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Fullerene derivatives act as inhibitors of leukocyte common antigen based on molecular dynamics simulations†

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Fullerene-based molecules are being studied as potential inhibitors of protein tyrosine phosphatases due to their unique properties and low toxicity. However, the underlying molecular mechanism remains elusive. In this study, molecular dynamics (MD) simulations in conjunction with molecular docking calculations were utilized to investigate the binding effects of C_{60} , $C_{60}(NH_2)_{30}$, and $C_{60}(OH)_{30}$ on the enzymatic activity of CD45 (a receptor-like protein tyrosine phosphatase). Our results show that all the investigated molecules can be docked into the region between D1 and D2 domains of CD45, and stabilize the protein structure. The average number of residues that directly interact with the $C_{60}(NH_2)_{30}$ is two more than that of $C_{60}(OH)_{30}$. F819 and F820 (located in the loop connects $\alpha 3$ and $\beta 12$), resulting in different effects of $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ on protein activity. Detailed MD simulation analyses show that transformation of the interaction network caused by $C_{60}(NH_2)_{30}$ is completely different from that of the control simulation due to the misfolding of $\alpha 3$. Furthermore, the movement of D1 active pocket and KNRY motif are most severely impaired by docking with $C_{60}(NH_2)_{30}$. Our simulation results illustrate that fullerene derivatives modified with amino groups exhibit conspicuous tumor inhibition to protein tyrosine phosphatases, and can act as effective inhibitors. Our results give insight into the inhibitory effects of fullerene-based molecules on protein tyrosine phosphatases and providing a theoretical basis for the design of effective inhibitors.

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Introduction

The fast developing field of nanotechnology has led to applications in different Frontier technologies, such as medicine, catalysts, chemicals and coatings. However, the effects of nanomaterials on the structure and activity of proteins remain mostly unclear.^{1,2} In order to facilitate their biological applications, such as gene delivery,³ cellular imaging,⁴ tumor therapy,⁵ and biological experimental technology,⁶ it is essential to understand the interactions between nanomaterials and proteins.

Carbon-based nanomaterials, one of the most prevalent types of nanomaterials, such as fullerene, graphene and carbon nanotubes, have attracted considerable attention because of their excellent chemical and physical properties.^{7–10} Fullerene, with the 7.1 Å wide cage, contains 20 hexagons and 12 pentagons, and all the double bonds within its geodesic structure are conjugated.¹¹ The presence of both five- and six-membered rings might introduce more specific reactive sites for covalent derivatization,^{12,13} and an anchor point for non-covalent interactions, respectively. Several investigators have developed fullerene-based inhibitors for the therapy of HIV-1P for AIDS.^{14–16} Additionally, fullerene-based molecules have also been found to bind and inhibit the activity of glutathione-S-transferase (GST),^{17,18} where subsequent docking simulations revealed that C_{60} bound at the interface of GST dimer. Blocking this binding site can suppress the function of GST, thus lead to cytotoxicity to cells. Park *et al.* have used MD simulations to show that fullerenes with an average diameter of 0.7–0.9 nm could bind onto the entrance of a potassium channel, leading to the blocking of K^+ ions entering into the protein channel.¹⁹ Further theoretical analyses (based on molecular docking and MD simulations) suggest that the specific binding of fullerenes mainly depend on the structure of different K^+ channels,²⁰ where fullerenes trigger rearrangement of proteins and inhibit the enzyme activity by allosteric effect. Recently, NMR chemical

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† Electronic supplementary information (ESI) available: Fig. S1: the energy of $C_{60}(NH_2)_{30}/C_{60}(OH)_{30}$ in a water box during 20 ns simulation, Fig. S2: the grid of interactions (gray region) between $C_{60}(NH_2)_{30}$, $C_{60}(OH)_{30}$ and CD45, Fig. S3 shows docking poses of CD45 + C_{60} , CD45 + $C_{60}(NH_2)_{30}$, and CD45 + $C_{60}(OH)_{30}$, Fig. S4 shows comparison of the segment (C764 to D766, P800 to I824) of CD45 + C_{60} , CD45 + $C_{60}(NH_2)_{30}$, and CD45 + $C_{60}(OH)_{30}$, Fig. S5 shows the vdW energy between every residue of binding pocket and C_{60} , $C_{60}(NH_2)_{30}$, $C_{60}(OH)_{30}$, Table S1 shows the calculated pK_a values of all histidines, Table S2 shows the probability of residues that directly interact with $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$, Table S3 shows the full name and the abbreviation of amino acids. See DOI: 10.1039/c7ra13543b



shift perturbation analysis identified the binding position of C_{60} in solution, which is localized at the catalytic site of the protein and with high specificity.²¹

In the aspect of physical properties, the pristine fullerene is highly hydrophobic and can exhibit strong hydrophobic interactions to hydrophobic binding pocket of various protein targets, which may make C_{60} act as effective inhibitor for these proteins. Surface modification of fullerene increases their poor solubility and low biocompatibility, leading to new functional properties for a large variety of applications ranging from imaging,²² drug delivery²³ and bio-sensing,²⁴ cancer and gene therapy,^{25,26} and tissue engineering,²⁷ *etc.* More studies have been reported that many fullerene-based molecules with different surfaces and architectures could be used as actives or (potential actives) with low toxicity to interfere or alter the native functions of proteins,^{28,29} where interactions between fullerene-based actives and proteins have been identified both experimentally^{30–33} and computationally.^{34–36}

Comprehensive analyses of interactions between a protein and fullerene-based nanomaterial molecules by simple models can help understanding its potential toxicity, a topic still under debate.^{37,38} MD simulations can provide accurate information with respect to interactions between fullerene-based molecules to proteins, and can explain the mechanism of effects of surface chemistry on inhibition efficiency as well.

Recently, some investigators have shown that fullerene-based molecules exhibit inhibitory effects on protein tyrosine phosphatases.³⁹ Leukocyte common antigen cd (CD45) is a receptor-like protein tyrosine phosphatase (RPTP) and has been found in all nucleated hematopoietic cells.^{40,41} T and B cells express large amounts of CD45 on their surfaces, indicating that CD45 plays an essential role in antigen-induced activation of T and B cells.⁴¹ CD45 controls immune response by dephosphorylating a number of signal molecules.^{40,42} All CD45 isoforms share a same cytoplasmic region (707 amino acids), which includes a membrane proximal PTP D1 domain and a membrane distal PTP D2 domain. Experimental data that corroborate this model have been reported.^{43,44} There are extensively complementary surfaces and interactions between D1 and D2 domains, and thus maintain relative orientation and stability of CD45. The energy required to destroy the interactions between the two monomers is ~ 16 kcal mol^{−1}. Because of these multiple factors, the occurrence of slightly structural changes of D2 can affect the conformational stability of D1 through inter-molecular and intra-molecular interactions. In other words, the D1 and D2 domains establish extensive association, so that one requires another to maintain the reasonable structure of CD45.

