



Cite this: *RSC Adv.*, 2018, 8, 5902

Manganese(II) enhanced fluorescent nitrogen-doped graphene quantum dots: a facile and efficient synthesis and their applications for bioimaging and detection of Hg²⁺ ions†

Li Yang,^a Aimiao Qin,^{ID}*^a Shuoping Chen,^a Lei Liao,^a Jiangke Qin^b and Kaiyou Zhang^a

Manganese ion (Mn²⁺) bonded nitrogen-doped graphene quantum dots (Mn(II)-NGQDs) with water solubility have been successfully synthesized by a simple, one-pot hydrothermal carbonization, using sodium citrate, glycine and manganese chloride as raw materials. The photoluminescence (PL) characteristics of Mn(II)-NGQDs were studied in detail. The resulting Mn(II)-NGQDs show a remarkably enhanced PL intensity and quantum yield (QY = 42.16%) compared with the product without Mn(II)-doped (named as NG, QY = 27.06%) and the product doped with other metal ions. The Mn(II)-NGQDs not only display low toxicity and high cellular uptake efficiency for fluorescence live cell imaging in biological evaluations but also exhibit a fast, highly selective and sensitive fluorescence quenching effect toward Hg²⁺ ions, with a detection limit of 3.4×10^{-8} mol L⁻¹.

Received 4th November 2017

Accepted 31st January 2018

DOI: 10.1039/c7ra12133d

rsc.li/rsc-advances

Introduction

Graphene quantum dots (GQDs) have drawn much attention in the wide fields of catalysis, energy conversion, optoelectronics, bioimaging probes and biosensors.^{1–5} Very recently, GQDs have been intensively studied as new fluorescent materials in bioimaging and biosensors, because of their unique optical properties, low toxicity, water solubility and excellent biocompatibility.⁶ GQDs bearing oxygen functionalities such as carboxyl, epoxy and hydroxyl groups indicate obvious advantages in water solubility, optical and electronic properties.^{7–10} GQDs possess high crystallinity and their crystal structure resemble regular hexagonal lattice for the layered graphene.¹¹ GQDs can flexibly combine with target drug molecules due to the unique two-dimensional (2D) structure and abundant surface functional groups, thus they have been attracted much attention as a kind of novel light-emitting nanomaterial for versatile theranostic biomedical applications, including diagnostic imaging-guided therapy, fluorescence biosensing-based tumor detection, drug delivery, and synergistic cancer therapy.^{12,13}

To date, a myriad of strategies for fabricating GQDs and graphene derivatives have been explored, such as pyrolysis synthesis,¹⁴ electrochemical synthesis,¹⁵ microwave-assisted solid-phase synthesis,¹⁶ hydrothermal synthesis¹⁷ and *etc.* Among these synthesis approaches, the carbonization of citric acid to prepare GQDs were considered to be one of promising method, since it can provide stable quantum dots with high yields.^{18,19} However, there are a series of problems for the direct carbonization of citric acid to face, such as complicated preparation process, harsh reaction conditions (such as strong acid/alkali using), accurate pH control and tedious postprocessing, which may lead to great restrictions for the further biomedical applications of QDs.^{20–22}

To extend and improve the physical and chemical properties of GQDs, different modification methods have been applied, including surface functional groups modification,²³ heteroatoms doping^{24,25} and formation of composite.^{26,27} For example, the ways of functionalization with surface groups and heteroatoms doping (such as N,S) may significantly transform the optical properties, quantum yields, chemical activities and electronics structure of GQDs,^{28,29} which would provide interesting luminescent properties, such as pH-dependent and excitation-dependent photoluminescence (PL) behaviors.^{30,31} The interaction of GQDs and metal ions or rare earth elements^{32,33} in solution may lead to the excitation/emission spectrum suppression, amplification or shift, which has been used for high selectivity and sensitivity detection of different ions and specificity molecules. Han *et al.* reported the magnetic-fluorescent imaging behavior based on manganese(II)-carbon

^aKey Lab New Processing Technology for Nonferrous Metals & Materials, Ministry of Education, Guangxi Key Laboratory in Universities of Clean Metallurgy and Comprehensive Utilization for Non-ferrous Metals Resources, College of Materials Science & Engineering, Guilin University of Technology, Guilin, China. E-mail: 592491245@qq.com

^bCollege of Chemistry and Pharmaceutical Science, Guangxi Normal University, Guilin, China

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c7ra12133d



dots, indicating that the metal bonded GQDs could be good fluorescent probes for bioimaging.³⁴

In this paper, in order to explore the doping effect of different metal ions on the PL behaviors of GQDs, we chose sodium citrate and glycine as carbon and nitrogen sources respectively to synthesize multifunctional GQDs for the first time. By using the facile and efficient hydrothermal method, Mn²⁺ bonded nitrogen-doped graphene quantum dots (Mn(II)-NGQDs) with water solubility and multifunction were successfully produced, which is the first case of Mn(II)-NGQDs with Mn²⁺ enhanced fluorescence *via* a confinement effect between Mn²⁺ and functional groups. The Mn(II)-NGQDs exhibit a strong emission with high quantum yield, and display a highly selective and sensitive fluorescence quenching response to Hg²⁺. Moreover, the Mn(II)-NGQDs show low cytotoxicity and are suitable for fluorescence live cell imaging, which would be used for bio-imaging application.

Experimental

Reagents and chemicals

Glycine, sodium citrate, MnCl₂·4H₂O, MnSO₄, Mn(NO₃)₂, Mn(CH₃COO)₂, Ni(NO₃)₂·6H₂O, Pb(NO₃)₂, AgNO₃, Zn(NO₃)₂·6H₂O, CuCl₂·2H₂O, FeSO₄·7H₂O, FeCl₃·6H₂O, Co(NO₃)₂·6H₂O, CaCl₂, CdCl₂, hexamethylenetetramine (HMTA) and other chemicals were of analytical grade and used as received without further purification. Milli-Q water (18.2 MΩ) was used for all of the experiments. Stock solutions of Hg²⁺, Ag⁺, Al³⁺, Ba²⁺, Cr³⁺, Ca²⁺, Cd²⁺, Co²⁺, Cu²⁺, Fe²⁺, Fe³⁺, Mg²⁺, Mn²⁺, Zn²⁺, Ni²⁺ and Pb²⁺ ions were prepared by dissolving corresponding metal salts in Milli-Q water.

Characterization

Transmission electron microscopy (TEM) was carried out on a JEOL-2010 TEM at 200 kV. Fourier transform infrared (FTIR) spectra were recorded using a FTIR2500 spectrometer (KBr disk). Raman spectra were collected on a LabRam-1B Raman spectroscope equipped with a 632.8 nm laser source. X-ray diffraction spectra (XRD) were determined on a Holland PANalytical X'Pert PRO X-ray diffractometer with Cu Kα (λ = 1.54056 Å) as radiation source in the 2θ range of 5–80°. X-ray photoelectron spectroscopy (XPS) measurements were made on Kratos AXIS UltraDLD (Kratos Analytical Ltd.) with mono Al Kα radiation (hν = 1487.71 eV) at a power of 75 W. Emission spectra were collected using a Varian Cary 100 spectrometer. Fluorescence life time of the products were determined using a FLS980 fluorometer (Edinburgh Instruments Ltd.). Absorption spectra were recorded on a UV-3600 UV-vis-NIR spectrophotometer (Shimadzu). Fluorescence quantum yield (QY) was detected using quinine sulfate as the standard (QY = 54% in 0.1 M H₂SO₄). Confocal laser-scanning microscopy (CLSM) images were recorded on a Zeiss LSM 710 CLSM (Zeiss LSM710, Germany). The zeta potential of Mn(II)-NGQDs and NG were measured by a Zetasizer Nano ZS90 (Malvern Instruments).

