under salt–alkali stress and nitrogen deposition. We aimed firstly to clarify the role of AMF during the adaptation of the *Leymus chinensis* seedlings to salt–alkali stress, and then we further explored the response of the plant–AMF to salt/alkali stress and its interactive effects with nitrogen deposition (different concentrations of NH_4^+ / NO_3^-) from the perspective of osmotic adjustment and ion balance.

Materials and methods

Plant culture, inoculation, and stress treatments

The AMF (*Glomus mosseae*, BGC HEB02) used for inoculation was provided by the Institute of Plant Nutrition and Resources, Beijing Academy of Agriculture and Forestry Sciences, China. A pot experiment was conducted from April to July in 2015 at Northeast Forestry University, Harbin City, Heilongjiang Province, China. A randomized block design with three factors was used in this experiment: AMF treatment (non-AMF as the control), nitrogen treatments [0 mM; N1 (10 mM NH_4NO_3) and N2 (10 mM NH_4Cl : NH_4NO_3 = 3 : 1)], and salt–alkali stress (0, 100, and 200 mM $NaCl/NaHCO_3$). Each nitrogen level had +AMF

or –AMF treatment, and each +AMF or –AMF treatment had salt or alkali stress treatment. The pH values of the salt stress were 6.28 and 6.30, and were 10.18 and 10.39 for the alkali stress. Each treatment was conducted in 4 replicates and eight seedlings were included in each pot.

Leymus chinensis seeds were collected from the Grassland Ecosystem Field Station, Institute of Grassland Science in Northeast Normal University, Jilin Province, China (123°44'E, 44°44'N) in July 2013. The seeds were firstly surface sterilized with 10% sodium hypochlorite for 10 min, and then were sown in the plastic pots. Each pot was 15 cm in upper diameter, 12 cm in lower diameter, 15 cm in depth, and filled with 2 kg of an autoclaved soil mixture (soil : sand 3 : 1, v/v). Before this step, the substrate was autoclaved at 121 °C and 240 kPa pressure for 2 h to ensure that all possible mycorrhizal propagules and other microorganisms had been destroyed. Each pot was inoculated with either 20 g of the inoculant for the inoculation treatment (+AMF) or 20 g of sterilized inoculant for the non-inoculation treatment (–AMF). We also added 30 mL of filtered inoculant (through a 0.25 μm filter membrane), free from mycorrhizal propagules, to the –AMF treatment to maintain the same microorganism biota. The inoculants were added 2 cm below the seeds. The stress treatments were carried out by adding the corresponding solutions (as described above) 120 days after sowing, and the control treatment was irrigated with 300 mL of distilled water. The seedlings were harvested 7 days after the treatments.

Growth index and mycorrhizal colonization measurements

The harvested seedlings were firstly washed with distilled water and then the roots and shoots were separated. The shoots were vacuum-dried at 65 °C to a constant weight and the dry weights were then determined.

For the evaluation of AMF colonization, a fraction of the roots was carefully washed, cut into 1 cm long segments, cleaned with 10% KOH solution, and stained with 0.05% trypan blue in lactophenol.²¹ The AMF colonization was then quantified according to the formula described by Wu *et al.*:²²

$$\text{AMF colonization (\%)} = (\text{root length infected / root length observed}) \times 100$$

Determination of inorganic ions and organic solutes

Dry samples of *Leymus chinensis* seedlings (50 mg) were treated with 10 mL deionized water at 100 °C for 1 h and the extract was taken to determine any free inorganic ions and organic acids. An atomic absorption spectrophotometer (TAS-990, Purkinje General, Beijing) was used to determine the Na^+ , K^+ , free Mg^{2+} , and free Ca^{2+} content. The NO_3^- , Cl^- , $H_2PO_4^-$, and SO_4^{2-} amounts were determined by ion chromatography (DX-300 ion chromatographic system; AS4A-SC ion-exchange column). For the analysis of the organic acids, measurements were also undertaken using ion chromatography (DX-300 ion chromatographic system; ICE-AS6 ion-exclusion column). The proline and total soluble sugar amounts were measured using



ninhydrin and anthrone, respectively, according to Bao *et al.*²³ and Zhu *et al.*²⁴ Betaine was extracted with 80% methanol for 20 min at 70 °C and determined using the method established by Greive and Grattan.²⁵

Statistical analysis

The data were analyzed using the statistical software SPSS 13.0 (SPSS Inc, Chicago, IL, USA). The data were subjected to an analysis of variance (one-, two- or three-way ANOVA) with the inoculation treatment, salt/alkali stress, nitrogen treatment, and their interactions as the sources of variation. *Post hoc* comparisons with Duncan's multiple range tests were used to identify the differences between groups with $P < 0.05$ as the significance cut-off.

Results and discussion

Root colonization and seedling growth

The root mycorrhizal colonization was decreased significantly by the nitrogen treatments under salt-alkaline stress ($P < 0.01$). Without the nitrogen treatment and stress conditions, the root mycorrhizal colonization can reach a maximum value of 97%. Compared to the N1 treatment, the colonization rate under the N2 treatment was much lower. Moreover, both salt and alkali stresses affected the mycorrhizal colonization, and the inhibition action of alkali stress was much stronger. Under the highest salinity stress concentration (200 mM), the colonization rate reached nearly 90%, but only reached 63% under alkalinity stress at the same stress intensity (Tables 1 and 2).