Due to the encouraging effects of fullerene-based molecules to CD45, here, we studied the structural dynamics of CD45 complexed with C_{60} , $C_{60}(OH)_{30}$, and $C_{60}(NH_2)_{30}$ by performing molecular docking and MD simulations. The effects of fullerene-based molecules on the enzymatic activity of CD45 were also investigated. Our simulation results show that all the three fullerene-based compounds (C_{60} , $C_{60}(OH)_{30}$, and $C_{60}(NH_2)_{30}$) can be docked into the region between D1 and D2 domains of CD45, leading to the deformation of the originally existed stable

interactions between D1 and D2 domains. D1 and D2 domains establish compact connection to the extent, where even subtle changes will destroy the maintenance of their proper structures. Thus, the active sites of D1 and D2 domains can be deformed by binding with fullerene-based molecules, in which deformation caused by binding with $C_{60}(NH_2)_{30}$ to CD45 is the most serious among the three fullerene-based molecules. To our knowledge, this is the first report to investigate how fullerene-based molecules modified with different functional groups interfere or alter the functions of proteins.

Materials and methods

Systems preparation

The native structure of protein tyrosine phosphatase (PDB: 1YGR⁴⁵) was obtained from Protein Data Bank (<http://www.pdb.org/pdb>). The structure of protein was prepared by Discover Studio version 2.5,⁴⁶ which includes residues repair and geometry optimization. We also used Discover Studio 2.5 to predict protein ionization. The calculated pK_a of all histidines are shown in Table S1.† Since the pK_a of each histidine is lower than the isoelectric point (7.59), all histidines were treated with unprotonated model. N- and C-terminals were methylamidated and acetylated, respectively.

In the current work, we simulated the structures of CD45 docked with pristine fullerene and two derivatives modified with hydroxyl and amino groups, respectively. The pristine fullerene was built by Material Studio 5.5.⁴⁷ Chemical groups of $-OH$ and $-NH_2$ were evenly attached to carbon atoms of the pristine fullerene, where the ratio between hydroxyl groups (or amino groups) and carbon atoms was set to 1 : 2, namely, $C_{60}(OH)_{30}$ and $C_{60}(NH_2)_{30}$.

Then geometry optimizations were performed for the three fullerene-based materials by the Forcite module⁴⁷ with Dreiding force field⁴⁸ and Gasteiger charges in Material Studio 5.5 (the maximum iterations were set to 50 000 and the convergence was set to 5.0×10^{-6}). The ultra fine quality was used for the energy calculation. The final structures were used as the initial structures for the following docking simulation.

Molecular docking

Recently, numerous docking protocols have been used for generating initial coordinates for complexes containing proteins and nanomaterials. Here, PatchDock⁴⁹ was used for docking of the three fullerene-based molecules (C_{60} , $C_{60}(OH)_{30}$ and $C_{60}(NH_2)_{30}$) to CD45, which works by maximizing the complementarity of the surface shapes of the two molecules, while minimizing the number of steric clashes. To accurately characterize the binding modes of the three fullerene-based molecules to CD45, FireDock⁵⁰ was further used to re-score the three fullerene-containing protein complexes (CD45 + C_{60} , CD45 + $C_{60}(OH)_{30}$, CD45 + $C_{60}(NH_2)_{30}$). For each complex, two thousand docking runs were performed and ranked by the docking score. The top 50 docking poses of C_{60} , $C_{60}(OH)_{30}$, $C_{60}(NH_2)_{30}$ between CD45 are shown in Fig. S3.† Binding mode



with the highest docking score of the three complexes was saved as the initial structures for the following MD simulations.

Molecular dynamics (MD) simulation

Simulations setup. All the complexes of CD45 + C₆₀, CD45 + C₆₀(OH)₃₀, CD45 + C₆₀(NH₂)₃₀ were embedded into a rectangular TIP3P⁵¹ water box with water molecules within 2.5 Å of the protein/nanomaterials eliminated. Chloride counter ions were used to neutralize the systems. For comparison, we also built a control simulation, namely, apo-state CD45 (CD45 without docking with any fullerene-based molecules). The final box sizes of the four systems are 80 × 80 × 105 Å³/85 × 85 × 106 Å³/85 × 80 × 105 Å³/85 × 80 × 105 Å³, for CD45/CD45 + C₆₀/CD45 + C₆₀(OH)₃₀/CD45 + C₆₀(NH₂)₃₀, respectively.

Particle Mesh Ewald (PME)⁵² method was applied to calculate electrostatics interactions, with a cutoff of 12 Å for both long-range electrostatic and van der Waal (vdW) interactions. Hydrogen atoms were maintained by SHAKE algorithm, and a time step of 1 fs was applied for all the MD runs. Langevin dynamics⁵³ and Nosé-Hoover Langevin piston⁵⁴ were adopted to control the system temperature (at 310 K) and pressure (at 1 bar), respectively.

The MD simulations were carried out with NAMD software package,⁵⁵ where the proteins (CD45) were simulated with CHARMM27 force field⁵⁶ and the fullerene-based molecules were performed with Dreiding force field. Visualization and data collection were done by VMD.⁵⁷

Systems minimization, equilibration, and production. In order to fully equilibrate these systems, 10 000 steps of energy minimization were performed. Then, a three steps protocol was used for the equilibration of the systems: (1) 500 ps-equilibration in NPT ensemble in the absence of positional restraints of water molecules at 310 K; (2) a further 500 ps-equilibration by only restricting the heavy atoms of C₆₀/C₆₀(NH₂)₃₀/C₆₀(OH)₃₀; (3) a 500 ps-equilibration without any positional constraints. Finally, 100 ns unrestrained full atomistic MD simulations were carried out for the four equilibrated systems (CD45, CD45 + C₆₀, CD45 + C₆₀(NH₂)₃₀, and CD45 + C₆₀(OH)₃₀).

Results and discussion

To investigate the binding effects of fullerene-based molecules to tyrosine phosphatase CD45, molecular docking and 100 ns unrestrained MD simulations were performed for the four systems: apo-state CD45, CD45 + C₆₀, CD45 + C₆₀(NH₂)₃₀, and CD45 + C₆₀(OH)₃₀.

The rationality of C₆₀(NH₂)₃₀ and C₆₀(OH)₃₀

According to experimental data, the highest water solubility and biocompatibility are obtained when the number of hydroxyl groups is 20–30.^{58,59} The reason we choose the ratio (1 : 2) is the following: (i) the results obtained in many laboratories show that polyhydroxylated fullerenes, C₆₀(OH)_{*n*} (*n* = 20–30) water soluble and exert mainly antioxidant activity in biological systems.^{60,61} In addition, the fluorescence detection of water

soluble fullerene derivatives under different pH values shows that when pH = 6.5–7.5, it is a neutral state.² Thus, the ionization of C₆₀(OH)₃₀ and C₆₀(NH₂)₃₀ do not occur at the pH of our work (~7.4).

