


Cite this: *RSC Adv.*, 2018, 8, 311

Recent advances in green reagents for molecularly imprinted polymers

Xi Wu, ^{†*a} Jiajun Du,^{†b} Mengyao Li,^a Lintao Wu,^a Chun Han^a and Feng Su^a

Molecularly imprinted polymers (MIPs) are tailor-made materials with special binding sites. They have been extensively used in various areas because of its maturity. MIP is usually prepared by using a variety of "traditional" organic substances. In recent years, green reagents, such as room temperature ionic liquids (RTILs) and deep eutectic solvents (DESSs), have been increasingly used in the preparation of MIPs. These new green reagents are applied for various purposes, *i.e.* to reduce pollution, to lower the cost and to improve the performance of materials. The RTILs- and DESSs-based MIPs have attracted more and more attention because of their superior performance and interesting properties. We hereby propose to review recent advances in the application of RTILs and DESSs to MIPs. The current status and perspectives of RTILs- and DESSs-based MIPs are discussed in this brief review.

Received 7th October 2017
Accepted 14th December 2017

DOI: 10.1039/c7ra11047b

rsc.li/rsc-advances

1. Introduction

After decades of development, molecularly imprinted technology (MIT) has become a mature technique to prepare polymers with specific binding sites at the molecular level.^{1,2} This makes molecularly imprinted polymers (MIPs), tailor-made polymers, have excellent recognition ability to templates and their structural analogues. MIPs show many outstanding properties, such as low cost, good mechanical property, chemical stability and resistance to a wide range of pH, solvents, and temperatures. Hence, MIPs have been found applicable in many areas such as chemistry and biology for several years, and widely used in pretreatment of complicated samples in a variety of ways.^{3,4} MIPs can be synthesized by a variety of substances. Generally speaking, MIPs synthesis protocols must contain following essential ingredients: template, functional monomer, cross-linker, porogen (solvent) and initiator. There are two typical ways to prepare MIPs on the basis of interactions (covalent/non-covalent) between template and functional monomer.^{5,6} Owing to less restrictions and flexible application, non-covalent imprinting is far more popular than covalent imprinting. Normal procedures for the preparation of MIPs can be described as polymerization of functional monomer and cross-linker around a template in porogen (solvent). The general scheme for preparation of MIPs was shown in Fig. 1. To get MIPs with good performance in a particular environment, a good synthesis scheme is a prerequisite. Obviously, the first

question to consider for MIPs preparation is the type of formulation components for MIPs. Up till now, the number of functional monomers, cross-linkers and porogens used in the preparation of MIPs is relatively limited.⁷ By contrast, the type of templates is multifarious, which includes ions, organic molecules, biomacromolecules and so on. It is implied that exploration of new polymer system is very important to enlarge the application fields of MIPs.

Room temperature ionic liquids (RTILs) can be considered as molten salts with melting points close to room temperature.⁸ It is a new type of reagents with unique and interesting characteristics, such as non-volatility, non-flammability, high ion density, good ionic conductivity, and enhanced dispersibility in inorganic/organic solvents. Hence, RTILs have received extensive attention from chemistry research workers. For example, RTILs can be applied in sample preparation,⁹ chromatography,¹⁰ capillary electrophoretic separation¹¹ and reaction media.¹² As a result, RTILs become popular in the molecular imprinting field. At present, owing to their low vapour pressure and high boiling point that facilitates their recycling, RTILs are qualified as green solvents.¹³ However, some reports show that most RTILs have hazardous toxicity and poor biodegradability.^{14,15} In addition, RTILs are very expensive. These drawbacks limit the application of RTILs to some extent.

To overcome the shortcomings of RTILs, deep eutectic solvents (DESSs) have emerged¹⁶ as a new generation of green solvents. A DES is usually composed of two components through hydrogen bond interaction.¹⁷ The two components are defined as hydrogen bond acceptor (HBA) and hydrogen bond donor (HBD), respectively. Especially since choline chloride (ChCl) has the advantages of low toxicity, biodegradability, being cheap and readily available, it has become one of the most popular HBAs among existing DESSs.^{18–20} For a ChCl-based

^aDepartment of Chemistry, Changzhi University, Changzhi 046011, China. E-mail: wuxi040252768@163.com; Fax: +86-355-2178113; Tel: +86-355-2178113

^bDepartment of Medical Information, Chinese PLA General Hospital, 100853, Beijing, China

[†] These authors contributed equally to this work.



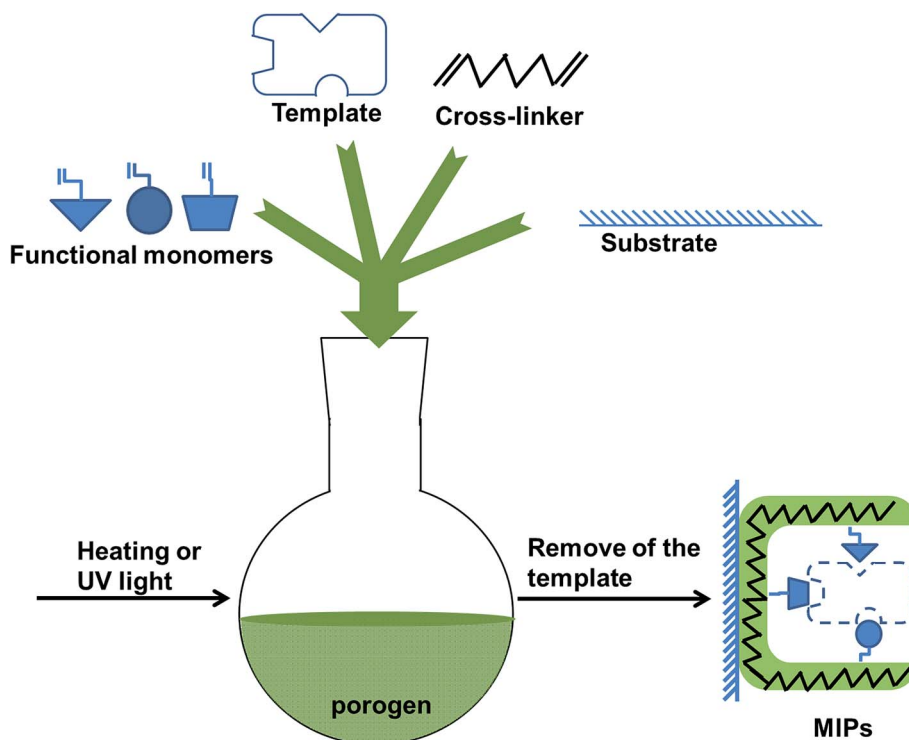


Fig. 1 The general scheme for preparation of MIPs.

DES, the most common HBDs are urea, amines, amides, alcohols and organic acids. Furthermore, it is worth pointing out that the DES preparation process is simple and environment-friendly. In a word, DESs share characteristics with RTILs and are more inexpensive and safe. Hence, application of DESs in the field of chemistry has been a hot research topic. Until now, only a small number of investigations on the application of DESs in molecular imprinting have been performed.

Green reagents as RTILs and DESs have been increasingly employed in MIP synthesis protocols and play several different roles in the MIPs preparation process. In current studies, RTILs and DESs can be used as functional monomer, porogen, pore template and so on. Relevant MIPs show excellent performance in many ways. For example, MIPs based on RTILs or DESs have excellent recognition ability to template in aqueous environment.^{21,22} To find green reagents as replacements of conventional organic solvents has become trends of MIT research. Hence, we focus on the application of RTILs and DESs in preparing MIPs in this brief review. Furthermore, we also focus our interest on the performance of RTILs and DESs-based MIPs in this paper. The assessment of MIPs performance is a significant work and there are many evaluation methods of imprinted materials. For example, the morphologies of MIPs are usually characterized by scanning electron microscopy (SEM) and transmission electron microscopy (TEM). The selectivity, specificity and binding capacity of the imprinted materials can be investigated *via* static equilibrium adsorption experiments and absorption kinetic experiments. In present review, the studies on green reagents-based MIPs and current trends in the area are discussed.

2. RTILs as functional monomer for MIPs

During the process of preparation of MIPs for a specific template, selection of functional monomer is very critical to getting good imprinting effect. So far as we know, small organic molecules, metal ions and biomolecules can be potential templates. However, the type of functional monomer is quite limited. Hence, the investigation of new functional monomers is important for the future development of MIPs. Researches in recent years show that RTILs can interact with multiple molecules such as organic compounds and biomacromolecules by hydrogen bonding, anion-exchange, electrostatic, hydrophobic and π - π interactions.^{23,24} Regarded as “designer solvents”, RTILs offer a large degree of flexibility, which makes them suitable for different purposes in MIT. Related researches have shown that RTIL-based MIPs have significant performance in aqueous media.

Wang *et al.* proposed a new method of using vinyl-imidazolium RTIL as a functional monomer for synthesis of chlorsulfuron MIPs.²¹ In this study, 1-vinyl-3-butylimidazolium chloride ([VBIM]Cl) was first taken as a unique functional monomer as the authors claimed. The novel MIP material was synthesized by bulk polymerization and had shown excellent selectivity and good adsorption/desorption for template. The binding selectivity of the prepared MIPs was evaluated by competitive adsorption using the mixture solution composed of chlorsulfuron and its analogues. The new MIP material showed excellent selectivity of 47.2% for chlorsulfuron, which was higher than that for the analogues. Moreover, it is worth



mentioning that low concentration template can be detected by the new material in aqueous medium.

Similarly, a high capacity hollow porous dummy MIPs was fabricated using RTIL as functional monomer by Shi *et al.*²⁵ The hollow MIPs were prepared using benzoic acid as dummy template, 1-vinyl-3-methylimidazolium chloride ([VMIM]Cl) as functional monomer for recognition of salicylic acid in *Actinidia chinensis* extract. The highest adsorption capacity (29.75 mg g^{-1}) and imprinting factor (IF) (5.61) of the hollow MIPs can be achieved with template : functional monomer : cross-linker at 1 : 4 : 20. The effect of copolymerization with “traditional” functional monomer (4-vinylpyridine, 4-VP) was also tested. The adsorption capacity of template on MIPs decreased to 22.61 mg g^{-1} when 4-VP was used as functional monomer. In other words, [VMIM]Cl was more efficient than “traditional” functional monomer in this study. The results proved that the resulting MIPs can selectively extraction of trace salicylic acid from complex fruit sample more effectively than MIPs prepared *via* a conventional formula.

Furthermore, four specific ionic liquids were prepared and used as functional monomers for synthesis of synephrine MIPs by Fan *et al.*²⁶ The molecular structure of the novel functional monomers named Brønsted acidic imidazolium-typed RTILs was shown in Fig. 2. Obviously, the significant difference among the four RTILs is the side-chain (carboxylic acid group) length. The results showed that MIP using 1-vinyl-3-carboxybutylimidazolium bromide ([COOHavim]Br) as the monomer had the best adsorption capacity. The authors suggest that interactions between synephrine and RTIL functional monomers are not only the π - π interactions between the phenyl group (synephrine) and imidazolyl (RTILs) but also the hydrogen bond interactions between the amine/hydroxyl group (synephrine) and the carboxylic acid group (RTILs). Moreover, the MIPs prepared with RTIL functional monomers showed good adsorption properties for synephrine in methanol aqueous solution, which is significant for enlarging the applications of MIP in aqueous medium.

The effect of different functional groups on RTILs was also investigated by Row *et al.*²⁷ In their study, the RTIL-modified porous polymers were prepared for effective separation of

three tanshinones from *Salvia miltiorrhiza* Bunge. The preparation process mainly consists of two steps: first, blank polymer was synthesized by using 4-(chloromethyl) styrene as monomer, divinylbenzene (DVB) as cross linker, heptane as porogen and polyvinylpyrrolidone (PVP) ethanol solution as dispersant, respectively. Then, taking 9,10-phenanthrenequinone and RTILs as the template and functional monomers, the IL-modified polymers were obtained by surface imprinting method. Five RTILs were used to modify porous polymers and the chemical structures of all RTILs were shown in Fig. 3. The interactions between three tanshinones and the five resulting polymers, imidazole polymer, methylimidazole polymer, carboxyl-imidazole polymer, amino-imidazole polymer, cyano-imidazole polymer, were investigated. The results showed that the carboxyl-imidazole polymer possessed the best performance. Under optimized conditions, 0.35 mg g^{-1} of cryptotanshinone, 0.33 mg g^{-1} of tanshinone I, and 0.27 mg g^{-1} of tanshinone IIA were obtained from plant extract. This research provides further evidence that functional groups on RTILs have played a significant role in the formation of template-functional monomer complex.