Synthesis of Mn(II)-NGQDs and NG

In typically synthesis, a mixture of glycine (0.0451 g), sodium citrate solution (12 mL, 0.1 mol L⁻¹), MnCl₂ solution (1 mL, 0.1 mol L⁻¹) and Milli-Q water (37 mL) was added into a flask with stirring, to form a clear reaction solution. The solution was then transferred into a 100 mL Teflon-lined autoclave, and then heated to 180 °C for 10 h. After that, the autoclave was cooled down naturally to room temperature. The obtained bright transparent solution was centrifuged at 10 000 rpm for 10 min to remove the deposit. Subsequently, the supernatant of Mn(II)-NGQDs solution (6.52 mg mL⁻¹, ESI⁺) was dialyzed (molecular weight cut off, 1000 Da) against water for 48 h. Afterwards, the pure and solid-state Mn(II)-NGQDs white powder was obtained by vacuum freeze drying. Finally, the resulted Mn(II)-NGQDs were dispersed in water as a stock solution for further characterization and detection.

As a contrast, other common metal ions doped in NG (M-NG, M = Ag⁺, Al³⁺, Ba²⁺, Cr³⁺, Ca²⁺, Cd²⁺, Co²⁺, Cu²⁺, Fe²⁺, Fe³⁺, K⁺, Mg²⁺, Zn²⁺, Ni²⁺ or Pb²⁺), were also prepared using the similar method as Mn(II)-NGQDs, except for using different metal salts. In addition, the NG without metal doping was also prepared using the above experimental procedure.

Fluorescence detection of Hg²⁺ ion

In a typical process, 200 μL of Hg²⁺ solution with various concentrations (0–5 × 10⁻⁶ mol L⁻¹) and 200 μL of Mn(II)-NGQDs (6.52 mg mL⁻¹) solution were mixed together and diluted to 5 mL with the HMTA buffer solution (pH = 7.35). After kept at room temperature for 10 min, the fluorescence emission spectra of the solution were collected under an excitation wavelength of 370 nm. As a contrast, the sensing capability of NG toward Hg²⁺ have been investigated by a same detection procedure. The procedure of Mn(II)-NGQDs detection of Hg²⁺ in Milli-Q water were also carried out.

Cellular toxicity test

The cytotoxicity of Mn(II)-NGQDs was evaluated by the MTT assay. HepG2 human liver cancer cells were harvested (obtained from Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences; the cell density was adjusted to 10⁵ cells per mL) and seeded in a 96-well plate overnight for 24 h. Then, the Mn(II)-NGQDs solution was added into each well with increasing concentrations from 0 to 300 μg mL⁻¹. After that, the cells were cultivated for 4 and 24 h, respectively. And then, 10 μL of 5 mg mL⁻¹ MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetra-zolium bromide) solution was added to each cell well. After incubation for 4 h, the culture medium was discarded, and 100 μL of dimethyl sulfoxide (DMSO) was added. The obtained mixture was shaken for 15 min in the dark at room temperature, and the absorbance of each well was measured at wavelength of 570 nm using an enzyme linked immunosorbent assay (Tecan Infinite M1000) reader with pure DMSO as a blank. In addition, HepG2 human liver cancer cells without Mn(II)-NGQDs treating were used as a control to test their relative cell viability.



Live cell imaging

HepG2 human liver cancer cells and Raw264.7 mouse macrophages (obtained Shanghai, China) were cultured in Dul-becco's modified eagle medium (DMEM) supplemented with PBS (pH = 7.4) in 10% and 100 mg mL⁻¹ streptomycin and 100 units per mL penicillin solution in 5% CO₂ at 37 °C. The cells were grown on the bottom of 35 mm glass culture dishes to the density of 80% confluence and then incubated with Mn(II)-NGQDs at 37 °C for 3 h. Then, the cells were washed three times with PBS. Images of the cells were immediately captured at ambient temperature on a laser scanning confocal microscopy using a digital camera.

Results and discussion

Characterization of Mn(II)-NGQDs

The synthetic procedure for the fabrication of fluorescent Mn(II)-NGQDs is illustrated in Fig. 1. The morphology and microstructure of the Mn(II)-NGQDs were confirmed by the typical TEM image. As shown in Fig. 2a, the Mn(II)-NGQDs were well dispersed and uniform in size and shape. The corresponding size distribution histogram was obtained by random counting 100 particles (Fig. S1, ESI[†]), indicating that the size of Mn(II)-NGQDs was about 1.0–5.5 nm with an average diameter of approximately 4.5 nm. By contrast, the NG consisted of a large amounts of irregular nanosheets and a small amount of QDs (Fig. 2b, ESI[†]). Obviously, the bonding of Mn²⁺ can change the morphology of NG effectively. The HRTEM image of the Mn(II)-NGQDs shows their well-resolved lattice fringes, which indicates that they possess relatively good crystallinities. The lattice spacings of Mn(II)-NGQDs (the inset in Fig. 2a) and NG

(the inset in Fig. 2b) are 0.24 and 0.26 nm respectively, which are assigned to the (100) and (020) plane of graphitic carbon, respectively.³⁵

According to the XRD patterns of Mn(II)-NGQDs and NG (Fig. S2, ESI[†]), both of Mn(II)-NGQDs and NG exhibit two broad peaks centered at 9.2° (2θ) and 31.5° (2θ), corresponding to the (001) and (002) plane of graphitic nitride carbon,^{36,37} respectively. Compared with NG, the broad peak at 31.5° (2θ) of Mn(II)-NGQDs is weakened obviously, indicating that in Mn(II)-NGQDs, the Mn²⁺ ions bond with the functional groups at the edge of NG, and may help to divide the large graphene sheets to small graphene domains.

Fig. 3a shows the FTIR spectra of Mn(II)-NGQDs and NG. The broad band at 3406 cm⁻¹ corresponds to the stretching vibration of O–H and N–H. The strong and broad absorption band ranging from 1500 cm⁻¹ to 1750 cm⁻¹ with a center at 1580 cm⁻¹ corresponded to the stretching vibrations of C=O and C=C in the aromatic ring. The bands at 1407 cm⁻¹ and 830 cm⁻¹ were assigned to the C–H bending and C–H stretching, respectively. The bands at 1258 cm⁻¹ and 1078 cm⁻¹ were attributed to the stretching vibrations of O–H and C–O–C, respectively.^{16,38,39} The small peaks at 1495 cm⁻¹ and 1449 cm⁻¹ corresponded to the NH₃⁺ symmetric bending and C–N stretching vibration, respectively,^{39,40} which confirmed that nitrogen atoms have been successfully doped into the Mn(II)-NGQDs and NG. However, compared to NG, the C–O stretching vibration at 1078 cm⁻¹ and NH₃⁺ symmetric bending at 1495 cm⁻¹ in Mn(II)-NGQDs disappear, indicating the existence of Mn²⁺ ions which have bonded to the NG. Moreover, the C–N stretching vibration of Mn(II)-NGQDs at 1449 cm⁻¹ increases obviously, indicates that more nitrogen elements are preserved after the coordination effect of Mn²⁺ ions. Fig. 3b shows the

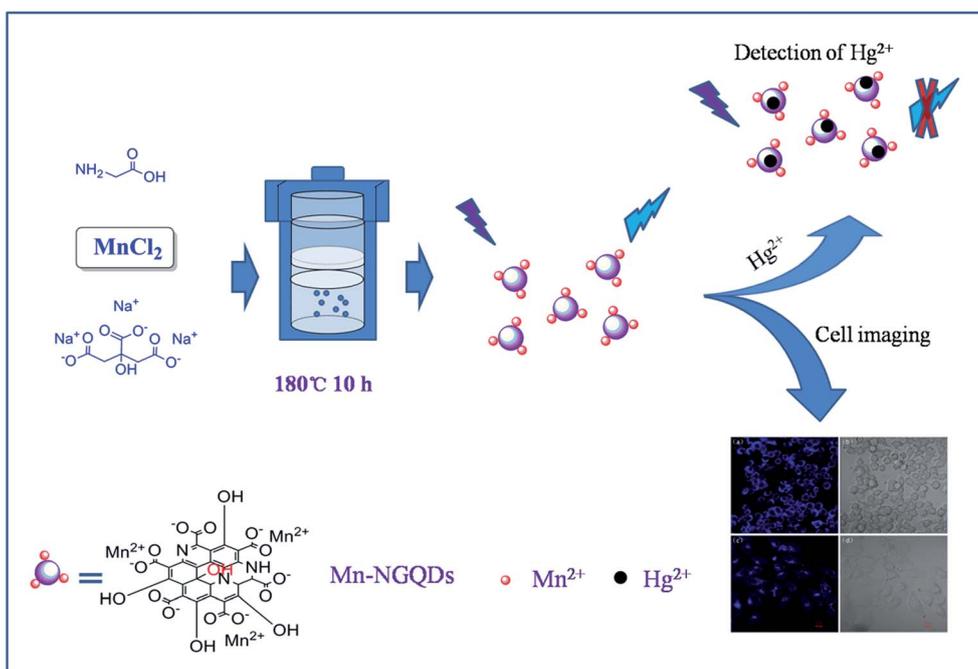


Fig. 1 Schematic illustration of the synthesis of Mn(II)-NGQDs and cells imaging and Hg²⁺ detection.