We carried out a 20 ns MD simulation of C₆₀(NH₂)₃₀/C₆₀(OH)₃₀ in a water box, and the energy was shown in the Fig. S1.† It indicates that the model is stable. We have drawn a grid of interactions between C₆₀(NH₂)₃₀/C₆₀(OH)₃₀ and protein. It can be seen from the Fig. S2† that only three amino groups (two hydroxyl groups) are not involved in the interaction with proteins, which suggests that reducing the concentration of functional groups can weaken the interaction and make it ineffective. Increased concentration, fullerene derivatives are unstable at pH = 7.4.² In addition, combined with the data in the Table S2 and Fig. S5,† the interaction energy between the –NH₂ and CD45 significantly higher than that of –OH. Thus, if a part of the –NH₂ is replaced by –OH, the coexistence of the two functional groups can also weaken the interaction energy with the protein, thus reducing its inhibitory effect.

Binding modes of fullerene-based molecules to CD45

We used PatchDock⁴⁹ to generate the initial coordinates of the complexes of CD45 and fullerene-based molecules. The top 50 docking poses of CD45 + C₆₀, CD45 + C₆₀(NH₂)₃₀, and CD45 + C₆₀(OH)₃₀ are shown in Fig. S3.† All the fullerene-based molecules (C₆₀, C₆₀(NH₂)₃₀ and C₆₀(OH)₃₀) can be docked into the region between the membrane proximal PTP D1 domain and the membrane distal PTP D2 domain (herein after referred as D1 and D2 domain, respectively) of CD45. Interestingly, for these three docked complexes, almost ninety percent of the binding poses are concentrated in the same region, which also includes that with highest score. Thus, we identified the binding pose with the highest docking score as the binding mode of the CD45-fullerenes complexes. As a result, the top 1 docking pose was saved as the initial structure for the 100 ns MD simulations for each complex.

The equilibrium of the investigated systems can be described by root mean square deviations (RMSD), calculated as following,

$$\text{RMSD}(t) = \sqrt{\frac{\sum_{i=1}^N |\vec{r}_i(t) - \langle \vec{r}_i \rangle|^2}{N}} \quad (1)$$

where $\vec{r}_i(t)$ represents the position of atom *i* at time *t*, $\langle \vec{r}_i \rangle$ represents the average position of atom *i* at all simulation times.

Fig. 1 shows the time evolutions of the root mean square deviations (RMSDs) of C₆₀[−], C₆₀(NH₂)₃₀[−], C₆₀(OH)₃₀-bound CD45 during 100 ns MD simulations. The RMSD values of C₆₀, C₆₀(NH₂)₃₀, and C₆₀(OH)₃₀ are 0.15 Å, 0.91 Å and 0.75 Å, respectively, indicating that the selected docking pose of C₆₀ in CD45 is the most stable one. Although the fluctuation of C₆₀(NH₂)₃₀ is the largest among the three molecules, it does not affect the stability of the docking structure with the average RMSD value lower than 1 Å. These results indicate that the three bound-state complexes are stable during 100 ns MD



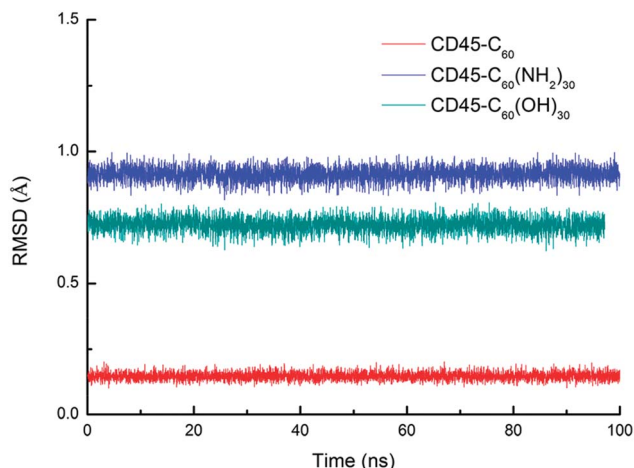


Fig. 1 Time evolutions of RMSDs of C_{60} (red), $C_{60}(NH_2)_{30}$ (blue), $C_{60}(OH)_{30}$ (green) in bound-state complexes. The low RMSDs (<1 Å) indicate that the three docked complexes are stable during 100 ns MD simulations.

simulations. Furthermore, the convergences of the three bound-state and the control (apo-state CD45) simulations are verified by the RMSD changes of $C\alpha$ atoms of CD45. As shown in Fig. 2, the average RMSD values of $C\alpha$ atoms of CD45 docked with C_{60} , $C_{60}(NH_2)_{30}$, and $C_{60}(OH)_{30}$ are 2.25 Å, 2.65 Å, 2.48 Å, respectively. In addition, these three bound-state complexes are equilibrated within ~ 20 ns. However, the control simulation is equilibrated after ~ 60 ns with the value of RMSD stabilized at around 3.25 Å. The lower RMSDs and shorter equilibrating simulation time of the bound-state complexes (compared with the control group) mean that the docking of fullerene-based molecules makes CD45 more stable.

The final structures obtained from 100 ns MD simulations are shown in Fig. 3. Owing to the different surface properties of fullerene-based molecules, the compositions of their binding

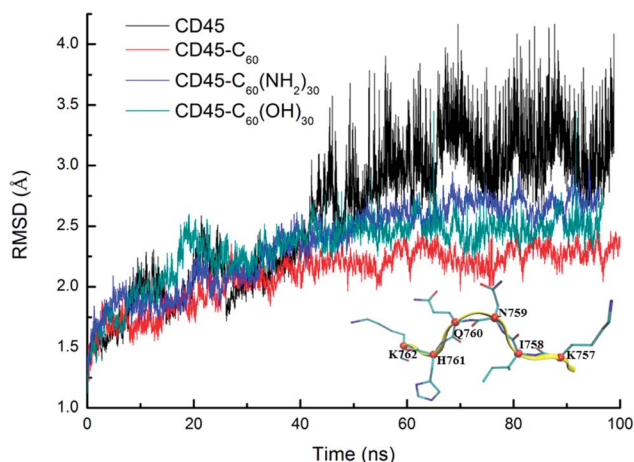


Fig. 2 Time evolutions of RMSDs of $C\alpha$ atoms of CD45 docked with C_{60} (red), $C_{60}(NH_2)_{30}$ (blue), and $C_{60}(OH)_{30}$ (green) during 100 ns MD simulations. To give a comparison, a control group of apo-state CD45 is also shown in black line. We randomly selected a segment of CD45 (K757–K762, $\beta 9$) to indicate the location of $C\alpha$ atoms of every residues, which is colored by red ball.