Table 1 Mycorrhizal colonization rate and seedling dry weight of *Leymus chinensis* under nitrogen deposition and salinity conditions^a

Treatment	Mycorrhizal colonization rate (%)	Dry weight (g per plant)
CK – AM	0.00 ± 0.00a	0.34 ± 0.03a
CK + AM	97.33 ± 1.33d	0.49 ± 0.03b
CK + AM + N1	78.33 ± 1.20c	0.36 ± 0.02a
CK + AM + N2	71.67 ± 1.76b	0.33 ± 0.02a
S1 – AM	0.00 ± 0.00a	0.27 ± 0.01a
S1 + AM	92.00 ± 1.00d	0.37 ± 0.04b
S1 + AM + N1	69.33 ± 0.67c	0.31 ± 0.01a
S1 + AM + N2	64.33 ± 0.67b	0.28 ± 0.01a
S2 – AM	0.00 ± 0.00a	0.24 ± 0.02a
S2 + AM	88.67 ± 0.67d	0.31 ± 0.04b
S2 + AM + N1	63.67 ± 1.33c	0.29 ± 0.02b
S2 + AM + N2	59.33 ± 2.96b	0.28 ± 0.02 ab
AM	***	***
S	***	*
AM*S	***	N.S.
N1	***	**
N1*AM*S	***	N.S.
N2	***	***
N2*AM*S	N.S.	N.S.

^a The different letters indicate significant differences between the treatments (Tukey's test $P < 0.05$). *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$, N.S. = not significant.

Table 2 Mycorrhizal colonization rate and seedling dry weight of *Leymus chinensis* under nitrogen deposition and alkalinity conditions^a

Treatment	Mycorrhizal colonization rate (%)	Dry weight (g per plant)
CK – AM	0.00 ± 0.00a	0.34 ± 0.03a
CK + AM	97.33 ± 1.33d	0.49 ± 0.03b
CK + AM + N1	78.33 ± 1.20c	0.36 ± 0.02a
CK + AM + N2	71.67 ± 1.76b	0.33 ± 0.02a
A1 – AM	0.00 ± 0.00a	0.25 ± 0.01a
A1 + AM	79.33 ± 1.76d	0.25 ± 0.02a
A1 + AM + N1	58.67 ± 2.03c	0.26 ± 0.01a
A1 + AM + N2	50.67 ± 0.67b	0.27 ± 0.01a
A2 – AM	0.00 ± 0.00a	0.20 ± 0.02a
A2 + AM	63.33 ± 1.67d	0.22 ± 0.03a
A2 + AM + N1	51.33 ± 0.67c	0.20 ± 0.01a
A2 + AM + N2	44.00 ± 1.00b	0.20 ± 0.01a
AM	***	**
A	***	***
AM*A	***	*
N1	*	***
N1*AM*A	*	*
N2	***	**
N2*AM*S	**	**

^a The different letters indicate significant differences between the treatments (Tukey's test $P < 0.05$). *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$, N.S. = not significant.

The dry weight of the *L. chinensis* seedlings was significantly inhibited with the increasing the salt-alkali stress concentration, and alkali stress also further inhibited the dry weight ($P < 0.05$, Tables 1 and 2). The plants inoculated with the AMF had significantly higher seedling weights under both stress and non-stress conditions. The nitrogen treatments (N1 and N2) decreased the dry weight compared with the inoculated seedlings. The N1 and N2 treatments made no difference to the seedling dry weight under salt stress ($P > 0.05$). In addition, when alkali stress reached 100 mM, no significant change was observed among the four treatments (Table 2).

It is known that salt stress can affect plants, AMF, and their interactions.^{26,27} In the present study, the mycorrhizal colonization decreased with the increasing salt stress intensity, and decreased much more markedly under alkali stress (Table 2), indicating that salt-alkali stress inhibited the AMF growth. Some other reports have also supported this viewpoint.²² The main reason is that salt stress inhibits spore germination or hyphal growth.²⁸ However, the much greater inhibition of alkali stress is perhaps due to high pH stress. Under such conditions, the plant root was subjected to much more damage, and the root also released many more chemical components into the soil, and then affected the mycorrhizal colonization. The specific reason requires further study. Our research also showed that nitrogen treatments could decrease the mycorrhizal colonization. One of the main reasons might be that the much higher concentration of nitrogen changed the function of the AMF. In addition, the inhibitory effect of NH_4^+ -N was much stronger, and it might be that NO_3^- -N is more easily absorbed by *L. chinensis*, and the ammonium had a toxic effect to some extent, which also needs further research.



Previous studies have demonstrated that plant growth can be inhibited under both salt and alkali stresses because of ion-excess effects.^{9,29} However, the plant must spend much more energy to cope with the high pH under alkali stress. Thus, the dry weight of *L. chinensis* under alkali stress was lower than that under salt stress. Moreover, the inoculated seedlings had higher dry weights under salt stress, showing that AMF could increase the salt tolerance of *L. chinensis*. Similar results have also been reported by others.^{30,31} However, alkali stress did not increase the seedling dry weights, which was also because of the high pH and much more damaging effects, and needs further research.

Inorganic ion (cation and anion) content

With the increasing intensity of the salt-alkali stress, the Na⁺ content in the shoot increased significantly ($P < 0.05$), and this effect was observed more markedly under alkali stress (Fig. 1A and B). AMF colonization decreased the Na⁺ content under stress conditions, especially under higher concentration stress. When the salt concentration reached 200 mM, the Na⁺ content of the seedlings inoculated with AMF was decreased by 18.1% and 18.8% compared with that of the non-inoculated seedlings under salt and alkali stresses. Both nitrogen treatments increased the Na⁺ content at each salinity/alkalinity level in the AMF colonization treatments ($P < 0.05$), and the N1 and N2 treatments showed no significant differences, except for at 200 mM salt stress. The K⁺ content in the shoot decreased with increasing salt and alkali stresses, and these changes were much greater under alkali stress ($P < 0.05$). Except for at 100 mM salt stress, AMF colonization increased the K⁺ content under salt-alkali stress conditions. The nitrogen treatments decreased the K⁺ content under salt-alkali stresses ($P < 0.05$), and the N1 and N2 treatments also showed no significant differences (Fig. 1C and D). In addition, AMF colonization did not affect the Mg²⁺ and Ca²⁺ content and, furthermore, the N treatments also had almost no influence on the Mg²⁺ and Ca²⁺ content (Fig. 1E–H).

