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Synthesis of an unusual quinazoline alkaloid: theoretical and experimental investigations of its structural, electronic, molecular and biological properties[†]

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An unusual quinazoline alkaloid (**1**) was obtained when 2-aminobenzaldehyde was refluxed with pyrrolidine in ethanol for 12 h. The synthesized compound was characterized using spectral data analysis augmented with X-ray and literature precedent. Single crystal analysis depicted four conformations differing slightly in bond angles and bond lengths. Compound **1** crystallizes in a triclinic crystal system with a $P\bar{1}$ space group having two molecules within the unit cell. The experimentally obtained parameters were compared to those obtained theoretically, which depicted a good agreement. Using the DFT/B3LYP/6-31G (d,p) level of theory, HOMO–LUMO energy gap, molecular electrostatic potential (MEP), vibrational (IR) and NMR analyses were carried out. The HOMO–LUMO energy gap allowed the calculation of chemical hardness, chemical inertness, electronegativity and the electrophilicity index of the molecule, which depicted its potential kinetic stability and reactivity. Prediction of activity spectra of the target compound revealed that compound **1** possesses notable antineoplastic activity with $P_a = 0.884$. The molecule was therefore evaluated against various cancerous cell lines in an *in vitro* SRB assay which depicted that compound **1** possesses the highest growth inhibition activity against THP-1 cell lines with an IC_{50} of 7 μ M.

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Introduction

Alkaloids are a very important class of compounds as far as their role in metabolism and activity is concerned.¹ They are among the oldest drugs used for the treatment of diseases² and are still used as promising therapeutic agents.^{3,4} Deep sea environments are nowadays the focus of research for discovering newer alkaloids with potential therapeutic activities.^{5–7} New alkaloids are also being produced *via* chemical synthesis and natural product modifications with some of them being more active than the naturally obtained ones.^{8,9} In general alkaloids possess a range of biological activities which include analgesic, antiviral, anti-malarial, antineoplastic, antimicrobial, anti-inflammatory, antioxidant, antifungal, antibacterial, hemoglobinization

agents of human leukemia cells, effects on the CNS, estrogenic effects *etc.*^{10–24} Based on these literature reports of alkaloids and our previous work on the isolation, and synthesis of bioactive compounds,^{25–37} we carried out the synthesis of an unusual self-condensation product of 2-aminobenzaldehyde by a method reported by Seidal and coworkers.³⁸ In order to get deeper insights into the molecular, structural and biological properties of the synthesized compound, we resorted to spectral, X-ray and DFT analysis. The experimentally obtained parameters were in close agreement to those obtained theoretically. Molecular electrostatic potential and frontier molecular orbital analysis were performed which permitted the calculation of HOMO–LUMO energy gap and related parameters. Based on PASS results, the molecule was evaluated against four different human cancer cell lines to explore its biological potential.

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† Electronic supplementary information (ESI) available. CCDC 1814904 contains the supplementary crystallographic data for **1**. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c8ra00138c

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Results and discussion

Reduction of 2-nitrobenzaldehyde using iron powder in glacial acetic acid : ethanol : water system (2 : 2 : 1) under ultrasonic irradiation conditions for 1.5 h at 30 °C yielded the corresponding 2-aminobenzaldehyde. Refluxing 2-aminobenzaldehyde in ethanol with pyrrolidine yielded a reaction mixture containing an unusual self-condensation quinazoline alkaloid (**1**) (Fig. 1).

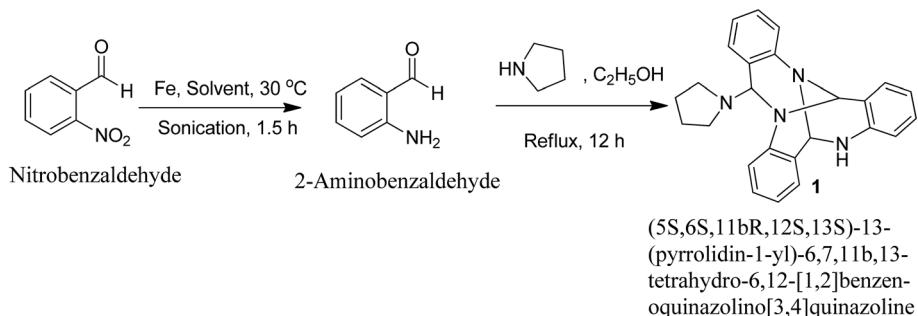


Fig. 1 Synthesis of an unusual quinazoline alkaloid (**1**).

Pure **1** was obtained when the reaction mixture was subjected to normal phase silica gel column chromatography. Its structure was elucidated while analyzing the spectral data in light of literature. The reaction proceeds *via* a series of nucleophilic additions of basic nitrogen of pyrrolidine ring on the highly electrophilic aldehyde group of 2-aminobenzaldehyde to give an intermediate which again condenses with two molecules of 2-

aminobenzaldehyde to yield compound **1** (Fig. 2). The product was characterized using LC-MS, IR and NMR data analysis. LC-MS depicted *m/z* at 381 assignable to $[M + H]^+$ corresponding to its molecular formula $C_{25}H_{24}N_4$. 1H NMR depicted the presence of 12 aromatic protons assignable to three aromatic rings in the molecule whose presence was further supported by the appearance of 18 ^{13}C signals in the aromatic region of the