pockets are different in a large extent. We consider that the residues within 6 Å of fullerene-based molecules can directly interact with them. The average number of residues that interact directly with C_{60} is ~ 20 , with hydrophobic amino acids predominantly, such as P765, L805, Y1001, V1175 and A1178, *etc.* Amphipathic and hydrophilic residues, such as K762, K771, R812, E894, E1167, E1168 and so on, play main roles in the formation of hydrophilic interactions with $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$. Nevertheless, the average number of residues that interact with $C_{60}(NH_2)_{30}$ is ~ 22 , slightly more than that of $C_{60}(OH)_{30}$ (~ 20). Table S4† list the probability of these residues interacts with these two fullerene derivatives during the last 80 ns simulations. From this table, the compositions of the binding pockets of $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ are almost identical except for F819 and F820, which are highlighted in Fig. 3b and c. $C_{60}(NH_2)_{30}$ can interact strongly with phenylalanine because nitrogen atoms are more electrophilic than oxygen atoms. It is noteworthy that, although F816 and F819/F820 are the same types of amino acids, there is a probability of 100% for both $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$. The main reason is the difference in the initial position of F816 and F819/F820. As shown in Fig. S4,† F819/F820 is outwardly offset in crystal structure without docking any molecule. When docked with $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$, F819/F820 formed stronger interaction with the former, resulting in its proximity to $C_{60}(NH_2)_{30}$ as a part of its binding pocket. However, in the crystal structure, F816 is located near the center of the binding pocket. The initial distances between F816 and $C_{60}(NH_2)_{30}/C_{60}(OH)_{30}$ are about 5 Å, resulting in a strong hydrophobic interaction, so it is difficult to escape during the simulation process. F819 and F820 are located in the loop (from residue N814 to I824) between $\alpha 3$ helix and $\beta 12$ strand, which connects the active pocket (from residue V826 to G836) of CD45 D1 domain. The flexibility of this loop is severely restricted by the strong interactions formed by F819/F820 and $C_{60}(NH_2)_{30}$, which further affects the movement of the active pocket (detailed analysis will be discussed in the section “Conformational changes of the active site”).

In addition, the interaction probability of C764 and S817 that directly interact with $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ are also different. For C764, as a sulfur-containing amino acid, the sulfhydryl groups on it are more likely to interact with amino groups. For S817, as shown in Fig. S4,† due to the intense interaction between F819/F820 and $C_{60}(NH_2)_{30}$, this loop undergoes a conformational change that results in S817 being far away from $C_{60}(NH_2)_{30}$, reducing the probability of its interaction with $C_{60}(NH_2)_{30}$. The differences between the binding models of $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ cause their different effects on CD45.

Interactions between fullerene-based molecules and CD45

For detailed analyses of the interactions between fullerene-based molecules and CD45, we firstly counted the probability of the number of H-bonds formed between C_{60} , $C_{60}(NH_2)_{30}$, $C_{60}(OH)_{30}$ and CD45. Here, the distance between a hydrogen donor and an acceptor no larger than 3.5 Å and the supplementary angle formed by the donor, hydrogen, and acceptor no



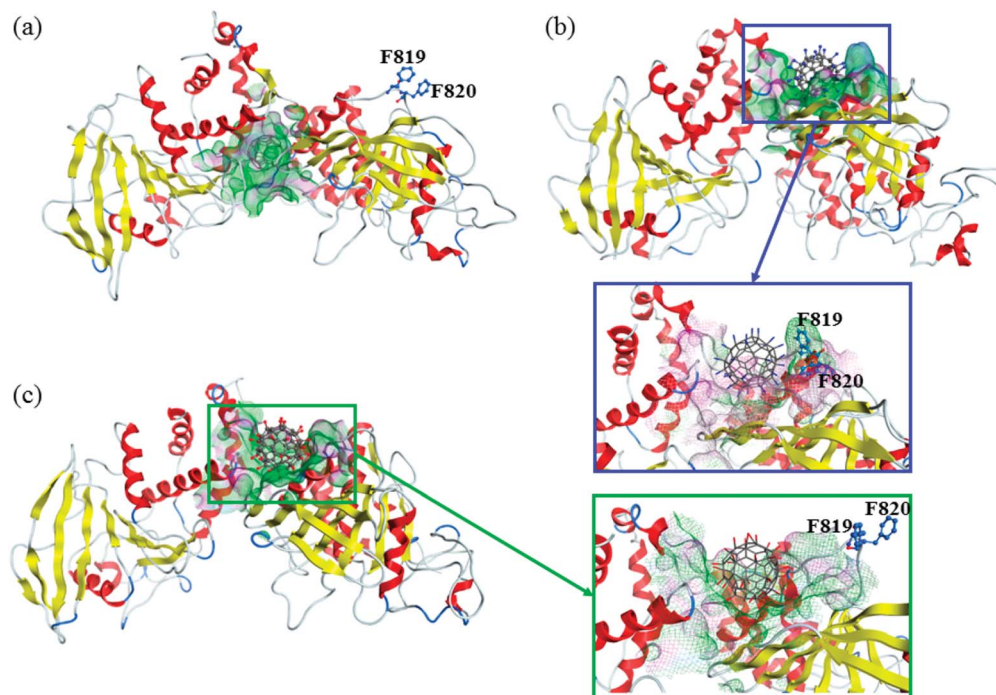


Fig. 3 The resulted bound structures of (a) CD45 + C_{60} , (b) CD45 + $C_{60}(NH_2)_{30}$, and (c) CD45 + $C_{60}(OH)_{30}$. Water molecules are excluded for clarity. The surfaces of their binding pockets are colored by H-bonding regions (purple), hydrophobic regions (green), and polar regions (blue). Residues F819 and F820 are highlighted.

larger than 35° are used as the criterion for identifying a H-bond. As shown in Fig. 4, due to the highly hydrophobic property of C_{60} , the probability of forming H-bonds (including forming 1, 2 and 3 H-bonds) is only 12% during the 100 ns MD simulation. Nevertheless, the probability of forming H-bonds between $C_{60}(NH_2)_{30}$, $C_{60}(OH)_{30}$ and CD45 are 86% and 97%, respectively. Whereas, the probability to form three or more H-bonds between $C_{60}(NH_2)_{30}$ and CD45 is only 35%, which is significantly lower than that between $C_{60}(OH)_{30}$ and CD45

(65%), moreover, the average numbers of H-bonds formed between $C_{60}(NH_2)_{30}$, $C_{60}(OH)_{30}$ and CD45 are 2 and 3, respectively. In summary, the ability to form H-bonds between $C_{60}(NH_2)_{30}$ and CD45 is slightly weaker than that of $C_{60}(OH)_{30}$.