With the increasing intensity of the salt stress, the Cl[−] content in the shoot increased significantly, and AMF colonization decreased the Cl[−] content under salt stress ($P < 0.05$; Fig. 2A). However, no significant change was observed in the Cl[−] content under either alkali stress or AMF colonization with alkali stress ($P > 0.05$; Fig. 2B). In addition, the nitrogen treatments increased the Cl[−] content in the *L. chinensis* seedlings. Under the alkali stress treatment, AMF colonization and nitrogen treatments both had no obvious significant effect on the H₂PO₄[−] and SO₄^{2−} content ($P > 0.05$; Fig. 2D and F). Moreover, the H₂PO₄[−] and SO₄^{2−} content only increased significantly at 100 mM salt stress with the N2 treatment ($P < 0.05$; Fig. 2C and E). The NO₃[−] content in the shoot decreased with increasing salt and alkali stresses, and AMF colonization increased the NO₃[−] content, but it only reached a significant level at CK and 200 mM salt stress treatment ($P < 0.05$; Fig. 2G). In addition, the N treatments have almost no influence on the NO₃[−] content (Fig. 2G and H).

Plants growing in salt-alkali soil generally suffer from two mainly distinct stresses: ionic stress and osmotic stress. Na⁺ is

one of the most dominating toxic ions in salinized and alkalized soil.³² It can disrupt the structure of many macromolecules and their normal physiological metabolism. In addition, ionic imbalance is also a distinguishing feature caused by the influx of superfluous Na⁺.^{33,34} The Na⁺ ions enter the plant cells through a high-affinity K⁺ transporter (HKT) and non-selective cation channels.³⁵ Thus, for most plants living in salt-alkali environments, Na⁺ often greatly accumulates in the vacuoles and also inhibits K⁺ absorption.⁷ In our results, the Na⁺ concentration sharply increased and the K⁺ concentration was simultaneously decreased under both salt and alkali stresses, with both the +AMF and −AMF treatments. However, the change was much greater under alkali stress, indicating that the high pH caused by alkali stress exerts much more damaging effects on the seedlings (higher Na⁺ and lower K⁺), which rely on a transmembrane proton gradient. In addition, this result was in contradiction with observations of other halophytes, such as *Kochia sieversiana* and *Chloris virgate*, that are also widespread in the Songnen salt-alkali grassland in Northeastern China,^{29,36} and showed no competitive inhibition between the two ions in the two species, indicating that different plants may have distinct pathways for the absorption and transfer of K⁺ and Na⁺, and need further research. Our results also showed that Mg²⁺ and Ca²⁺ increased under both salt stress and alkali stress, but their amounts were very low and their roles in ionic balance and osmotic adjustment were tiny.

It is widely recognized that AM symbiosis is a pivotal component in helping plants cope with adverse environments such as salt-alkali stress.³⁷ It is evident from the present study that AM seedlings showed lower Na⁺ and higher K⁺ compared with the −AM plants, especially under the highest concentration of stress (200 mM, Fig. 1). These results show that the role of the mycorrhizal fungi in alleviating salt stress is mainly due to the inhibition of toxic Na⁺ absorption and transportation. The lower levels of Na⁺ in the AM seedlings may also be explained by the dilution effect because of plant growth enhancement.²⁷ In addition, potassium is one of the most important essential elements for plant growth as it plays a key role in plant metabolism. The higher K⁺ concentration and its direct effect of establishing a higher K⁺/Na⁺ in the mycorrhizal plants under salt-alkali stress also relieved the negative impacts by the ionic balance of the cytoplasm or the Na efflux from the plant.¹⁵ Similar results have also been reported for other plants (with or without AM) under salt stress conditions, such as wheat and *Trigonella foenum-graecum*.³⁸ Studies carried out by Giri *et al.*¹⁵ have also indicated that the increased accumulation of K⁺ reduced the translocation of Na⁺ in the shoots of AMF-colonized *acacia* plants grown in saline soil.

Moreover, nitrogen deposition increased the Na⁺ content and also decreased the K⁺ content within the mycorrhizal inoculation treatments of the *L. chinensis* seedlings, especially under the N2 treatment (Fig. 1). The main reason for this finding is that excessive nitrogen in the soil induces a nutrient imbalance and also changes the distribution and transportation of ions. The ammonium may have a toxic effect to some extent, and the specific mechanism needs further research. The above results also clearly show that nitrogen deposition (different