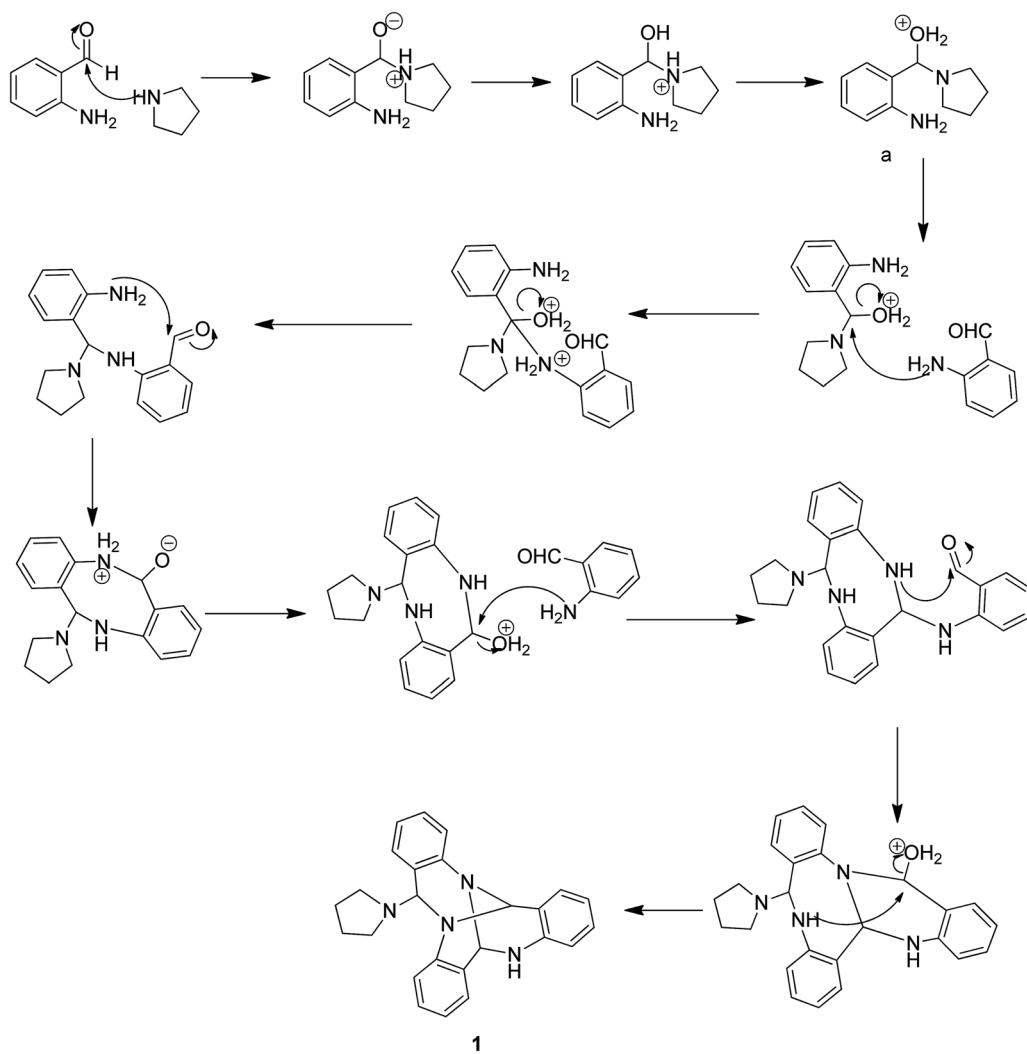


Fig. 2 Mechanism involved in the formation of quinazoline (1).

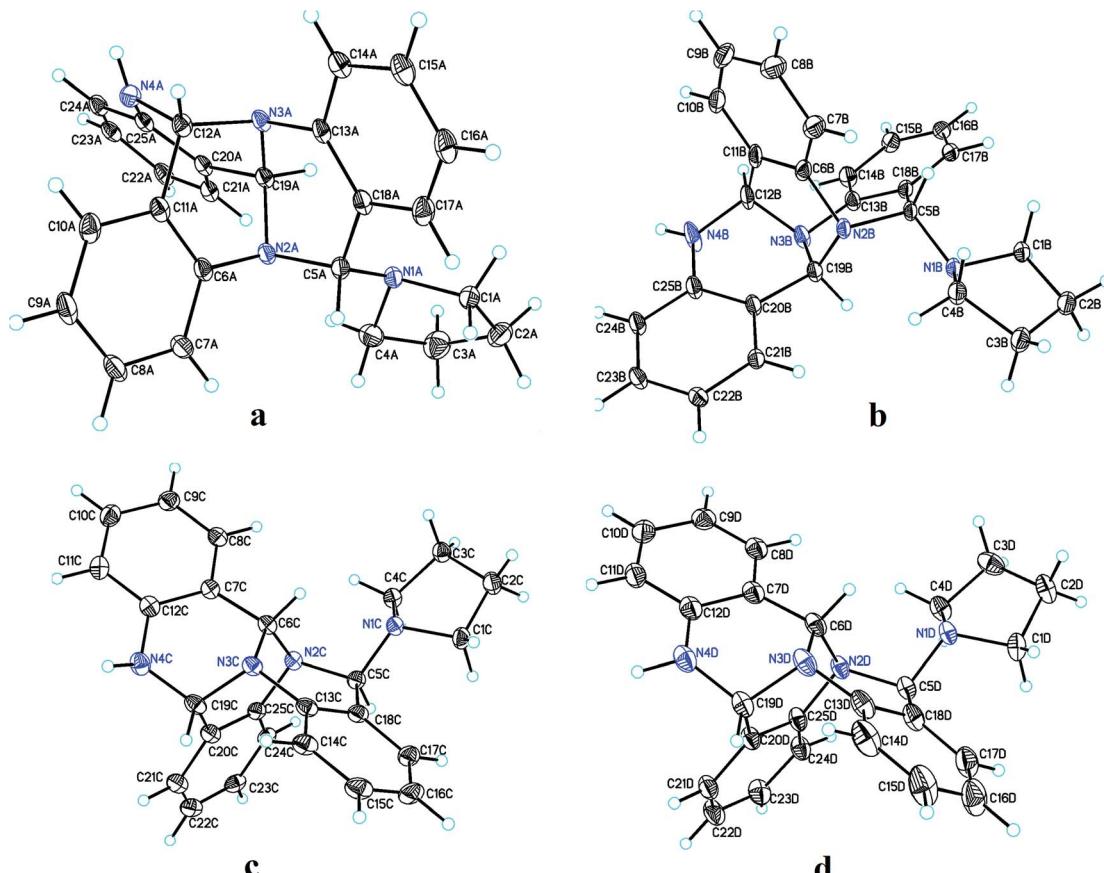


Fig. 3 Four different conformations (a–d) of compound 1 as shown by single crystal X-ray analysis.

Table 1 Crystal data and structure refinement of compound 1

Identification code	Shelx
Empirical formula	C ₁₀₁ H ₉₇ Cl ₃ N ₁₆
Formula weight	1641.29
Temperature	153 (2) K
Wavelength	1.54184 Å
Crystal system	Triclinic
Space group	<i>P</i> 1
Unit cell dimensions	<i>a</i> = 12.8967 (4) Å <i>b</i> = 13.9278 (3) Å <i>c</i> = 23.2246 (7) Å 4093.1 (2) Å ³
Volume	4093.1 (2) Å ³
<i>Z</i>	2
Density (calculated)	1.332 mg m ⁻³
Absorption coefficient	1.497 mm ⁻¹
<i>F</i> (000)	1732
Crystal size	0.345 × 0.275 × 0.233 mm ³
Theta range for data collection	3.492 to 76.684°
Index ranges	-16 ≤ <i>h</i> ≤ 16, -11 ≤ <i>k</i> ≤ 17, -29 ≤ <i>l</i> ≤ 28
Reflections collected	29 828
Independent reflections	16 636 [<i>R</i> (int) = 0.0333]
Completeness to theta = 67.684°	99.8%
Absorption correction	Gaussian
Max. and min. transmission	1.000 and 0.322
Refinement method	Full-matrix least-squares on <i>F</i> ²
Data/restraints/parameters	16 636/12/1111
Goodness-of-fit on <i>F</i> ²	1.045