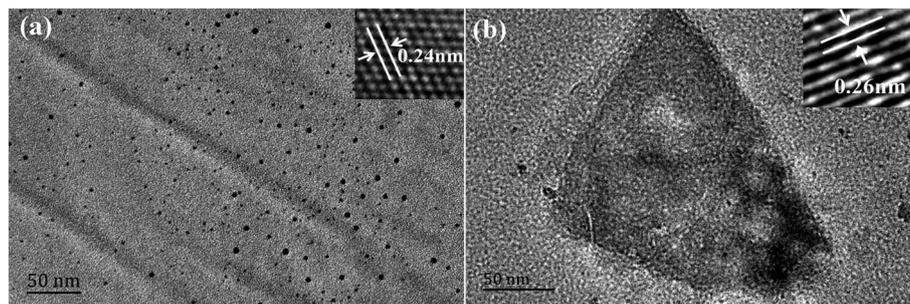


Fig. 2 TEM and HRTEM images of (a) Mn(II)-NGQDs and (b) NG.

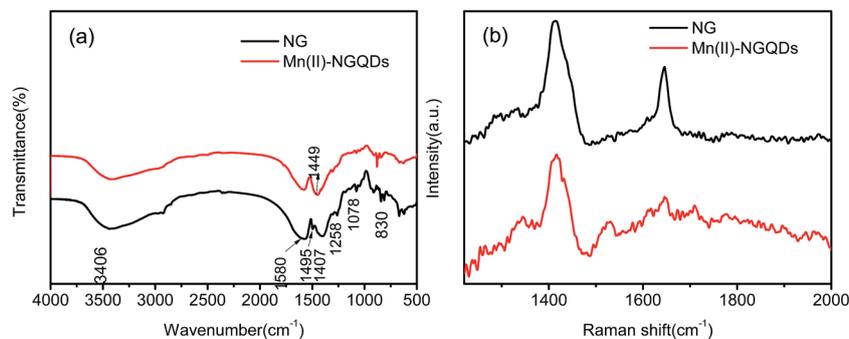


Fig. 3 (a) FT-IR spectra of Mn(II)-NGQDs and NG; (b) Raman spectrum of Mn(II)-NGQDs and NG.

Raman spectra of Mn(II)-NGQDs and NG. Both of Mn(II)-NGQDs and NG demonstrate a G band at 1645 cm^{-1} , which relates to the vibration of the sp^2 hybridization of graphitic carbon, and a D band at 1411 cm^{-1} , which corresponds to the carbon lattice distortion. Compared to NG, Mn(II)-NGQDs show a more weak G band at 1645 cm^{-1} and a larger intensity ratio $I_{\text{D}}/I_{\text{G}}$ increase from 1.29 to 2.23. Similar to the previous analysis,^{41–43} more structural defects, larger sp^2 domains and smaller sp^2 fragments are expected to be present in the Mn(II)-NGQDs. It is indicated that the bonding of Mn^{2+} can change the morphology of NG.

Furthermore, more information of the surface functional groups of Mn(II)-NGQDs and NG is further provided by XPS analysis (Fig. 4). The full-range spectra (Fig. 4a) show that, both of Mn(II)-NGQDs and NG contain C, N and O elements, and there is about 0.21% Mn (atomic fraction) in Mn(II)-NGQDs (Table S1, ESI†). Fig. 4b shows the C 1s XPS spectrum of the Mn(II)-NGQDs, which ranged from 280 to 292 eV. There are four adjacent peaks at 284.3 eV, 284.9 eV, 285.9 eV and 288.1 eV, which can be attributed to C–C/C=C, C–N, C–O and C=O bonding respectively.⁹ The N 1s spectrum of Mn(II)-NGQDs (Fig. 4d) exhibits three peaks at 398.9 eV, 399.2 eV and 400.9 eV, which are associated with C–N–C, N–(C)₃ and N–H groups.^{11,39,40} The O 1s spectra of Mn(II)-NGQDs (see Fig. S3a, ESI†) and NG (see Fig. S3b, ESI†) show two peaks at 531.0 eV and 531.8 eV, which are associated with O–H and C–O–C groups, respectively.¹⁷ These spectra reveals that nitrogen element has successfully introduced into the Mn(II)-NGQDs and NG, and Mn(II)-NGQDs and NG has been functionalized with some groups,

such as –COOH, –OH and N–H group. The high resolution XPS spectrum of Mn 2p (Fig. 4b) shows a dominant peak at 641 eV assigned to Mn^{2+} , indicating that the oxidation state of Mn on the surface of Mn(II)-NGQDs remains unchanged.³⁴ Compared to the XPS spectrum of NG, the C 1s (Fig. 4e) and N 1s (Fig. 4f) spectra of Mn(II)-NGQDs in the peak shape are very similar to those of NG, which means that the introduction of Mn^{2+} ions cannot affect the carbon-based species. However, the atomic fraction (Table S1, ESI†) of N in Mn(II)-NGQDs (4.24%) is dramatically increasing compared with that of NG (2.2%), which may indicate that more N–H groups are preserved by the coordination effect of Mn^{2+} ion. The above results clearly reveal that Mn(II)-NGQDs and NG have been successfully produced.

Optical properties

In order to investigate the optical properties of the as-prepared Mn(II)-NGQDs, the excitation, emission and UV-vis absorption spectra were studied and the results are given in Fig. 5a. The UV-vis absorption spectrum of Mn(II)-NGQDs shows two main absorption peaks: one peak centered at 260 nm is attributed to π – π^* transition of C–C band, and the other centered at 340 nm can be ascribed to the n – π^* transition of C–N and C=O groups.⁴⁴ NG and other M-NG also display the two peaks which are similar to Mn(II)-NGQDs (Fig. S4, ESI†). As shown in Fig. 5a, the maximum excitation wavelength (370 nm) of Mn(II)-NGQDs gets a red shift than the UV-vis absorption band (340 nm), which can be ascribed to a distribution of the different surface energy traps of the carbon dots. The emission wavelength ($\lambda_{\text{em}} = 443$) of Mn(II)-NGQDs has no obvious change, compared with NG,