To date, researches are very active on the imprinting of small organic molecules and a large number of relevant studies are published every year. By contrast, preparation of MIPs for large molecules such as polypeptide, enzyme and proteins has been seen as one of the toughest challenges in MIT. One of the reasons is lack of suitable functional monomer for large molecules. The introduction of RTILs is an alternative way to solve this problem. Thymopentin (TP5) is considered as a bio-macromolecule that consists of five amino acids. It is difficult to extract and purify TP5 from complex biological samples. Moreover, there are many different kinds of RTILs and it is difficult to find the best RTIL functional monomer for a certain template. In this case, with the help of computational simulation, Wang *et al.* proposed an approach to synthesize TP5 magnetic MIPs by surface-initiated ATRP polymerization.²⁸ TP5, ethylene glycol dimethacrylate (EDMA), CuCl, Fe_3O_4 -Br and *N,N,N',N',N''*-pentamethyldiethylene triamine (PMDTA) were used as template, cross-linker, catalyst, initiators and ligand, respectively. Four RTILs, 1-vinyl-3-butyl imidazolium chloride

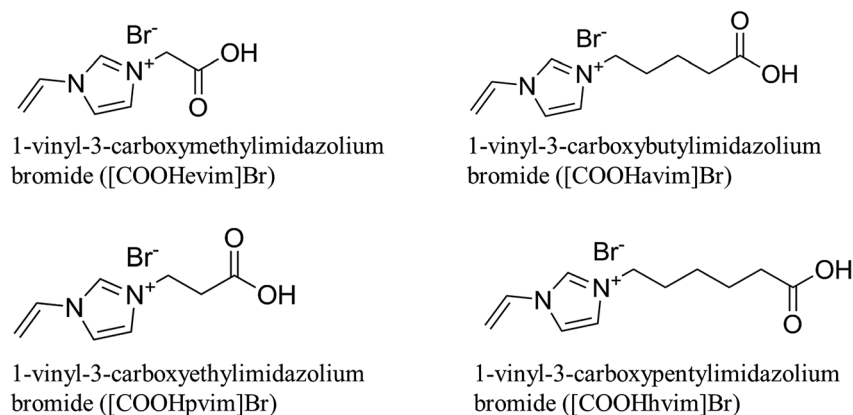


Fig. 2 The molecular structure of the new RTIL functional monomers.²⁶



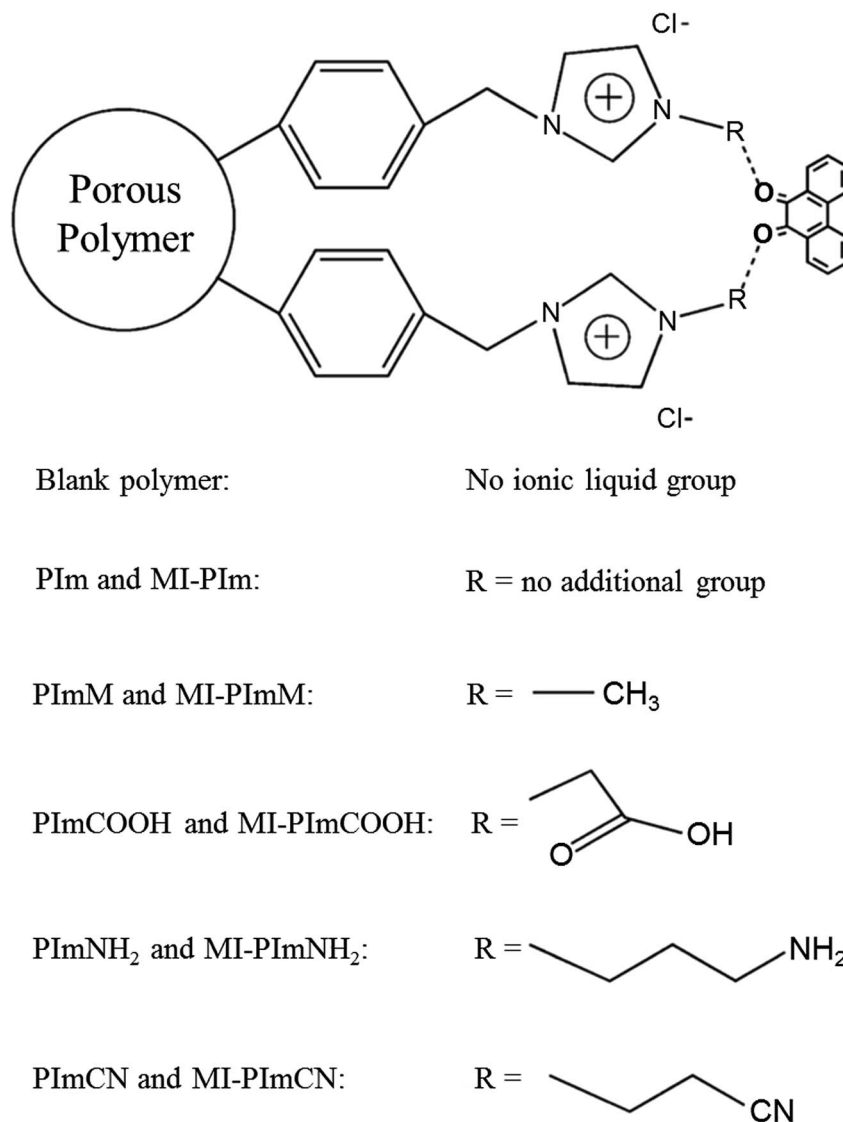


Fig. 3 The structures of all RTILs and the relevant porous polymers. Reprinted from ref. 27.

([VBIM]Cl), 1-vinyl-3-propyl imidazolium chloride ([VPIM]Cl), 1-vinyl-3-ethyl acetate imidazolium chloride and 1-vinyl-3-ethanamide imidazolium chloride, were investigated as candidate functional monomers. In this study, to find the best scheme of imprinting protocol, molecular dynamics (MD) simulations which was frequently used for computational design of MIPs was employed for researching the interactions between template and functional monomer.²⁹ The interaction energies of all candidate functional monomers with TP5 were obtained by MD simulation. In the simulation, 1 : 1 template-monomer complex in water was taken as a simulated system to reduce computation load. The binding energy (ΔE) of template with each RTIL functional monomer was obtained according to the following formula:

$$\Delta E = E(\text{template-functional monomer}) - E(\text{template}) - E(\text{functional monomer}) \quad (1)$$

In the formula, $E(\text{template-functional monomer})$, $E(\text{template})$ and $E(\text{functional monomer})$ were potential energy of the simulated system, potential energy of the template and potential energy of the functional monomer, respectively. The results showed that the binding energy of TP5 with [VBIM]Cl, [VPIM]Cl, 1-vinyl-3-ethanamide imidazolium chloride and 1-vinyl-3-ethyl acetate imidazolium chloride was -16.02 , -18.80 , -22.18 and -24.37 , respectively. It demonstrated that 1-vinyl-3-ethyl acetate imidazolium chloride had higher interaction energy with template. It also proved that 1-vinyl-3-ethyl acetate imidazolium chloride was the best functional monomer for TP5. The optimized template-functional monomer structure was shown in Fig. 4. Then, the TP5 magnetic MIPs were prepared according to the results of simulation. The results showed that the IF of TP5 magnetic MIPs can reach 1.76. The morphological structures of magnetic materials were characterized by TEM and the magnetic MIPs had regular cavities and



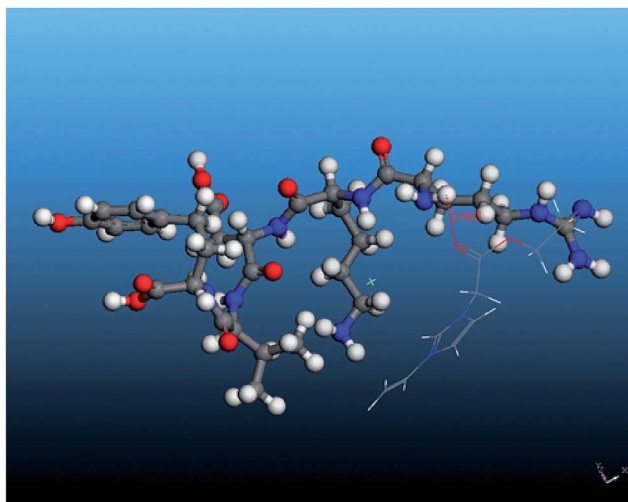


Fig. 4 The optimized structure of the template-functional monomer complex (1 : 1) formed between TP5 and RTIL. Reprinted from ref. 28.

small size. It means that the RTIL-based magnetic MIPs have good specific recognition to TP5.

Furthermore, L-phenylalanine imprinted microspheres were prepared by the same group.³⁰ In this study, a new amino acid RTIL named 1-butyl-3-vinylimidazolium amino hydrocinnamic acid ([BVIM][Phe]) was synthesized and used not only as a functional monomer but also as a dummy template. Similar to the prior research,²⁸ a theoretical computational method was taken to find the template-functional monomer complex with the highest binding energy. In this study, surface-initiated RAFT polymerization was employed for the preparation of PheIL-MIPs. For the comparison of selectivity of the obtained PheIL-MIPs, the traditional L-Phe MIPs were prepared with 4-VP as the functional monomer under the optimized condition of PheIL-MIPs. The amount of L-Phe bound to PheIL-MIPs and Phe-MIPs was approximately 53 and 28 $\mu\text{mol g}^{-1}$ and the

corresponding IFs were approximately 2.5 and 1.4, respectively. The results demonstrated that the proposed PheIL-MIPs has good affinity and selectivity for L-phenylalanine by using [BVIM][Phe] as functional monomer and dummy template.

RTILs were introduced to prepare bovine serum albumin (BSA) MIPs with an effective molecular interaction between RTILs and BSA by Liu *et al.*³¹ In this study, a MWCNTs@BSA-MIPILs film was synthesized by sol-gel technology. Alkoxy-functionalized RTIL 1-(3-trimethoxysilyl propyl)-3-methyl imidazolium chloride ([TMSPMIM]Cl) was used as monomer and multiwall carbon nano-tubes (MWCNTs) was taken as substrate. The synthesis scheme of the MWCNTs@BSA-MIPILs was shown in Fig. 5. The polymer showed good binding ability to BSA and its analogues under optimal conditions. The IFs for BSA, human serum albumin (HAS) and bovine hemoglobin (Bhb) were 5.84, 2.24 and 1.04 at pH 9.9, respectively. It is worth reminding that the prepared MWCNTs@BSA-MIPILs film is pH-responsive. The result means that the functionalized RTIL can not only be used as functional monomer but also hydrolysis catalyst in sol-gel system.

Qian *et al.* designed a new RTIL named 1-vinyl-3-amino formylmethyl imidazolium chloride ([VAFMIM]Cl) and used it to prepare BSA molecular imprinted hydrogels (MIHs).³² In this study, RTIL acted both as the co-monomer and stabilizer. More specifically, [VAFMIM]Cl has two functional groups (amide group and imidazolium ring) with a positive charge which can interact with the functional groups of BSA. Hence, it was utilized as monomer along with *N*-isopropylacrylamide (NIPA) in the study. Furthermore, the influence of [VAFMIM]Cl on the stability of BSA was investigated by circular dichroism. It is indicated that Cl^- in [VAFMIM]Cl is responsible for the stability of BSA. Moreover, the results also indicated that the MIHs composed of RTIL exhibited better recognition ability compared to the conventional ones. The IF value of MIH based on [VAFMIM]Cl could reach 2.66, whereas, the MIH based on the conventional monomers, 2-(dimethylamino)ethyl-methacrylate (DMAEMA), has a relatively lower IF value (1.56).

