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Controlled chemical synthesis of CaO₂ particles coated with polyethylene glycol: characterization of crystallite size and oxygen release kinetics

 Arghavan Rastinfard,^a Masoumeh Haghbin Nazarpak^{b*} and Fathollah Moztarzadeh^a

Incorporating oxygen generating substances into tissue constructs is one of the methods for increasing cell survival under hypoxic condition. Calcium peroxide from alkaline earth metal peroxide groups has been proposed as an appropriate compound for supplying sufficient oxygen to cells within the scaffolds. The high surface-to-volume ratio of CaO₂ particles can lead to an increase in oxygen generation ability over a particular time interval. One objective of this study was to decrease calcium peroxide crystallite size *via* changing the local supersaturation level by controlling the reactant addition rate during coprecipitation synthesis. Therefore, the effect of hydrogen peroxide addition rate on the crystallite size, particle size distribution, and CaO₂ purity, which have not been studied before, were discussed. It was found that the crystallite size of CaO₂ depends upon the reactant addition rate, and the crystallite size decreased from 111 to 37 nm when the addition rate increased from 0.29 to 0.88 ml min⁻¹. Furthermore, after determining the hydrogen peroxide addition rate, a facile method was developed to prepare CaO₂ particles with polyethylene glycol coating for the first time. The aim of PEG coating was to improve redispersion of the precipitated CaO₂ particles. Thermogravimetric analysis was used to measure the content of adsorbed PEG on the surface of the particles. Eventually, the oxygen release profile of PEG-coated calcium peroxide particles demonstrated that the release kinetics is dependent on the pH value of the exterior medium, diffusion through the calcium carbonate product layer, and molar ratios of PEG to calcium.

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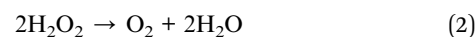
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1. Introduction

Though substantial progress has been achieved in improving the physical environment required for cell growth, engineering an ideal structural scaffold is a persistent challenge.¹ It is typically owing to the insufficient or inevitably delayed vascularization of the scaffold.² That, in turn, hampers oxygen and nutrient delivery and removal of waste products³ because oxygen deficiency arises when cells are beyond 100–200 microns away from blood vessels.⁴ Moreover, tissue trauma can bring about tissue hypoxia and prevent oxygen diffusion due to the destruction of the vascularization network.¹ Eventually, hypoxic condition established as a result of reasons mentioned above can induce cellular necrosis and apoptosis.⁵

The oxygen generating biomaterials, by administering sufficient oxygen for cells, eliminate the onset of hypoxia within tissue constructs from the time of implantation to vascularization,⁶ leading to an increase in cell survival before the

vascularization.⁷ Recently, different types of peroxide compounds such as liquid hydrogen peroxide and inorganic peroxides, *e.g.*, calcium peroxide (CaO₂), magnesium peroxide (MgO₂), and sodium percarbonate (Na₂CO₃·1.5H₂O₂), have been proposed as oxygen generating biomaterials.⁵ Oxygen release from the solid peroxides, contrary to liquid hydrogen peroxide directly decomposing to molecular oxygen, is a two-step process.⁸ Therefore, solid inorganic peroxides can induce more sustained oxygen release compared to liquid hydrogen peroxide.⁵ Inorganic peroxides form hydrogen peroxide as a result of hydrolysis when exposed to water and then liberate oxygen gas by decomposition of hydrogen peroxide.⁹ For example, calcium peroxide releases oxygen *via* the following reactions:¹⁰



However, in alkaline condition (pH = 12–13), CaO₂ is directly converted to molecular oxygen instead of H₂O₂ and no measurable H₂O₂ is released because of increasing the disproportionation of H₂O₂ to O₂ [eqn (2)] by increasing the pH

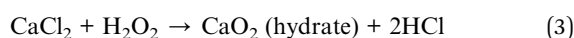
^aDepartment of Biomedical Engineering, Amirkabir University of Technology, P.O. Box: 15875-4413, Tehran 159163-4311, Iran