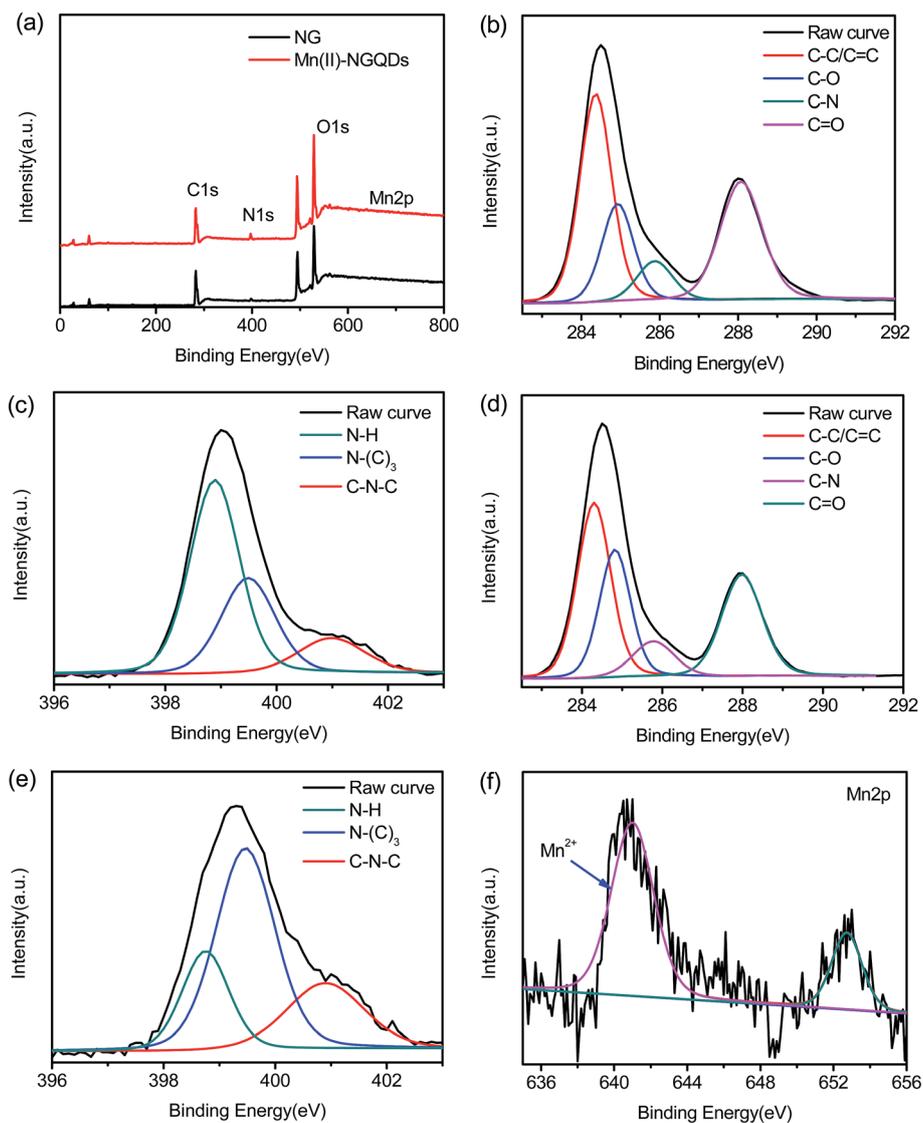


Fig. 4 (a) XPS survey spectra of Mn(II)-NGQDs and NG; (b) C 1s and (c) N 1s region of Mn(II)-NGQDs, (d) C 1s and (e) N 1s region of NG; XPS high resolution of (f) Mn 2p.

but the fluorescence intensity of Mn(II)-NGQDs shows much stronger than that of NG. Fig. 5b shows that the emission spectra of Mn(II)-NGQDs have an excitation independent feature as the excitation wavelength gradually increases from 350–400 nm, with the maximum excitation wavelength and the maximum emission wavelength at 370 and 443 nm, respectively. The excitation independent phenomenon were also observed in previous reported,^{19,22} which may be due to the uniform size of Mn(II)-NGQDs and the unitary PL process of $n-\pi^*$ transition of the aromatic N in Mn(II)-NGQDs. Fig. 5c and d show the emission spectra and QY of Mn(II)-NGQDs with different concentrations of Mn^{2+} ion in reaction system under the excitation wavelength of 370 nm. The results indicate that the strongest synergistic effect with functional groups in reaction systems appears at the concentration Mn^{2+} ion of $2 \times 10^{-3} \text{ mol L}^{-1}$. The highest fluorescence QY of the obtained Mn(II)-NGQDs was determined to be 42.16%, which is much

higher than that of NG (27.06%) and the previous report of N-GQDs (28.10%).³⁹ This implied that the induced Mn^{2+} into NG significantly enhanced the fluorescence QY of NG.

To further confirm that the enhanced PL indeed arises from the Mn^{2+} -assisted process, the control experiments were carried out by utilizing other Mn salts to replace manganese chloride, including manganese acetate, manganese sulfate and manganese nitrate, the results show that they can still enhance the fluorescence (Fig. S5a, ESI[†]), indicating that Mn^{2+} plays a key role in the enhanced PL from the Mn(II)-NGQDs. Furthermore, holding any other factors constant, citric acid was chosen to replace citric sodium as carbon precursor to prepare Mn(II)-NGQDs and NG for comparison. The comparison experiment results show that, choosing citric acid as precursor for high quality fluorescent quantum dots is unsuitable (Fig. S5b, ESI[†]). This indicates that citric sodium precursor also plays an important role in the improved PL of Mn(II)-NGQDs.



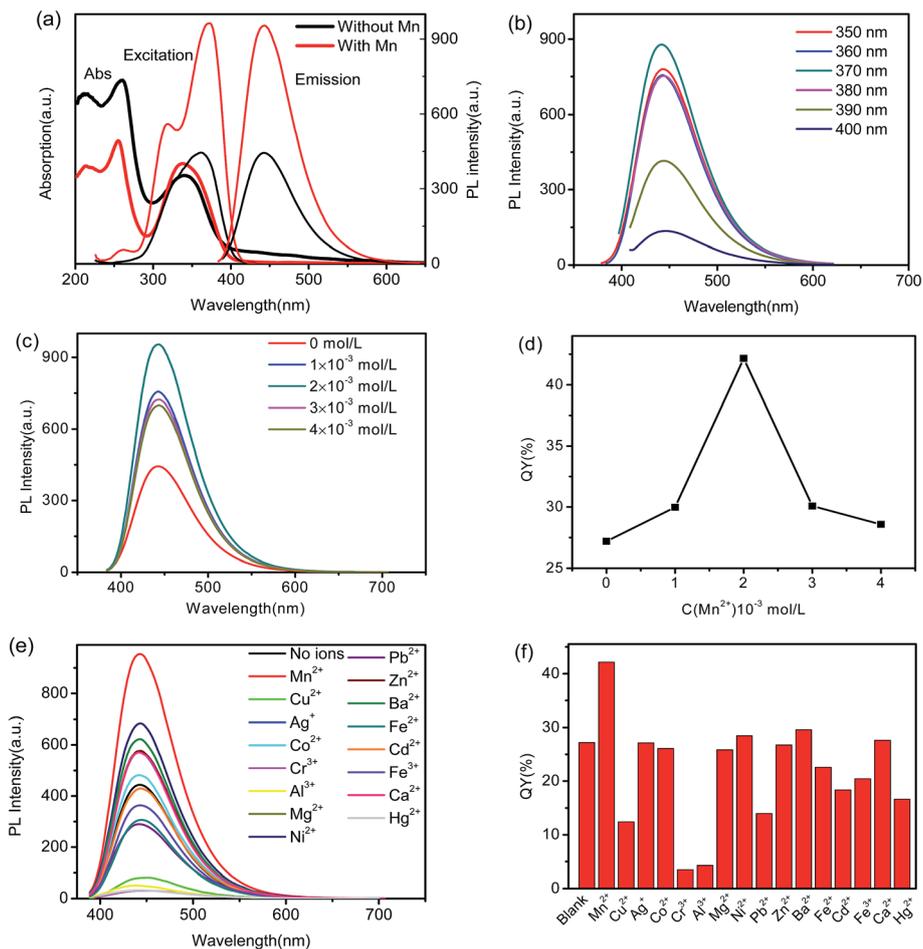


Fig. 5 (a) UV-Vis absorption and PL spectra ($\lambda_{\text{ex}} = 370$ nm) of Mn(II)-NGQDs and NG; (b) PL spectra of Mn(II)-NGQDs recorded for progressively longer excitation wavelengths from 350 to 400 nm; (c) the effect of Mn^{2+} doping concentrations on the PL spectra ($\lambda_{\text{ex}} = 370$ nm) of Mn(II)-NGQDs; (d) the effect of Mn^{2+} doping concentrations on the QY of Mn(II)-NGQDs; (e) the effect of common metal ions doping on the PL spectra of NG; (f) the effect of common metal ions doping on the QY of NG.

