


Cite this: *RSC Adv.*, 2018, 8, 12588

CO₂–C evolution rate in an incubation study with straw input to soil managed by different tillage systems

X. S. Li,^a H. F. Han,^a ^{ab} T. Y. Ning^a and R. Lal^b

A laboratory incubation experiment was conducted to assess the impact of straw input on CO₂–C emissions. After the winter wheat (*Triticum aestivum* L.) growing season, soil samples were collected from the 0–20 cm soil layer. The experiment was conducted on a brown loam soil, classified as a Udoll according to the U. S. soil taxonomy. Treatment levels consisted of three tillage practices: sub-soiling (ST), no-till (NT) and the conventional tillage (CT), two straw management (with and without input of straw), three temperature (25, 30 and 35 °C), and three moisture regimes (55%, 65% and 75% of field moisture capacity or FMC). The results showed that the rate of straw decomposition was the highest in the soil under NT management. The relationship between rate of cumulative CO₂–C and straw decomposition was significant under NT ($R^2 = 0.52$). The soil CO₂–C release under incubation was significantly higher with than without the input of straw ($R^2 = 0.95$). Furthermore, soil respiration increased with increases in incubation temperature and FMC. At 75% FMC, the rate of CO₂–C release increased by 21.9 mg kg^{−1} d^{−1} at 30 °C and 32.0 mg kg^{−1} d^{−1} at 35 °C compared with that at 25 °C. At 35 °C, the rate of CO₂–C release increased by 43.6 mg kg^{−1} d^{−1} at 65% FMC and 51.2 mg kg^{−1} d^{−1} at 75% FMC incubation than that of at 55% FMC under ST. The degree of control on the CO₂–C evolution rate during the pre-incubation period and with higher incubation temperature and FMC was better for the soil from NT than that from CT and ST, and better yet for the soil from ST than that from CT.

Received 23rd January 2018
Accepted 24th March 2018

DOI: 10.1039/c8ra00708j

rsc.li/rsc-advances

1. Introduction

The North China Plain (NCP) is one of China's major agricultural areas (3.3×10^5 km²) with a population of approximately 130 million.¹ Winter wheat (*Triticum aestivum* L.) and summer maize (*Zea mays*) are the predominant grain crops in the NCP. Maintaining increasing soil organic carbon (SOC) stock and reducing CO₂–C emissions are essential for the long-term sustainability of agriculture in this area. CO₂–C emissions from farmland soil is closely related to the straw decomposition rate.^{2,3} Therefore, the challenge lies in adopting management systems that would result in increasing SOC stock while reducing CO₂–C emissions.

SOC is certainly the source of soil CO₂–C emissions, but also the main existing C form in the farmland C bank. Thus, rational soil management is the key to enhance soil C sequestration.^{4,5} Tillage accelerates soil organic matter (SOM) oxidation by improving soil aeration, increasing the contact between soil and crop residues, and exposing aggregate-protected organic matter

to microbial attack.⁶ Crop residues are an important source of potential depletion of SOC stocks and partially as the source of CO₂–C emissions offset the environmental suitability.⁷ Different incubation temperatures and field moisture capacity (FMC) conditions may alter the soil C release rate under different tillage treatments.⁸ In general, gaseous fluxes are highly significantly ($P < 0.01$) or significantly ($P < 0.05$) related to temperature either at the soil surface or at 5 cm depth under both conventional tillage (CT) and no-till (NT) systems.⁹ The incubation temperatures and FMC might be crucial factors in diurnal variation of gaseous emissions under soil incubation.^{10,11}

The nutrients released from straw decomposition play an important role in soil fertility. It is widely accepted that straw retention provides a good environment for enhancing microbial activity and SOC stock by improving soil physical and chemical properties.^{2,12} Therefore, the study of the effects of straw retention on soil fertility, soil respiration, and dynamic changes in soil quality are of great interest in addressing global issues. However, the results of incubation studies are strongly influenced by straw retention, soil temperature and soil moisture which are well documented as fundamental parameters in model simulations and predictions of soil respiration.¹³ Controlling the CO₂–C emissions and using the straw effectively are a priority researchable themes. Modeling the influences of

^aCollege of Agronomy, State Key Laboratory of Crop Biology, Shandong Key Laboratory of Crop Biology, Shandong Agricultural University, Tai'an 271018, Shandong, P. R. China. E-mail: hhf@sdau.edu.cn; Tel: +86 538 8242653

^bCarbon Management and Sequestration Center, School of Environment and Natural Resources, The Ohio State University, Columbus, OH 43210, USA



farmland soil with varying C-substrates, incubation temperatures, and soil moisture regimes can be useful in identifying the most appropriate strategies of tillage methods and management options. In addition, on-going research experiments are focused on the effects of straw retention with NT or CT on soil C sequestration but not on sub-soiling (ST).^{1,2,14,15} Thus, there are few, if any, reports available regarding the mechanisms of the evolution of CO₂-C emissions from soil in relation to different organic substrates, incubation temperatures, and soil moisture conditions for different tillage methods.¹⁶

The present study was conducted to determine whether there is a best management practice (BMPs, *i.e.*, rational combination of tillage management with straw input), that can be used to identify strategies or methods of mitigating CO₂-C evolution. The present study is based on a 12 years ongoing soil management experiment involving different tillage systems in a wheat (*Triticum aestivum* L.)-maize (*Zea mays*) cropping system in NCP, China. The objective of this study was to understand how crop residues, incubation temperature, and soil moisture regime affect soil respiration under long-term management using diverse tillage systems.