^bNew Technologies Research Center (NTRC), Amirkabir University of Technology, Tehran 15875-4413, Iran. E-mail: mhaghbin@gmail.com



value.¹¹ Calcium peroxide has been preferred over other inorganic peroxides since the oxygen release from CaO₂ is slower than that from sodium percarbonate.^{10,12} Moreover, CaO₂ enables higher oxygen production capacity compared to MgO₂ due to the higher purity of commercial CaO₂.¹²

Calcium peroxide, contrary to other alkaline earth metal peroxides such as SrO₂ and BaO₂, cannot be synthesized through direct reaction of molecular oxygen with calcium oxide.¹³ Thus, two other methods have been proposed for the preparation of CaO₂. In the first method, CaO₂ is synthesized by the dehydration of CaO₂·8H₂O in water at 50 °C and then vacuum drying of the filtered precipitate at 100 °C.⁹ The latter method consists of the reaction of dilute hydrogen peroxide with calcium oxide hydrate suspension⁹ or the reaction of calcium salt solutions (chloride or nitrate) with ammonia and hydrogen peroxide [eqn (3) and (4)].^{9,14}



The growth of the surface-to-volume ratio of CaO₂ particles in synthesis at nanoscale can lead to increase in oxygen generation ability over a specific time interval. Khodaveisi *et al.*¹⁵ showed that the reaction rate of nanosized CaO₂ was more rapid than that of microsized CaO₂ with silver nanoparticles. It is attributed to rapid decomposition rate of CaO₂ nanoparticles, which induces a higher concentration of H₂O₂ for the degradation of silver nanoparticles. However, the faster hydrolysis of CaO₂ nanoparticles and burst release of oxygen from these particles create hyperoxic conditions and alkaline pH within the engineered scaffold.¹⁶ Therefore, for overcoming this problem and providing sufficient time for early revascularization, oxygen release must take place over time in a controlled and continual manner^{16,17} by incorporating the particles into tissue constructs.⁵ Some methods have been devised for incorporating the solid peroxides into biomaterials such as encapsulating particles into PCL nanofibers,^{18,19} a 3-D scaffold of PLGA,¹⁶ PDMS disks,⁶ films of PLGA,¹⁷ and PTMC microspheres.²⁰ The main problems with the incorporation of inorganic nanoparticles into the hydrophobic polymer are their high surface area, slight miscibility with organic solvents, and a low affinity with hydrophobic polymers; thus, they are agglomerated and will have a non-uniform distribution in the polymer matrix.^{21,22} Hence, it is of the utmost importance that polymers such as poly(ethylene glycol) (PEG),²³ poly(vinyl alcohol),²⁴ and dextran²⁵ are used as a surface stabilizer of the nanoparticles. The adsorption of the stabilizer onto the nanoparticles prevents them from irreversible agglomeration over the precipitation process and forms a stable dispersion *via* the increased steric repulsion.²⁶ Polyethylene glycol has been used in a number of studies on the stabilization of the nanoparticles, since it possesses stability against reduction, oxidation, and decomposition by acid, base, and hydrogen peroxide.²⁷ Additionally, nanoparticles with adsorbed PEG molecules can be soluble in various solvents due to their amphiphilic features.²⁸

In the previous studies, various approaches have been used for controlling calcium peroxide size in the nanoscale. For example, nanosized calcium peroxide has been obtained previously by using calcium nitrate as a precursor,²⁹ polyethylene glycol with low molecular weight as a stabilizer¹⁵ and different dispersants such as PVP and PVA.^{30,31} However, in the present study, decreasing calcium peroxide crystallite size was achieved by controlling the addition rate of hydrogen peroxide and using diethylene glycol monomethyl ether as a stabilizer. As the results demonstrated, the presence of the stabilizer was not sufficient for producing nanosized CaO₂, and the rate of H₂O₂ addition strongly affected the final diameter and particle size distribution. Moreover, this study reported a facile method to prepare CaO₂ particles coated with PEG through the synthesis of particles directly in the PEG medium for the first time. Since the amount of PEG used for preparation had a noticeable impact on the amount of adsorbed PEG on the surface of the particles and redispersion of the precipitated CaO₂ particles, we investigated the effect of PEG/calcium ratio on the amount of the adsorbed molecules. Ultimately, the oxygen release profile, which depends on the shrinking-core model for CaO₂ hydrolysis and the pH of the medium, was evaluated.