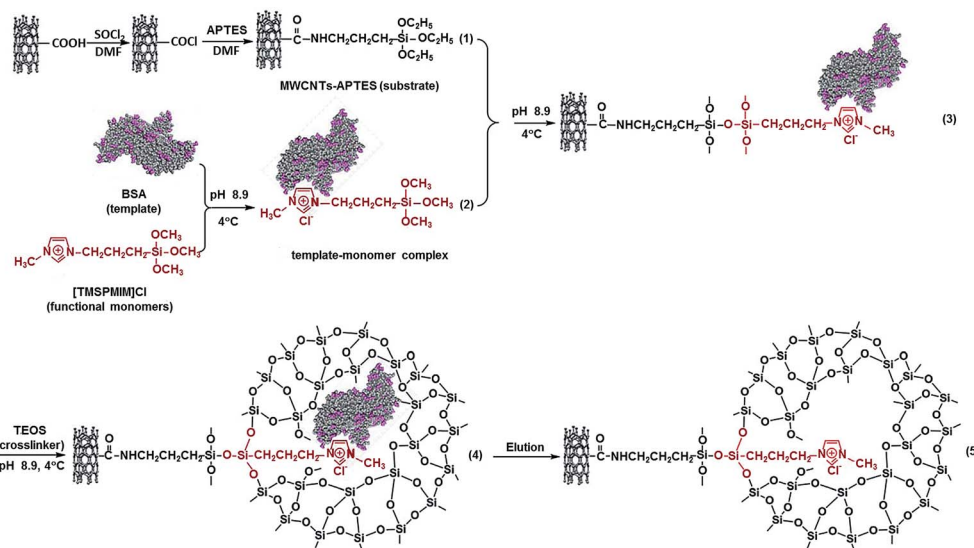


Fig. 5 The synthesis scheme of the MWCNTs@BSA-MIPILs. Reprinted from ref. 31.



Furthermore, the lysozyme-MIPs were prepared by Wang and co-workers³³ with [VBIM]Cl as functional monomer, lysozyme as template, *N,N'*-methylenebisacrylamide (MBA) and MWCNTs as cross-linker and substrate. To prepare lysozyme-imprinted polymer, [VBIM]Cl, 1-vinyl-3-octyl imidazolium chloride ([VOIM]Cl) and vinyl imidazole were studied as candidate functional monomers in this work. The results demonstrated that the [VBIM]Cl-based MIPs had a higher IF in all tested MIPs. The above results imply that RTILs can be used as functional monomer to improve recognition performance of MIPs and it will have a wonderful prospect in the field of large molecule imprinting. More reports about successful applications of RTILs in MIPs are shown in Table 1.

3. RTILs as porogens for MIPs

Porogens play an important role in a scheme for MIPs. For preparation of MIPs, all the components in the scheme should be dissolved in porogens at first and template-monomer complex will form by covalent or non-covalent interactions. It means that the porogens are chosen according to their solubility and its effect on the template-monomer complex formation. Interestingly, MIPs prepared with RTILs as porogens have several special characteristics.

According to the previous study,²¹ MIPs prepared with RTIL as functional monomer have impressive performance in aqueous medium. Yan *et al.* proposed a new method to prepare dicofol MIPs with a novel synthesized RTIL (1-allyl-3-methyl imidazolium bromide, [AMIM]Br) as porogen.⁴⁴ The results showed that RTIL played a key function in improving performance of MIPs. On one hand, the Cl[−] of template could be interacted with the imidazole with positive charge of RTIL by electrostatic interaction. Meanwhile, cross linking could take place between RTIL and functional monomer by using DVB. Hence, RTIL can help to form the specific binding sites in the process of MIPs preparation and the degree of shrinking or swelling of MIPs can be reduced. It is verified that the RTIL-MIPs possess superior properties in polar conditions, and efficient and selective extraction of dicofol from vegetable sample can be achieved by the proposed method. The results showed that the recoveries of dicofol at the spiked levels of 2.3, 23.2, and 232.5 ng g^{−1} ranged from 86.6% to 101.9% with RSDs of less than 6.5%. Liu *et al.* also confirmed that shrinking or swelling of MIPs could be reduced by using RTIL as porogens.⁴⁵ They developed a new MIPs chiral stationary phase for HPLC. The MIP monoliths were prepared with *R*-mandelic acid as template, 4-VP as functional monomer, EDMA as cross-linker, metal ions as metal pivot and a mixed solvent as porogen. The porogenic system contains 1-butyl-3-methyl imidazolium tetrafluoroborate ([BMIM][BF₄]), sulfoxide and dimethylformamide. The authors noted that the use of porogen without RTIL resulted in an MIP monolith with cracks. It indicates that the RTILs have significant influence on MIPs morphology.

It is clear that, as a new porogen, RTILs can influence the MIPs system. However, most studies failed to make a thorough exploration of the effect mechanisms of RTIL. Taking propranolol as a model template, McCluskey *et al.* made

a beneficial trial to explain the functions of the RTILs in the process of synthesis MIPs.⁴⁶ Four RTILs, [BMIM][BF₄], 1-butyl-3-methyl imidazolium hexafluorophosphate ([BMIM][PF₆]), 1-hexyl-3-methyl imidazolium hexafluorophosphate ([HMIM][PF₆]), 1-octyl-3-methyl imidazolium hexafluorophosphate ([OMIM][PF₆]) and one traditional organic solvent, CHCl₃, were taken as porogens for propranolol MIPs and the effect of the RTILs on MIP characters and performance was investigated. The results showed that MIPs prepared with CHCl₃ (MIP_{CHCl₃}) have higher imprinting selectivity (IF = 4.64) than MIPs prepared with [BMIM][PF₆] (MIP_{PF₆}) (IF = 1.98). The differences between NIP/MIP_{CHCl₃} and NIP/MIP_{PF₆} in physicochemical properties were investigated by means of several analysis methods, such as Brunauer-Emmett-Teller (BET) specific surface area, BET pore volume, positron annihilation lifetime spectroscopy (PALS) diameter of small/large pores, relative number of small/large pores and zeta potential. They found some interesting phenomenons in this study. For example, the polymer morphology can be influenced by template only in the traditional organic solvent systems. The BET specific surface areas of MIP_{CHCl₃} and NIP_{CHCl₃} were 306 and 509 m² g^{−1}. In contrast, the surface areas of MIP and NIP were essentially identical in the RTIL systems (185 and 180 m² g^{−1}). Furthermore, the investigation demonstrated that the swelling ratios of MIPs prepared by [BMIM] and [HMIM] based systems were 34% and 33%, respectively. Meanwhile, the [OMIM][PF₆]-prepared MIP showed only 6% swelling. The thermal stability of the polymers was increased in the order of [HMIM] ≈ [OMIM] > [BMIM]. It means that the swelling ratio of MIPs decreased and the thermal stability of MIPs increased with increasing alkyl chain length of RTILs. In addition, rebinding studies show that the MIP prepared with OMIM system has a higher IF (2.1) than the MIP prepared with HMIM system (IF = 1.1).

Moreover, Liu *et al.* proposed a new porogenic system based on RTIL to increase the affinity of MIPs.⁴⁷ The porogen was a ternary mixture of [BMIM][BF₄], dimethyl sulfoxide and macromolecular crowding agent (polymethyl methacrylate) chloroform solution. In this study, with oleanic acid as template, 4-VP as functional monomer, EDMA as cross-linker, a MIP monolith was prepared in the novel porogenic solvent. The type of RTILs is one important polymerization variable and the influence of [BMIM][BF₄], [BMIM][PF₆], 1-butyl-3-methyl imidazolium hydrogen sulfate ([BMIM][HSO₄]) and 1-octyl-3-methyl imidazolium tetrafluoroborate ([OMIM][BF₄]) on the performance of MIPs were investigated. The results were shown in Fig. 6. It is clear that the MIPs prepared with [BMIM][BF₄]-based porogen has the best affinity to the template. Compared with MIPs made with the [BMIM][BF₄] porogen system, the selectivity of the [OMIM][BF₄] porogen-based MIPs is limited. It is suggested that the length of the cation alkyl chain may have a significant effect on the interactions between the template and functional monomer. Moreover, it should be noted that the anions in the four RTILs also have an influence on the selectivity of the MIPs. The MIP with [HSO₄][−] in place of [BF₄][−] in [BMIM][BF₄] showed no selectivity for the template (IF = 1.06). The results showed that the IF value of the polymers decreased in the order of [HSO₄][−] < [PF₆][−] < [BF₄][−].



Table 1 Partial reports about the application of RTIL functional monomers in MIPs

RTIL	Protocol of preparing MIPs	Comments	Ref.
1-(Triethoxysilyl) propyl-3-aminopropylimidazole bromide (SilprImN)	Dummy template: BPA Functional monomer: SilprImN Cross-linker: TEOS Supported material: activated silica gel Solvent: tetrahydrofuran/methanol	By a sol-gel process, the MIPs was prepared for the simultaneous determination of nine organochlorine pesticides in environmental and food samples	34
1-Allyl-3-ethylimidazolium bromide ([AEIM]Br)	Dummy template: phenylephrine Co-monomer: [AEIM]Br/MAA Cross-linker: EDMA Solvent: chloroform Dispersant: PVP aqueous solution	A dummy molecularly imprinted microspheres (DMIMs) were synthesized for the determination of clenbuterol and clorprenaline in urine by aqueous suspension polymerization	35
1-Allyl-3-ethylimidazolium hexafluorophosphate ([AEIM][PF ₆])	Dummy template: 4,4-dichlorobenzhydrol Co-monomer: [AEIM][PF ₆]/MAA Cross-linker: EDMA Magnetic material: Fe ₃ O ₄ @oleic acid Solvent: chloroform Dispersant: PVP aqueous solution	By aqueous suspension polymerization, a magnetic molecularly imprinted microspheres was synthesized for determination of organochlorine pesticides in environmental water	36
1-Vinyl-3-ethyl acetate imidazolium chloride ([VEAIM]Cl)	Template: thymopentin Functional monomer: [VEAIM]Cl Cross-linker: MBA Ligand: PMDETA Catalyst: CuCl Solvent: water	By surface-initiated ATRP in aqueous solution, a molecularly imprinted membranes (MIMs) was proposed for isolation and purification of crude thymopentin. The MD simulations were performed for simulating the optimized imprinting protocol	37
1-Butyl-3-vinyl imidazolium α -aminohydrocinnamic acid salt ([BVIM][Phe])	Dummy template/functional monomer: [BVIM][Phe] Co-monomer: 4-VP Supported material: PDVB microspheres Cross-linker: EDMA Solvent: acetonitrile–water	By a RAFT approach, IL-based L-Phe MIPs was prepared for determination of L-Phe in human serum. The polymerizable IL, [Bvim][Phe], was synthesized and used as dummy template and functional monomer. The MD simulations were utilized for evaluating the template–functional monomer complex	38
1-Allyl-3-methyl imidazolium bromide ([AMIM]Br)	Dummy template: FHO Co-monomer: [AMIM]Br/acrylamide Cross-linker: DVB Solvent: acetonitrile	By precipitation polymerization, an IL-based MIPs was utilized as a selective adsorbent for extraction and determination of CFO in urine	39
1-Allyl-3-methyl imidazolium X, (X = X = Cl [−] , PF ₆ [−] , CF ₃ SO ₃ [−] , BF ₄ [−]) ([AMIM]Cl, [AMIM][BF ₄], [AMIM][PF ₆], [AMIM][CF ₃ SO ₃])	Template: BSA Functional monomer: [AMIM][X](X = Cl [−] , PF ₆ [−] , CF ₃ SO ₃ [−] , BF ₄ [−]) Cross-linker: NNMB Substrate: AAm modified MWCNTs Solvent: phosphate buffered solution Catalyst: TEMED	By free radical polymerization and surface imprinting technique, a biocompatible MWCNTs@BSA-MIPILs with satisfactory selective recognition ability for BSA was prepared. Four ILs with different anion species as monomers were investigated and the materials prepared with PF ₆ [−] and CF ₃ SO ₃ [−] based IL monomers were confirmed to have the better performance	40
3-Propyl-1-vinyl imidazolium bromide ([C3VIM]Br)	Template: amoxicillin Functional monomer: MWCNTs@[C3VIM]Br Cross-linker: EDMA Solvent: methanol–water	In this paper, IL was grafted on MWCNT surface by using ionic exchange strategy and the resulting MWCNTs@IL was taken as monomer to prepare MWCNTs@MIP. Then, the MWCNTs@MIP was coated on dendritic Pt–Pd nanoparticle to constructed the electrochemical sensor. The obtained sensor showed sensitive and selective response to amoxicillin	41
1-Allyl-3-ethyl imidazolium bromide ([AEIM]Br) 1-allyl-3-butyl imidazolium chloride ([ABIM]Cl), 1-allyl-3-hexyl imidazolium chloride ([AHIM]Cl), 1-allyl-3-octyl imidazolium chloride ([AOIM]Cl)	Template: phenolic acid Functional monomer: [AEIM]Br, [ABIM]Cl, [AHIM]Cl, [AOIM]Cl Cross-linker: EDMA Solvent: ethanol/H ₂ O, isopropanol/H ₂ O, <i>n</i> -butanol/H ₂ O, <i>n</i> -hexanol/H ₂ O	A new material, ionic liquid-based molecularly imprinted anion-exchange polymer was synthesized and used for separation of phenolic acids from <i>Salicornia herbacea</i> L. extract. Four ILs and four solvent systems were studied and the [AEIM]Br and Ethanol/H ₂ O were considered as the optimal monomer and porogen, respectively	42



Table 1 (Contd.)