2. Materials and methods

2.1. Experimental site

The field experiment was located at Tai'an (Northern China, 36°10'19" N, 117°9'03" E; altitude 134.0 m), which is a typical grain producing area in NCP. This experiment was established in a long-term tillage experiment site of Shandong Agricultural University. The average annual precipitation and temperature were 697 mm and 13.0 °C, respectively. The annual frost-free period was approximately 170–196 d and the annual sunlight duration was 2627 h. The soil is classified as Udoll according to the U. S. soil taxonomy,¹⁷ and it is a brown loam in texture. The soil textural composition consisted of 40% sand, 44% silt, and 16% clay. Some additional characteristics of the surface layer (0–20 cm) were as follows: pH = 6.8; soil bulk density = 1.43 g cm⁻³; SOM = 1.36%; soil total nitrogen = 0.13%; and soil total phosphorous = 0.13%.

2.2. Experimental design

The study was initiated in 2002, and consisted of three tillage practices: sub-soiling (ST), no-till (NT) and the conventional tillage (CT) as the control. The tillage treatments were laid out in a randomized block design (RBD) with three replications. The plot sizes for the main plots and subplots were 60 × 15 m and 15 × 15 m, respectively. The three tillage methods used in this experiment were implemented just before wheat (*Triticum aestivum* L.) was seeded. Maize was seeded directly in an unploughed seedbed. The CT treatment comprised of furrow ploughing to 25 cm depth, and ST of subsoiling to 40 cm depth. Both CT and ST plots were disked twice to 10 cm depth before seeding. Winter wheat (Jimai-22) was sown at a seed rate of 90 kg ha⁻¹, around October 12, 2012 and was harvested on June 13, 2013. The basal fertilizer applied before sowing comprised of 225 kg N ha⁻¹, 150 kg P₂O₅ ha⁻¹, and 105 kg K₂O ha⁻¹. 100 kg N

ha⁻¹ was used as a topdressing during the jointing stage, along with 160 mm of supplemental irrigation. Maize (Zhengdan-958) was sown on June 18, 2013 at a rate of 66 600 plants ha⁻¹ and was harvested on October 8, 2013. For maize, 120 kg N ha⁻¹, 120 kg P₂O₅ ha⁻¹, and 100 kg K₂O ha⁻¹ were used as a basal fertilizer, and 120 kg N ha⁻¹ was used as topdressing during the jointing stage.

2.3. Sampling and culture design

Soil samples were collected after the wheat harvest in June 2013; each sample was a composite from five random positions in each plot from the 0–20 cm soil layer. Soil samples were air dried, gently ground and passed through a 2 mm sieve.

A three-factor-factorial experiment was established comprising of soil samples obtained from three tillage systems, three incubation temperatures, three moisture regimes, and two residue treatments either with or without straw input. Fifty grams of soil was mixed with the 0.67 g of winter-wheat straw powder. The three incubation temperatures used were 25 °C, 30 °C, and 35 °C, and the three soil moisture regimes were 55%, 65%, and 75% of the maximum field moisture capacity.^{18,19} The soil moisture content of the composite sample was determined using a Theta Probe soil moisture sensor (Delta T Devices, UK).

Fifty grams of dried soil from each plot was placed into a 50 ml polyvinyl tube (30 mm internal diameter) after mixing with 0.67 g winter-wheat straw powder (at a rate of 5 g C kg⁻¹ soil). The control consisted of soil without the straw powder. The straw of winter wheat consisted of 507 g C and 18.5 g N kg⁻¹ (C : N ratio of 27.4).

A soil-filled tube was gently tapped by hand against the wall, which resulted in a soil bulk density of approximately 1.2 g cm⁻³ (typical value of the field conditions). The water content in the soil was adjusted to 55%, 65%, and 75% of the soil water-holding capacity and was maintained by the method of Mahmood-ul-Hasan.¹⁸ Soil-filled tubes with open tops were immediately transferred to the incubator. Each soil-filled tube was then placed into an enclosed 280 cm³ jar containing a vial with 10 ml 0.5 M NaOH for trapping the respired CO₂. These jars were incubated in the dark for up to 30 d under constant incubation temperatures and moisture regimes. The CO₂-trapped vials were replaced with new vials every 3 d.

2.4. Measurements

The straw decomposition rate was determined based on a suppose that adding straw does not affect the original SOC mineralization. The decomposition rate was computed by comparing the SOC content in the soil with and without straw input using the following equation:^{20,21}

$$\text{Decomposition rate} = (\text{CO}_2\text{-C}_t - \text{CO}_2\text{-C}_0)/C \times W \times 100\%$$

where CO₂-C_t is the amount of SOC released with straw input at time, and CO₂-C₀ is the amount of SOC released without straw; C is the SOC of straw and W is the straw dry matter.

The amount of CO₂-C released from each tube was determined by titrating the residual NaOH solution with 0.5 M HCl in



the presence of excess BaCl_2 to a phenolphthalein end point.¹⁹ The release rate of $\text{CO}_2\text{-C}$ for different treatments was calculated as the function of incubation duration in days. Three blank jars without soil but with a vial of NaOH solution were also included as control during the incubation period. The soil respiration rate was calculated as the difference in the amount of HCl consumed. The $\text{CO}_2\text{-C}$ evolution rate and cumulative $\text{CO}_2\text{-C}$ evolution of different treatments were calculated as the function of incubation duration in days.

2.5. Statistical analysis

In order to describe the correlation among soil respiration rate and the straw decomposition rate for different incubation temperatures, moistures and straw inputs, several multivariate statistical analyses were applied with a minimum number of parameters.⁶ The analysis consisted of three main steps. First, outliers were eliminated, because they did not follow a normal distribution when assessed using statistical tests. Second, the normality of residues was analyzed by using SPSS 19.0 statistical software and was classified by a cluster analysis to eliminate duplications of highly correlated properties. Third, a factor analysis was applied to obtain a small number of homoscedasticity (orthogonal properties). The figures were drawn using Sigma Plot 10.0 and Excel 2003. The standard deviation (SD) and least significant difference (LSD) were calculated to compare the treatment means. LSD was used to identify differences between treatments at $P < 0.05$.