2. Materials and methods

2.1. Materials

Hydrogen peroxide (H₂O₂, 30 wt%), calcium chloride dihydrate (CaCl₂·2H₂O, 99%), ammonia solution (NH₃, 25 wt%), sodium hydroxide (NaOH, ≥98%), diethylene glycol monomethyl ether (C₅H₁₂O₃, ≥98%), and polyethylene glycol 4000 (HO(C₂H₄O)_{*n*}H) were purchased from Merck (Germany).

2.2. Synthesis of calcium peroxide particles

In this study, the synthesis of calcium peroxide was performed according to the method established by Philipp *et al.*¹⁴ However, a few modifications were made to the technique. Calcium peroxide was synthesized *via* the precipitation reaction of a calcium chloride solution with hydrogen peroxide (second method) [eqn (3) and (4)]. In this stage, the addition rate of hydrogen peroxide was studied to characterize the optimized flow rate for producing a smaller particle size of crystalline calcium peroxide. The reaction time was one hour. Initially, 0.01 mol calcium chloride dihydrate was dissolved in 15 ml of distilled water, and then the solution temperature was maintained at 70 °C using an oil bath. This temperature was selected due to the decreasing degree of hydration of the precipitated calcium peroxide.¹⁴ Subsequently, 20 ml of diethylene glycol monomethyl ether was added to the stirring solution. The agitation speed was 375 rpm throughout the process. Then 8 ml of aqueous ammonia (25% w/w) was added to the above solution. In the next step, 7 ml of hydrogen peroxide (about 1 : 7 molar ratio of CaCl₂/H₂O₂) was added at different flow rates (from 0.29 to 3.5 ml min⁻¹) by a syringe. Finally, 13 ml of sodium hydroxide solution was added dropwise to the peroxide suspension. The beige-colored precipitates were separated by centrifugation. Subsequently, the resultant precipitate was



washed three times with sodium hydroxide solution and several times with distilled water until the pH value of 10 for the supernatant was achieved. In the final stage, the resultant precipitate was dried at 80 °C for 2 hours in an oven. A flowchart of the synthesis method is shown in Fig. 1.

2.3. Synthesis of PEG-coated calcium peroxide particles

PEG-coated CaO₂ particles were prepared by direct synthesis of particles in the PEG medium. The two samples named CP-PEG1 and CP-PEG2 were prepared as follows. The CP-PEG1 and CP-PEG2 particles were prepared by the addition of PEG, instead of diethylene glycol monomethyl ether, as a steric stabilizer and inhibitor of irreversible agglomeration of nanoparticles throughout the precipitation, with 0.17 and 0.34 molar ratio of PEG/calcium to the solution, respectively. Moreover, a H₂O₂ flow rate of 0.88 ml min⁻¹ and drying temperature of 45 °C were chosen for this stage. Other experimental parameters were identical to those described in the previous section.

2.4. Characterization

The phase analysis of the as-prepared CaO₂ was evaluated by an X-ray diffractometer (Equinox 3000). The powders were scanned over the range of $2\theta = 5\text{--}118^\circ$ using CuK α radiation (wavelength = 1.54187 Å, 30 mA, 40 kV). X'Pert HighScore Plus software was used to analyze the XRD spectra and calculate the full width at half maximum (FWHM) of the peaks.

The surface morphology of the synthesized CaO₂ nanoparticles and the difference between the crystallite and particle

size of the nanoparticles were also characterized using FESEM (MIRA3 TESCAN).

The size distribution of particles was determined by the dynamic light scattering (DLS) method. The DLS technique measures the intensity fluctuations in the scattered light due to the Brownian motion of the particles,³² and then the size of the particles can be calculated from the rate of these fluctuations.³³ The measurements were taken using a Nano ZS (red badge) ZEN 3600 device from Malvern Co, with ethanol as the dispersant and the detection angle of 173°. In this study, the span of distribution as a measure of the width of particle size distribution (PSD) was calculated by eqn (5):

$$\text{span} = \frac{D90 - D10}{D50} \quad (5)$$

where *D*10, *D*50, and *D*90 are the particle sizes that 10, 50, and 90% (v/v) of the number of particles are at or below these sizes, respectively.