RTIL	Protocol of preparing MIPs	Comments	Ref.
Mono-6A-deoxy-6-(1-vinyl imidazolium)- β -cyclodextrin tosylate	Template: C-terminal peptides of Cyt C Functional monomer: mono-6A-deoxy-6-(1-vinylimidazolium)- β -cyclodextrin tosylate Cross-linker: MBA Magnetic substrate: Fe_3O_4 @ SiO_2 -MPS Solvent: phosphate buffer	A novel IL-based functional monomer was synthesized and used for the preparation of Fe_3O_4 @EMIPs by using the combination of epitope and surface imprinting approach. The novel material has a good selectivity, adsorption capacity and recognition ability on Cyt C	43

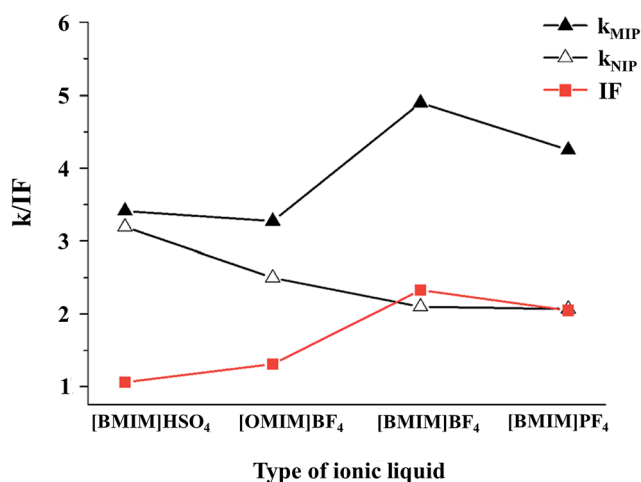


Fig. 6 Effect of ionic liquid type on the imprinting factor of the UA-MIP monolith. Reprinted from ref. 47.

Furthermore, Wu *et al.* fabricated a MIP for extraction of four fluoroquinolones from real aqueous samples by using a similar porogenic system.² Comparing with the conventional porogens, the RTIL porogenic system has more advantages. The resulting MIPs exhibited satisfactory selective extraction ability for the target molecules (IF = 3.22) in aqueous samples. The MIPs prepared by acetonitrile and protoporphyrin–chloroform porogenic system showed lower selective extraction ability in acetonitrile and chloroform and the corresponding IF values were 1.76 and 2.34. These studies are further evidence that RTIL-based porogens give special properties to MIPs.

According to studies by McCluskey *et al.*, porogens have a significant effect on the process of MIPs preparation.^{48,49} Taking *trans*-aconitic acid and cocaine as templates, the conventional porogens, acetonitrile and chloroform, and the RTIL porogens, [BMIM][BF₄] and [BMIM][PF₆], were investigated. Two polymerization methods, bulk polymerization and precipitation polymerization, were performed at 5 °C and 60 °C. The results showed that the polymers were not formed by using conventional porogens at low temperature no matter what polymerization method was taken. In contrast, the polymers can be yielded by both polymerization methods with RTIL porogens at low temperature. At high temperature, the rate of MIPs polymerization in RTILs is significantly faster than in conventional porogens. Relevant results were shown in Table 2. The results indicated that the polymerization process of MIPs

was accelerated by using RTILs as porogen. Subsequently, the impacts of porogens on polymer morphology were also studied and the detailed process is shown in Fig. 7. Under the combined influence of templates, porogens and polymerization methods, the polymers show different morphology characteristics. In the end, this study found that MIPs prepared with RTILs porogens at low temperature have better selective extraction ability.

Interestingly, a study proposed by Liu *et al.* also supports McCluskey's conclusion.⁵⁰ In Liu's study, a MIP monolith with good permeability was obtained by using ketoprofen or naproxen as template and [BMIM][BF₄]/DMSO mixture as porogen. At the same content of porogenic solvent, the RTIL-based MIP monolith has much better column permeability than the MIP monolith prepared with conventional porogens (isooctane/toluene, 20/80, v/v), which suggested that RTIL produced significant effect on morphology of MIPs. Moreover, they found that the polymerization of IL-based MIP monolith was completed in a short time (<1.5 h). It is proved that RTIL can accelerate the rate of polymerization in the preparation of MIPs. Influence of RTIL porogen on the performance of MIP monolith was investigated as well. In this work, high column efficiency of the monolith was achieved. The capacity factor and IF of the RTIL-based MIP monolith for ketoprofen achieved 10.40 and 8.64, respectively. More similar studies on the application of RTIL porogen in the synthesis of MIPs were shown in Table 3.

4. RTILs used for other purposes in MIT

In the field of MIT, imprinting of macromolecules has always been regarded as a challenging problem. This is because macromolecules, such as protein, DNA and enzymes, have the characteristics of structural complexity and conformational flexibility. Hence, surface imprinting technology was proposed to solve the problem.⁶⁰ In this strategy, a substrate with effective immobilization capacity is a key to obtaining surface MIPs with high performance and RTILs could play an important role in this field. Qian *et al.* used BSA as model template to explore the effect of RTIL functionalized material as substrate in surface imprinting technology.⁶¹ RTIL functionalized Fe_3O_4 nanoparticles (Fe_3O_4 @IL) were prepared by a simple method and used as substrate. The scheme of the synthesis Fe_3O_4 @IL@MIP material is shown in Fig. 8. The results show that the resulting magnetic MIP nano-materials have good selectivity and specificity for BSA. The Fe_3O_4 @IL@MIP showed good selectivity



Table 2 Polymerization times for *trans*-aconitic acid and cocaine. Reprinted from ref. 49

Porogen	Reaction temperature [°C]	Volume porogen [mL]	Reaction time [h]	
			<i>trans</i> -Aconitic acid	Cocaine
CH ₃ CN	5	5	— ^a	— ^b
		25	— ^a	— ^b
	60	5	6	— ^b
		25	18	— ^b
CHCl ₃	5	5	— ^b	— ^a
		25	— ^b	— ^a
	60	5	— ^b	6
		25	— ^b	18
[BMIM][BF ₄]	5	5	0.75	0.5
		25	2	2
	60	5	2	2
		25	8	4
[BMIM][PF ₆]	5	5	0.5	0.75
		25	2	2
	60	5	2	2
		25	8	4

^a No reaction. ^b Not performed.

towards BSA with IF value of 3.33 and adsorption capacity value of 50.6 mg g⁻¹. This investigation indicated that immobilization of macromolecules using RTIL functionalized substrate in surface imprinting technology would be helpful in preparing MIPs with high capacity and selectivity.

However, one shortcoming in the application of MIPs is their poor recognition performance to strong hydrophilic templates when using conventional functional monomers such as methacrylic acid (MAA), methyl methacrylate (MMA) and 4-VP. To address this disadvantage, a water-compatible MIP was synthesized by Zhao *et al.*⁶² In this work, an RTIL functionalized graphene was prepared using RTIL (1-(3-aminopropyl)-3-methyl imidazolium bromide) and graphene at first. Then, the MIP film was prepared on the surface of RTIL functionalized graphene which was used as functional monomer and substrate. At last, the composite material was used as electrochemical sensor for imidacloprid sensing. The detailed process is shown in Fig. 9. The resulting imprinted RTIL polymer exhibited high adsorption capacity and good selectivity for imidacloprid in vegetable and fruit samples. The results showed that the peak currents of imidacloprid displayed small change (95.4–107.8%) in the presence of 10-fold interfering substances, indicating that the obtained composite material had good selectivity. It means that some RTILs can interact with hydrophilic molecules and it also indicates the RTIL-based MIPs can have a good performance in aqueous medium.

MIPs coated quantum dots (QDs) is a new kind of material and combines the advantages of the two materials, such as good selectivity and high sensitivity. However, several important problems need to be solved in order to obtain MIPs coated QDs with satisfactory performance. For example, the surface of QDs is lack of appropriate functional groups to form firm chemical bond with MIPs and the application of QDs is limited because

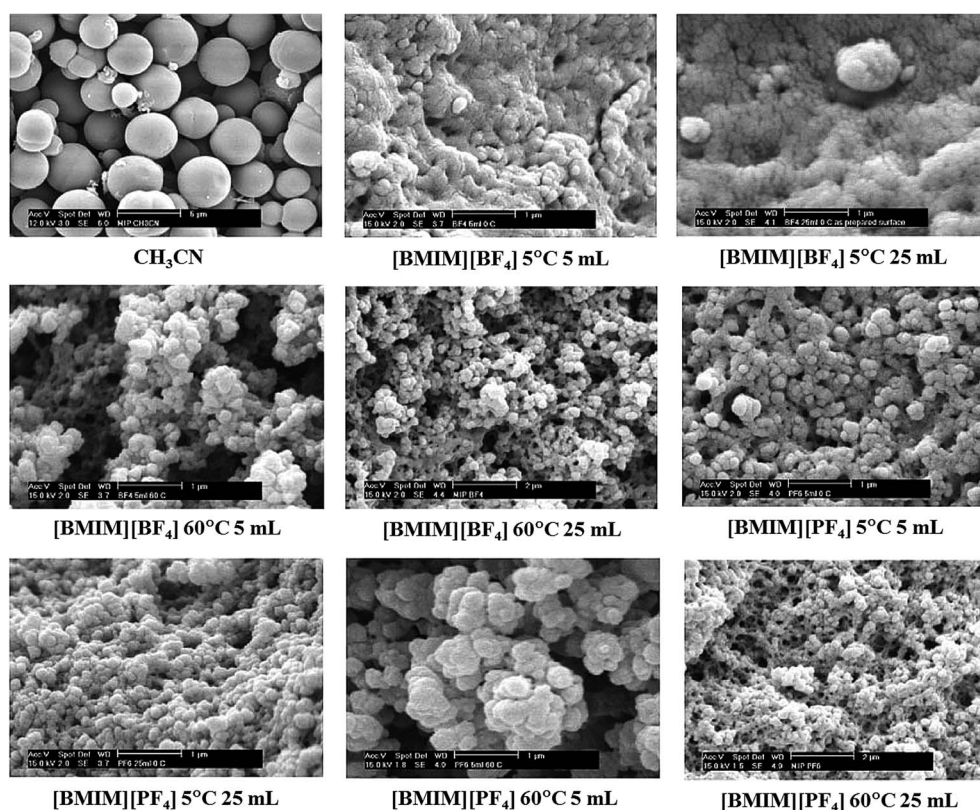


Fig. 7 SEM images of *trans*-aconitic acid MIPs prepared using different porogens, temperatures, and volumes. Reprinted from ref. 49.