3. Results

3.1. Percentage increase in $\text{CO}_2\text{-C}$ evolution with straw input

A significant interaction between tillage methods and straw input on soil respiration was observed throughout the incubation period (S1). Compared with the treatments without straw, input of straw significantly ($P < 0.05$) increased the soil respiration rate. In the treatment without straw input, the rate of soil $\text{CO}_2\text{-C}$ evolution peaked on the 6th day and remained at the steady state until the 24th day. (Fig. 1A). In comparison, the treatment with straw input reached the maximum $\text{CO}_2\text{-C}$ evolution on the 9th day and remained at the steady state rate until the 27th day of incubation (Fig. 1B). Regardless of the straw input, soil $\text{CO}_2\text{-C}$ evolution rate was the lowest under NT (Fig. 1A and B).

ST reduced the $\text{CO}_2\text{-C}$ evolution rate in the first incubation period, and the $\text{CO}_2\text{-C}$ evolution rate gradually increased during the later incubation period, with or without the input of straw (Fig. 1C and D). However, NT increased the percentage of the $\text{CO}_2\text{-C}$ evolution rate without straw input during the late incubation period (Fig. 1C). As the soil C substrate was enhanced with the straw input, the process of soil respiration intensity was also enhanced. Soil from NT significantly reduced the percentage of $\text{CO}_2\text{-C}$ evolution rate ($P < 0.01$) at all the times, and that under ST significantly reduced the $\text{CO}_2\text{-C}$ evolution rate during the first 15 d. Meanwhile, the percentage of $\text{CO}_2\text{-C}$ evolution rate increased at the end of the incubation period and was approximately 14.9% of that under CT (Fig. 1D).

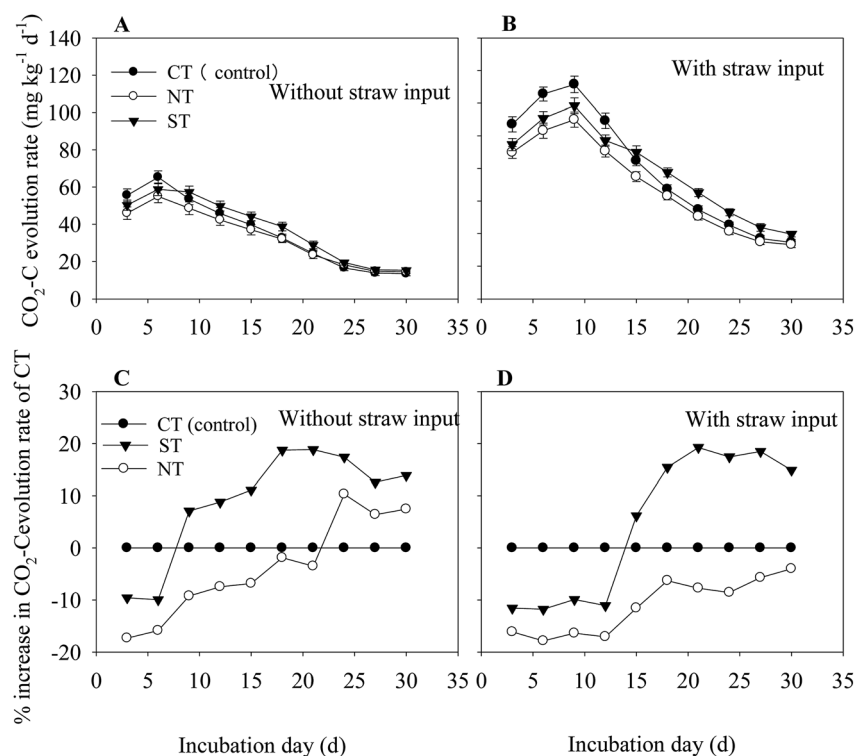


Fig. 1 Rate of $\text{CO}_2\text{-C}$ evolution in soils under the input of with (A), and without (B) straw, and the percentage increase in $\text{CO}_2\text{-C}$ evolution under straw input compared with the treatment of without straw input (C) and with straw (D). Bars in (A)–(C) are \pm standard errors of means ($n = 3$), which come from the standard deviation of systematic error. CT: conventional tillage; ST: sub-soiling; NT: no tillage.



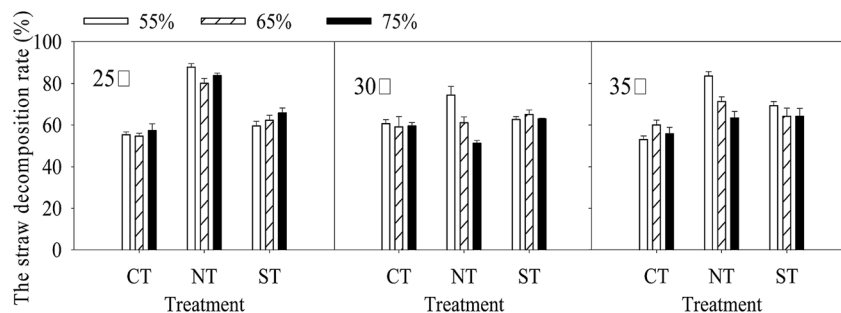


Fig. 2 The straw decomposition rate after different incubation temperature and FMC. Bars represent standard error ($n = 3$), which come from the standard deviation of systematic error. CT: conventional tillage; ST: sub-soiling; NT: no tillage.

3.2. Straw decomposition rate

Within the same incubation temperature and FMC regime, the straw decomposition rate (Fig. 2) was the highest under NT followed by that under ST. Within the same incubation temperature, straw decomposition rate under NT peaked at 55% FMC. Within the same moisture regime, straw decomposition rate was the highest at 30 °C under CT and at 35 °C under ST. After incubation at 25 °C, straw decomposition rate under CT and ST were the highest under 75% FMC. The evolution of $\text{CO}_2\text{-C}$ was the highest under NT at 55% FMC. Within the condition of 25 °C and 55% FMC, the straw decomposition rate increased by 58.8% and 47.6% under NT compared with that under CT and ST, respectively.