The as-synthesized Mn(II)-NGQDs also exhibit a good stability (Fig. S5c, ESI[†]). The PL spectra of the Mn(II)-NGQDs are almost the same with the fresh Mn(II)-NGQDs after 7 days of storage under ambient conditions. In addition, the synergy process of Mn^{2+} ion with functional groups would also be influenced by the reaction temperature (Fig. S5d, ESI[†]) and time, the optimum hydrothermal temperature and time were considered as 180 °C and 10 hours, respectively.

Fig. 5e and f show the PL spectra and QYs of various M-NG. All of the other M-NG display a weaker emission behavior compared to Mn(II)-NGQDs (Fig. 5e). Fig. 5f indicates that the QYs of seven kinds of M'-NG ($M' = \text{Ni}^{2+}, \text{Mg}^{2+}, \text{Zn}^{2+}, \text{Ba}^{2+}, \text{Co}^{2+}, \text{Ag}^+$ and Ca^{2+}) are similar to that of NG, while other seven kinds of M''-NG ($M'' = \text{Cu}^{2+}, \text{Cr}^{2+}, \text{Cd}^{2+}, \text{Fe}^{2+}, \text{Fe}^{3+}, \text{Al}^{3+}$ and Pb^{2+}) have much lower QYs than that of NG.

Why only the introduction of Mn^{2+} ion into NG can obviously enhance their fluorescence? The mechanism is conjectured as the follows: in the hydrothermal reaction of glycine and sodium citrate, parts of carboxyl groups of sodium citrate may not be carbonized and retain on the surface of NG. These carboxyl groups have high reactivity for further condensation and

carbonization, that is why a large amount of irregular graphene nanosheets and some agglomeration were observed in NG samples without Mn^{2+} doping. These irregular graphene nanosheets and large particles would reduce the fluorescent emission behavior. However, in the reaction process of Mn(II)-NGQDs, Mn^{2+} ions can bond with functional groups such as carboxyl groups on the surface of NG. These bonded Mn^{2+} ions not only make NG lose reactivity for further condensation and carbonization, but also prevent the grain growth and agglomeration of the particles of NG. Therefore, the Mn(II)-NGQDs show well dispersed QDs with uniform size and shape, and exhibit a high QY than that of NG. It is notable that, compared to Mn^{2+} , other metal ions (such as $\text{Fe}^{2+}, \text{Co}^{2+}, \text{Ni}^{2+}, \text{Mg}^{2+}, \text{Zn}^{2+}, \text{Ba}^{2+}, \text{Ag}^+$ and Ca^{2+}) do not display such confinement effect to control the shape and grain growth of NG. TEM images of NG doping with other metal ions display similar morphology to pure NG, some large irregular sheets and small QDs can be observed (Fig. S6, ESI[†]). All of these M-NG display weaker fluorescent emission than Mn(II)-NGQDs. This confirms our conjecture and indicates that Mn^{2+} plays a key role in the synthesis of Mn(II)-NGQDs with enhanced fluorescent properties.



Detection of Hg²⁺ ion

The surface characterization of Mn(II)-NGQDs clearly shows the feasibility of application as a fluorescent probe for metal ions detection, due to the homogeneous size distribution and excellent stable fluorescence properties. In this work, HMTA solution was chosen as the buffer to detect Hg(II) since it has been widely used in literature.⁴⁵ Fig. 6a shows the fluorescence response of Mn(II)-NGQDs in the presence and absence of $4 \times 10^{-6} \text{ mol L}^{-1}$ Hg²⁺ in different pH values. From Fig. 6a, it can be observed that the fluorescence intensity of Mn(II)-NGQDs displays a remarkably quenching effect with the addition of Hg²⁺ in a pH range of 5.5 to 8.7. Moreover, the pH value of the solution affects not only the fluorescence intensity of Mn(II)-NGQDs but also the quenching efficiency. The fluorescence

intensity of Mn(II)-NGQDs and quenching efficiency increase rapidly when the pH value increase from 5.5 to 8.0, and reach the maximum at the pH = 7.35 (Fig. 6a). However, an obvious decrease in quenching efficiency is observed when the pH value is higher than 7.35. Thus, it is clear that the quenching efficiency will decay in the acidic or alkaline conditions, and it is easier for Hg²⁺ ions to combine with Mn(II)-NGQDs and form a stable complex in neutral solution, which may be due to the weak protonated process of functional groups in the Mn(II)-NGQDs. Considering the highest quenching efficiency, the following experimental conditions are all set in a pH of 7.35.

To discuss the sensitivity of Mn(II)-NGQDs as fluorescent probe for Hg²⁺ detection, the emission spectra of Mn(II)-NGQDs ($\lambda_{\text{ex}} = 370 \text{ nm}$) in the presence of different concentrations of