Table 3 Some applications of RTIL porogens in preparation of MIPs

RTIL	Protocol of preparing MIPs	Comments	Ref.
1-Butyl-3-methyl imidazolium hexafluorophosphate ([BMIM][PF ₆])	Dummy template: α -chloro-DDT Functional monomer: acrylamide (AM) Cross-linker: DVB Solvent: acetonitrile-Toluene-[BMIM][PF ₆]	Taking [BMIM][PF ₆] as the auxiliary solvent, a RTIL-based MIPs was synthesized by precipitation polymerization. The RTIL-based polymer was utilized as SPE sorbents and it has better selectivity and adsorption capacity to dicofol than conventional SPE sorbents in aqueous medium	51
	Template: sulfamoxol Functional monomer: MAA Cross-linker: EDMA Supported material: modified silica monolith Solvent: acetonitrile-toluene-[BMIM][PF ₆]	In this study, a silica monolithic support was synthesized in capillary by the sol-gel method at first. Then, the RTIL-based MIPs were coated on the surface of silica monolith. The proposed material was evaluated by CEC. The results demonstrated that the permeability of the RTIL-based MIPs monolithic column was improved and the selectivity of monolithic column was better than MIPs monolithic column without RTIL	52
	Template: dibutyl phthalate (DBP) Functional monomer: MAA Cross-linker: EDMA Solvent: chloroform-[BMIM][PF ₆]	The MIPs was synthesized by using [BMIM][PF ₆] as an auxiliary porogen. The results showed that the RTIL-based polymer had excellent enrichment efficiency and selectivity for template	53
	Template: dichlorvos Functional monomer: MAA Cross-linker: TRIM Solvent: acetonitrile-toluene-[BMIM][PF ₆]	In this work, a dichlorvos MIPs was prepared by bulk polymerization technique. The shrinking or swelling ratio of polymers decreased by using RTILs as auxiliary porogen. The microstructure of MIPs can keep stable and it is significant to maintain the specific recognition property of MIPs	54
1-Butyl-3-methyl imidazolium tetrafluoroborate ([BMIM][BF ₄])	Dummy template: testosterone Functional monomer: 3-(triethoxysilyl)propyl isocyanate Solvent: [BMIM][BF ₄]	By sol-gel nano casting and sacrificial spacer MIT, the molecularly imprinted silica was obtained. Briefly, the covalent monomer-template complex was synthesized at first. Then, the monomer-template complex was added into a mixture of TEOS and RTIL and the new material was prepared by sol-gel method. The results indicated that [BMIM][BF ₄] had good porogen properties and solvation qualities	55
	Template: Corilagin Functional monomer: 4-VP Cross-linker: EDMA Solvent: DMF-DMSO-[BMIM][BF ₄]	A RTIL-mediated MIPs was prepared and evaluated as a SPE sorbent for the pretreatment of corilagin. The results showed that the new MIPs have good specific adsorption capacity for template	56
	Template: norfloxacin Functional monomer: MAA Cross-linker: EDMA Solvent: DMF-DMSO-[BMIM]BF ₄	By the proposed formula, a RTIL-mediated MIPs was prepared and was used as HPLC monolith and SPE sorbent. The study showed that the proposed polymer exhibited excellent ability of selective recognition to quinolones	57
	Template: ketoprofen Functional monomer: AAm-AMPS Cross-linker: EDMA Solvent: DMSO-[BMIM][BF ₄]	A thermo responsive MIP monolith was prepared and evaluated. In this work, the RTIL-based porogen was used to increase the solubility of AMPS. The sensibility of MIP to temperature was also investigated. The novel MIPs has the best specific adsorption capacity for ketoprofen in the transition temperature of 35 °C. It was more than 75 times that at 25 °C	58
	Template: methyl gallate Functional monomer: 4-VP Cross-linker: EDMA Solvent: DMSO-DMF-[BMIM][BF ₄]	An imprinted monolithic HPLC column was prepared with good performance and permeability by using a RTIL-based ternary porogenic system. In this paper, a polar molecule was taken as template and the RTIL-based monolithic column showed higher imprinting efficiency	59



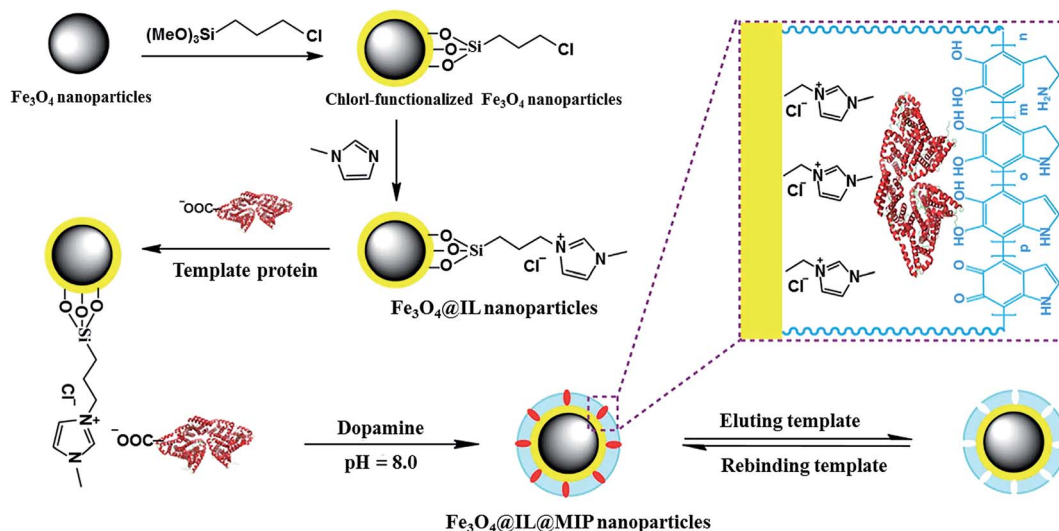


Fig. 8 Schematic illustration of the synthesis $\text{Fe}_3\text{O}_4\text{@IL@MIP}$ nanoparticles. Reprinted from ref. 61.

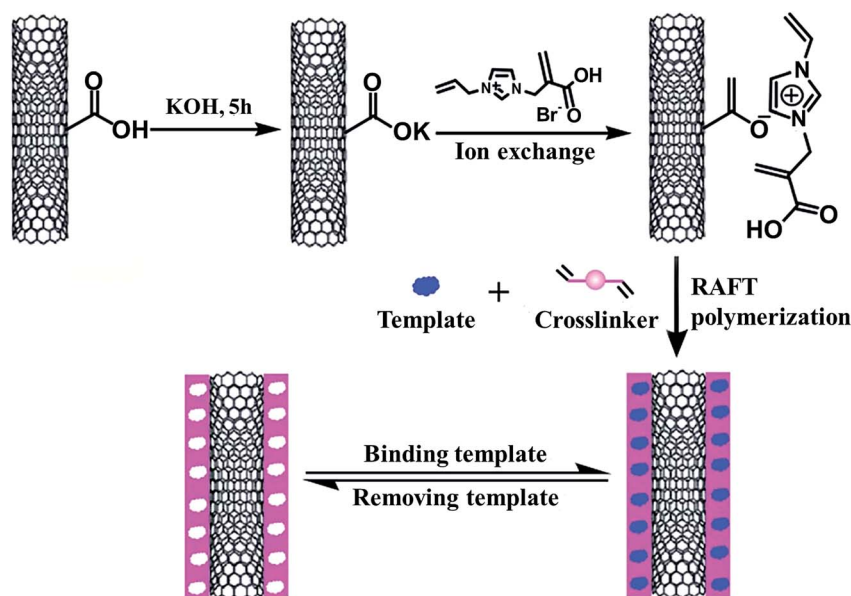


Fig. 9 Schematic illustration of preparation of water-compatible surface-imprinted RTIL polymer coated MWNTs. Reprinted from ref. 62.

of its low fluorescence stability. Hence, a series of MIPs based on RTIL-modified QDs were proposed by Wang *et al.*^{63–66} In these researches, the core strategy is that RTILs with carbon-carbon double bonds bind to the surface of CdSe/ZnS QDs. Two RTILs, 1-vinyl-3-octyl imidazolium hexafluorophosphate ([VOIM][PF₆]), 1-vinyl-3-butyl imidazolium hexafluorophosphate ([VBIM][PF₆]), were investigated. The results showed that related RTIL-modified QDs could form binding groups with MIPs^{63–65} or carboxylated graphene oxide (GO).⁶⁶ The MIPs coated IL-modified QDs material with a stable structure can be obtained by the proposed strategy. Furthermore, the fluorescent stability of QDs was also improved because of high thermal and chemical stability of RTILs.

Furthermore, RTILs can be used as pore template in sol-gel processing.⁶⁷ In this technique, silicate matrix is assembled in

proper solvent (pore template) and a highly cross-linked polymer with specific shape and size cavities could be obtained. Moreover, some studies have shown that the shrink or swell of MIPs could be avoided to some extent by using RTILs as solvent.^{68,69} It is important for functional groups in the polymer to maintain relative position. Wang and co-workers⁷⁰ proposed an RTIL-mediated, non-hydrolytic sol-gel method to prepare MIP silica-based hybrid monoliths for chiral separation of racemic naproxen. In this research, a hydrophobic RTIL ([BMIM][PF₆]) was taken as pore template for non-hydrolytic sol-gel (NHSG) processing and the synthesized product provided a stable solid support for subsequent polymerization. This method avoided the shrink of sol-gel and provided a polymer network with stable structure. As a result, the silica-based hybrid MIP has good selectivity. A resolution value of



8.82 for the separation of (*S*)-naproxen and (*R*)-naproxen could be obtained. Subsequently, the same method was applied to preparation of molecularly imprinted silica-based hybrid monoliths and the resulting material was used for chiral separation of zolmitriptan by capillary electrochromatography (CEC).⁷¹ In this study, [BMIM][BF₄] was utilized to prevent gel shrinkage and also used as pore template. The results showed that the porosity and selectivity of hybrid monoliths were obviously improved. These two researches implied that the RTIL-mediated NHSG strategy is a hopeful method for the preparation of imprinted hybrid materials with high performance.

As mentioned above, some RTILs have hazardous toxicity and poor biodegradability.^{14,15} This means that RTILs might result in environmental problem in the course of preparation and application of RTILs. To effectively monitor the RTILs in environment, a RTIL surface MIP was prepared by Gao *et al.*^{72,73} In these works, the alkyl imidazolium chloride RTILs, 1-butyl-3-methyl imidazolium chloride ([C₄MIM]Cl, *i.e.* [BMIM]Cl) and 1-octyl-3-methyl imidazolium chloride ([C₈MIM]Cl, *i.e.* [OMIM]Cl) were taken as templates. The prepared MIPs were used as solid phase extraction material. The results show that the RTIL surface MIPs have excellent selective adsorption of RTILs ([C_{*n*}MIM]Cl, *n* = 2, 4, 6, 8, 12, 16) in complex environmental samples. Obviously, [C₄MIM]Cl and [C₈MIM]Cl could be regarded as template analogues in such cases. Template analogue technique is generally used to overcome some limiting factors. For example, actual template compound can be very expensive, heat- or light-sensitive or insoluble in polymerisation porogens. Moreover, it can also enhance the affinity of functional monomer to template. An innovative method of preparation for L-phenylalanine (L-Phe) MIPs was proposed by Yang *et al.*⁷⁴ In this study, an oil-soluble amino acid IL, 1-butyl-3-methyl imidazolium amino hydrocinnamic acid salt ([BMIM][Phe]), was introduced as template and a surface MIPs was prepared for the selective recognition of L-Phe. The affinity of functional monomer (4-VP) to [BMIM][Phe] and 4-VP to L-Phe were investigated and it was found that the affinity of 4-VP to [BMIM][L-Phe] was much higher. The data showed that MIPs prepared with [BMIM][Phe] were three times selective than the corresponding MIPs prepared with 4-VP. It means that the template analogue technique converted the hydrophilic template imprinting process from water to the organic phase.

Reversible addition-fragmentation chain transfer (RAFT) polymerization was a method of living or controlled radical polymerization and the technique is commonly used to preparation of MIPs. Studies found that the rate of bimolecular termination during the RAFT polymerization was reduced by using RTILs as reaction medium, indicating that the rate of free radical polymerization can be significantly increased.⁷⁵ Even more remarkably, by using reverse atom transfer radical polymerization (ATRP) and RTILs, the molecular weight distribution of MIPs is more concentrated.⁷⁶ Based on these methods, Liu and co-workers⁷⁷ proposed a method based on RAFT polymerization for preparing carprofen MIP monoliths. With the aid of [BMIM][BF₄]-based ternary porogen, the obtained MIP monoliths had excellent permeability and satisfactory column

efficiency. Furthermore, by morphology analysis, it is verified that the structure of this MIP monolith is uniform.