The correlation between the cumulative $\text{CO}_2\text{-C}$ evolution and straw decomposition rate varied under different treatments. A significant negative correlation between the cumulative $\text{CO}_2\text{-C}$ evolution rate and straw decomposition rate was observed under NT ($P < 0.05$). However, no significant relationship was observed between the cumulative $\text{CO}_2\text{-C}$ evolution rate and straw decomposition rate under CT and ST.

3.3. Soil $\text{CO}_2\text{-C}$ evolution rate under straw input

The rate of $\text{CO}_2\text{-C}$ release from the soil with the straw input differed among different incubation temperatures and moisture regimes. The overall trend was a reduction in the rate after the initial increase before attainment of the steady state. The $\text{CO}_2\text{-C}$ respiration rate was the highest under CT during the first 12 d. In addition, the $\text{CO}_2\text{-C}$ respiration rate was higher under ST than it was under CT, and it was the lowest under NT at all the times.

Regarding to the effects of environmental variables, the interactive effect of incubation temperature and moisture regime significantly affected the soil respiration rate throughout the incubation period in treatments with straw input ($P < 0.01$). The $\text{CO}_2\text{-C}$ respiration rate was the lowest under NT at all times. Compared with CT, the soil respiration rate under ST significantly increased in treatments with higher temperature and moisture regimes ($P < 0.01$). The higher the temperature and the moisture regime during incubation, the higher was the soil respiration rate (Fig. 3A–I).

In soil with 65% and 75% FMC, the soil respiration rate significantly increased with the increase in incubation temperature. Under 65% FMC incubation, the soil respiration rate under ST was increased by 27.8% and 58.8% at incubation temperatures of 30 °C and 35 °C, respectively, compared with that at 25 °C; the incremental increase in $\text{CO}_2\text{-C}$ evolution under ST was the lowest among the three treatments. Under the same incubation temperature but different moisture regimes, the soil respiration rate increased as FMC increased. At 35 °C, the respiration rate for soil from ST with FMC of 65% and 75% was increased by 37.9% and 82.5%, respectively, compared with that under 55% FMC during the latter part of incubation.

At 75% FMC and 35 °C, the peak time of the respiration rate for soil under ST exceeded 3–5 d, compared with that for 30 °C and 25 °C. Therefore, higher incubation temperatures and moisture regimes increased the soil respiration rate. On the contrary, soil under NT had a significantly lower $\text{CO}_2\text{-C}$ evolution rate than soil under CT and ST at all incubation time. However, the $\text{CO}_2\text{-C}$ evolution rate under ST was significantly lower than that under CT for the earlier incubation time under higher temperature and moisture regimes.

3.4. Percentage increase in soil cumulative $\text{CO}_2\text{-C}$ evolution with straw input

The cumulative $\text{CO}_2\text{-C}$ evolution, as affected by tillage, also changed with straw input and incubation time (Fig. 4A–D). Without straw input, the cumulative $\text{CO}_2\text{-C}$ evolution from soil under ST and NT was generally lower than that under CT during the first 15 d. Thereafter, the cumulative $\text{CO}_2\text{-C}$ evolution under ST gradually exceeded that under CT (Fig. 4A and C). The cumulative $\text{CO}_2\text{-C}$ evolution was significantly ($P < 0.05$) low under NT and was the lowest among the three treatments. With straw input, the cumulative $\text{CO}_2\text{-C}$ evolution in soil under ST and NT was significantly lower than that under CT throughout the incubation period, and the rate under NT was lower by 13.3% compared with that under CT by the end of the incubation period (Fig. 4B and D).

3.5. Soil cumulative $\text{CO}_2\text{-C}$ evolution under straw input

The cumulative $\text{CO}_2\text{-C}$ evolution, as affected by soil incubation temperature and moisture regimes, also changed with the



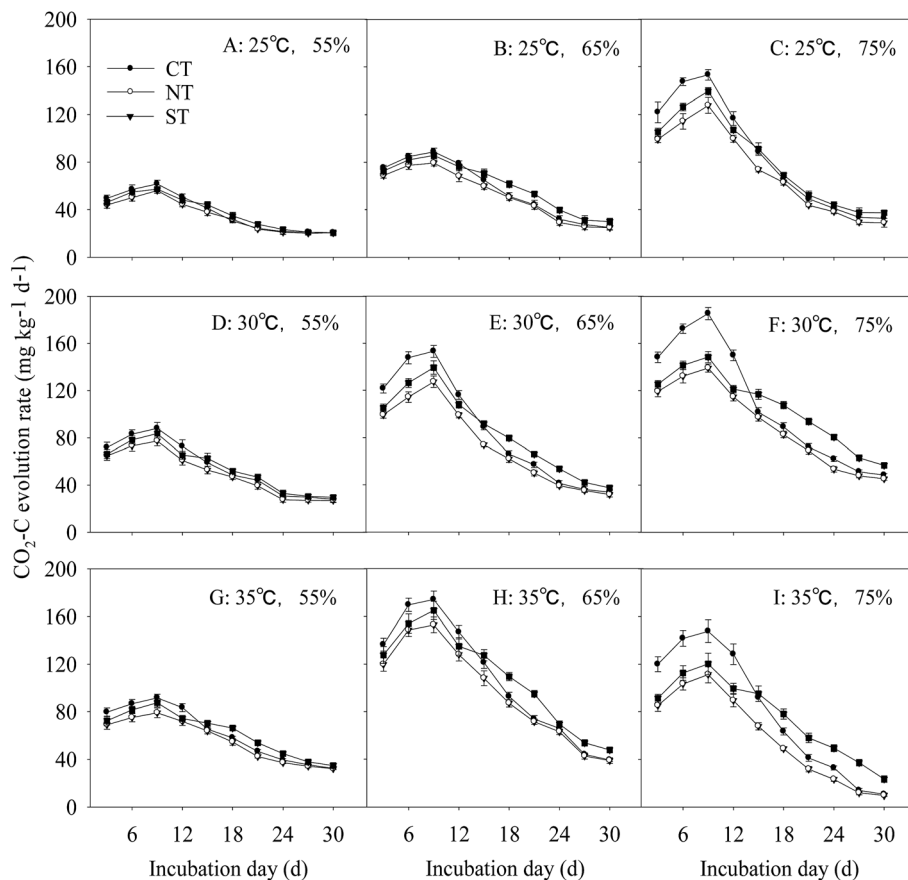


Fig. 3 Rate of soils $\text{CO}_2\text{-C}$ evolution in different incubation temperatures and FMC under straw input. A to I: the soil treatments with different incubation temperatures and FMC. Bars in (A)–(I) are \pm standard errors of means ($n = 3$), which come from the standard deviation of systematic error. CT: conventional tillage; ST: sub-soiling; NT: no tillage.