The thermogravimetric behavior of the PEG-coated calcium peroxide particles, pure CaO₂, and pure PEG was investigated with a Pyris Diamond thermal analyzer (PerkinElmer Co., USA) in a N₂ gas atmosphere, with a 10 °C min⁻¹ heating rate, and in the temperature range of 25 to 550 °C.

The FTIR spectra of the CP-PEG2 particles and pure PEG were recorded on a Thermo Nicolet Nexus 670 spectrometer in pressed KBr discs. The spectra were obtained in an absorption mode in the 400–4000 cm⁻¹ region.

The oxygen release from CP-PEG1, CP-PEG2, CP2 (calcium peroxide synthesized with H₂O₂ addition rate of 0.58 ml min⁻¹), and CP3 (calcium peroxide synthesized with H₂O₂ addition rate

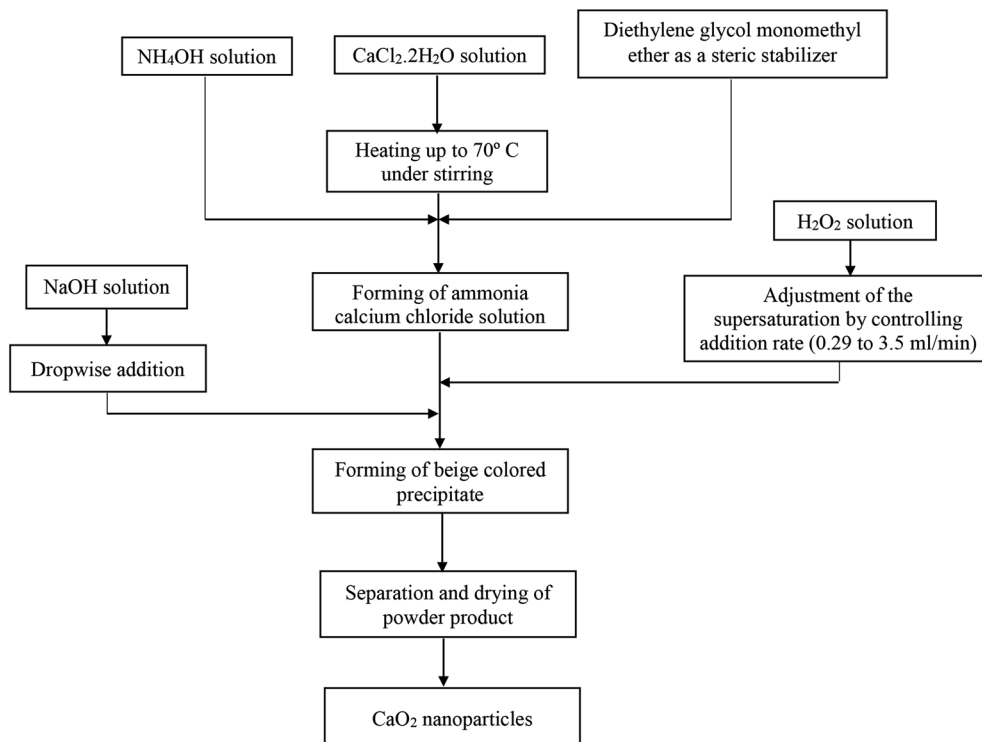


Fig. 1 Flowchart of the synthesis of calcium peroxide nanoparticles.



of 0.88 ml min^{-1}) samples immersed in distilled water (1 g l^{-1}) was quantified by a portable oxygen meter (MO128, Mettler-Toledo, Switzerland). The pH evaluation was performed using a pH meter (WP-80, TPS). The dissolved oxygen equilibrium concentration in the distilled water without the presence of CaO_2 particles was taken into account as a control.

2.5. Statistical analysis

All results were described as mean \pm standard deviation. One-way ANOVA followed by post-hoc LSD tests were performed to specify whether differences existed between the released oxygen levels from CP-PEG1, CP-PEG2, and control samples. A p -value < 0.05 was considered a significant statistical difference.