molecule. Three downfield ¹H NMR singlets at δ 5.96, 5.29 and 4.52 ppm correspond to 3 CH groups present in the molecule which was supported by three carbon NMR resonances at δ 84, 71 and 64 ppm in its DEPT-NMR. Presence of upfield resonances at 51 and 23 ppm on the downside in DEPT NMR were assignable to four CH₂ groups of the pyrrolidine moiety which was supported by upfield resonances corresponding to 8 protons in ¹H NMR (see ESI†). The actual structure was however arrived at using single crystal X-ray analysis (Fig. 3). Pure crystals of 1 were obtained by its crystallization from chloroform and ethanol at room temperature. The molecule crystallizes in a triclinic system with *P*1 space group having 2-units in one unit cell. An X-ray ortep structure of compound 1 depicted four different conformations as shown in (Fig. 3). The crystal refinement of the molecule is shown in Table 1. The four conformers have slight variations among bond lengths and bond angles. Only a few significant differences with regard to bond lengths have been mentioned in Table 2. The characteristic feature of crystal packing view (Fig. 4) is the hydrogen bonding network in the form of N···H–N, Cl···H–C. The molecules are interconnected by means of these hydrogen bonds. Another interesting hydrogen bond is the one involving Cl···H–C (Table 3). Since crystallization of the molecule has been carried out in CHCl₃:EtOH solvent, it is likely that one of CHCl₃ molecule per unit cell has intruded inside during the process of crystallization providing additional stability to the unit cell.



Table 2 Comparative analysis of experimentally obtained bond lengths of four conformers with that obtained theoretically (notable ones have been shown, for all refer to ESI file)

Bond	Exp. bond length (Å) of Conf. A	Exp. bond length (Å) of Conf. B	Exp. bond length (Å) of Conf. C	Exp. bond length (Å) of Conf. D	Theo. bond length (Å)
N(4)–H(4)	0.80	0.88	0.86	0.96	1.0138
C(2)–C(3)	1.496	1.531	1.527	1.566	1.5385
C(3)–C(4)	1.518	1.520	1.523	1.485	1.5335
C(6)–C(7)	1.399	1.383	1.511	1.512	1.420
C(11)–C(12)	1.525	1.514	1.399	1.399	1.487
C(21)–C(22)	1.388	1.385	1.377	1.386	1.398

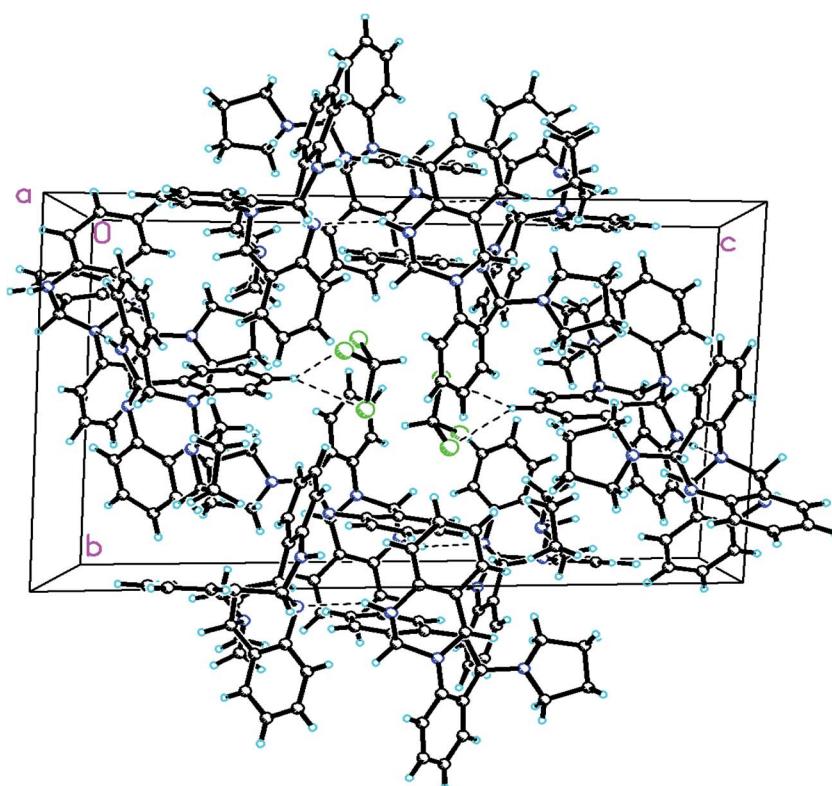


Fig. 4 A perspective view of the crystal packing in the compound 1.

DFT studies

Theoretical calculations using DFT were performed. This computational method is emerging as an important quantum, chemical tool for studying various chemical problems and is because this versatile method determines energy based on electron density instead of wave-function. The DFT calculations were carried out with a hybrid functional B3LYP (Becke's three parameter hybrid functional) at 6-31G (d,p) level of theory using Gaussian 09 package. The geometries were optimized without imposing any molecular symmetry constraints.

Optimized structures

The computational chemistry studies in this work aimed at determination of geometric parameters for compound 1 and the same were compared with that of the experimentally

obtained X-ray data. Table 2 summarizes the selected experimentally obtained (single crystal X-ray analysis) and theoretically calculated parameters (selected bond lengths) using DFT methods. A good approximation of bond lengths while doing optimization using DFT was observed to the experimentally obtained values.

Molecular electrostatic potential surface analysis

MEPS is a plot of electrostatic potential mapped on to constant potential electron density surface. These surfaces help in predicting reactivity sites towards positively and negatively charged reactants, in studies of biological recognition of one molecule by another as in drug receptor and enzyme substrate interactions as well as hydrogen-bonding interactions. MEP surfaces reveal the size shape and variation of electron density, electronegativity, partial charges and the sites of chemical reactivity with in the



molecule.^{39–43} **Fig. 5** shows MEP surface for compound **1**, calculated using DFT/B3LYP/6-31G (d,p) level of theory. The pictorial representation with rainbow colour scheme of electrostatic potential for compound **1** lies in the range of -5.422 a.u to 5.422 a.u. The darkest red regions indicate regions with high electron density (negative potential) which invite electrophilic attacks while as the darkest blue regions indicate regions with low

electron density (positive potential) which allow nucleophiles to approach easily. As can be seen in the MEP for compound **1** the region surrounding the secondary N-atom seems to the darkest blue that is one with lowest electron density, a site more vulnerable to nucleophilic attack. A plausible explanation in this regard is that the lone pair of electrons on N-atom enters in conjugation with the phenyl moiety thereby decreasing the electron density around that N-atom.