incubation time (Fig. 5A–I). The cumulative $\text{CO}_2\text{-C}$ evolution was the lowest under NT at all times and was lower under ST than under CT during the early incubation period. As the incubation time increased, the cumulative $\text{CO}_2\text{-C}$ evolution changed with the change in incubation temperatures and moisture regimes.

For soil under the same FMC and different incubation temperatures, the cumulative $\text{CO}_2\text{-C}$ evolution increased as incubation temperature increased, revealing significant ($P < 0.05$) differences in 65% and 75% FMC. With 65% FMC, the cumulative $\text{CO}_2\text{-C}$ evolution was lower under ST and NT than that under CT during the first 21 d under 25 °C and 35 °C. At 30 °C, the cumulative $\text{CO}_2\text{-C}$ evolution was lower under ST and NT than that under CT at all times. At 75% FMC, the cumulative $\text{CO}_2\text{-C}$ evolution was significantly ($P < 0.01$) lower under ST and NT than that under CT. Incubating the soil under ST at 30 °C and 35 °C increased the cumulative $\text{CO}_2\text{-C}$ evolution by 30.2% and 43.8%, respectively, compared with that at 25 °C. The cumulative $\text{CO}_2\text{-C}$ evolution for soil under NT at 30 °C and 35 °C increased by 25.5% and 36.8%, respectively, than that at 25 °C. Under the same incubation temperature and different FMC, the cumulative $\text{CO}_2\text{-C}$ evolution increased as FMC increased.

Under the condition of 65% and 75% FMC, the cumulative $\text{CO}_2\text{-C}$ evolution increased significantly with the increase in

incubation temperature. At 35 °C incubation, the cumulative $\text{CO}_2\text{-C}$ evolution for soil under ST at 65% and 75% FMC increased by 74.0% and 86.9%, respectively, compared with that at 55% FMC. At 35 °C incubation, cumulative $\text{CO}_2\text{-C}$ evolution for soil under NT at 65% and 75% FMC decreased by 71.8% and 75.7%, respectively, compared with that at 55% FMC. The incremental increase of cumulative $\text{CO}_2\text{-C}$ evolution for soil under CT was the greatest, and that under NT was the least among the three treatments. At the end of the incubation period, the cumulative $\text{CO}_2\text{-C}$ evolution for soil under ST and NT at 75% FMC was lower than that of the soil under CT by 2.2% and 17.6%, respectively.

4. Discussions

This study was conducted under laboratory conditions to better understand the mechanisms that control $\text{CO}_2\text{-C}$ evolution in relation to changes in straw input, temperature, and soil moisture for soil samples obtained from sites under different long-term tillage systems. The short-term incubation period could decrease the bias caused by the loss of soil structure under *in situ* field conditions.^{22,23}