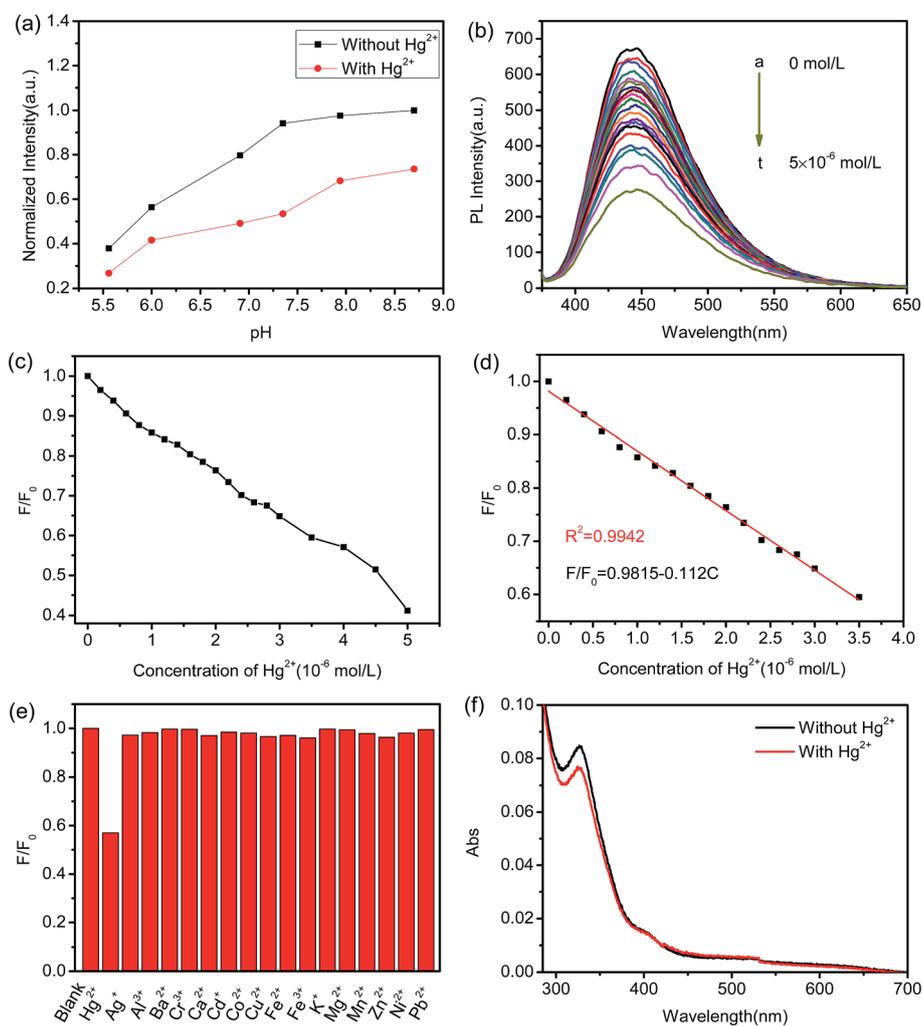


Fig. 6 (a) Fluorescence responses of Mn(II)-NGQDs to different pH value of HMTA buffer under the absence and presence of $4 \times 10^{-6} \text{ mol L}^{-1}$ Hg²⁺ ions; (b) emission spectra ($\lambda_{\text{ex}} = 370 \text{ nm}$) of Mn(II)-NGQDs in the presence of different concentrations of Hg²⁺ (0 – $5.0 \times 10^{-6} \text{ mol L}^{-1}$) in HMTA buffer solution (pH = 7.35). (a) $0 \times 10^{-6} \text{ mol L}^{-1}$, (b) $0.2 \times 10^{-6} \text{ mol L}^{-1}$, (c) $0.4 \times 10^{-6} \text{ mol L}^{-1}$, (d) $0.6 \times 10^{-6} \text{ mol L}^{-1}$, (e) $0.8 \times 10^{-6} \text{ mol L}^{-1}$, (f) $1.0 \times 10^{-6} \text{ mol L}^{-1}$, (g) $1.2 \times 10^{-6} \text{ mol L}^{-1}$, (h) $1.4 \times 10^{-6} \text{ mol L}^{-1}$, (i) $1.6 \times 10^{-6} \text{ mol L}^{-1}$, (j) $1.8 \times 10^{-6} \text{ mol L}^{-1}$, (k) $2.0 \times 10^{-6} \text{ mol L}^{-1}$, (l) $2.2 \times 10^{-6} \text{ mol L}^{-1}$, (m) $2.4 \times 10^{-6} \text{ mol L}^{-1}$, (n) $2.6 \times 10^{-6} \text{ mol L}^{-1}$, (o) $2.8 \times 10^{-6} \text{ mol L}^{-1}$, (p) $3.0 \times 10^{-6} \text{ mol L}^{-1}$, (q) $3.5 \times 10^{-6} \text{ mol L}^{-1}$, (r) $4.0 \times 10^{-6} \text{ mol L}^{-1}$, (s) $4.5 \times 10^{-6} \text{ mol L}^{-1}$, (t) $5.0 \times 10^{-6} \text{ mol L}^{-1}$; (c) the relationship between the F/F_0 and Hg²⁺ concentrations (c: 0 – $5.0 \times 10^{-6} \text{ mol L}^{-1}$); (d) plots of intensity ratio of F/F_0 versus the concentrations of Hg²⁺ (0 – $3.5 \times 10^{-6} \text{ mol L}^{-1}$); (e) fluorescence responses of Mn(II)-NGQDs to the different metal ions in HMTA buffer solution (pH = 7.35). The concentration of each metal ion is $4.0 \times 10^{-6} \text{ mol L}^{-1}$. F_0 and F correspond to the fluorescence intensities of Mn(II)-NGQDs at 440 nm with the 370 nm excitation wavelength in the absence and presence of metal ions, respectively; (f) the UV absorption spectra of Mn(II)-NGQDs without Hg²⁺ and in the presence of Hg²⁺ ($4.0 \times 10^{-6} \text{ mol L}^{-1}$).



Hg^{2+} ion (in the range of 0 to $5.0 \times 10^{-6} \text{ mol L}^{-1}$) were recorded and shown in Fig. 6b. A gradually decrease emission intensity at 443 nm is observed with the increase concentration of Hg^{2+} ion, implying that the fluorescence of Mn(II)-NGQDs solution is sensitive to Hg^{2+} . As shown in Fig. 6c, the quenching efficiency (F/F_0) displays a good linear response to the concentration of Hg^{2+} in the range of 0 to $3.5 \times 10^{-6} \text{ mol L}^{-1}$ (Fig. 6d). The linear equation is calculated to be $F_0/F = 0.9815 - 0.112 [\text{Hg}^{2+}]$ ($R^2 = 0.9942$), where F_0 and F are emission intensities at 443 nm in the absence and presence of Hg^{2+} ion, respectively. Under this experiment condition, the limit of detection (LOD) for Hg^{2+} is calculated to be $3.4 \times 10^{-8} \text{ mol L}^{-1}$ by the $3\delta/S$ (δ is the standard deviation of the lowest signal and S is the slope of linear calibration plot), which is much lower than most of other reported of common fluorescent probes (see Table S2, ESI†).^{46–50}

The selectivity of the Mn(II)-NGQDs toward Hg^{2+} was also explored. Different metal ions such as Cu^{2+} , Mn^{2+} , Ag^+ , Pb^{2+} , K^+ , Ba^{2+} , Zn^{2+} , Co^{2+} , Hg^{2+} , Al^{3+} , Fe^{2+} , Fe^{3+} , Ni^{2+} , Mg^{2+} , Cd^{2+} and Hg^{2+} ions were added into the probe solution respectively, and the solution was analyzed after incubation of 10 min. The results in Fig. 6e manifest that these metal ions except Hg^{2+} ion have almost no interference on the fluorescence intensity of Mn(II)-NGQDs, the fluorescence intensity is apparently quenched only by Hg^{2+} ion. Therefore, the fluorescence sensor based on Mn(II)-NGQDs shows a higher selectivity for Hg^{2+} detection over the other metal ions. Similarly, the effects of non buffer (Milli-Q water) on the detection of Hg^{2+} were performed, the results show that Mn(II)-NGQDs also exhibit a highly selective and sensitive capability to Hg^{2+} in Milli-Q water (Fig. S7, ESI†), which is similar to that in the HMTA buffer.

To clarify whether the fluorescence quenching is a static or dynamic quenching process, the fluorescence decay characterization was conducted. Fig. S8 (ESI†) show the fluorescence decay profile of Mn(II)-NGQDs in the absence and presence of Hg^{2+} ions, respectively. The fluorescence decay profile of Mn(II)-NGQDs follows a single exponential fitting that indicated the presence of only one emitting species with lifetimes. However, the fluorescence decay profile of NG follows a tri-exponential fitting (Fig. S8, Table S3, ESI†), which indicates the presence of three emitting species with different lifetimes. It is possible due to the mixture of NG with nanosheets and QDs, which resulting a complex different lifetimes emission.⁵¹ The average lifetime of Mn(II)-NGQDs is 7.58 ns, which is shorter than that of NG (8.88 ns) and also support our point that the doping with Mn^{2+} can influence the morphology of NG. The lifetime data in Table S3 (ESI†) shows that the average lifetime of Mn(II)-NGQDs have not significantly change with the concentration increase of Hg^{2+} ions. It is indicated that the static quenching takes place during complex formation between Hg^{2+} ions and Mn(II)-NGQDs.^{52,53}

The quenching effect can be attributed to the chelating reaction between the functional groups (such as carboxyl, hydroxyl and amino groups) on the surface of Mn(II)-NGQDs and Hg^{2+} ion, which can decelerate the radiative recombination of the exciton of $n \rightarrow \pi^*$ transition through an effective electron transfer process, leading to the fluorescent quenching.⁵⁴ In order to further understand the binding between Mn(II)-NGQDs

and Hg^{2+} , the UV absorption spectra of Mn(II)-NGQDs without Hg^{2+} and in the presence of Hg^{2+} ($4.0 \times 10^{-6} \text{ mol L}^{-1}$) were recorded (Fig. 6f). The absorption peaks at 340 nm (due to the $n \rightarrow \pi^*$ transition) for Mn(II)-NGQDs is remarkably declined in the presence of Hg^{2+} , demonstrating the surface states of the quantum dots can be influenced by the binding of Mn(II)-NGQDs with Hg^{2+} . This means that compared to other metal ions, Hg^{2+} ion shows a more powerful affinity and faster chelating toward the functional groups on the surface of the Mn(II)-NGQDs. In addition, the photoinduced electron transfer from conduction band to the state of Hg^{2+} would be located between the conduction band and valence band of Mn(II)-NGQDs, causing the fluorescence quenching. These results also indicate that the fluorescence quenching of Mn(II)-NGQDs by Hg^{2+} is a static quenching process.