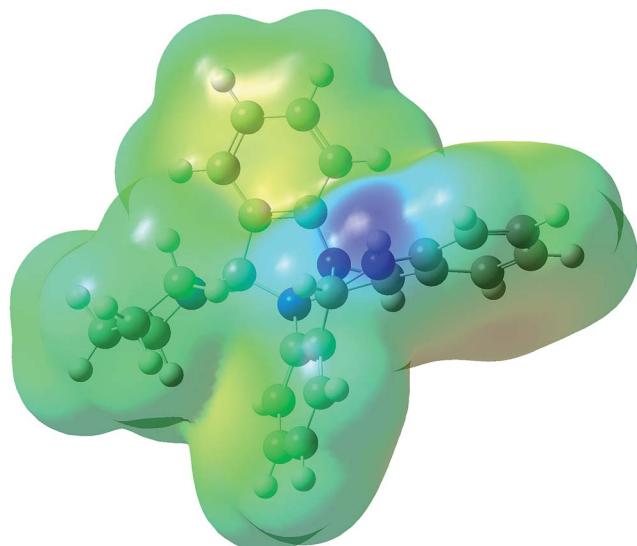


Fig. 5 MEP plot of compound **1** calculated using DFT/B3LYP/6-31G (d,p) level of theory.

Frontier orbital analysis

Frontier molecular orbitals play important role in the determination of electric and optical properties electronic transitions, kinetic stability.⁴⁴ FMO's of compound **1** were calculated using the Gaussian 09 package at the B3LYP/6-31G (d,p) level of theory. The results are shown in **Fig. 6**. As can be seen from the figures HOMO and LUMO are located or spread over the regions containing the N-atoms and delocalised over one of the benzene ring with adjacent NH moiety. In the target compound **1**, HOMO–LUMO energy gap is 0.094 eV. Using HOMO–LUMO energy gap important relative reactivity descriptors like hardness (η), chemical potential (μ), electronegativity (χ), ionization energy (I) and electron affinity (A) were calculated and are defined as follows:^{45–49}

$$\eta = \frac{(I - A)}{2}, \quad \mu = \frac{-(I + A)}{2}, \quad \chi = \frac{(I + A)}{2}$$

where A and I represent electron affinity and ionization potential of the molecule, which are in turn obtained from HOMO and

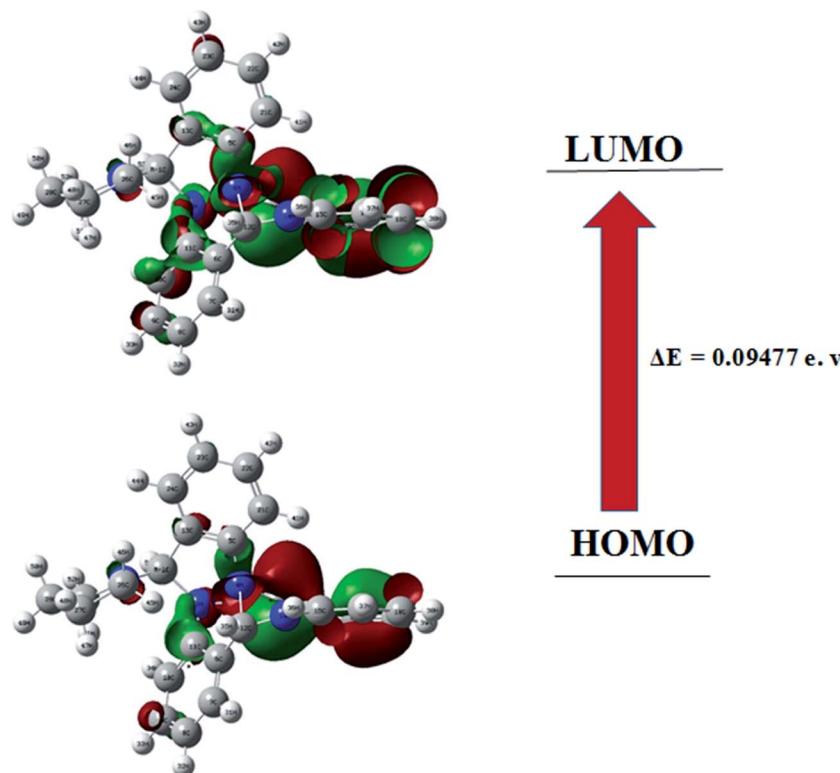


Fig. 6 HOMO–LUMO plots of compound **1**.



Table 3 Hydrogen bonds for compound 1 [Å and °]

D-H···A	d(D-H)	d(H···A)	d(D···A)	∠(DHA)
N(4B)-H(4B)···N(3A)	0.88(5)	2.34(5)	3.143(4)	153(4)
N(4D)-H(4D)···N(3C)	0.96(6)	2.25(6)	3.125(4)	151(4)
C(5D)-H(5DA)···Cl(1T)	1.00	2.96	3.718(7)	133.1

LUMO energies as $I = -E_{\text{HOMO}}$ and $A = -E_{\text{LUMO}}$ as per Janak theorem⁵⁰ and Perdew *et al.*⁵¹ As can be seen the HOMO-LUMO energy gap in compound 1 is just 0.094 eV which indicates its highly reactive nature. Another important factor is the global electrophilicity index (ω), a global reactivity index that is related to chemical hardness and chemical potential, introduced by Parr *et al.*⁴⁹ represented as $\omega = \mu^2/2\eta$ and was found to be 0.012 eV indicating a poor electrophilic nature of 1. All the above parameters have been calculated for compound 1 using B3LYP/6-31G (d,p) level of theory and are presented in Table 4 as shown:

NMR analysis

Experimentally ^1H and ^{13}C NMR of the title compound was evaluated in CDCl_3 solvent using TMS as an internal standard. Theoretically it was calculated in polarisable continuum model with CHCl_3 as solvent using GIAO method⁵² involving B3LYP/6-31G (d,p) level of theory. Both theoretical and experimental ^1H and ^{13}C NMR spectral data are presented in Table 5. Experimental ^{13}C NMR of 1 revealed the presence of 23 resonances assignable to its 25 carbons atoms because of symmetrical nature of pyrrolidine ring. However its theoretical ^{13}C NMR spectrum displayed all the 25 resonances for 25 carbon atoms. Similarly ^1H NMR revealed the presence of 24 protons out of which 12 correspond to aromatic protons of three phenyl rings, three protons as CH groups, one as NH and 8 protons assignable to the pyrrolidine moiety. In ^1H NMR three CH protons appeared at 4.52, 5.29 and 5.96 ppm and calculated using DFT at 5.4, 5.8 and 6.2 ppm respectively. Similarly eight protons of the pyrrolidine ring were observed in the range 1.25–3.32 ppm and calculated at 1.37–4.1 ppm for DFT which again revealed a fair agreement. The remaining 12 aromatic protons observed in the range 6.6–7.30 ppm were calculated at 6.6–8.2 ppm for DFT.