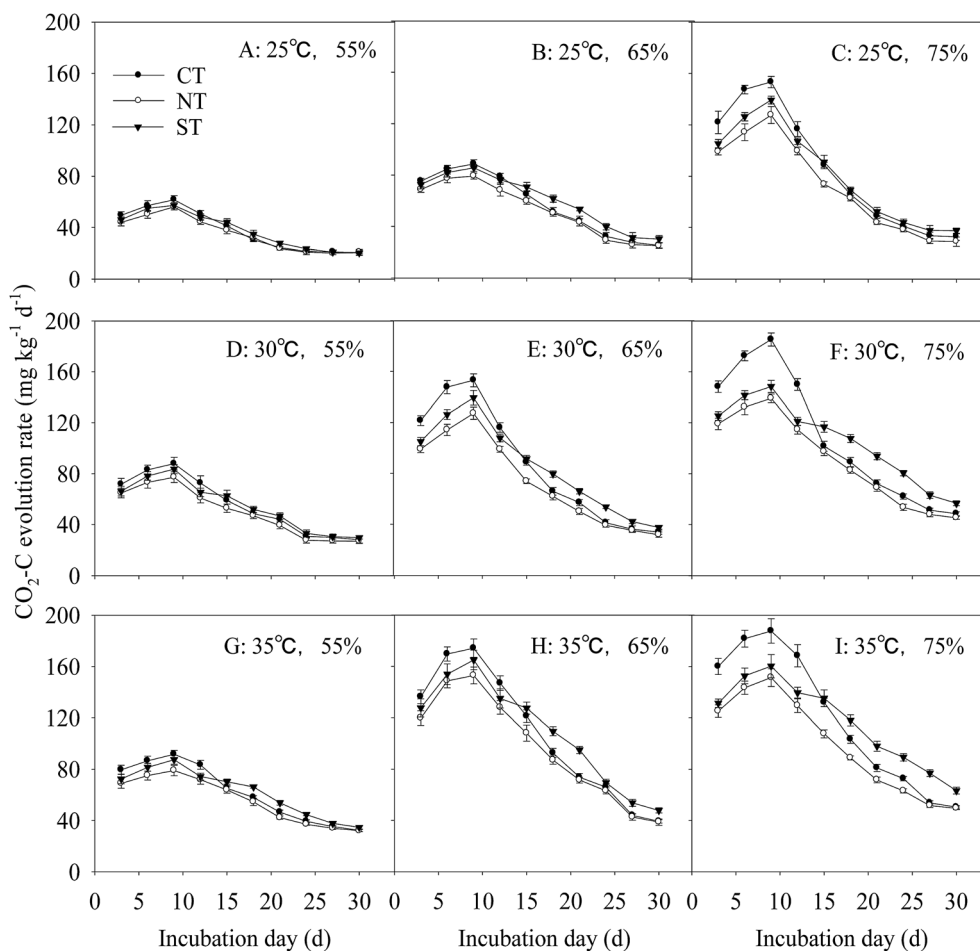


Fig. 4 Cumulative $\text{CO}_2\text{-C}$ evolution in soils under different incubation temperatures and FMC. Bars in (A)–(I) are \pm standard errors of means ($n = 3$), which come from the standard deviation of systematic error. CT: conventional tillage; ST: sub-soiling; NT: no tillage.

4.1. Tillage management systems and straw input effects on soil $\text{CO}_2\text{-C}$ evolution rate

A review of the available literature showed that straw input significantly increase soil respiration rate.^{24,25} However, the data presented in this study did not demonstrate a significant relationship between the soil $\text{CO}_2\text{-C}$ evolution rate and straw decomposition rate, except in soil under NT. This trend of negative correlations observed in this study can be attributed to the assessment of soil respiration using an incubation experiment for determining the rate of straw decomposition. Furthermore, an interaction was observed between tillage and straw input on the soil $\text{CO}_2\text{-C}$ evolution rate and straw decomposition rate.²⁶ In another study, a 30-d micro system test also demonstrated the significant effects of different incubation temperatures, moisture regimes, and organic material added to incubation soil had effect on the respiration rate under different tillage methods.^{27,28}

Similar results were reported from field studies conducted by Han *et al.*,² indicating that ST and NT methods can reduce the soil $\text{CO}_2\text{-C}$ evolution rate compared with that by CT. This trend is attributed to the low level of soil disturbance under ST and NT compared with that under CT, and it leads to frequent

disruption and breakdown of soil aggregates.¹¹ Thus, the rate of soil $\text{CO}_2\text{-C}$ evolution is faster in soil under CT than that under the conservation tillage. Similar results were also reported who conducted the field studies.^{8,29} Furthermore, the reduction in the soil $\text{CO}_2\text{-C}$ evolution rate under tillage with incubation time suggests that the impact of BMPs can be identified even from a short-term laboratory incubation.

Several studies have reported that input of straw substrate increases the soil $\text{CO}_2\text{-C}$ evolution rate^{2,10} due to the effects of tillage on the soil microbial community^{8,25} and on soil desiccation.³⁰ Microbial communities quickly proliferate with the start of incubation under favorable incubation temperature and moisture regimes.³¹ Thus, both the soil $\text{CO}_2\text{-C}$ evolution rate and cumulative $\text{CO}_2\text{-C}$ evolution increase significantly in the initial incubation period. A rapid proliferation of microbial communities might have subsequently allocated more substrates to their proliferation and growth than to respiration thereby decreasing the $\text{CO}_2\text{-C}$ evolution in soil with straw compared with those without straw input, especially during the earlier incubation period. Thereafter, the soil respiration rate and cumulative $\text{CO}_2\text{-C}$ evolution reached a steady state equilibrium.



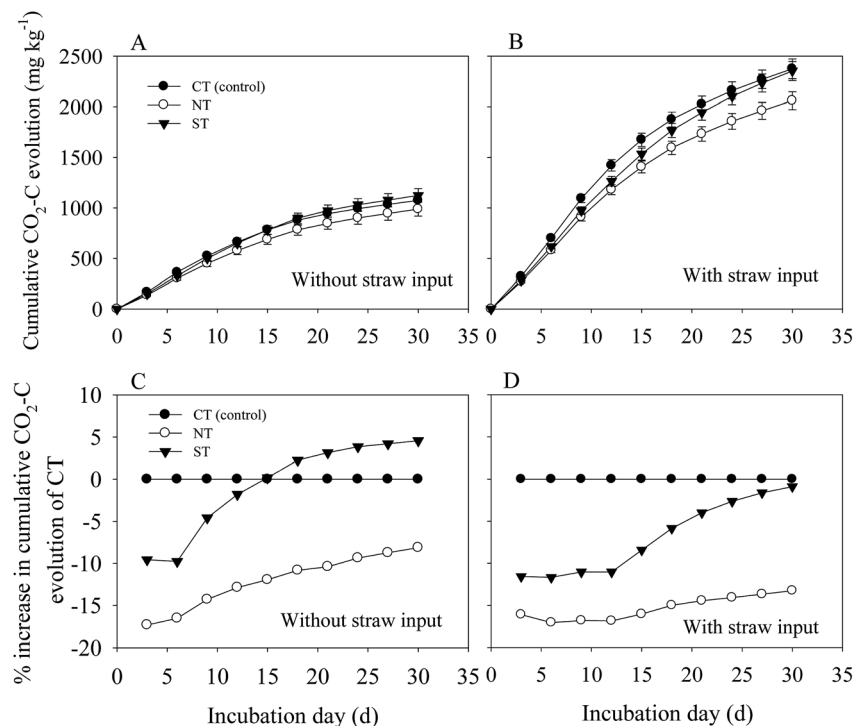


Fig. 5 Cumulative CO₂-C evolution in soils under the input of with and without straw, and the percentage increase in CO₂-C evolution of with straw input compared with without straw input soil (control). Bars in (A)–(D) are \pm standard errors of means ($n = 3$), which come from the standard deviation of systematic error. CT: conventional tillage; ST: sub-soiling; NT: no tillage.