3. Results and discussion

3.1. XRD analysis

Fig. 2 shows the X-ray diffraction patterns of the synthesized samples with different addition rates of hydrogen peroxide. The XRD patterns of the as-prepared CaO_2 at all reactant addition rates completely matched the simulated XRD of the orthorhombic CaO_2 structure by Zhao *et al.*³⁴ It is concluded that the synthesized calcium peroxide structure was orthorhombic. As indicated in Fig. 2, no extra diffraction peaks corresponding to $\text{Ca}(\text{OH})_2$ and CaO were detected, but the XRD patterns showed the peaks attributable to the (1 0 4) and (1 1 6) planes of calcite. Comparison between the (1 0 4) peak of calcite at $2\theta = 29.14^\circ$ revealed that the increase in the H_2O_2 addition rate from 0.29 to 0.88 ml min^{-1} did not significantly affect the relative intensity. Nevertheless, as shown in Table 1, the relative intensity of the (1 0 4) peak significantly increased from 0.21 to 0.64 when the addition rate of H_2O_2 increased from 0.88 to 3.5 ml min^{-1} , respectively. Therefore, the faster addition of H_2O_2 affected the purity of the CaO_2 synthesized; the H_2O_2 addition rate ranged

Table 1 Intensity ratio of the (1 0 4) peak corresponding to calcite for various samples

Addition rate of H_2O_2 (ml min^{-1})	Intensity ratio of the (1 0 4) peak of calcite
0.29	0.23
0.58	0.20
0.88	0.21
2.3	0.29
3.5	0.64

from 0.29 to 0.88 ml min^{-1} was a suitable condition for the synthesis of CaO_2 with a low quantity of impurities.

3.2. Crystallite size and strain

Broadening of diffraction peaks is commonly associated with instrumental effects, crystallite size, and lattice strain. According to the material, crystallite may mean a domain or grain. A domain is the coherently diffracting part of the specimen. In the nanocrystalline particles with very low grain sizes, the domain does not form a substructure; consequently, crystallite, domain, and grain are identical.³⁵ The instrument-corrected broadening of CaO_2 diffraction peaks due to crystallite size and lattice strain was calculated by eqn (6):

$$\beta^2 = \beta_M^2 - \beta_S^2 \quad (6)$$

where β_S is the measured breadth of the line of the standard material (instrumental broadening) and β_M is the measured breadth of the diffraction line.³⁶

The average crystallite size was estimated by the Williamson-Hall (W-H) eqn (7):

$$\beta \cos \theta = \frac{k\lambda}{D} + 4\epsilon \sin \theta \quad (7)$$

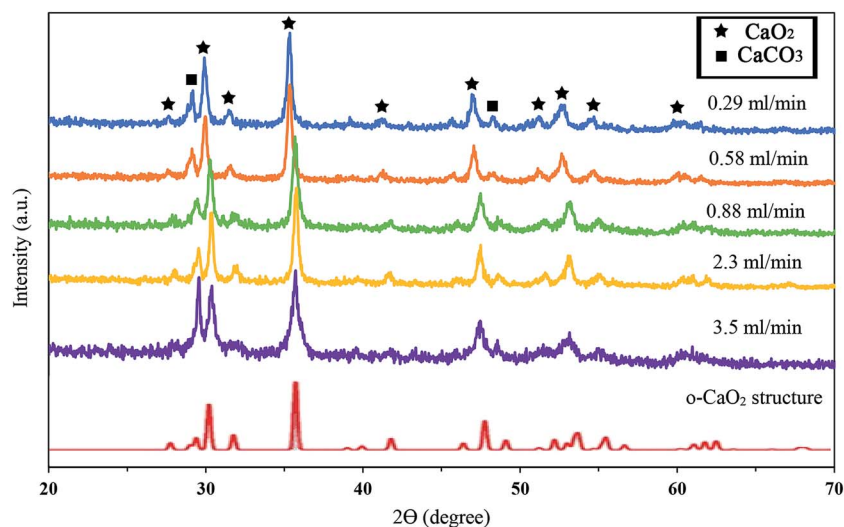


Fig. 2 XRD patterns of synthesized CaO_2 with different addition rates of hydrogen peroxide and the simulated XRD spectra of the orthorhombic CaO_2 structure.



where λ is the wavelength of CuK α radiation, θ is the Bragg's angle, D is the average crystallite size, k is the shape factor, and ε is the strain in the material. $\beta \cos \theta$ was plotted against $4 \sin \theta$ for the specimens with a lower percentage of impurity, which is evident in Fig. 3. It is clear that crystallite size and lattice strain were extracted from the y-intercept and the slope of the fitted line respectively.³⁷ The crystallite size and lattice strain of the particles are summarized in Table 2.