Due to protein denaturation at higher temperature, protein imprinting is a difficult task in the field of MIT. How to retain the structure of protein at high temperature is a troublesome problem. It is reported that the biocompatible RTIL, choline dihydrogen phosphate (Chol Dhp), was used as a stabilizer during the process of lysozyme-imprinted microspheres preparation.⁷⁸ It was confirmed that the structural stability of lysozyme can be insured in the polymerization system with 5% RTIL as the stabilizer at 75 °C. The stabilizing effect of Chol Dhp on protein structure is probably the result of its function of hydrophobic and chelate.

Furthermore, RTILs can serve multiple purposes at the same time during the preparation of MIPs. An approach to prepare protein imprinted microspheres was proposed by Zhang *et al.*⁷⁹ In this work, an amphiphilic ionic liquid, 1-dodecyl-3-methylimidazolium chloride ([DMIM]Cl), was not only used as stabilizer, but also used as emulsifier. In other words, [DMIM]Cl was used to prevent protein denaturation and applied to form an oil-in-water emulsion system at the same time. There are many examples that RTILs play several different roles in the process of MIPs preparation. These studies introduced the diversity of RTILs in the field of MIT.

In summary, RTILs have brought many benefits to MIPs. For instance, the polymerization of MIPs was achieved with less volatile organic compound solvents, offering economic advantages as well as environmental advantages. Moreover, it is proved that the polymerization process of RTILs-based MIPs could be completed in a short time and that means less energy consumption. Furthermore, the development of RTILs-based MIPs has been giving impetus to the development of protein macromolecule imprinted technology. Besides, the swelling ratio of MIPs can be reduced by using RTIL as porogens. Therefore, the research of RTILs-based MIPs has the character of flexibility and diversity. It is a potential research field that has a wide application view.

5. The application of DES in the preparation of MIPs

DESs are another type of green solvents and it is also classified as one kind of RTILs by some researchers.⁸⁰ Compared with RTILs, DES has the advantages of low toxicity⁸¹ and low price. Moreover, DES is easy to prepare by mixing two or three inexpensive components in a certain proportion. The corresponding components were respectively defined as the HBA and the HBD, since the interaction between them is hydrogen bonds.¹⁷ It is worth noting that a broad group of compounds is likely candidates for the HBA or the HBD. This variety of DES provides more flexible choice for various applications, including drug dissolution,⁸² metal oxides dissolution,²⁰ gas absorption,⁸³ synthesis of nanoparticles,⁸⁴ organic synthesis⁸⁵ and synthesis of nanoparticles.⁸⁶ In view of the fact that DES share similar properties with RTILs, the application of DES for MIPs has just begun to receive considerable attention.



Like RTILs, DES performed varied functions in the preparation of MIPs. Liu *et al.* proposed a novel magnetic DES-MIPs for bovine hemoglobin (BHb) by using DES as functional monomer.²² This is the first study to report the application of DES in MIT. In this study, the DES was prepared by heating the mixture of choline chloride and methacrylic acid (molar ratio 1 : 2) at 90 °C and the prepared DES was a colorless and uniform liquid. The Fe₃O₄ nanoparticles were used as magnetic provider. It was obtained by the solvothermal approach and modified with acrylic acid. BHb, *N,N*-methylenebisacrylamide (MBAA) and ammonium persulfate (APS)/*N,N,N',N'*-tetramethylethylenediamine (TEMED) were used as template, cross-linker and initiator, respectively. Magnetic DES-MIPs were polymerized at room temperature to prevent heat denaturation of the protein. The detailed preparation procedure is shown in Fig. 10. The reported preparation strategy has the advantages of being simple, low toxicity and low cost. The protein adsorption experiments have shown that the IF of the magnetic DES-MIPs achieved 4.57 under optimum conditions. It is implied that the new material has a good specific recognition property. In order to further investigate the specificity of the new material, the competitive batch rebinding test was performed. As BSA is of similar size with BHb, it was taken as a competitive protein in competitive batch rebinding tests. Moreover, real sample (calf blood) adsorption experiment was carried out to evaluate the practicability of magnetic DES-MIPs. The adsorption capacity of magnetic DES-MIPs for BHb still kept at a high value (150.52 mg g⁻¹). The results demonstrated that the synthesized magnetic DES-MIPs have absorption with good selectivity on the BHb.

Hydrophilic resins are a class of adsorbents with the advantage of low cost, good porosity, diverse functional groups. It has good adsorption properties for target molecules in aqueous environments. However, poor specific molecular recognition ability restricts its application range. Conversely, specific molecular recognition ability is the crucial advantage of

MIPs. Liang *et al.* developed a molecularly imprinted phloroglucinol-formaldehyde-melamine resin (MIPFMR) by combining hydrophilic resins with MIT.⁸⁷ As mentioned before, MIPs synthesized in organic solvents have limited compatibility with water and poor molecular recognition of template in aqueous solution. To solve this problem, ethylene glycol-choline chloride DES was used as reaction medium in this study. Compared with materials synthesized in common reaction medium (alcoholic solvent systems), the MIPFMR prepared in DES shows better adsorption capability for the templates, clorprenaline and bambuterol. Although DES has the characteristics of a polar solvent, the adsorptive capacity of MIPFMR was increased. The authors suggest that DES have little impact on the template-monomers hydrogen bonds.

In the studies of DES, ChCl is the most popular HBA and most of the related studies are based on ChCl-based DES. However, ChCl is still relatively expensive. The searching for HBAs which are inexpensive, readily available and of low toxicity has been an important subject in the field of DES. Row *et al.*⁸⁸ developed a new type of DES by using betaine and ethylene glycol as HBA and HBD. In this research, DES was synthesized by betaine, ethylene glycol and water with the mole ratio of 1 : 2 : 1. The preparation process of DES-modified MIPs mainly contains three steps. First, the functional monomer 3-amino-propyltriethoxysilane (APTES)-MAA was fabricated under a certain condition. Then, the template (levofloxacin or tetracycline) and the monomer were added to methanol for forming the template-functional monomer complex. At last, the DES, cross-linker (EDMA), initiator (2,2-azobisisobutyronitrile, AIBN) and tetraethoxysilane (TEOS) were added to the template-functional monomer complex solution and the mixture was synthesized at 60 °C for 24 h. The synthesis scheme of DES-modified MIPs is shown in Fig. 11. The selective adsorption capacity of the levofloxacin DES-MIPs and tetracycline DES-MIPs were evaluated and the results show DES-modified MIPs

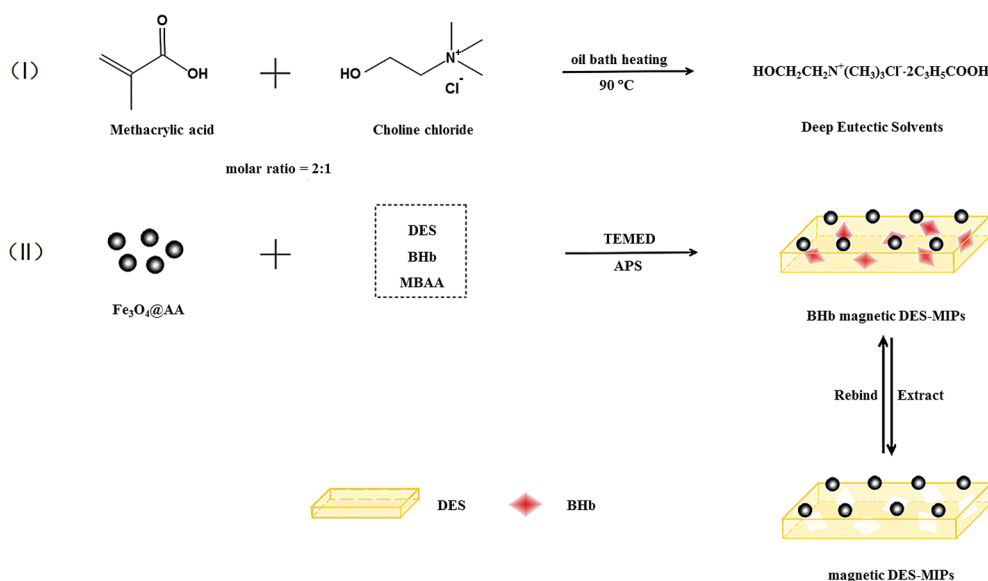


Fig. 10 Schematic illustration of the procedure for the magnetic DES-MIPs. Reprinted from ref. 22.



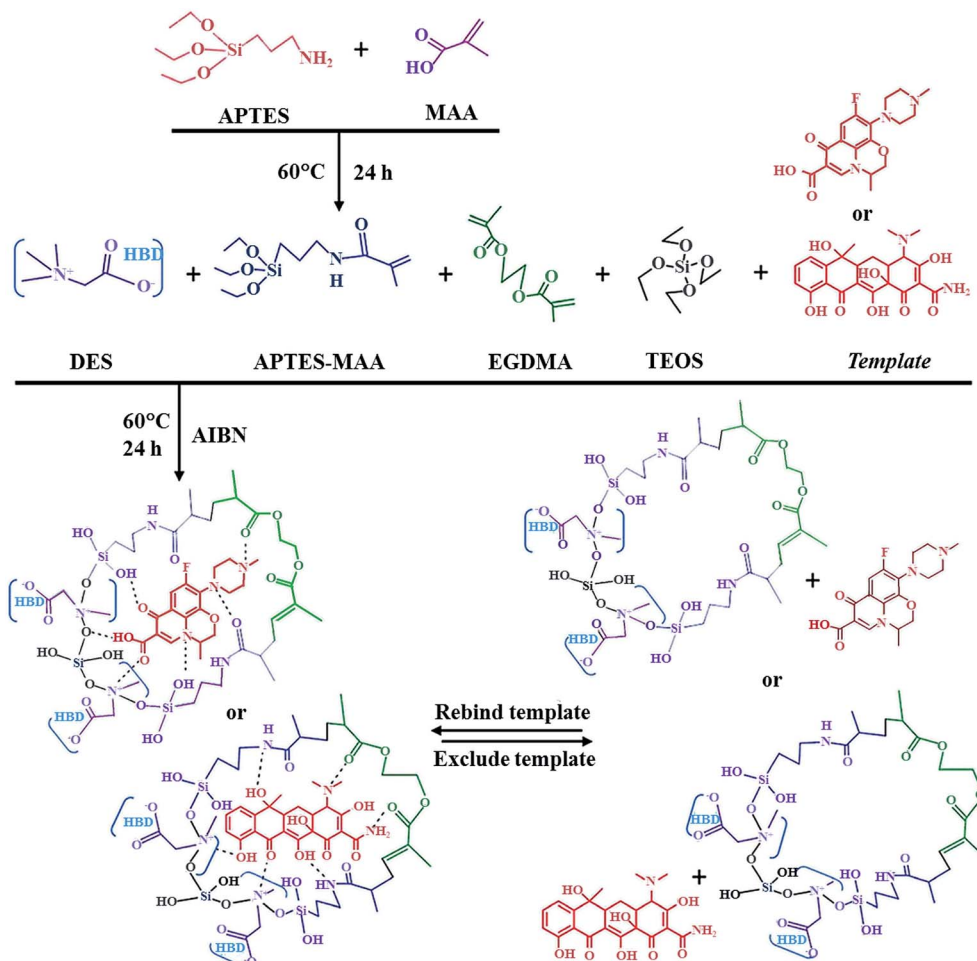


Fig. 11 Schematic illustration of the proposed materials formation. Reprinted from ref. 88.

have better adsorption ability than the conventional MIPs and the NIPs. Furthermore, the relevant DES-MIPs showed the highest selectivity recovery for levofloxacin (94.5%) and tetracycline (93.3%) from millet extract with mixture antibiotics, and could remove the interferent effectively. The results suggest that the performance of MIPs could be improved by using DES during MIPs preparation.