Due to differences in the straw decomposition rate under different treatments, the effects of straw on the soil respiration rate under different treatments also varied. The straw decomposition rate was higher under NT and ST than under CT. These results differ from a research experiment conducted by Jiang³² which demonstrated that the input of maize straw under NT decomposed at a slower rate than that under other tillage method. However, straw retention can significantly increase soil CO₂ release,³³ as straw provides C substrates and increases soil porosity³⁴ which accelerates the release of CO₂-C. However, the process of C absorption and release by soil with the input of straw requires further research.

4.2. Temperature and moisture effects on soil CO₂-C evolution rate

The relationship of the soil CO₂-C evolution rate with temperature and moisture involves complex interactions dependent on the relative limitation of variable microbial activity and soil substrate supply. These interactions among environments, and thus the relationships may vary accordingly. Similar to the results of an infrared warming experiment³⁵ and a cropland study,³⁶ soil respiration increased as temperature increased in the present study. However, some reports showed that the soil CO₂-C evolution rate might be decoupled from temperature under certain soil moisture levels, resulting in a soil CO₂-C evolution rate that is unaffected by temperature under water stress. For example, Chen³⁷

and Balogh *et al.*²⁸ observed that the soil CO₂-C evolution rate decreased with increasing soil moisture content. However, the results of the present study show that the soil CO₂-C evolution rate increased as soil moisture content increased. The discrepancy between the aforementioned results and those of the present study might be due to differences in the examined soils regarding their chemical, physical, and microbial properties. The decrease in the soil CO₂-C evolution rate at lower soil moisture has been attributed to soluble substrate limitation, whereas at higher soil moisture level—especially at saturation—the soil CO₂-C evolution rate was primarily limited by oxygen. However, the high moisture and temperature incubation could be due to the persistently high microbial activity, as this was the optimal moisture level and high microbial activity might override the influence of substrate limitation.

In general, previous studies documented that the response of the soil CO₂-C evolution rate to warming³⁵ and moisture regimes³⁶ is important for reducing the greenhouse effect and improving soil fertility.³⁸ With increasing global warming and a rapidly falling groundwater table, understanding the response of soil and straw input to high incubation temperature and drought stress is critical. The effect of the farmland soil CO₂-C evolution rate on soil fertility, crop production, and warming are major global issues. Our understanding of the soil CO₂-C evolution rate still needs to be improved by examining different rates of straw input and a wider range of incubation temperatures and moisture regimes.



4.3. Variations in soil CO₂-C evolution rate with incubation time

A laboratory incubation study by Li *et al.*²⁰ demonstrated that the soil CO₂-C evolution rate increased with incubation time, which was ascribed to a change in substrate quality from labile to recalcitrant. The data from the present study for relatively short-term incubation experiment show that the soil CO₂-C evolution rate changes with incubation time, *i.e.*, a temporal effect of soil tillage on soil C dynamics was observed. The present study has demonstrated that the effect of soil tillage on the soil CO₂-C evolution rate increased with incubation time initially but declined during later incubation stages at different soil temperature and moisture regimes, as has also been shown by other studies.³⁵ Over the 30 d incubation period, with 65% FMC, the cumulative CO₂-C evolution was lower under ST and NT than under CT during the first 21 d under 25 °C and 35 °C. At 30 °C, cumulative CO₂-C evolution was lower under ST and NT than under CT at all times. The underlying mechanisms were ascribed to substrate depletion:¹⁷ the longer the incubation time, the more time the microbes had to consume labile C, leaving less of it in the soil. This occurrence was likely due to more confounding factors and less incubation time involved in the present incubation experiment. At the end of the incubation period, cumulative CO₂-C evolution for soil under ST and NT at 75% FMC was lower than that of soil under CT by 2.2% and 17.6%, respectively. The data presented herein showing that the effect of soil tillage changed with incubation time suggests a strong need for extrapolating the data on C dynamics from short term laboratory conditions to large scale field-studies.

5. Conclusions

The data from incubating a long-term tillage soil under two straw management, three temperature levels and three moisture regimes and measuring straw decomposition and soil CO₂-C evolution rate and cumulative CO₂-C evolution throughout the incubation showed that:

(1) Soil management by NT could increase the straw decomposition rate more than that by ST and CT due to differences in incubation temperatures and moisture regimes. During incubation, under ST the soil CO₂-C evolution rate decreased at an earlier stage and increased during the later stage;

(2) Lower incubation temperature and moisture content decreased the CO₂-C evolution rate and cumulative CO₂-C evolution;

(3) Straw input significantly increased CO₂-C evolution rate;

(4) Thus, the data obtained from the present study indicate that, under higher incubation temperatures and moisture regimes, ST and NT management can significantly control CO₂ release compared with that by CT.

These results imply that the response of the CO₂-C evolution rate to future global warming may be affected by changes in soil management practices. In NT and ST, global warming and high FMC may cause less soil CO₂-C evolution, but relatively more CO₂-C evolution may be emitted in CT. Considering that ST and

NT management can significantly control CO₂-C evolution, the rise in temperature and the high FMC would mean less soil CO₂-C evolution to the atmosphere through BMPs in the future.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was financially supported in part by the National Nature Science Foundation of China (No. 31471453; 31771737), and by the Special Fund for Agro-scientific Research in the Public Interest of China (No. 201503117).