Furthermore, we also calculated the interaction energies between C_{60} , $C_{60}(NH_2)_{30}$, $C_{60}(OH)_{30}$ and CD45. These results are listed in Table 1. Owing to the pure hydrophobicity of C_{60} , the electrostatic energy between it and CD45 is 0 kcal mol $^{-1}$, and the vdW energy is -71.95 ± 3.42 kcal mol $^{-1}$, which is the strongest among the three systems. We break down the vdW energy into every residue, the result is shown in Fig. S5.† Although the average number of residues that interact with C_{60} is two less than with $C_{60}(NH_2)_{30}$, the vdW energy between each residue and C_{60} is higher than that of $C_{60}(NH_2)_{30}$. So the total vdW energy of C_{60} and CD45 is more stronger. For the other two fullerene-based molecules, the electrostatic energy between CD45 and $C_{60}(NH_2)_{30}$ is -12.08 ± 0.23 kcal mol $^{-1}$, slightly less than that of $C_{60}(OH)_{30}$ complexed CD45 (-15.46 ± 0.71 kcal mol $^{-1}$), which is consistent with the hydrogen

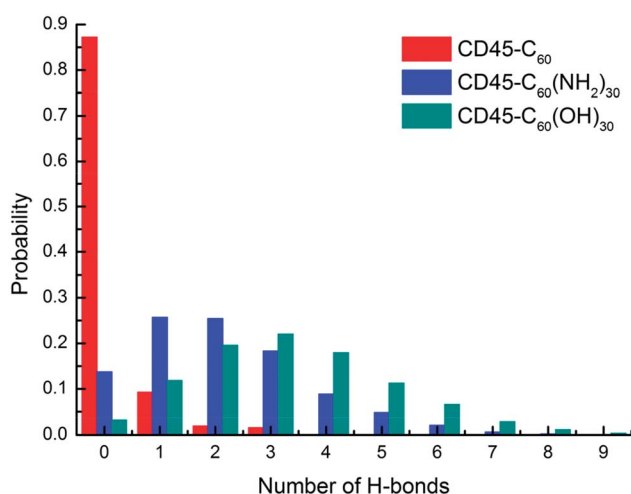


Fig. 4 Probability of the number of H-bonds formed between C_{60} (red), $C_{60}(NH_2)_{30}$ (blue), $C_{60}(OH)_{30}$ (green) and CD45 during 100 ns MD simulations.

Table 1 Interaction energies between CD45 and fullerene-based molecules, including vdW and electrostatic energies (kcal mol $^{-1}$)

Fullerene-based molecules	vdW	Elec.	Total
C_{60}	-71.95 ± 3.42	0	-71.95 ± 3.42
$C_{60}(NH_2)_{30}$	-51.38 ± 3.42	-12.08 ± 0.23	-63.46 ± 3.14
$C_{60}(OH)_{30}$	-42.54 ± 2.70	-15.46 ± 0.71	-58.00 ± 1.93



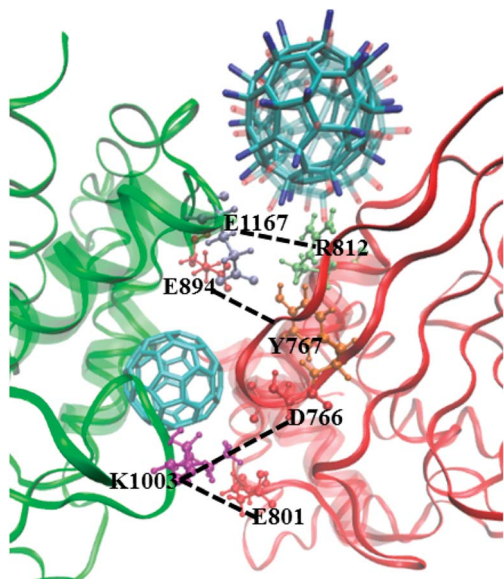


Fig. 5 Complementary surfaces between D1 (red) and D2 (green) segments of CD45. The $\alpha 3$ and $\alpha 6$ helices of D1 and the $\alpha 4$ and $\alpha 5$ helices of D2 are highlighted by transparent NewCartoon representation. The critical residues involved in intra-molecular interactions between D1 and D2 are represented by Licorice and CPK models. C_{60} , $C_{60}(NH_2)_{30}$, and $C_{60}(OH)_{30}$ complexed CD45 are shown with Licorice representation.

bonding results obtained earlier. However, the vdW energies between CD45 and $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ are -51.38 ± 3.42 and -42.54 ± 2.70 kcal mol $^{-1}$, respectively, probably due to the relatively greater average number of residues that interact directly with $C_{60}(NH_2)_{30}$. Totally, the interaction energy between CD45 and C_{60} is the strongest among the three systems, while weaker interactions is shown of system of $C_{60}(NH_2)_{30}$ -CD45.

Deformation of the conformations between D1 and D2

As reported in previous work,⁴⁰ the cytoplasmic region of CD45 contains two homologous protein tyrosine phosphatase

domains, the membrane proximal PTP D1 domain and the membrane distal PTP D2 domain. There are extensively complementary surfaces between D1 and D2 domains, including the residues of the $\alpha 3$ and $\alpha 6$ helices of D1 and the $\alpha 4$ and $\alpha 5$ helices of D2. D1 and D2 domains pack tightly against each other by an extensive network of interactions, consisting of H-bonds, van der Waals interactions, and salt bridges. Critical residues such as D766, Y767, E801, R812 (in D1 domain) and E894, K1003, E1167 (in D2 domain) play important roles in maintaining a stable inter-domain orientation between D1 and D2 domains. Among these residues, R812 and E1167 are highly conserved in most RPTP. Schematic of the complementary surfaces between D1 and D2 domains and these critical residues are shown in Fig. 5.

Our docking results show that both fullerene-based molecules can be docked into the region between D1 and D2 domains. Therefore, binding of fullerene-based molecules to CD45 can seriously impair the network of the original interactions. We elaborated the interactions by analyzing distances between critical residues in D1 and D2 domains and the results are shown in Fig. 6. Generally speaking, two H-bond formable residues can form strong H-bond or salt bridge interactions when their distance is less than ~ 3.5 Å. The strength of the interaction decreases with the increase of distance, and completely disappears when the distance reaches up to ~ 6 Å. Fig. 6a shows the time evolutions of the distance between R812 in D1 and E1167 in D2. We can see that the distance of the control simulation (black line) keep stable around 3.5 Å, then it is destroyed and cannot be recovered until the ending of simulation. The distance of the system of CD45 + C_{60} is averaged at ~ 6 Å after ~ 30 ns simulation, which is similar to that of the control simulation, indicating that the interaction formed between R812 and E1167 is not significantly affected by the binding of C_{60} . However, the distances keep stable at around 3 Å throughout the simulations for both the systems of CD45 + $C_{60}(NH_2)_{30}$ (blue lines) and CD45 + $C_{60}(OH)_{30}$ (green line). These results mean that R812 and E1167 can form strong and stable

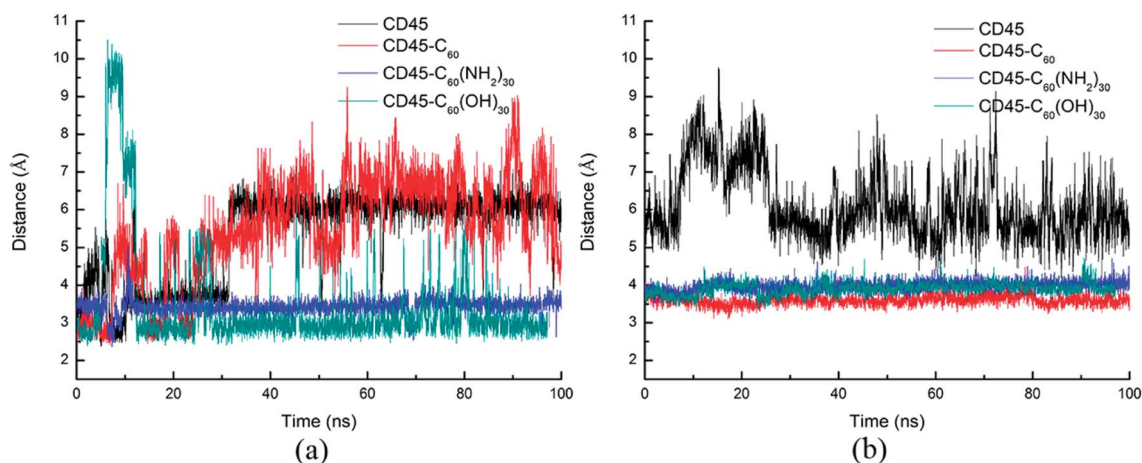


Fig. 6 Time evolutions of the distances between critical residues (a) R812 in D1 and E1167 in D2, (b) Y767 in D1 and E894 in D2 during 100 ns MD simulations, where the interactions can be considered disappear when the distance is up to 6 Å.