Moreover, an extraction method using DES-based MIPs as sorbent for the determination of levofloxacin green bean extract was proposed by the same team.⁸⁹ In this study, the preparation method of DES-MIPs was similar with literature.⁸⁸ The resulting materials also show excellent performance of adsorption ability and selectivity. DES-MIP showed the highest selectivity recovery (95.2%) for levofloxacin the complex matrix. It is proved that DES-modified MIP is still stable for real samples. Furthermore, the influence of various ChCl-based DESs and 1-methylimidazole-based RTILs on the performance of MIPs was also investigated by Row's team.⁹⁰ Taking rutin, scoparone and quercetin in as multi-template, γ -aminopropyltriethoxysilane methacrylic (KH-550-MAA) as multiple functional monomers, EDMA as cross-linker and AIBN as initiator, the DES/RTIL modified hybrid MIPs (HMIPs) were prepared. In this article, three DESs (ChCl-ethylene glycol, ChCl-glycerol and ChCl-1,4-

butanediol) and three RTILs (1-ethyl-3-methyl imidazolium bromide ([EMIM]Br), 1-butyl-3-methyl imidazolium bromide ([BMIM]Br) and 1-hexyl-3-methyl imidazolium bromide ([HMIM]Br)) were used for modifying the HMIPs. The results show that the modified HMIPs have good recovery ability and recognition performance toward templates in *Herba Artemisiae Scopariae*. As described in the study, DES-HMIPs have higher recoveries than RTILs-HMIPs and the ChCl-glycerol modified HMIPs have the best performance among DES-HMIPs with the recoveries of 92.27% (rutin), 87.51% (scoparone) and 80.02% (quercetin), respectively. It is implied that DES has great advantages when applied to MIT. In addition to the functions mentioned above, DES has played multiple roles in the preparation of MIPs. In Row's latest communication,⁹¹ DES was synthesized by using choline chloride-caffeic acid-ethylene glycol with a special mole ratio and it was utilized as a functional monomer and template. By the proposed approach, the DES-MIPs were obtained with good recognition ability for the polyphenols in real samples. The research showed that caffeic acid in DES has a great influence on selective recognition ability of MIPs.

Obviously, with the development of MIT, DESs have begun to be applied in the preparation of MIPs because of their particular



advantages. DES-based MIPs have good compatibility with water and high molecular recognition of template in aqueous solution. Furthermore, it was proved that DESs could improve the selectivity and affinity of MIPs. However, the DES-based MIPs research is only at the start stage and some relevant mechanisms are still unclear. For this reason, a vast amount of further research is needed to explore the potential value of DES-based MIPs.

6. Conclusion

At present, RTIL-based MIPs and DES-based MIPs have become an active and exciting research area in MIT. The investigations on these new materials have got some progressions up to now. RTIL-based MIPs and DES-based MIPs overcome many shortcomings of traditional MIPs. For example, RTILs and DESs can contribute to improving the effect of macromolecules MIPs. The shrinking or swelling of MIPs is reduced by using RTIL as porogens. Furthermore, it has been proved that RTIL-based MIPs and DES-based MIPs can maintain significant performances in aqueous medium. Comparing with traditional MIPs, the new materials have better specific recognition property. However, studies on the new materials are relatively few, especially DES-based MIPs. The effect mechanism of RTIL and DES in the synthetic process of MIPs has not been studied thoroughly. However, it can be expected that RTIL-based MIPs and DES-based MIPs have a broad application prospects and more relevant reports will be presented in the future.

List of abbreviations

4-VP	4-Vinylpyridine	KH-550-MAA	γ -Aminopropyltriethoxysilane-methacrylic
AAm	Acrylamide	L-Phe	L-Phenylalanine
AIBN	2,2-Azobisisobutyronitrile	MAA	Methacrylic acid
AMPS	2-Acrylamide-2-methyl propanesulfonic acid	MBA	<i>N,N'</i> -Methylenebisacrylamide
APS	Ammonium persulfate	MBAA	<i>N,N'</i> -Methylenebisacrylamide
APTES	3-Aminopropyltriethoxysilane	MD	Molecular dynamics
ATRP	Atom transfer radical polymerization	MIPFMR	Molecularly imprinted phloroglucinol-formaldehyde-melamine resin
BET	Brunauer-Emmett-Teller	MIPs	Molecularly imprinted polymers
BHb	Bovine hemoglobin	MIT	Molecularly imprinted technology
BPA	Bisphenol A	MMA	Methyl methacrylate
BSA	Bovine serum albumin	MWCNTs	Multiwall carbon nanotubes
CEC	Capillary electrochromatography	NHSG	Non-hydrolytic sol-gel
CFO	(<i>Z</i>)-3-(Chloromethylene)-6-fluorothiochroman-4-one	NNMBA	<i>N,N'</i> -Methylenebis (acrylamide)
ChCl	Choline chloride	NIPA	<i>N</i> -Isopropylacrylamide
Chol Dhp	Choline dihydrogen phosphate	PALS	Positron annihilation lifetime spectroscopy
CTA-2	(4-Cyanopentanoic acid)-4-dithiobenzoate	PMDETA	<i>N,N,N',N',N''</i> -Pentamethyldiethylene triamine
Cyt C	Cytochrome C	PVP	Polyvinylpyrrolidone
DESs	Deep eutectic solvents	QDs	Quantum dots
DVB	Divinylbenzene	RAFT	Reversible addition-fragmentation chain transfer
EDMA	Ethylene glycol dimethacrylate	RTILs	Room temperature ionic liquids
FHO	(<i>Z</i>)-6-Fluoro-3-(hydroxymethylene)-thiochroman-4-one	SEM	Scanning electron microscopy
GO	Raphene oxide	TEM	Transmission electron microscopy
HBA	Hydrogen bond acceptor	TEMED	<i>N,N,N',N'</i> -Tetramethylethylenediamine
HBD	Hydrogen bond donors	TEOS	Tetraethoxysilane
		TP5	Thymopentin
		[ABIM]Cl	1-Allyl-3-butyl imidazolium chloride
		[AEIM]Br	1-Allyl-3-ethyl imidazolium bromide
		[AEIM][PF ₆]	1-Allyl-3-ethyl imidazolium hexafluorophosphate
		[AHIM]Cl	1-Allyl-3-hexyl imidazolium chloride
		[AMIM]Br	1-Allyl-3-methyl imidazolium bromide
		[AOIM]Cl	1-Allyl-3-octyl imidazolium chloride
		[BMIM][BF ₄]	1-Butyl-3-methyl imidazolium tetrafluoroborate
		[BMIM]Br	1-Butyl-3-methyl imidazolium bromide
		[BMIM]Cl	1-Butyl-3-methyl imidazolium chloride
		[BMIM]	1-Butyl-3-methyl imidazolium hydrogen sulfate
		[HSO ₄]	
		[BMIM][PF ₆]	1-Butyl-3-methyl imidazolium hexfluorophosphate
		[BMIM][Phe]	1-Butyl-3-methyl imidazolium aminohydrocinnamic acid salt
		[BVIM][Phe]	1-Butyl-3-vinyl imidazolium aminohydrocinnamic acid
		[C3VIM]Br	3-Propyl-1-vinyl imidazolium bromide
		[COOHavim]	1-Vinyl-3-carboxybutyl imidazolium bromide
		Br	
		[COOHevim]	1-Vinyl-3-carboxymethyl imidazolium bromide
		Br	
		[COOHhvim]	1-Vinyl-3-carboxypentyl imidazolium bromide
		Br	
		[COOHpvim]	1-Vinyl-3-carboxyethyl imidazolium bromide
		Br	
		[DMIM]Cl	1-Dodecyl-3-methyl imidazolium chloride
		[EMIM]Br	1-Ethyl-3-methyl imidazolium bromide
		[HMIM]Br	1-Hexyl-3-methyl imidazolium bromide
		[HMIM][PF ₆]	1-Hexyl-3-methyl imidazolium hexfluorophosphate
		[OMIM][BF ₄]	1-Octyl-3-methyl imidazolium tetrafluoroborate



[OMIM]Cl	1-Octyl-3-methyl imidazolium chloride
[OMIM][PF ₆]	1-Octyl-3-methyl imidazolium hexafluorophosphate
SilprImN	1-(Triethoxysilyl) propyl-3-aminopropyl imidazole bromide
[TMSPMIM]Cl	1-(3-Trimethoxysilyl propyl)-3-methyl imidazolium chloride
[VAFMIM]Cl	1-Vinyl-3-aminoformylmethyl imidazolium chloride
[VBIM]Cl	1-Vinyl-3-butyl imidazolium chloride
[VBIM][PF ₆]	1-Vinyl-3-butyl imidazolium hexafluorophosphate
[VEAIM]Cl	1-Vinyl-3-ethyl acetate imidazolium chloride
[VMIM]Cl	1-Vinyl-3-methyl imidazolium chloride
[VOIM]Cl	1-Vinyl-3-octyl imidazolium chloride
[VOIM][PF ₆]	1-Vinyl-3-octyl imidazolium hexafluorophosphate

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work is sponsored by the Fund for Shanxi Key Subjects Construction and supported by National Natural Science Foundation of China (21402012).

References

- Z. H. Wei, X. Wu, B. Zhang, R. Li, Y. P. Huang and Z. S. Liu, *J. Chromatogr. A*, 2011, **1218**, 6498–6504.
- X. Wu and L. T. Wu, *J. Sep. Sci.*, 2015, **38**, 3615–3621.
- Z. G. Xu, C. Y. Song, Y. L. Hu, J. Wang and G. K. Li, *Talanta*, 2011, **85**, 97–103.
- W. Boonjob, Y. L. Yu, M. Miro, M. A. Segundo, J. H. Wang and V. Cerda, *Anal. Chem.*, 2010, **82**, 3052–3060.
- R. Schirhagl, *Anal. Chem.*, 2014, **86**, 250–261.
- K. J. Shea and D. Y. Sasaki, *J. Am. Chem. Soc.*, 1991, **113**, 4109–4120.
- L. X. Chen, X. Y. Wang, W. H. Lu, X. Q. Wu and J. H. Li, *Chem. Soc. Rev.*, 2016, **45**, 2137–2211.
- T. Welton, *Chem. Rev.*, 1999, **99**, 2071–2084.
- S. Gao, H. Jin, J. You, Y. Ding, N. Zhang, Y. Wang, R. Ren, R. Zhang and H. Zhang, *J. Chromatogr. A*, 2011, **1218**, 7254–7263.
- D. W. Armstrong, L. He and Y. S. Liu, *Anal. Chem.*, 1999, **71**, 3873–3876.
- C. C. Liu, Q. L. Deng, G. Z. Fang, X. Feng, H. Qian and S. Wang, *Anal. Bioanal. Chem.*, 2014, **406**, 7175–7183.
- M. Bohm, A. A. Tietze, P. Heimer, M. Chen and D. Imhof, *J. Mol. Liq.*, 2014, **192**, 67–70.
- A. Martin-Calero, V. Pino and A. M. Afonso, *Trends Anal. Chem.*, 2011, **30**, 1598–1619.
- A. Romero, A. Santos, J. Tojo and A. Rodriguez, *J. Hazard. Mater.*, 2008, **151**, 268–273.
- N. V. Plechkova and K. R. Seddon, *Chem. Soc. Rev.*, 2008, **37**, 123–150.
- A. P. Abbott, G. Capper, D. L. Davies, R. K. Rasheed and V. Tambyrajah, *Chem. Commun.*, 2003, **8**, 70–71.
- Q. H. Zhang, K. D. O. Vigier, S. Royer and F. Jerome, *Chem. Soc. Rev.*, 2012, **41**, 7108–7146.
- A. P. Abbott, G. Capper, D. L. Davies, K. J. McKenzie and S. U. Obi, *J. Chem. Eng. Data*, 2006, **51**, 1280–1282.
- M. Karimi, S. Dadfarnia, A. M. H. Shabani, F. Tamaddon and D. Azadi, *Talanta*, 2015, **144**, 648–654.
- A. P. Abbott, D. Boothby, G. Capper, D. L. Davies and R. K. Rasheed, *J. Am. Chem. Soc.*, 2004, **126**, 9142–9147.
- L. Guo, Q. L. Deng, G. Z. Fang, W. Gao and S. Wang, *J. Chromatogr. A*, 2011, **1218**, 6271–6277.
- Y. J. Liu, Y. Z. Wang, Q. Z. Dai and Y. G. Zhou, *Anal. Chim. Acta*, 2016, **936**, 168–178.
- S. F. Yuan, Q. L. Deng, G. Z. Fang, M. F. Pan, X. R. Zhai and S. Wang, *J. Mater. Chem.*, 2012, **22**, 3965–3972.
- R. K. Desai, M. Streefland, R. H. Wijffels and M. H. M. Eppink, *Green Chem.*, 2014, **16**, 2670–2679.
- H. Y. Xiang, M. J. Peng, H. Li, S. Peng and S. Y. Shi, *J. Pharm. Biomed. Anal.*, 2017, **133**, 75–81.
- J. P. Fan, Z. Y. Tian, S. Tong, X. H. Zhang, Y. L. Xie, R. Xu, Y. Qin, L. Li, J. H. Zhu and X. K. Ouyang, *Food Chem.*, 2013, **141**, 3578–3585.
- M. L. Tian, W. T. Bi and K. H. Row, *Anal. Bioanal. Chem.*, 2011, **399**, 2495–2502.
- C. L. Wang, X. L. Hu, P. Guan, L. W. Qian, D. F. Wu and J. Li, *Int. J. Polym. Anal. Charact.*, 2014, **19**, 70–82.
- X. Wu, Y. P. Huang and Z. S. Liu, *Curr. Org. Chem.*, 2016, **20**, 459–468.
- J. Li, X. L. Hu, P. Guan, D. M. Song, L. W. Qian, C. B. Du, R. Y. Song and C. L. Wang, *J. Sep. Sci.*, 2015, **38**, 3279–3287.
- M. M. Liu, J. Y. Pi, X. J. Wang, R. Huang, Y. M. Du, X. Y. Yu, W. F. Tan, F. Liu and K. J. Shea, *Anal. Chim. Acta*, 2016, **932**, 29–40.
- L. W. Qian, X. L. Hu, P. Guan, B. Gao, J. Li, C. L. Wang and Y. M. Tang, *Talanta*, 2014, **121**, 56–64.
- S. F. Yuan, Q. L. Deng, G. Z. Fang, J. H. Wu, W. W. Li and S. Wang, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2014, **960**, 239–246.
- X. Y. Gao, M. F. Pan, G. Z. Fang, W. Jing, S. Y. He and S. Wang, *Anal. Methods*, 2013, **5**, 6128–6134.
- H. Y. Yan, S. T. Liu, M. M. Gao and N. Sun, *J. Chromatogr. A*, 2013, **1294**, 10–16.
- F. X. Qiao, M. M. Gao and H. Y. Yan, *J. Sep. Sci.*, 2016, **39**, 1310–1315.
- C. L. Wang, X. L. Hu, P. Guan, D. F. Wu, L. W. Qian, J. Li and R. Y. Song, *J. Pharm. Biomed. Anal.*, 2015, **102**, 137–143.
- J. Li, X. L. Hu, P. Guan, X. Y. Zhang, L. W. Qian, R. Y. Song, C. B. Du and C. L. Wang, *RSC Adv.*, 2015, **5**, 62697–62705.
- Y. N. Yuan, S. R. Liang, H. Y. Yan, Z. Y. Ma and Y. X. Liu, *J. Chromatogr. A*, 2015, **1408**, 49–55.
- H. Y. Ding, R. F. Chen, M. M. Liu, R. Huang, Y. M. Du, C. Huang, X. Y. Yu, X. H. Feng and F. Liu, *RSC Adv.*, 2016, **6**, 43526–43538.
- G. M. Yang and F. Q. Zhao, *Electrochim. Acta*, 2015, **174**, 33–40.