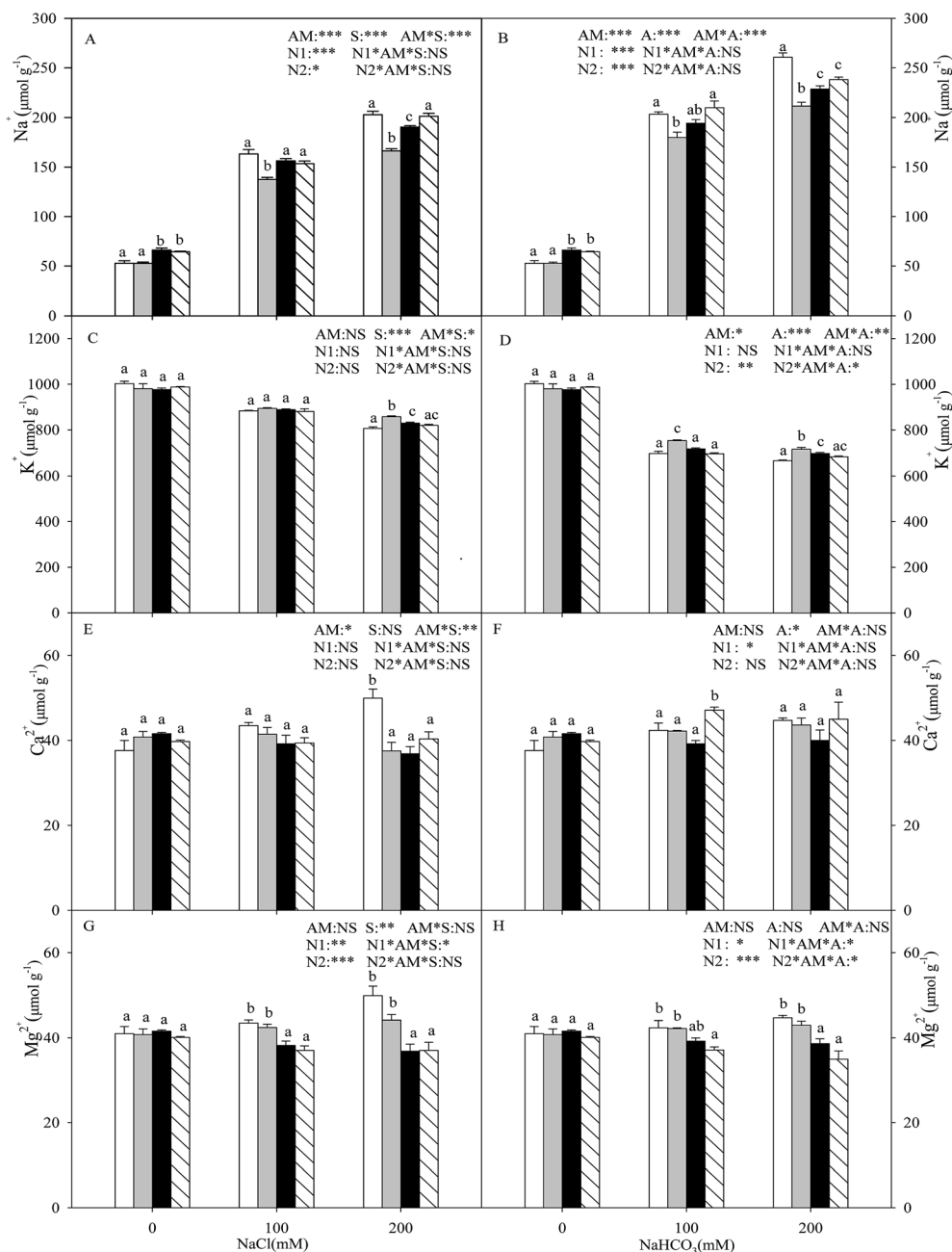


Fig. 1 Na⁺ (A and B), K⁺ (C and D), Ca²⁺ (E and F), and Mg²⁺ (G and H) content in *Leymus chinensis* seedlings [non AM (□), AM (■), AM + N1 (■), AM + N2 (▨)] under nitrogen deposition, salinity (A, C, E, and G), and alkalinity (B, D, F, and H) conditions. The bars represent mean ± S.E. (*n* = 4). The different letters indicate significant differences between the treatments (Tukey's test *P* < 0.05). ****P* < 0.001, ***P* < 0.01, **P* < 0.05, NS = not significant.

concentrations of NH₄⁺/NO₃⁻ decreased the salt-alkali tolerance of the *L. chinensis* seedlings by reducing the suitability of the colonization.

Ionic imbalance within plants has been reported to be mainly caused by the influx of superfluous Na⁺.³³ Most plants always accumulate the main inorganic anions, such as Cl⁻, at this point to cope with adverse environments. In our study, Cl⁻ and SO₄²⁻ were accumulated in the *L. chinensis* seedlings under salt stress. However, all of the anions showed a decreasing trend under alkali stress (Fig. 2A). Our results verified that salt stress

and alkali stress had different physiological effects on the plants, and the high pH might inhibit the absorption of inorganic anions such as Cl⁻, NO₃⁻, and SO₄²⁻. Similar results were also reported for other plants (both crops and grasses), such as the tomato,³⁹ seabuckthorn⁴⁰, and *Lathyrus quinquenervius*.¹¹

In general, superfluous Cl⁻ can also be toxic to plant growth in saline areas.⁴¹ This problem can also be resolved by using arbuscular mycorrhiza to some extent, which can decrease the Cl⁻ uptake.⁴² In our results, AMF colonization decreased the Cl⁻ content under salt stress, which is consistent with the previous