Table 4 Calculated energy values for compound 1 using B3LYP/6-31G (d,p) level of theory

Parameter	Compound 1
Energy (au)	-1186.014
Dipole moment (debye)	3.499
E_{HOMO} (eV)	-0.139
E_{LUMO} (eV)	-0.045
$E_{\text{HOMO-LUMO}}$ (eV)	0.094
$E_{\text{HOMO-1}}$ (eV)	-0.175
$E_{\text{LUMO+1}}$ (eV)	-0.005
$E_{(\text{HOMO-1})-(\text{LUMO+1})}$ (eV)	0.169
Hardness (η)	0.047
Chemical potential (μ)	0.092
Electronegativity (χ)	-0.092
Electrophilicity index (ω)	0.012

Table 5 Comparsion of theoretical and experimental NMR data of compound 1

Experimental (δ ppm)	^1H NMR		^{13}C NMR	
	Theoretical	Experimental (δ ppm)	Theoretical	Experimental (δ ppm)
7.30–6.70 (m, 12H)	8.10 (s, 1H)	146.08	133.86	
5.96 (s, 1H)	7.60 (s, 1H)	144.23	132.90	
5.28 (d, $J = 1.5$ Hz, 1H)	7.38 (s, 2H)	140.08	118.62	
4.82 (br s, 1H)	7.19 (s, 2H)	130.20	114.61	
4.52 (s, 1H)	7.10 (s, 2H)	129.59	113.18	
3.32 (m, 2H)	6.98 (s, 1H)	129.12	112.86	
2.85 (m, 2H)	6.78 (s, 1H)	128.73	112.63	
2.05–1.92 (m 4H)	6.72 (s, 1H)	128.48	111.98	
	6.65 (s, 1H)	128.31	110.86	
	6.59 (s, 1H)	127.88	110.58	
	6.17 (s, 1H)	124.97	109.86	
	5.83 (s, 1H)	124.74	108.72	
	5.39 (s, 1H)	124.56	108.11	
	4.10 (s, 1H)	123.93	107.89	
	3.53 (s, 1H)	123.32	106.40	
	3.06 (s, 1H)	123.09	105.42	
	2.62 (s, 1H)	119.98	104.73	
	1.89 (s, 1H)	117.44	97.81	
	1.71 (s, 1H)	84	94.84	
	1.66 (s, 1H)	71.04	88.20	
	1.38 (s, 1H)	64.12	76.45	
		51.43	45.55	
		51.43	36.62	
		23.76	19.07	
		23.76	18.70	

IR analysis

Theoretically a total of 153 normal modes of vibration were obtained for compound 1. However in the desired region between 500 and 4000 cm^{-1} only 130 such modes were observed. The calculations have been performed within the harmonic approximation for vibrations. Comparision of selected theoretical and experimental modes is presented in Table 6. A fair agreement is observed between theoretical and experimental IR data. Experimental FT-IR displayed bands in the region 3400–2800 cm^{-1} , 1700–1000 and 1000–500 cm^{-1} . Bands in the region 3400–2800 cm^{-1} were assignable to N-H stretchings, C-H stretchings of all the aromatic rings, C-H stretchings in which C-atoms are connected to N-atoms and CH_2 stretchings of the pyrrolidine moiety. The bands between 1620–1000 cm^{-1} were assignable to C=C stretching vibrations in aromatic rings, H-C=C aromatic ring bend modes, CH_2 scissorings, CH_2 twistings and N-C stretching vibrations. However bands between 900–500 mostly correspond to the torsion modes *i.e.* distortions in dihedral angles mostly of HCCC, CCCC, NCCC and CNCC type.

PASS and biological evaluation

After having successfully optimized and calculated various reactivity descriptors for compound 1, using DFT employing the well-known B3LYP/6-31G (d,p) level of theory, we next studied its bioevaluation against various human cancer cell lines. Before carrying out the actual analysis, it was envisaged to



Table 6 Comparison of theoretical and experimental IR (cm^{-1}) data of compound 1 (selected modes only)

Experimental (cm^{-1})	Theoretical (cm^{-1})	Experimental (cm^{-1})	Theoretical (cm^{-1})	Experimental (cm^{-1})	Theoretical (cm^{-1})
3401	3224	1465	1472	960	967
3062	3061	1341	1336	856	857
2980	3028	1301	1314	795	792
2830	3006	1253	1257	750	753
2800	2986	1222	1225	685	689
1620	1613	1120	1126	620	617
1560	1555	1036	1031	580	577
1500	1501	1000	987	534	535
1490	1475	982	979	512	521

predict the pharmacological properties of the target compound (1), using an available online PASS.⁵³ PASS is an important tool that evaluates the biological activity of a molecule in relation to its structure. It assesses the druglikeness and toxicities of molecules like teratogenicity, carcinogenicity embryogenecity etc. The average accuracy of prediction is about 95% according to leave-one-out-cross validation (LOOCV) estimation and the probabilities, P_a (probable activity) and P_i (probable inactivity), are values that vary from 0.00 to 1.00, and generally $P_a + P_i \neq 1$ as these probabilities are calculated independently.⁵⁴ PASS analysis results for the title compound 1 have been enlisted in Table 7. Among the range of activities predicted, the most

notable is the antineoplastic activity with P_a of 0.884. To validate the pass results, the title compound was evaluated using sulforhodamine B (SRB) assay against human-leukemia (THP-1), colon (HCT-116), prostate (PC-3) and lung (A-549) cancer cell lines. It was observed that the title compound possesses a broad spectrum anticancer effect on the cancerous cells with THP-1 cell line as its most easy and soft target. Compound 1 displayed IC_{50} of 07, 12, 10, and 17 μM against THP-1, colon (HCT-116), prostate (PC-3) and human lung (A-549) cancer cell lines respectively. Since the lowest IC_{50} is observed against human leukemia cell lines (THP-1) cell-lines, it implies that the usual target of this compound may be leukemia cell lines. However it is equally true that leukemia cell lines are more-sensitive.