- 42 W. T. Bi, M. L. Tian and K. H. Row, *J. Chromatogr. A*, 2012, **1232**, 37–42.
- 43 X. Y. Zhang, N. Zhang, C. B. Du, P. Guan, X. M. Gao, C. Y. Wang, Y. F. Du, S. C. Ding and X. L. Hu, *Chem. Eng. J.*, 2017, **317**, 988–998.
- 44 H. Y. Yan, C. Yang, Y. Y. Sun and K. H. Row, *J. Chromatogr. A*, 2014, **1361**, 53–59.
- 45 L. H. Bai, X. X. Chen, Y. P. Huang, Q. W. Zhang and Z. S. Liu, *Anal. Bioanal. Chem.*, 2013, **405**, 8935–8943.
- 46 K. Booker, C. I. Holdsworth, C. M. Doherty, A. J. Hill, M. C. Bowyer and A. McCluskey, *Org. Biomol. Chem.*, 2014, **12**, 7201–7210.
- 47 C. Zhang, F. Li, S. X. Wang, Z. S. Liu and H. A. Aisa, *Anal. Methods*, 2015, **7**, 10256–10265.
- 48 K. Booker, M. C. Bowyer, C. I. Holdsworth and A. McCluskey, *Chem. Commun.*, 2006, **16**, 1730–1732.
- 49 K. Booker, M. C. Bowyer, C. J. Lennard, C. I. Holdsworth and A. McCluskey, *Aust. J. Chem.*, 2007, **60**, 51–56.
- 50 X. Y. Li, X. X. Chen, D. D. Zhong, Y. P. Huang and Z. S. Liu, *RSC Adv.*, 2014, **4**, 50662–50667.
- 51 H. Y. Yan, N. Sun, Y. H. Han, C. Yang, M. Y. Wang and R. J. Wu, *J. Chromatogr. A*, 2013, **1307**, 21–26.
- 52 J. X. He, G. Z. Fang, Y. C. Yao and S. Wang, *J. Sep. Sci.*, 2010, **33**, 3263–3271.
- 53 D. D. Zhong, Y. P. Huang, X. L. Xin, Z. S. Liu and H. A. Aisa, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2013, **934**, 109–116.
- 54 Z. X. Xu, G. Z. Fang and S. Wang, *Food Chem.*, 2010, **119**, 845–850.
- 55 C. Y. He, Y. Y. Long, J. L. Pan, K. A. Li and F. Liu, *Talanta*, 2008, **74**, 1126–1131.
- 56 C. Wang, X. J. Li, J. Yang, Y. X. Zhao, Z. S. Liu and H. A. Aisa, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2017, **1041**, 98–103.
- 57 X. L. Sun, J. He, G. R. Cai, A. Q. Lin, W. J. Zheng, X. Liu, L. X. Chen, X. W. He and Y. K. Zhang, *J. Sep. Sci.*, 2010, **33**, 3786–3793.
- 58 X. Sun, C. Y. Zhao, X. H. Wang, Y. P. Huang and Z. S. Liu, *Anal. Bioanal. Chem.*, 2014, **406**, 5359–5367.
- 59 X. M. Zhang, W. M. Yang, N. W. Wang, X. N. Ni, H. J. Wang, Z. Y. Gu, X. Y. Wu and W. Z. Xu, *Adv. Polym. Technol.*, 2015, **36**, 220–229.
- 60 X. Qu, F. Wang, Y. Sun, Y. Tian, R. Chen, X. Ma and C. Liu, *RSC Adv.*, 2016, **6**, 86455–86463.
- 61 L. W. Qian, J. X. Sun, C. Hou, J. F. Yang, Y. W. Li, D. Lei, M. X. Yang and S. F. Zhang, *Talanta*, 2017, **168**, 174–182.
- 62 L. J. Zhao, J. Yang, H. L. Ye, F. Q. Zhao and B. Z. Zeng, *RSC Adv.*, 2017, **7**, 4704–4709.
- 63 H. L. Liu, G. Z. Fang, C. M. Li, M. F. Pan, C. C. Liu, C. Fan and S. Wang, *J. Mater. Chem.*, 2012, **22**, 19882–19887.
- 64 G. Z. Fang, C. Fan, H. L. Liu, M. F. Pan, H. D. Zhu and S. Wang, *RSC Adv.*, 2014, **4**, 2764–2771.
- 65 Q. H. Wang, G. Z. Fang, Y. Y. Liu, D. D. Zhang, J. M. Liu and S. Wang, *Food Anal. Methods*, 2017, **10**, 2585–2592.
- 66 H. L. Liu, G. Z. Fang, H. D. Zhu, C. M. Li, C. C. Liu and S. Wang, *Biosens. Bioelectron.*, 2013, **47**, 127–132.
- 67 Y. Liu, M. J. Wang, Z. Y. Li, H. T. Liu, P. He and J. H. Li, *Langmuir*, 2005, **21**, 1618–1622.
- 68 Z. X. Xu, J. Zhou, D. Y. Zhao, X. G. Qiao and J. M. Yang, *J. Food Sci.*, 2010, **75**, C49–C54.
- 69 Z. X. Xu, J. M. Song, D. Y. Zhao, J. Zhou and X. G. Qiao, *Int. J. Polym. Anal. Charact.*, 2011, **16**, 67–77.
- 70 H. F. Wang, Y. Z. Zhu, X. P. Yan, R. Y. Gao and J. Y. Zheng, *Adv. Mater.*, 2006, **18**, 3266–3270.
- 71 H. F. Wang, Y. Z. Zhu, J. P. Lin and X. P. Yan, *Electrophoresis*, 2008, **29**, 952–959.
- 72 X. Gao, R. Li, G. F. Zhu and J. Fan, *Monatsh. Chem.*, 2015, **146**, 475–484.
- 73 X. Gao, J. Fan, G. F. Zhu, X. L. Wang and J. J. Wang, *J. Sep. Sci.*, 2013, **36**, 3277–3284.
- 74 L. F. Yang, X. L. Hu, P. Guan, J. Li, D. F. Wu and B. Gao, *J. Appl. Polym. Sci.*, 2015, **132**, 42485.
- 75 M. A. B. H. Susan, T. Kaneko, A. Noda and M. Watanabe, *J. Am. Chem. Soc.*, 2005, **127**, 4976–4983.
- 76 H. Y. Ma, X. H. Wan, X. F. Chen and Q. F. Zhou, *J. Polym. Sci., Part A: Polym. Chem.*, 2003, **41**, 143–151.
- 77 L. Ban, X. Han, X. H. Wang, Y. P. Huang and Z. S. Liu, *Anal. Bioanal. Chem.*, 2013, **405**, 8597–8605.
- 78 L. W. Qian, X. L. Hu, P. Guan, B. Gao, D. Wang, C. L. Wang, J. Li, C. B. Du and W. Q. Song, *Anal. Bioanal. Chem.*, 2014, **406**, 7221–7231.
- 79 N. Zhang, X. L. Hu, P. Guan, C. B. Du, J. Li, L. W. Qian, X. Y. Zhang, S. C. Ding and B. P. Li, *Chem. Eng. J.*, 2017, **317**, 356–367.
- 80 F. X. Chen, S. L. Xie, J. H. Zhang and R. Liu, *Mater. Lett.*, 2013, **112**, 177–179.
- 81 K. Radosevic, M. C. Bubalo, V. G. Srcek, D. Grgas, T. L. Dragicevic and I. R. Redovnikovic, *Ecotoxicol. Environ. Saf.*, 2015, **112**, 46–53.
- 82 H. G. Morrison, C. C. Sun and S. Neervannan, *Int. J. Pharm.*, 2009, **378**, 136–139.
- 83 X. Li, M. Hou, B. Han, X. Wang and L. Zou, *J. Chem. Eng. Data*, 2008, **53**, 548–550.
- 84 H. G. Liao, Y. X. Jiang, Z. Y. Zhou, S. P. Chen and S. G. Sun, *Angew. Chem., Int. Ed.*, 2008, **47**, 9100–9103.
- 85 Z. Chen, B. Zhou, H. Cai, W. Zhu and X. Zou, *Green Chem.*, 2009, **11**, 275–278.
- 86 H. G. Liao, Y. X. Jiang, Z. Y. Zhou, S. P. Chen and S. G. Sun, *Angew. Chem.*, 2008, **120**, 9240–9243.
- 87 S. R. Liang, H. Y. Yan, J. K. Cao, Y. H. Han, S. G. Shen and L. G. Bai, *Anal. Chim. Acta*, 2016, **951**, 68–77.
- 88 X. X. Li and K. H. Row, *RSC Adv.*, 2017, **7**, 16997–17004.
- 89 X. Li and K. H. Row, *Anal. Sci.*, 2017, **33**, 611–617.
- 90 G. Z. Li, W. S. Ahn and K. H. Row, *J. Sep. Sci.*, 2017, **39**, 4465–4473.
- 91 N. Fu, X. Liu, L. Li, B. Tang and K. H. Row, *J. Sep. Sci.*, 2017, **40**, 2286–2291.