Cellular toxicity and cell fluorescent imaging

In order to evaluate the potential biological imaging application of Mn(II)-NGQDs, the biocompatibility of Mn(II)-NGQDs was evaluated *via* incubating HepG2 cells with different concentrations of Mn(II)-NGQDs for 4 and 24 h, respectively. The survival quantity of HepG2 cells was calculated based on MTT cell-viability assay. As show in Fig. 7, the cell viabilities of HepG2 cells are all close to 100% in the tested period with the concentration of Mn(II)-NGQDs from 0 to $300 \mu\text{g mL}^{-1}$, and there are almost no changes of the cells viability values of HepG2 cells incubated by Mn(II)-NGQDs with various concentrations for 4 and 24 h. It is obvious that the Mn(II)-NGQDs have no cytotoxicity to HepG2 cells. In addition, the zeta potential of Mn(II)-NGQDs and NG was also explored (Fig. S9, ESI†). Compared with NG (-15.2 eV), Mn(II)-NGQDs exhibit a more negative potential value (-19.7 eV), which indicates that Mn(II)-NGQDs will be more stable and efficiently internalized by cells.

The utilization of Mn(II)-NGQDs in cellular imaging was also investigated by confocal laser scanning microscopy (CLSM) observation. As shown in Fig. 8, the obvious blue fluorescent signal can be observed in the cytoplasm of Raw264.7 (Fig. 8a)

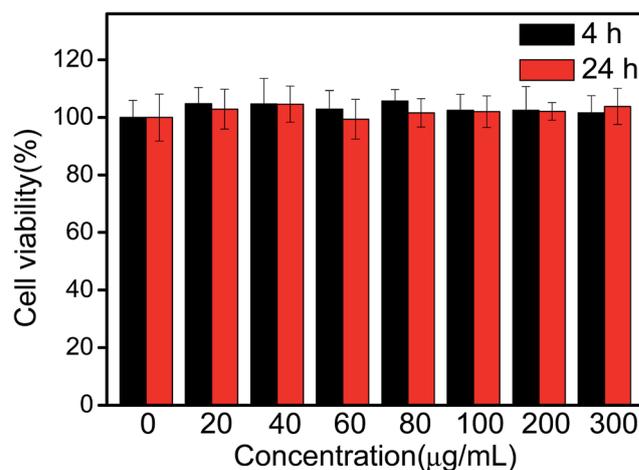


Fig. 7 Cell viability of HepG2 cells after incubation with Mn(II)-NGQDs (0–300 $\mu\text{g mL}^{-1}$) for 4 and 24 h, respectively.



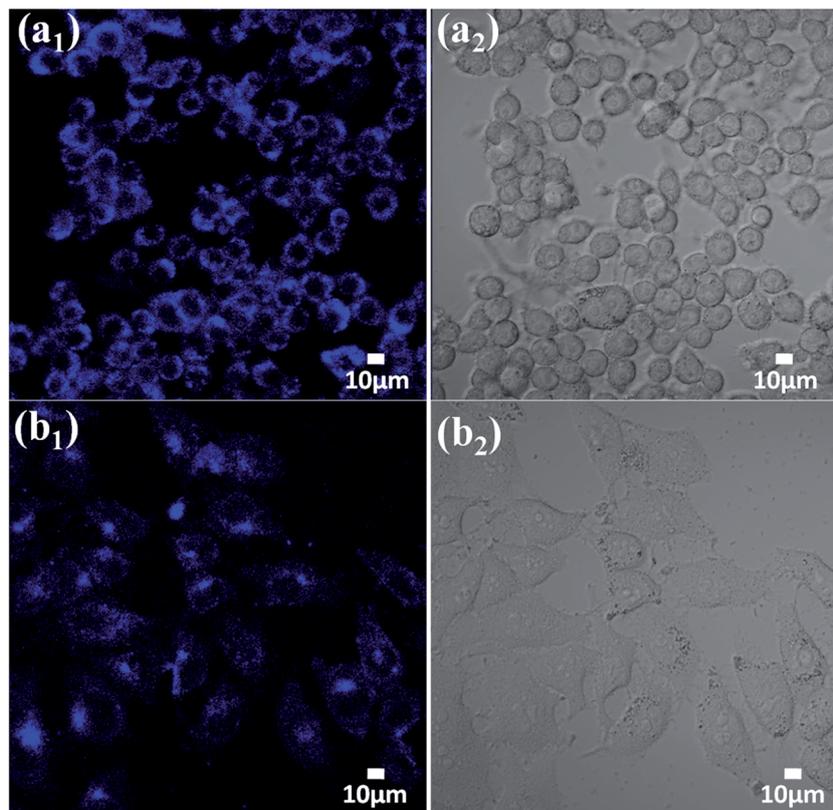


Fig. 8 CLSM images and bright field of Raw264.7 mouse macrophages (a_1 ; a_2) and HepG2 human liver cancer cells (b_1 ; b_2) incubated with Mn(II)-NGQDs ($40 \mu\text{g mL}^{-1}$) for 3 h by excitation at 405 nm.

and HepG2 cells (Fig. 8b₁) incubated with Mn(II)-NGQDs at a concentration of $40 \mu\text{g mL}^{-1}$ for 3 h, indicating that the Mn(II)-NGQDs can be efficiently internalized by Raw264.7 and HepG2 cells and facilely enter into the cells for efficiency cell imaging.^{55,56} Moreover, both the Raw264.7 (Fig. 8a₂) and HepG2 cells (Fig. 8b₂) incubated with Mn(II)-NGQDs still kept their normal morphology and strongly adhered onto cell plates, revealing the good biocompatibility of Mn(II)-NGQDs. Thus, the Mn(II)-NGQDs can be served as an efficient and non-toxic and fluorescence bioimaging probe for various biomedical applications.

Conclusion

In conclusion, we have demonstrated a simple and feasible synthesis strategy to prepare fluorescent Mn(II)-NGQDs through hydrothermal carbonization of glycine and sodium citrate with Mn^{2+} as modifier. The resulted Mn(II)-NGQDs display the morphology with uniform size and good dispersion, and show enhanced fluorescent compared with NG and NG doped with other common metal ions. The Mn(II)-NGQDs can be utilized directly as a fluorescent probe for highly sensitive and selective detection of Hg^{2+} ions. Confinement effect and radiative recombination mechanism were used to explained the fluorescence enhancement and quenching, respectively. The water-soluble Mn(II)-NGQDs also show low cytotoxicity, superior brightness and high photostability for fluorescence live cell

imaging. This work may provide a strategic approach for preparing high-quality fluorescent GQDs to meet the requirement of multifunctional application.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was financially supported by the National Natural Science Foundation of China (No. 51564009, 51468011) and the Natural Science Foundation of Guangxi Province (No. 2015GXNSFDA139035).

References

- 1 J. Liu, H. Xu, Y. Xu, Y. Song, J. Lian, Y. Zhao, L. Wang, L. Huang, H. Ji and H. Li, *Appl. Catal., B*, 2017, **207**, 429–437.
- 2 A. Subramanian, Z. Pan, G. Rong, H. Li, L. Zhou, W. Li, Y. Qiu, Y. Xu, Y. Hou, Z. Zheng and Y. Zhang, *J. Power Sources*, 2017, **343**, 39–46.
- 3 D. H. Kim and T. W. Kim, *Nano Energy*, 2017, **32**, 441–447.
- 4 L. Wang, W. Li, B. Wu, Z. Li, D. Pan and M. Wu, *Chem. Eng. J.*, 2017, **309**, 374–380.
- 5 L. Chen, G. Yang, P. Wu and C. Cai, *Biosens. Bioelectron.*, 2017, **96**, 294–299.