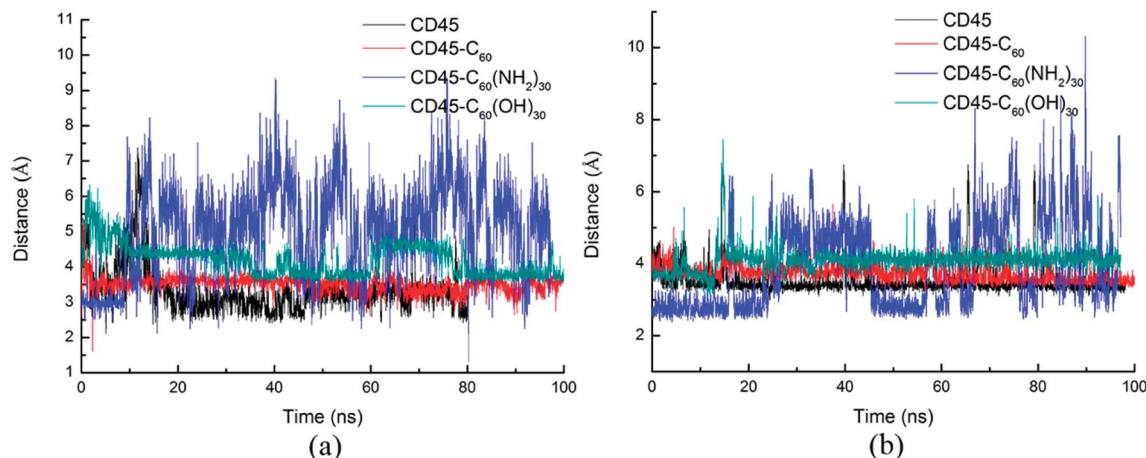


Fig. 7 Time evolutions of the distances between critical residues (a) D766 in D1 and E1003 in D2, (b) E801 in D1 and E1003 in D2 during 100 ns MD simulations.

interactions by binding with $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ in CD45. The distance between Y767 in D1 and E894 in D2 is shown in Fig. 6b. For the control simulation, there are strong variations of the distance in the early stage of the simulation, whereas keep stable at around 6 Å after ~30 ns MD simulation, indicating that weak interaction exists between the two residues. Significantly, both of these distances are reduced to 4 Å and maintained extreme stable by binding with C_{60} , $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ in CD45, indicating that these two residues can form strong interactions mediated by the fullerene-based molecules.

Fig. 7 shows the distances between D766 and E1003, and E801 and E1003, respectively. Similar tendency can be observed from Fig. 7a and b. For control simulation (black line), the distances between D766 and E1003 (in Fig. 7a), and E801 and E1003 (in Fig. 7b) keep stable at 3 Å and 3.5 Å, respectively. For

the bound-state complexes of CD45 + C_{60} and CD45 + $C_{60}(OH)_{30}$, the average distances only increase by ~0.1 Å and ~0.5 Å, respectively. These results mean that the interactions are not significantly affected and remain stable throughout the simulations. Nevertheless, when CD45 binding with $C_{60}(NH_2)_{30}$, these two distances become very unstable and fluctuate between 3 Å and 7 Å, implying that the interactions between D766 and E1003, and E801 and E1003, are significantly impaired by the binding of $C_{60}(NH_2)_{30}$ in CD45 (blue lines in Fig. 7a and b). For a profound comprehension of the mechanisms of fullerene-based molecules to the intermolecular interactions between D1 and D2 domains, a schematic diagram is shown in Fig. 8. Compared with the control simulation (Fig. 8a), the transformation of the interaction networks of CD45 + C_{60} (Fig. 8b) and CD45 + $C_{60}(OH)_{30}$ (Fig. 8d) are not

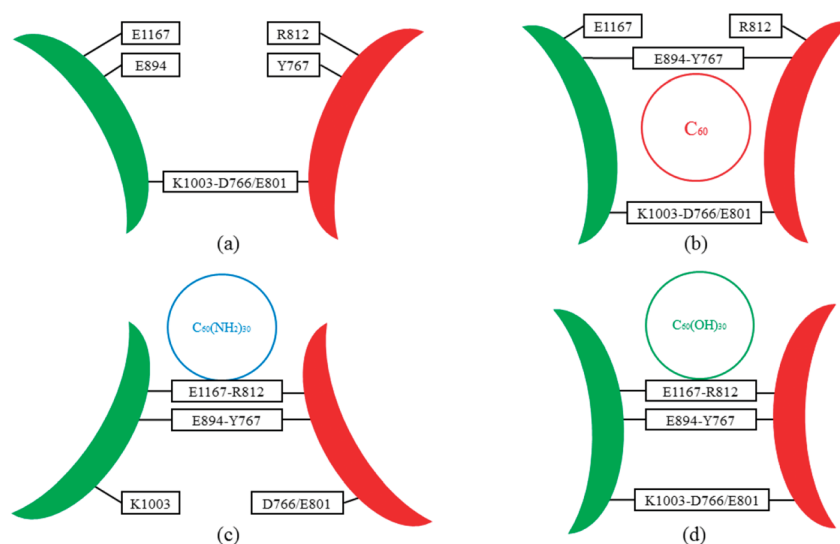


Fig. 8 Schematic representation of intermolecular interaction networks of (a) CD45, (b) CD45 + C_{60} , (c) CD45 + $C_{60}(NH_2)_{30}$, and (d) CD45 + $C_{60}(OH)_{30}$. The D1 and D2 domains are represented in red and green arcs, respectively. The binding sites of C_{60} , $C_{60}(NH_2)_{30}$, and $C_{60}(OH)_{30}$ are represented by circles. The critical residues in D1 and D2 domains are highlighted in the square frames. They form interactions when they are in the same square frame. On the contrary, there is no interaction when the two residues are not in the same square frame.