References

- 1 W. X. Dong, C. S. Hu, S. Y. Chen and Y. M. Zhang, *Nutr. Cycling Agroecosyst.*, 2009, **83**, 27–37.
- 2 H. F. Han, T. Y. Ning, Z. J. Li and H. M. Cao, *Exp. Agr.*, 2017, **53**, 118–130.
- 3 Q. R. Liu, X. H. Liu, C. Y. Bian, C. J. Ma, K. Lang, H. F. Han and Q. Q. Li, *Sci. World J.*, 2014, **7**, 1–8.
- 4 R. Lal, *Nutr. Cycling Agroecosyst.*, 2008, **81**, 113–127.
- 5 A. Legrand, C. Gaucherel, J. Baudry and J. M. Meynard, *Agron. Sustainable Dev.*, 2011, **31**, 515–524.
- 6 L. J. Guo, Z. S. Zhang, D. D. Wang, C. F. Li and C. G. Cao, *Biol. Fertil. Soils*, 2014, **51**, 65–75.
- 7 F. Monforti, E. Lugato, V. Motola and J. F. Dallemand, *Renewable Sustainable Energy Rev.*, 2015, **44**, 519–529.
- 8 E. Lellei, E. Kovacs, Z. Botta, T. Kalapos, B. Emmett and C. Beier, *Eur. J. Soil Biol.*, 2011, **47**, 247–255.
- 9 S. Z. Tian, T. Y. Ning, S. Y. Chi, Y. Wang, B. W. Wang, H. F. Han, C. Q. Li and Z. J. Li, *Acta Ecol. Sin.*, 2012, **32**, 879–888.
- 10 S. R. Chen, J. Li, P. L. Lu, Y. H. Wang and Q. Yu, *Chin. J. Ecol.*, 2004, **15**, 1552–1560.
- 11 S. Z. Tian, Y. Wang, N. Li, T. Y. Ning, B. W. Wang and Z. J. Li, *Soil Tillage Res.*, 2012, **121**, 57–62.
- 12 T. Kautz, C. López Fandob and F. Ellmer, *Appl. Soil. Ecol.*, 2006, **33**, 278–285.
- 13 Y. Luo and X. Zhou, *Soil respiration and the environment*, Academic Press, an imprint of Elsevier, 2006, ISBN: 9780120887828.
- 14 Y. H. Wei, X. Zhao, Y. L. Zhai, E. P. Zhang, F. Chen and H. L. Zhang, *Tran. CSAE*, 2013, **29**, 87–95.
- 15 X. W. Chen, N. Wang, X. H. Shi, X. P. Zhang, A. Z. Liang, S. X. Jia, R. Q. Fan and S. C. Wei, *Acta Ecol. Sin.*, 2013, **33**, 2676–2683.
- 16 W. P. Zhou, D. F. Hui and W. J. Shen, *PLoS One*, 2014, **9**, e92531.
- 17 Soil Survey Staff, *Keys to Soil Taxonomy*, 1998, 8th edn.
- 18 M. Mahmood-ul-Hasan, *Pak. J. Sci. Ind. Res.*, 2002, **45**, 145–150.
- 19 X. G. Li, X. M. Shi, D. J. Wang and Z. Wei, *Biol. Fertil. Soils*, 2012, **48**, 597–602.



- 20 Q. X. Wen, *Soil Organic Matter Research*, Agricultural Science Press, Beijing, 1984, pp. 285–295.
- 21 X. B. Zhang, M. G. Xu and W. J. Zhang, *Sci. Agric. Sin.*, 2011, **24**, 5013–5020.
- 22 A. Chatterjee and G. D. Jenerette, *Geoderma*, 2011, **163**, 171–177.
- 23 E. Lellei, E. Kovacs, Z. Bot, T. Kalapos, B. Emtamett and C. Beier, *Eur. J. Soil Biol.*, 2011, **47**, 247–255.
- 24 X. S. Li, N. Wu, L. Liu, Y. P. Feng, X. Xu, H. F. Han, T. Y. Ning and Z. J. Li, *Chin. J. Ecol.*, 2015, **6**, 1765–1771.
- 25 Y. Wang, H. J. Li, B. Han, Z. Q. Shi, T. Y. Ning and Z. J. Li, *Acta Ecol. Sin.*, 2007, **27**, 3384–3390.
- 26 S. Z. Tian, T. Y. Ning, Y. Wang, H. J. Li, W. L. Zhong and Z. J. Li, *Chin. J. Ecol.*, 2010, **21**, 373–378.
- 27 C. J. Hu, G. H. Liu, B. J. Fu, T. T. Jin, L. Guo and C. H. Su, *Acta Ecol. Sin.*, 2009, **29**, 4700–4709.
- 28 J. Balogh, K. Pinter, S. Foti, D. Cserhalmi, M. Papp and Z. Nagy, *Soil Biol. Biochem.*, 2011, **43**, 1006–1013.
- 29 S. Muhammad, T. Müller and R. G. Joergensen, *Biol. Fertil. Soils*, 2006, **43**, 93–101.
- 30 M. L. Kakumanu, C. L. Cantrell and M. A. Williams, *Soil Biol. Biochem.*, 2013, **57**, 644–653.
- 31 D. M. Nobili, M. Contin, C. Mondini and P. C. Brookes, *Soil Biol. Biochem.*, 2001, **33**, 1163–1170.
- 32 X. D. Jiang, S. Y. Chi, Y. Wang, T. Y. Ning and Z. J. Li, *Trans. Chin. Soc. Agric. Eng.*, 2009, **25**, 247–251.
- 33 J. Pramod, G. Nikita, B. L. Lakaria, A. K. Biswas and A. S. Rao, *Soil Tillage Res.*, 2012, **121**, 57–62.
- 34 S. Z. Tian, T. Y. Ning, S. Y. Chi, Y. Wang, B. W. Wang, H. F. Han, C. Q. Li and Z. J. Li, *Acta Ecol. Sin.*, 2012, **32**, 879–888.
- 35 C. Tu, F. D. Li, Y. F. Qiao, N. Zhu, C. K. Gu and X. Zhao, *J. Integr. Agric.*, 2017, **16**, 967–979.
- 36 L. T. Liu, C. S. Hu, P. P. Yang, Z. Q. Ju, J. E. Olesen and J. W. Tang, *Agric. For. Meteorol.*, 2015, **207**, 38–47.
- 37 S. T. Chen, Y. Zhang, Z. H. Hu and X. S. Shen, *Environ. Sci. Technol.*, 2012, **33**, 1476–1483.
- 38 X. Yan, F. G. Zhang, L. Yang, T. Li, Y. Y. Che and S. J. Deng, *Eur. J. Soil Biol.*, 2018, **85**, 23–29.