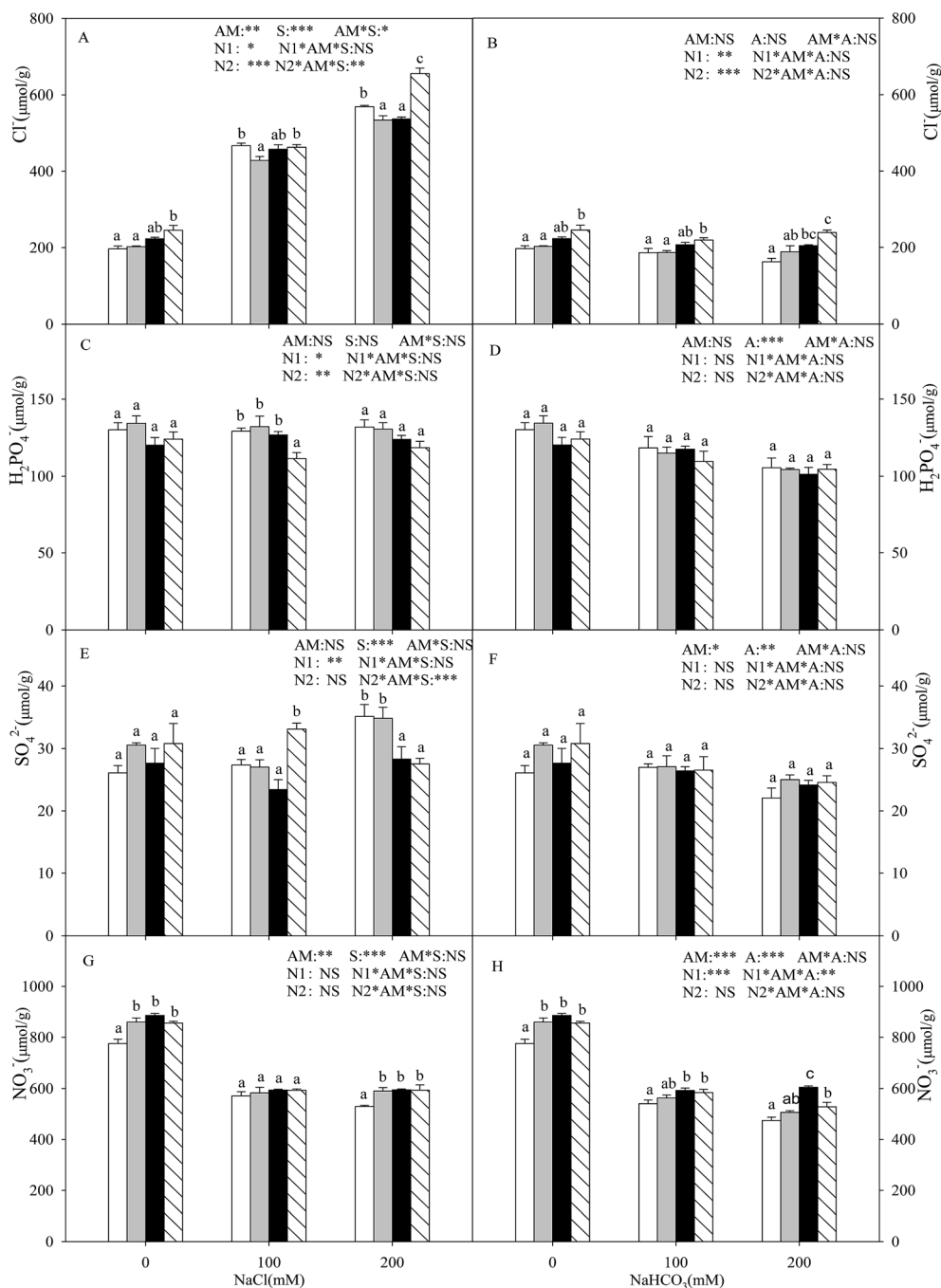


Fig. 2 Cl^- (A and B), H_2PO_4^- (C and D), SO_4^{2-} (E and F), and NO_3^- (G and H) content in *Leymus chinensis* seedlings [non AM (\square), AM (\blacksquare), AM + N1 (\blacksquare), AM + N2 (\square)] under nitrogen deposition, salinity (A, C, E, and G), and alkalinity (B, D, F, and H) conditions. The bars represent mean \pm S.E. ($n = 4$). The different letters indicate the significant differences between the treatments (Tukey's test $P < 0.05$). *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$, NS = not significant.

report on lettuce and onions,⁴³ but there was no significant difference between the –AM and mycorrhizal inoculation treatments in *L. chinensis* seedlings under alkali stress because of the severe influence of the high pH. In addition, the result was in contradiction with observations reported for citrus plants, spring wheat, and winter barley.^{44,45} They found that the Cl^- content increased due to the mycorrhizal colonization, and the main reason is perhaps the carbon drain imposed by the

mycorrhizal hyphae on plants, and then the enhancement of the translocation of Cl^- from the saline soil. The differences in the plant species may be a reason for these differences and for their specific causes, both of which should be further investigated. Moreover, similar changes were observed for Cl^- and Na^+ when the *L. chinensis* seedlings were inoculated with mycorrhizal, further illustrating the negative impact of nitrogen deposition, especially the ammonium nitrogen. Interestingly, *L.*



chinensis seedlings can also maintain constant ionic balance under nitrogen treatment.

Organic solute content

With the increasing intensity of the salt-alkali stress, the proline content in the shoots of the *L. chinensis* seedlings increased significantly, and this effect was observed more markedly under salt stress ($P < 0.05$, Fig. 3A and B). AMF colonization decreased the proline content under stress conditions, especially under the highest concentration stress (200 mM), where the proline content decreased by 65.8%. The application of either nitrogen treatment increased the proline content at each salinity/alkalinity level (Fig. 3A and B). The soluble sugar content showed a similar tendency to the proline (Fig. 3C and D). In addition, AMF colonization only decreased the soluble sugar content to 19.6% at 200 mM alkali stress. The N treatments had no influence on the soluble sugar content of the

inoculated seedlings. The soluble sugar content only increased at alkali stress under the N2 treatment. AM colonization did not affect the MDA content under salt and alkali stresses (Fig. 3E and F). The N treatments increased the MDA content, much more markedly with the N2 treatment.

Malic acid, citric acid, acetate acid, and oxalate acid were detected in the shoots of the *L. chinensis* seedlings under salt and alkali stresses (Fig. 4). The citric acid content increased under alkali stress (Fig. 4B), and AMF colonization and N treatments did not affect the citric acid content under salt-alkali stress. The malic acid content showed a similar tendency to that of citric acid. AMF colonization decreased the malic acid content by 31.5% under 200 mM alkali stress, but did not affect the content at other stress concentrations (Fig. 4D and E). In addition, the N treatments have almost no influence on the malic acid content. AMF colonization also decreased the acetate acid content according to Fig. 4E and F. However, unlike these