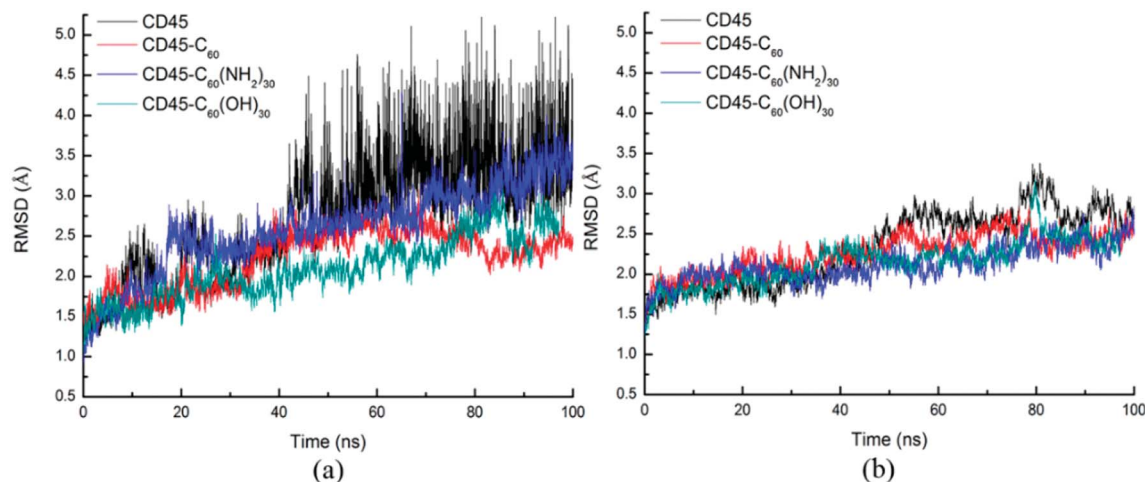


Fig. 9 Time evolutions of RMSDs of the $C\alpha$ atoms of (a) D1 and (b) D2 domains complexed with C_{60} (red), $C_{60}(NH_2)_{30}$ (blue), and $C_{60}(OH)_{30}$ (green) during 100 ns MD simulations. The control simulation without any fullerene-based molecules is colored in black lines.

significant. Nevertheless, the binding of $C_{60}(NH_2)_{30}$ in CD45 (Fig. 8c) completely changes the original interaction mode between D1 and D2 domains. The main reason is that F819 and F820 form strong interactions with $C_{60}(NH_2)_{30}$, resulting in R812 in helix $\alpha 3$ to approach D2 domain and the misfolding of $\alpha 3$. Meanwhile, the loop region, which contains from R763 to Y767, is more flexible and detrimental to interact with K1003 in D2 domain (detailed analysis will be discussed in the section “Conformational changes of the active site”).

Conformational changes of D1 and D2 domains

D1 and D2 domains establish extensive association to the extent that one requires another for maintenance of its proper structure. Thus the deformation of the intermolecular interactions between D1 and D2 domains will cause structural changes of D1 and D2 domains.

Fig. 9 shows the time evolutions of RMSDs of the $C\alpha$ atoms of D1 (Fig. 9a) and D2 (Fig. 9b) domains. For each of the four systems, the fluctuation of RMSD values of D1 domain are coincide with those of the whole CD45 (Fig. 3), indicating that conformational changes of CD45 mainly occur in D1 domain. Significantly, the RMSD values of D2 domain are much lower and more stable, either for the control simulation or the other three bound-state complexes. These results indicate that the binding of fullerene-based molecules to CD45 have more notable influences on D1 domain than D2 domain. Our conclusions also partly confirm that the D2 domain is not biologically active, consistent with the results in the literature.⁴²

Fig. 10 shows the root mean square fluctuation (RMSF) of every residue of CD45, CD45 + C_{60} , CD45 + $C_{60}(NH_2)_{30}$, and CD45 + $C_{60}(OH)_{30}$. There are several unusual active fragments in nature structure of CD45 (black line in this figure), such as residues E624, Y658, D796, S829, D1043, D1088 or E1103, indicating by the peak values in Fig. 10. These amino acids can flexibly regulate their position and orientation to accommodate substrate due to their high flexibility. However, the freedom of motion of these residues is severely restricted with the binding

of fullerene-based molecules, so that they are not flexible enough to bind substrate to achieve the catalytic function of CD45. As reported in the literature,⁴² the active pocket of CD45 D1 domain is composed by the residues of the PTP signature motif (from V826 to G836), and surrounded by three additional segments: the KNRY motif (from K655 to Y658), the WPD motif (from W794 to D766), and the Q-rich motif (from Q872 to Q876). Importantly, as shown in the inset in Fig. 10, the RMSF values of active pocket (mainly the residues from H827 to V832) are significantly reduced after binding with fullerene-based molecules, especially for $C_{60}(NH_2)_{30}$. The RMSF values of the loop region (N814–I824) are the lowest of the all investigated system, especially for F819 and F820 (RMSF are 0.928 and 0.838, respectively). The results indicate that the flexibility of the loop containing F819 and F820 is significantly reduced due to the

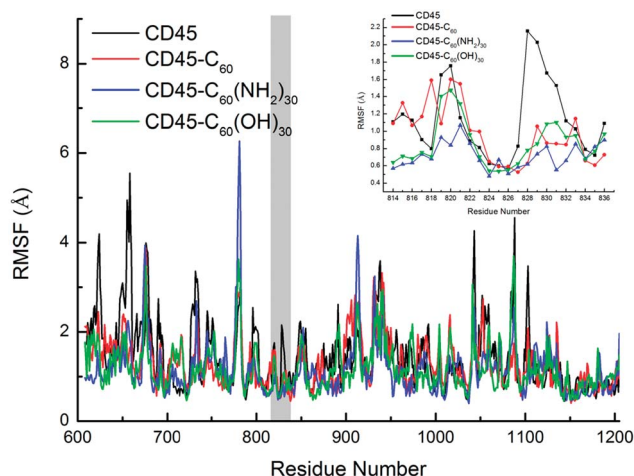


Fig. 10 Comparison between the RMSF plots of $C\alpha$ atoms for CD45 (black), CD45 + C_{60} (red), CD45 + $C_{60}(NH_2)_{30}$ (blue), and CD45 + $C_{60}(OH)_{30}$ (green) during the MD simulations. The residues of the loop region (N814–I824) and active pocket (V826–G836) are highlighted by gray area, and the corresponding RMSF values are clearly indicated by an inset.