Fig. 2 depicts the relationship between the position of the CaO₂ main peak and H₂O₂ addition rate. It can be noticed that the peak shifted from 35.29 to 35.65° on increasing H₂O₂ addition rate from 0.29 to 0.88 (ml min⁻¹). The peak shift to higher scattering angles can be due to the decreased crystallite size (Table 2). As shown in Table 2, the crystallite size of CaO₂ decreases from 111 to 37 nm as the addition rate increases from 0.29 to 0.88 ml min⁻¹. Therefore, the crystallite size depends on the reactant addition rate. The reactant supersaturation in solution is crucial to controlling the particle size. A high supersaturation can result in a reduction in critical nucleus radius. The relationship between the critical nucleus radius and supersaturation is proved by the classical nucleation theory. The critical nucleus size will decline with increasing supersaturation level according to the classical equation (Mullin 1972).³⁸ Therefore, the decreased critical nucleus size as a result of increasing supersaturation, the probability of nucleation compared to the probability of crystal growth will increase.³⁹ As a consequence, smaller crystallites are produced from the solution with more supersaturation.

Furthermore, a larger supersaturation boosts the nucleation rate.^{40,41} The production of a many nuclei as a result of high nucleation rate can induce a large surface area for growth, which in turn decreases the nucleus growth. As a consequence, smaller crystallites are produced from the solution with higher supersaturation due to reducing critical nucleus size and higher nucleation rate.

Fig. 4 shows the FE-SEM micrographs of CaO₂ nanoparticles synthesized using the addition rate of 0.88 ml min⁻¹. The images reveal the average particle size of 44 ± 14 nm, which is in good agreement with the crystallite size estimated by the Williamson–Hall (W–H) equation. Moreover, the images indicate the heterogeneous surface morphology of nanoparticles.

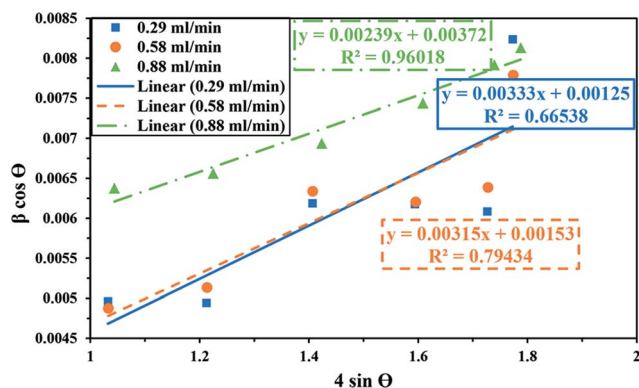


Fig. 3 W–H analysis of synthesized CaO₂ with different addition rates of hydrogen peroxide.

Table 2 Crystallite size, lattice strain and main peak position for different samples

Addition rate of H ₂ O ₂ (ml min ⁻¹)	Crystallite size (nm)	Lattice strain	Main peak position (°)
0.29	111	0.00333	35.29
0.58	90	0.00315	35.32
0.88	37	0.00239	35.65

3.3. Particle size distribution

The volume-mean particle size and particle size distribution (PSD) of the CaO₂ particles were also determined by DLS measurement. The volume particle size distribution is shown in Fig. 5. Each reactant addition rate gave unimodal PSDs with different peak modes. The volume-mean size for various H₂O₂ addition rates of 0.29, 0.58, and 0.88 ml min⁻¹ were 278.02, 133.49, and 134.81 nm, respectively. In comparison to the crystallite size calculated by XRD, the DLS showed a larger diameter. The larger particle diameter, which was assessed by the DLS technique, was due to measuring the hydrodynamic diameter instead of the physical diameter⁴² and the interaction forces in the solution.⁴³ Moreover, the DLS method is sensitive to the polydispersity of the particles, which means that the wide PSD curve at the reactant addition rate of 0.29 ml min⁻¹ (Fig. 5) can induce a crucial difference compared to the XRD crystallite size. Therefore, DLS mean diameter of CaO₂ particles (278.02 nm) was more than twice as large as the crystallite size (111 nm).