Table 7 PASS prediction for the activity spectrum of compound 1 with $P_a > 0.5$

P_a	P_i	Activity name
0.884	0.003	Antineoplastic enhancer
0.851	0.009	5 Hydroxytryptamine release stimulant
0.769	0.014	Nicotinic alpha-2-beta-2-receptor antagonist
0.728	0.026	Nicotinic alpha-6-beta-3-beta-4-alpha-5-receptor antagonist
0.707	0.007	Antihypoxic
0.676	0.014	Thioredoxin inhibitor
0.690	0.080	Phobic disorders treatment
0.605	0.003	Poly(ADP-ribose) polymerase inhibitor
0.637	0.052	Fibrinolytic
0.591	0.009	Atherosclerosis treatment
0.585	0.007	Stroke treatment
0.576	0.010	Octopamine antagonist
0.617	0.065	Antineurotic
0.576	0.027	(R)-6-Hydroxynicotine oxidase inhibitor
0.567	0.021	(S)-6-Hydroxynicotine oxidase inhibitor
0.559	0.030	1,4-Lactonase inhibitor
0.563	0.034	Chloride peroxidase inhibitor
0.579	0.054	Pseudolysin inhibitor
0.589	0.068	Glycosylphosphatidylinositol phospholipase D inhibitor
0.550	0.033	Alopecia treatment
0.594	0.089	Antieczematic
0.580	0.098	Testosterone 17 beta-dehydrogenase (NADP ⁺) inhibitor
0.503	0.077	Calcium channel (voltage-sensitive) activator
0.528	0.114	Nootropic
0.516	0.065	Phthalate 4,5-dioxygenase inhibitor
0.526	0.081	Kidney function stimulant

Conclusion

An unusual quinazoline alkaloid (1) was synthesized and characterized using IR and NMR spectral techniques. Single crystal analysis depicted four conformations differing slightly in bond angles and bond lengths. Compound 1 crystallizes in the triclinic crystal system with P-1 space group having two molecules within the unit cell. The experimentally obtained parameters were compared to that obtained theoretically which depicted a good agreement. Using DFT/B3LYP/6-31G (d,p) level of theory, HOMO-LUMO energy gap, molecular electrostatic potential surface (MEPS), vibrational and NMR analysis were carried out. HOMO-LUMO energy gap was established to carry out the calculation of chemical hardness, chemical inertness, electronegativity and electrophilicity index of the molecule, which depicted the potential kinetic stability and reactivity of the molecule. Prediction of activity spectra of the target compound was carried out which revealed that compound 1 possesses notable antineoplastic activity. The pass results were validated by evaluating the molecule against colon (HCT-116), leukemia (THP-1), prostate (PC-3) and human lung (A-549) cancer cell lines respectively in an SRB assay. It was observed that compound 1 possesses remarkable potency against THP-1 cell lines with IC_{50} of 7 μM .

Experimental

General considerations

2-Nitrobenzaldehyde, iron powder, pyrrolidine etc were procured from Sigma-Aldrich (USA) and used as received. The



FT-IR spectra of the compound was recorded on Agilent FT-IR spectrometer in KBr discs (4000–400 cm^{-1}). ^1H , ^{13}C NMR spectra were recorded on a Bruker Spectrospin DPX-400 NMR spectrometer at 400.13 and 100.47 respectively using TMS as an internal standard. Mass analysis was carried out using Nexera UHPLC at 130 MPa with SIL-30 AC Nexera autosampler coupled to an LC-MS 8030 tandem mass spectrometer manufactured by Shimadzu Corporation, Kyoto, Japan. The software SADABS was used for absorption correction and SHELXTL for space group, structure determination and refinements. All non-hydrogen atoms were refined anisotropically. All the computations have been carried out at DFT/B3LYP/6-31G (d,p) level of theory using GAUSSIAN 09 software.

Synthesis of 2-aminobenzaldehyde

2-Aminobenzaldehyde was synthesized by a procedure reported by Gamble *et al.*⁵⁵ To a suspension of 2-nitrobenzaldehyde (0.200 g, 1.32 mmol) in a mixture of glacial acetic acid (2 mL), ethanol (2 mL) and water (1 mL), reduced iron powder (0.280 g, 5.00 mmol) was added. The reaction mixture was exposed to ultrasonic irradiation for 1–2 h at room temperature with continuous TLC monitoring for the completion of reaction. The mixture was filtered to remove the iron residue. The filtrate was partitioned with 2 M KOH and the basic layer was further extracted with ethyl acetate (3 \times 25 mL). The combined organic extracts were washed with brine and water and dried under reduced pressure. The crude residue was then chromatographed over silica gel column chromatography using 30% EtOAc : Hexane as eluent to give 2-aminobenzaldehyde in 80% yield.

Synthesis of 1

A 100 mL round bottomed flask was charged with 2-aminobenzaldehyde (120 mg, 1 mmol), ethanol (2.5 mL) and pyrrolidine (0.027 mL, 0.33 mmol) and stirred under reflux conditions and the reaction monitored using TLC analysis till it was complete. After the completion of reaction the solvent of the mixture was evaporated under reduced pressure. The solid residue so obtained, was subjected to silica gel column chromatography and purified using 65 : 45 ethyl acetate : hexane as eluent. Compound **1** was obtained as pure yellowish solid.

Yield: 60%, mp: 183 °C, IR (KBr cm^{-1}): 3401, 3062, 2980, 2830, 2800, 1620, 1560, 1500, 1490, 1465, 1380, 1341, 1301, 1253, 1222, 1120, 1036, 1000, 982, 960, 856, 795, 750, 685, 620, 580, 534, 512, 500. ^1H NMR (400 MHz, CDCl_3) δ 7.30–6.70 (m, 12H), 5.96 (s, 1H), 5.28 (d, J = 1.5 Hz, 1H), 4.82 (br s, 1H), 4.52 (s, 1H), 3.32 (m, 2H), 2.85 (m, 2H), 2.05–1.92 (m 4H), ^{13}C NMR (100 MHz, CDCl_3) δ 146.08, 144.23, 140.08, 130.20, 129.59, 129.12, 128.73, 128.48, 128.31, 127.88, 124.97, 124.74, 124.56, 123.93, 123.32, 123.09, 119.98, 117.44, 84.00, 71.04, 64.12, 51.43 (2C), 23.76 (2C). (ESI-MS) at m/z = 381 for $[\text{M} + \text{H}]^+$.