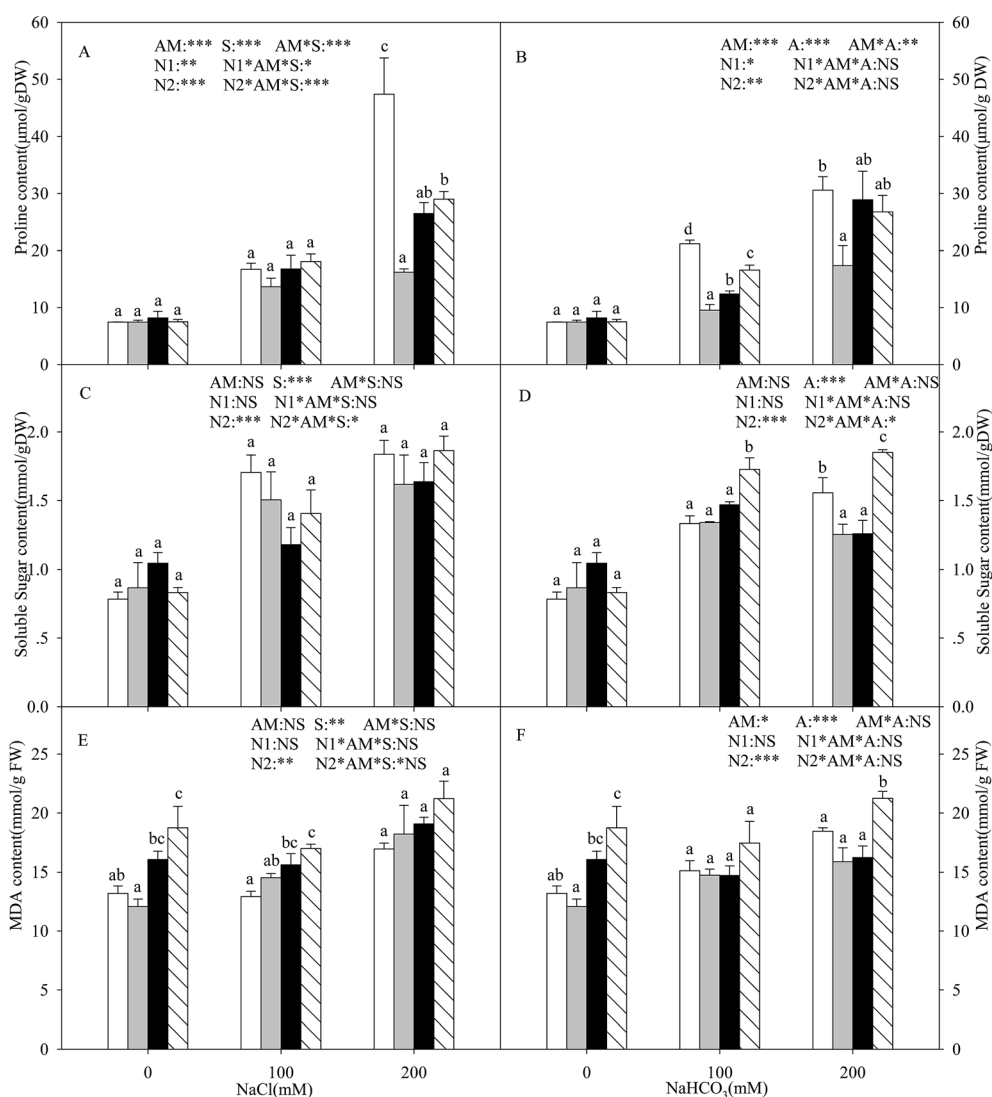


Fig. 3 Proline (A and B), soluble sugar (C and D), and MDA (E and F) content in *Leymus chinensis* seedlings [non AM (□), AM (■), AM + N1 (■), AM + N2 (▨)] under nitrogen deposition, salinity (A, C, E, and G), and alkalinity (B, D, F, and H) conditions. The different letters indicate significant differences between the treatments (Tukey's test $P < 0.05$). *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$, NS = not significant.