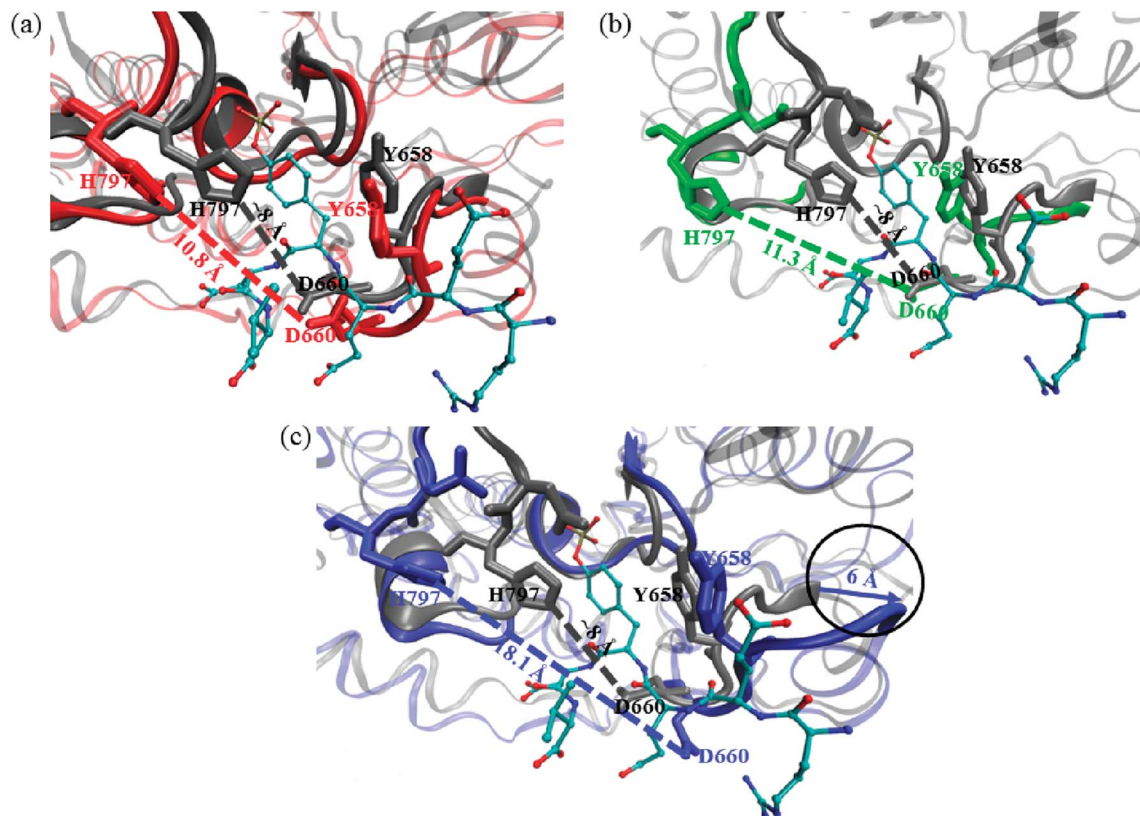


Fig. 11 Comparison of the active pocket of CD45 D1 domain in crystal structure (gray) with the structure bound with (a) C₆₀ (red), (b) C₆₀(NH₂)₃₀ (blue), and (c) C₆₀(OH)₃₀ (green). The critical residues, Y658, D660 and H797 are also represented in relative colors. The crystal structure of substrate is represented by CPK. The circular frame in (c) indicates that the KNRY motif moves outward by ~ 6 Å in the structure bound with C₆₀(NH₂)₃₀.

formation of the intense interactions with C₆₀(NH₂)₃₀, thereby inhibiting the movement of β 12, that is the active pocket. Moreover, the high RMSF values of E624, Y658/D660 (in KNRF motif), D796/H797 (in WPD motif) are also significantly reduced, accompanied by the binding of fullerene-based molecules.

Unexpectedly, the binding of C₆₀(NH₂)₃₀ greatly increases the RMSF values of the loop region (residues E779 to T782) between β 10 and β 11. Combining the data in Table S4,[†] the probability of the residues which located in β 10 and β 11 (C764, Y767, I769, K771, T787 and I789) interact with C₆₀(NH₂)₃₀ are greater than those interact with C₆₀(OH)₃₀. Furthermore, according to the data in Fig. S5b,[†] when C₆₀(NH₂)₃₀ is docked, the total vdW energy of the aforementioned residues (13.08 kcal mol⁻¹) is almost three times than that of C₆₀(OH)₃₀ (3.419 kcal mol⁻¹). As these residues need to maintain strong interaction with C₆₀(NH₂)₃₀, loop region needs to be more flexible to adjust the conformation of protein. We speculated that this is an indirect reason for the increase in the RMSF value of the loop.

We compared the crystal structure of CD45 bound with substrate (T-cell receptor: CD3 zeta ITAM-1 peptide) to our simulation structures, and the results are shown in Fig. 11. The aromatic ring of Y658 in the KNRY motif stabilizes the substrate by forming π - π interaction. However, it flips to perpendicular to the substrate by the binding of either for C₆₀, C₆₀(NH₂)₃₀ or C₆₀(OH)₃₀, which is extremely detrimental to the formation of

π - π interaction with substrate. As shown in Fig. 11c, D660 moved outward ~ 4.8 Å with the binding of C₆₀(NH₂)₃₀, while it did not move along with the combination of C₆₀, C₆₀(OH)₃₀. Furthermore, this positioning of these two oppositely charged residues, D660 and H797 (in WPD motif) determine the backbone configuration of the substrate. In the crystal structure, the distance between D660 and H797 is ~ 8 Å, which increases to 18.1 Å after the binding of C₆₀(NH₂)₃₀, whereas this distance increases only to 10.8 Å and 11.3 Å by the binding of C₆₀ and C₆₀(OH)₃₀. Importantly, compared with the KNRY motif in the crystal structure, it moves outward by ~ 6 Å in the structure bound with C₆₀(NH₂)₃₀.

In summary, the binding of fullerene-based molecules affects the movement of the KNRY, and the active pocket, in which the effect of C₆₀(NH₂)₃₀ on both is most severe.

Conclusions

Fullerene is being studied for their potential applications to be effective inhibitors for protein tyrosine phosphatases, where appropriate surface modification will enhance its inhibitory effect. However, very little is known about how fullerene-based molecules modified with different functional groups affect the activity of protein.

In this study, we utilized molecular docking and atomistic MD simulations to investigate the binding modes of CD45-



fullerenes complexes. Docking calculations of fullerene-based molecules (C_{60} , $C_{60}(NH_2)_{30}$, and $C_{60}(OH)_{30}$) to CD45 revealed that all the three fullerene-based molecules can be docked into the region between D1 and D2 domains. Due to the different surface properties, C_{60} was docked into the region mainly composed of hydrophobic and aromatic residues, while the binding positions of $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ are very similar, except that the average number of residues directly interact with the former is two more than that of the latter. These two amino acids, F819 and F820, lead to different effects of $C_{60}(NH_2)_{30}$ and $C_{60}(OH)_{30}$ on protein activity. Detailed MD simulation analyses of the interactions formed between D1 and D2 domains show that the behavior of $C_{60}(NH_2)_{30}$ is completely different from that of the control simulation due to the misfolding of $\alpha 3$. Furthermore, the movement of active pocket and KNRY motif of D1 domains are most severely impaired by docking with $C_{60}(NH_2)_{30}$. Succinctly, fullerene derivatives modified with amino group exhibit conspicuous tumor inhibition to CD45.

Our study and understanding of the fullerenes–protein interactions can be exploited in the view of design biologically-active hybrid materials for the inhibition of protein tyrosine phosphatases.

Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Conflicts of interest

The authors declare no competing financial interest.

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