X-ray diffraction study

Pure rod shaped crystals were selected after recrystallization of the final product (**1**) from CHCl_3 and ethanol solvent. The diffraction data on single crystal of compound **1** was collected

on a Bruker AXS SMART Apex CCD diffractometer using Mo-K α (0.71073 Å) radiations at 298 K. The software SADABS⁵⁶ was used for absorption correction and SHELXTL for space group, structure determination and refinements.^{57,58} All non-hydrogen atoms were refined anisotropically.

Density functional theory study

All the computations are carried out using GAUSSIAN 09 software.⁵⁹ The DFT modeling method, using the hybrid B3LYP⁶⁰ functional was used to calculate theoretical parameters for compound **1** with the basis set combination 6-31 G (d,p).⁶¹ Geometry optimization was carried out until global minima were achieved. The results of the optimized structure for compound **1** are reported in Table 3 and are compared with X-ray diffraction results.

SRB cytotoxic assay

SRB cytotoxic assay was used to screen the compounds for cell cytotoxicity as per the reported procedures.^{62,63} Various human cancer cell lines, human leukemia (THP-1, at a density of 7×10^3), human lung carcinoma cell line (A-549, at a density of 8×10^3 cells per mL per 100 μL per well), human prostate cancer cell line (PC-3, at a density of 8×10^3 cells per mL per 100 μL per well) and human colon cancer cell line (HCT-116, at a density of 1×10^4 cells per mL per 100 μL per well) used in this study were purchased from European collection of cell culture (ECACC) (USA) and seeded in flat-bottomed 96-well plates.

Conflicts of interest

The authors declare no conflict of interest.

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References

- 1 T. Aniszewski, *Sci. Legumes*, 1994, **1**, 1–24.
- 2 M. Wink, A short history of alkaloids, in. *Alkaloids: Biochemistry, Ecology, and Medicinal Applications*, ed. Roberts M. F., and Wink M., Plenum Press, New York, London, 1998, pp. 11–44.
- 3 S. Bhattacharya, C. Haertel, A. Maelicke and D. Montag, *PLoS One*, 2014, **9**, e89454.
- 4 Z. Li, Y. N. Geng, J. D. Jiang and W. J. Kong, *J. Evidence-Based Complementary Altern. Med.*, 2014, **2014**, 289264.
- 5 J. Peng, X. Y. Zhang, Z. C. Tu, X. Y. Xu and S. H. Qi, *J. Nat. Prod.*, 2013, **76**(5), 983–987.
- 6 B. Yang, H. M. Tao, X. F. Zhou, X. P. Lin and Y. H. Liu, *Nat. Prod. Res.*, 2013, **27**, 433–437.
- 7 X. W. Yang, G. Y. Zhang, J. X. Ying, B. Yang, X. F. Zhou and A. Steinmetz, *Mar. Drugs*, 2013, **11**(1), 33–39.

8 X. Guo, Y. Li, C. F. Li, H. M. Luo, L. Z. Wang and J. Qian, *Gene*, 2013, **527**(1), 131–138.

9 R. H. Pouwer, S. M. Deydier, P. V. Le, B. D. Schwartz, N. C. Franken and R. A. Davis, *ACS Med. Chem. Lett.*, 2014, **5**(2), 178–182.

10 C. Agyare, G. A. Koffuor, Y. D. Boakye and K. B. Mensah, *Pharm. Biol.*, 2013, **51**(4), 418–425.

11 A. A. Boligon, T. F. Kubica, D. N. Mario, T. F. de Brum, M. Piana and R. Weiben, *Acta Physiol. Plant.*, 2013, **35**(7), 2229–2239.

12 E. V. Costa, P. E. O. da Cruz, C. C. de Lourenco, V. R. D. Moraes, P. C. D. Nogueira and M. J. Salvador, *Nat. Prod. Res.*, 2013, **27**(11), 1002–1006.

13 J. Hu, X. D. Shi, J. G. Chen, X. Mao, L. Zhu and L. Yu, *Food Chem.*, 2014, **148**, 437–444.

14 J. F. Wang, X. P. Lin, C. Qin, S. R. Liao, J. T. Wan and T. Y. Zhang, *J. Antibiot.*, 2014, **67**(8), 581–583.

15 Y. Yang, W. J. Zuo, Y. X. Zhao, W. H. Dong, W. L. Mei and H. F. Dai, *Planta Med.*, 2012, **78**(17), 1881–1884.

16 C. Iannello, J. Bastida, F. Bonvicini, F. Antognoni, G. A. Gentilomi and F. Poli, *Nat. Prod. Res.*, 2014, **28**(10), 704–710.

17 L. Zhang, Z. Hua, Y. Song and C. Feng, *Fitoterapia*, 2014, **97**, 142–147.

18 A. O. Adeoye, B. O. Oguntimein, A. M. Clark and C. D. Hufford, *J. Nat. Prod.*, 1986, **49**, 534–537.

19 M. K. Ray, C. Balachandran, V. Duraipandian, P. Agastian, S. Ignacimuthu and A. Vijayakumar, *Med. Chem. Res.*, 2013, **22**, 823–830.

20 C. Dupont, E. Couillerot, R. Gillet, C. Caron, M. Zeches-Hanrot and J. F. Riou, *Planta Med.*, 2005, **71**, 489–494.

21 S. S. Nazrullaev, I. A. Bessonova and K. S. Akhmed khodzhaeva, *Chem. Nat. Compd.*, 2001, **37**, 551–555.

22 M. Goyal and D. Sasmal, *J. Ethnopharmacol.*, 2014, **151**, 536–542.

23 N. Wang, K. J. Wicht, E. Shaban and T. A. Ngoc, *MedChemComm*, 2014, **5**(7), 927–931.

24 M. Taki, K. Niitu, Y. Omiya, M. Fukuchi, M. Aburada and M. Okada, *Planta Med.*, 2003, **69**, 800–803.

25 S. H. Lone, K. A. Bhat, S. Rehman, R. Majeed, A. Hamid and M. A. Khuroo, *Bioorg. Med. Chem. Lett.*, 2013, **3**, 4931–4934.

26 S. Naseer, S. H. Lone, J. A. Lone, M. A. Khuroo and K. A. Bhat, *J. Chromatogr. B: Biomed. Sci. Appl.*, 2015, **989**, 62–70.

27 S. H. Lone, K. A. Bhat, R. Majeed, A. Hamid and M. A. Khuroo, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 1047–1051.