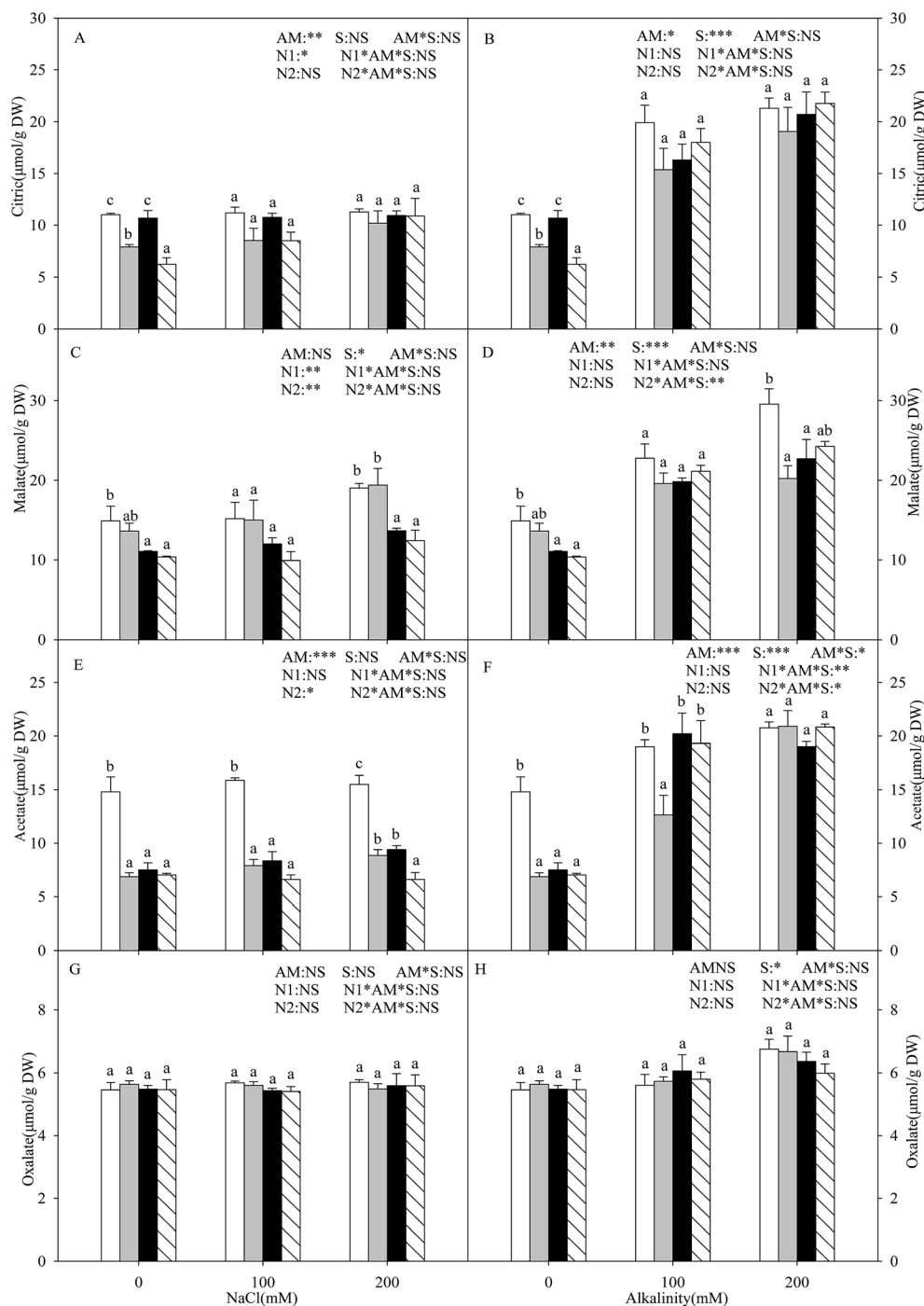


Fig. 4 Citric acid (A and B), malic acid (C and D), acetic acid (E and F), and oxalic acid (G and H) content in *Leymus chinensis* seedlings [non AM (□), AM (■), AM + N1 (■), AM + N2 (▨)] under nitrogen deposition, salinity (A, C, E, and G), and alkalinity (B, D, F, and H) conditions. The different letters indicate significant differences between the treatments (Tukey's test $P < 0.05$). *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$, NS = not significant.

three organic acid compositions, no change was observed in the oxalate acid content under both the AM or N treatments (Fig. 4G and H).

In salt-alkali environments, plants must face the negative impact of the lower water potential of the soil, which makes it difficult for the plants to acquire adequate water from the surrounding soil. Several organic solutes, such as proline,

soluble sugar, and MDA, are accumulated under these conditions. In general, proline accumulation is thought to be an adaptive feature under salt-alkali stress in both AM and non-AM plants.⁴⁶ Our results showed that the accumulation of proline in the *L. chinensis* seedlings increased with increasing salt-alkaline concentration (Fig. 3A and B). Moreover, the proline content was detected to decrease in the seedlings with



AMF inoculation. This differed to most previous reports, which found that AMF colonization could increase the proline content in plants such as soybean and wheat.⁴⁶ However, similar results were found for *Vicia faba* by Rabie and Almadini.⁴⁷ The possible reason is that the proline content can also be used for measuring the degree of plants that were injured, indicating that AMF colonization alleviates salt-alkali stress on the *L. chinensis* seedlings. The nitrogen deposition results also prove this viewpoint.

Organic acids also played a potential role as cell osmolytes in the osmotic adjustment.⁴⁸ In our study, we clearly found that salt stress did not affect the amounts of four organic acids (malic acid, citric acid, acetate acid, and oxalate acid), but alkali stress significantly enhanced them, indicating that organic acid accumulation is a specific response to high pH stress. Under alkali stress, superfluous Na⁺ was accumulated, but the Cl[−] content had no significant change. Thus, we consider that the accumulation of organic acids not only played an important osmotic role but also buffered excess toxic cations and maintained ionic balance. Similar viewpoints have also been reported by others.^{9,11} In addition, we also found that AMF colonization did not affect the organic acid content, except for acetate acid, but decreased most of them under alkali stress. Combined with the Na⁺ and Cl[−] results as described above, we conclude that there exists a correlation between the Na⁺ and organic acid content, and that *L. chinensis* seedlings can maintain ionic balance under both AM and non-AM conditions. In addition, the organic acid content showed an increasing trend in the AM seedlings that underwent the N treatments to some extent, especially the N2 treatment. The main reason for this finding is that excessive nitrogen in the soil causes a negative effect on the AMF, and the host plant needs to synthesize much more organic acid to resist the stress conditions, which needs further research.