- 6 L. Li, G. Wu, G. Yang, J. Peng, J. Zhao and J. J. Zhu, *Nanoscale*, 2013, **5**, 4015–4039.
- 7 T. F. Yeh, W. L. Huang, C. J. Chung, I. T. Chiang, L. C. Chen, H. Y. Chang, W. C. Su, C. Cheng, S. J. Chen and H. Teng, *J. Phys. Chem. Lett.*, 2016, **7**, 2087–2092.
- 8 Y. Zhang, H. Gao, J. Niu and B. Liu, *New J. Chem.*, 2014, **38**, 4970–4974.
- 9 T. F. Yeh, C. Y. Teng, S. J. Chen and H. Teng, *Adv. Mater.*, 2014, **26**, 3297–3303.
- 10 M. Zhang, L. Bai, W. Shang, W. Xie, H. Ma, Y. Fu, D. Fang, H. Sun, L. Fan, M. Han, C. Liu and S. Yang, *J. Mater. Chem.*, 2012, **22**, 7461.
- 11 A. H. Castro Neto, F. Guinea, N. M. R. Peres, K. S. Novoselov and A. K. Geim, *Rev. Mod. Phys.*, 2009, **81**, 109–162.
- 12 Z. Fan, S. Li, F. Yuan and L. Fan, *RSC Adv.*, 2015, **5**, 19773–19789.
- 13 Y. Zhou, X. Jing and Y. Chen, *J. Mater. Chem. B*, 2017, **5**, 6451–6470.
- 14 B. Shi, Y. Su, L. Zhang, M. Huang, X. Li and S. Zhao, *Nanoscale*, 2016, **8**, 10814–10822.
- 15 A. Muthurasu, P. Dhandapani and V. Ganesh, *New J. Chem.*, 2016, **40**, 9111–9124.
- 16 Q. Zhuang, Y. Wang and Y. Ni, *Luminescence*, 2016, **31**, 746–753.
- 17 T. V. Tam, N. B. Trung, H. R. Kim, J. S. Chung and W. M. Choi, *Sens. Actuators, B*, 2014, **202**, 568–573.
- 18 T. Alizadeh and M. Shokri, *Sens. Actuators, B*, 2016, **222**, 728–734.
- 19 Y. Dong, J. Shao, C. Chen, H. Li, R. Wang, Y. Chi, X. Lin and G. Chen, *Carbon*, 2012, **50**, 4738–4743.
- 20 W. Zhang and J. Gan, *Appl. Surf. Sci.*, 2016, **372**, 145–151.
- 21 H. X. Zhao, Y. C. Wang, L. Y. Zhang and M. Wang, *New J. Chem.*, 2015, **39**, 98–101.
- 22 S. Yang, J. Sun, C. Zhu, P. He, Z. Peng and G. Ding, *Analyst*, 2016, **141**, 1052–1059.
- 23 U. Baruah and D. Chowdhury, *Nanotechnology*, 2016, **27**, 145501.
- 24 S. Kundu, R. M. Yadav, T. N. Narayanan, M. V. Shelke, R. Vajtai, P. M. Ajayan and V. K. Pillai, *Nanoscale*, 2015, **7**, 11515–11519.
- 25 H. Xu, S. Zhou, L. Xiao, Q. Yuan and W. Gan, *RSC Adv.*, 2016, **6**, 36554–36560.
- 26 X. Zhu, G. Wu, N. Lu, X. Yuan and B. Li, *J. Hazard. Mater.*, 2017, **324**, 272–280.
- 27 S. Vempati, A. Celebioglu and T. Uyar, *Nanoscale*, 2015, **7**, 16110–16118.
- 28 G. S. Kumar, R. Roy, D. Sen, U. K. Ghorai, R. Thapa, N. Mazumder, S. Saha and K. K. Chattopadhyay, *Nanoscale*, 2014, **6**, 3384–3391.
- 29 S. H. Jin, D. H. Kim, G. H. Jun, S. H. Hong and S. Jeon, *ACS Nano*, 2013, **7**, 1239–1245.
- 30 K. Paek, H. Yang, J. Lee, J. Park and B. J. Kim, *ACS Nano*, 2014, **8**, 2848–2856.
- 31 Z. Gan, H. Xu and Y. Hao, *Nanoscale*, 2016, **8**, 7794–7807.
- 32 C. A. T. Toloza, S. Khan, R. L. D. Silva, E. C. Romani, D. G. Larrude, S. R. W. Louro, F. L. Freire and R. Q. Aucélio, *Microchem. J.*, 2017, **133**, 448–459.
- 33 D. Yu, X. Zhang, Y. Qi, S. Ding, S. Cao, A. Zhu and G. Shi, *Sens. Actuators, B*, 2016, **235**, 394–400.
- 34 C. Han, H. Xu, R. Wang, K. Wang, Y. Dai, Q. Liu, M. Guo, J. Li and K. Xu, *J. Mater. Chem. B*, 2016, **4**, 5798–5802.
- 35 X. Guo, C. F. Wang, Z. Y. Yu, L. Chen and S. Chen, *Chem. Commun.*, 2012, **48**, 2692–2694.
- 36 T. F. Yeh, F. F. Chan, C. T. Hsieh and H. Teng, *J. Phys. Chem. C*, 2011, **115**, 22587–22597.
- 37 H. Abdolmohammad-Zadeh and E. Rahimpour, *Sens. Actuators, B*, 2016, **225**, 258–266.
- 38 B. Karimi and B. Ramezanzadeh, *J. Colloid Interface Sci.*, 2017, **493**, 62–76.
- 39 Z. Yan, X. Qu, Q. Niu, C. Tian, C. Fan and B. Ye, *Anal. Methods*, 2016, **8**, 1565–1571.
- 40 L. Li, L. Li, C. Wang, K. Liu, R. Zhu, H. Qiang and Y. Lin, *Microchim. Acta*, 2014, **182**, 763–770.
- 41 J. Gliniak, J. H. Lin, Y. T. Chen, C. R. Li, E. Jokar, C. H. Chang, C. S. Peng, J. N. Lin, W. H. Lien, H. M. Tsai and T. K. Wu, *ChemSusChem*, 2017, **10**, 3260–3267.
- 42 E. Yazici, S. Yanik and M. B. Yilmaz, *Carbon*, 2017, **111**, 822–827.
- 43 Z. X. Gan, S. J. Xiong, X. L. Wu, C. Y. He, J. C. Shen and P. K. Chu, *Nano Lett.*, 2011, **11**, 3951–3956.
- 44 Y. Liu, C. Y. Liu and Y. Liu, *Appl. Surf. Sci.*, 2011, **257**, 5513–5518.
- 45 P. Hou, Y. Long, J. Zhao, J. Wang and F. Zhou, *Spectrochim. Acta, Part A*, 2012, **86**, 76–79.
- 46 Z. Xiaoyan, L. Zhangyi and L. Zaijun, *Spectrochim. Acta, Part A*, 2017, **171**, 415–424.
- 47 X. Zuo, H. Zhang, Q. Zhu, W. Wang, J. Feng and X. Chen, *Biosens. Bioelectron.*, 2016, **85**, 464–470.
- 48 Z. Wu, M. Feng, X. Chen and X. Tang, *J. Mater. Chem. B*, 2016, **4**, 2086–2089.
- 49 R. Zhang and W. Chen, *Biosens. Bioelectron.*, 2014, **55**, 83–90.
- 50 J. He, H. Zhang, J. Zou, Y. Liu, J. Zhuang, Y. Xiao and B. Lei, *Biosens. Bioelectron.*, 2016, **79**, 531–535.
- 51 S. Barman and M. Sadhukhan, *J. Mater. Chem. C*, 2012, **22**, 21832.
- 52 Y. Zhang, P. Cui, F. Zhang, X. Feng, Y. Wang, Y. Yang and X. Liu, *Talanta*, 2016, **152**, 288–300.
- 53 Y. Wang, Q. Chang and S. Hu, *Sens. Actuators, B*, 2017, **253**, 928–933.
- 54 X. Mao, H. Su, D. Tian, H. Li and R. Yang, *ACS Appl. Mater. Interfaces*, 2013, **5**, 592–597.
- 55 Z. Fan, Y. Li, X. Li, L. Fan, S. Zhou, D. Fang and S. Yang, *Carbon*, 2014, **70**, 149–156.
- 56 F. Yuan, L. Ding, Y. Li, X. Li, L. Fan, S. Zhou, D. Fang and S. Yang, *Nanoscale*, 2015, **7**, 11727–11733.