28 S. H. Lone and K. A. Bhat, *Tetrahedron Lett.*, 2015, **56**, 1908–1910.

29 I. Ayoob, S. H. Lone, M. Rahman, O. A. Zargar, R. Bashir, S. Rehman, M. A. Khuroo and K. A. Bhat, *ChemistrySelect*, 2017, **2**, 10153–10156.

30 S. Rehman, K. A. Bhat, S. H. Lone and F. A. Malik, *Arabian J. Chem.*, 2015, DOI: /10.1016/j.arabjc.2015.10.009.

31 S. Rehman, B. Rah, S. H. Lone, R. Rasool, S. Farooq, D. Nayak, N. A. Chikan, S. Chakraborty, A. Behl, D. M. Mondhe, A. Goswami and K. A. Bhat, *J. Med. Chem.*, 2015, **58**, 3432–3444.

32 S. H. Lone, K. A. Bhat, F. A. Malik and M. A. Khuroo, *Planta Medica International Open*, 2016, **3**, 51–54.

33 S. H. Lone, S. Rehman and K. A. Bhat, *Drug Res.*, 2017, **67**, 111–118.

34 B. A. Dar, S. H. Lone, W. A. Shah and K. A. Bhat, *Drug Res.*, 2016, **66**, 427–431.

35 I. Ayoob, Y. M. Hazari, S. H. Lone, S. Rehman, M. A. Khuroo, K. M. Fazili and K. A. Bhat, *ChemistrySelect*, 2017, **2**, 2965–2968.

36 S. H. Lone, K. A. Bhat, S. Naseer, R. A. Rather, M. A. Khuroo and S. A. Tasaduq, *J. Chromatogr. B: Biomed. Sci. Appl.*, 2013, **940**, 135–141.

37 S. H. Lone and K. A. Bhat, *Steroids*, 2015, **96**, 164–168.

38 M. T. Richers, I. Deb, A. Y. Platonova, C. Zhang and D. Seidel, *Synthesis*, 2013, **45**(13), 1430–1748.

39 K. Abhishek, K. S. Ambrish, G. Shashi, M. Neeraj, M. Avijit and B. Goutam, *J. Mol. Des.*, 2015, **1096**, 94–101.

40 M. N. Arshad, A. Bibi, T. Mahmood, A. M. Asiri and K. Ayub, *Molecules*, 2015, **20**, 5851–5874.

41 H. Zhou and J. Skolnick, *Mol. Pharm.*, 2012, **9**, 1775–1784.

42 E. Scrocco and J. Tomasi, *Adv. Quantum Chem.*, 1979, **11**, 115–193.

43 N. Okulik and A. H. Jubert, *J. Mol. Des.*, 2005, **4**, 17–30.

44 K. Fukui, T. Yonezawa and H. J. Shingu, *J. Chem. Phys.*, 1952, **20**, 722–725.

45 R. A. Costa, P. O. Pitt, M. L. B. Pinheiro, K. M. T. Oliveira, K. S. Salome, A. Barison and E. V. Costa, *Spectrochim. Acta, Part A*, 2017, **174**, 94–104.

46 R. A. Costa, K. M. T. Oliveira, E. V. Costa and M. L. B. Pinheiro, *J. Mol. Struct.*, 2017, **1145**, 254–267.

47 N. Özdemir, S. Dayan, O. Dayan, M. Dinçer and N. Kalaycioglu, *J. Atom. Mol. Phys.*, 2013, **11**(6), 707–723.

48 R. Parr, *Functional Theory of Atoms and Molecules*, Oxford University Press, New York, 1989.

49 R. Parr, L. Szentpaly and S. Liu, *J. Am. Chem. Soc.*, 1999, **121**, 1922–1924.

50 J. F. Janak, *Phys. Rev. B*, 1978, **18**, 7165.

51 J. P. Perdew, R. G. Parr, M. Levy and J. L. Balduz Jr, *Phys. Rev. Lett.*, 1982, **49**, 1691–1694.

52 R. Ditchfield, *J. Chem. Phys.*, 1972, **56**, 5688–5691.

53 A. Lagunin, A. Stepanchikova, D. Filimonov and V. Poroikov, *Bioinformatics*, 2000, **16**, 747–748.

54 R. Pramely and T. L. S. Raj, *J. Biochem. Technol.*, 2012, **3**, 375–379.

55 A. B. Gamble, J. Garner, C. P. Gordon, S. M. J. O’Conner and P. A. Keller, *Synth. Commun.*, 2007, **37**, 2777–2786.

56 Brucker SAINT Brucker AXS Inc., Madison Wisconsin, USA, 2002.

57 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 2008, **64**, 112–122.

58 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Adv.*, 2015, **71**, 3–8.

59 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota,



R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian 09, B. 01*, Gaussian, Inc., Wallingford CT, 2010.

60 (a) A. D. Becke, *J. Chem. Phys.*, 1993, **98**, 1372–1378; (b) P. J. Stephens, F. J. Devlin, C. F. Chabalowski and M. J. Frisch, *J. Phys. Chem. B*, 1994, **98**, 11623.

61 (a) A. D. McLean and G. S. Chandler, *J. Chem. Phys.*, 1980, **72**, 5639; (b) K. Raghavachari, J. S. Binkley, R. Seeger and J. A. Pople, *J. Chem. Phys.*, 1980, **72**, 650; (c) J. P. Blaudeau, M. P. McGrath, L. A. Curtiss and L. Radom, *J. Chem. Phys.*, 1997, **107**, 5016; (d) A. J. H. Wachters, *J. Chem. Phys.*, 1970, **52**, 1033; (e) P. J. Hay, *J. Chem. Phys.*, 1977, **66**, 4377; (f) K. Raghavachari and G. W. Trucks, *J. Chem. Phys.*, 1989, **91**, 1062; (g) R. C. Binning Jr and L. A. Curtiss, *J. Comput. Chem.*, 1990, **11**, 1206; (h) M. P. McGrath and L. Radom, *J. Chem. Phys.*, 1991, **94**, 511; (i) L. A. Curtiss, M. P. McGrath, J. P. Blaudeau, N. E. Davis, R. C. Binning Jr and L. Radom, *J. Chem. Phys.*, 1995, **103**, 6104.

62 R. Majeed, M. V. Reddy, P. K. Chinthakindi, P. L. Sangwan, A. Hamid, G. Chashoo, A. K. Saxena and S. Koul, *Eur. J. Med. Chem.*, 2012, **49**, 55.

63 G. Chashoo, S. K. Singh, D. M. Mondhe, P. R. Sharma, S. S. Andotra, B. A. Shah, S. C. Taneja and A. K. Saxena, *Eur. J. Pharmacol.*, 2011, **668**, 390.