Conclusion

In brief, the results of the present investigation show that salt stress and alkali stress differ greatly and the physiological adaptive strategy of *L. chinensis* seedlings to the two stresses is also different. AMF colonization can protect the seedlings against salt-alkali stress and nitrogen deposition by adjusting the osmotic and ion balance (by changing the inorganic cation-anion and organic solute content). In addition, excessive nitrogen exerts negative effects on the mycorrhizal colonization and the salt-alkali tolerance of *L. chinensis* seedlings. Therefore, AMF inoculation in *L. chinensis* seedlings could serve as a useful tool for alleviating salinity and alkalinity stress, and could be further applied to a practice such as restoring and reestablishing deteriorated salt-alkali grasslands, as found in northeastern China. In addition, our study also provides an important theoretical basis for the responses of *Leymus chinensis*-AMF symbionts to nitrogen deposition.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 M. Kanakidou, S. Myriokefalitakis, N. Daskalakis, G. Fanourgakis, A. Nenes, A. R. Baker, K. Tsigaridis and N. Mihalopoulos, *J. Atmos. Sci.*, 2016, **73**(5), 2039–2047.
- 2 M. A. Sutton and A. Bleeker, *Nature*, 2013, **494**(7438), 435–437.
- 3 A. F. Bouwman, D. P. Van Vuuren, R. G. Derwent and M. Posch, *Water, Air, Soil Pollut.*, 2002, **141**(1), 349–382.
- 4 W. D. Bowman, C. C. Cleveland, Í. Halada, J. Hreško and J. S. Baron, *Nat. Geosci.*, 2008, **1**(11), 767–770.
- 5 C. M. Jiang, W. T. Yu, Q. Ma, Y. G. Xu, H. Zou, S. C. Zhang and W. P. Sheng, *Environ. Pollut.*, 2013, **182**, 430–436.
- 6 H. F. Fan, C. X. Du and S. R. Guo, *Environ. Exp. Bot.*, 2013, **86**, 52–59.
- 7 R. Munns, *Plant, Cell Environ.*, 2002, **25**(2), 239–250.
- 8 A. K. Parida and A. B. Das, *Ecotoxicol. Environ. Saf.*, 2005, **60**(3), 324–349.
- 9 C. W. Yang, P. Wang, C. Y. Li, D. C. Shi and D. L. Wang, *Photosynthetica*, 2008, **46**(1), 107–114.
- 10 R. Guo, L. X. Shi and Y. F. Yang, *Soil Sci. Plant Nutr.*, 2009, **55**(5), 667–679.
- 11 J. T. Zhang and C. S. Mu, *Soil Sci. Plant Nutr.*, 2009, **55**(5), 685–697.
- 12 N. Aliasgharzadeh, S. N. Rastin, H. Towfighi and A. Alizadeh, *Mycorrhiza*, 2001, **11**(3), 119–122.
- 13 G. N. Al-Karaki, *Mycorrhiza*, 2000, **10**(2), 51–54.
- 14 H. R. Asghari, P. Marschner, S. E. Smith and F. A. Smith, *Plant Soil*, 2005, **273**(1–2), 245–256.
- 15 B. Giri, R. Kapoor and K. G. Mukerji, *Microb. Ecol.*, 2007, **54**(4), 753–760.
- 16 Z. Q. He, C. X. He, Z. B. Zhang, Z. R. Zuo and H. S. Wang, *Colloids Surf., B*, 2007, **59**(2), 128–133.
- 17 M. Echeverria, A. A. Scambato, A. I. Sannazzaro, S. Maiale, O. A. Ruiz and A. B. Menéndez, *Mycorrhiza*, 2008, **18**(6–7), 317–329.
- 18 J. X. Lin, Y. N. Wang, S. N. Sun, C. S. Mu and X. F. Yan, *Sci. Total Environ.*, 2017, **576**, 234–241.
- 19 G. S. Liu and D. M. Qi, *Acta Prataculturae Sinica*, 2004, **13**, 6–11.
- 20 Y. F. Yang, G. C. Liu and B. T. Zhang, *Acta Bot. Sin.*, 1995, **37**, 147–153.
- 21 J. M. Phillips and D. S. Hayman, *Trans. Br. Mycol. Soc.*, 1970, **55**(1), 158–163.
- 22 Q. S. Wu, Y. N. Zou and X. H. He, *Acta Physiol. Plant.*, 2010, **32**, 297–304.
- 23 S. D. Bao, *Analysis Methods for Soil and Agriculture Chemistry*, China Agr. Press, Beijing, 1981.
- 24 G. L. Zhu, X. W. Deng and W. N. Zuo, *Plant Physiol.*, 1983, **1**, 35–37.



- 25 C. M. Greive and S. R. Grattan, *Plant Soil*, 1983, **70**, 303–307.
- 26 A. A. Murkute, S. Sharma and S. K. Singh, *Hortic. Sci.*, 2006, **33**, 70–76.
- 27 H. Evelin, R. Kapoor and B. Giri, *Ann. Bot.*, 2009, **104**, 1263–1280.
- 28 R. Hajiboland, A. Aliasgharzadeh, S. F. Laiegh and C. Poschenrieder, *Plant Soil*, 2010, **331**, 313–327.
- 29 C. W. Yang, W. Q. Guo and D. C. Shi, *J. Agron.*, 2010, **102**(4), 1081–1089.
- 30 G. Feng, F. S. Zhang, X. L. Li, C. Y. Tian, C. Tang and Z. Rengel, *Mycorrhiza*, 2002, **12**, 185–190.
- 31 F. Jahromi, R. Aroca, R. Porcel and J. M. Ruiz-Lozano, *Microb. Ecol.*, 2008, **55**, 45–53.
- 32 E. Blumwald, *Curr. Opin. Cell Biol.*, 2000, **12**(4), 431–434.
- 33 R. Munns and M. Tester, *Annu. Rev. Plant Biol.*, 2008, **59**, 651–681.
- 34 X. Wang, S. Geng, Y. J. Ri, D. Cao, J. Liu, D. Shi and C. Yang, *Sci. Hortic.*, 2011, **130**(1), 248–255.
- 35 J. K. Zhu, *Curr. Opin. Cell Biol.*, 2003, **6**, 441–445.
- 36 C. W. Yang, J. N. Chong, C. N. Kim, D. C. Shi and D. L. Wang, *Plant Soil*, 2007, **294**, 263–276.
- 37 R. Porcel, R. Aroca and J. M. Ruiz-Lozano, *Agron. Sustainable Dev.*, 2012, **32**, 181–200.
- 38 H. Evelin, B. Giri and R. Kapoor, *Mycorrhiza*, 2013, **23**(1), 71–86.
- 39 X. P. Wang, S. J. Geng, Y. J. Ri, D. H. Cao, J. Liu, D. C. Shi and C. W. Yang, *Sci. Hortic.*, 2011, **130**, 248–255.
- 40 W. C. Chen, P. J. Cui, H. Y. Sun, W. Q. Guo, C. W. Yang, H. Jin, B. Fang and D. C. Shi, *Ind. Crops Prod.*, 2009, **30**(3), 351–358.
- 41 G. H. Xu, H. Magen, J. Tarchitzky and U. Kafkafi, *Adv. Agron.*, 1999, **68**, 97–150.
- 42 P. Zuccarini and P. Okurowska, *J. Plant Nutr.*, 2008, **31**(3), 497–513.
- 43 I. C. Cantrell and R. G. Linderman, *Plant Soil*, 2001, **233**, 269–281.
- 44 J. G. Bulwada, D. P. Stribley and P. B. Tinker, *New Phytol.*, 1983, **93**, 217–225.
- 45 J. H. Graham and J. P. Syversten, *New Phytol.*, 1984, **97**, 277–284.
- 46 M. Sharifi, M. Ghorbanli and H. Ebrahimzadeh, *J. Plant Physiol.*, 2007, **164**(9), 1144–1151.
- 47 G. H. Rabie and A. M. Almadini, *Afr. J. Biotechnol.*, 2005, **4**, 210–222.
- 48 J. X. Lin, C. S. Mu, Y. Wang and X. Li, *J. Anim. Plant Sci.*, 2014, **24**(3), 904–912.