As can be seen, DLS mean diameter of CaO₂ particles decreased from 278.02 to 133.49 nm when the reactant addition rate increased from 0.29 to 0.58 ml min⁻¹, which was consistent with the XRD results. However, a higher addition rate did not affect DLS mean diameter significantly. As shown in Table 3, the span of the distribution of the particles synthesized with the reactant addition rate of 0.58 ml min⁻¹ was 0.333, compared with 0.4363 for the reactant addition rate of 0.88 ml min⁻¹. Consequently, although the D10 particle size of the addition rate of 0.88 ml min⁻¹ was less than that of the addition rate of 0.58 ml min⁻¹, the variance in the particle size can lead to an identical DLS mean diameter.

The data in Table 3 represents the significant impact of H₂O₂ addition rate on the PSD of CaO₂. It is evident that a nonlinear relationship existed between the span of distribution and H₂O₂ addition rate, as by increasing the reactant concentration, first, a steep decrease and then a slight rise in span were observed. When the addition rate of H₂O₂ was slow, possibly due to the reactant concentration diminishing below the supersaturation level at the end of the growth stage, Ostwald ripening phenomenon occurred; in this phenomenon, the smaller particles dissolve and then precipitate onto the larger particles.^{37,41} Broadening of the size distribution as a result of stopping the growth process in the Ostwald ripening stage⁴⁴ can be achieved because the smaller particles are swallowed by the larger ones.^{45,46} As a consequence, the span, as a measure of the size distribution, was higher than that of the rest of the samples when the H₂O₂ addition rate was 0.29 ml min⁻¹.



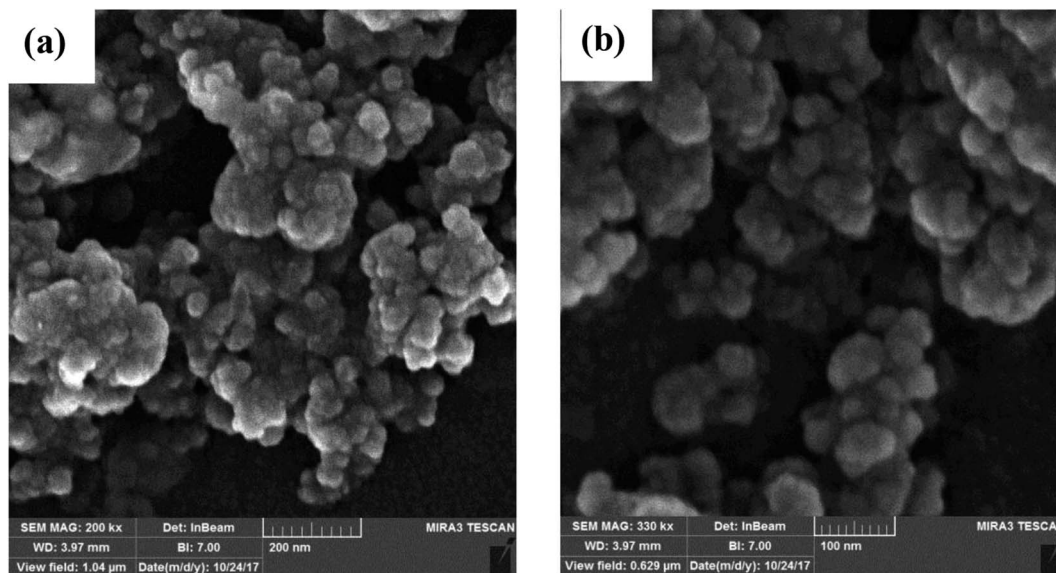


Fig. 4 FE-SEM micrographs of CaO_2 nanoparticles synthesized using an addition rate of 0.88 ml min^{-1} with different magnifications (a and b).

With the reactant addition rate increasing to 0.58 ml min^{-1} , higher local supersaturation and higher reagent concentration in the solution induced conditions in which the particles underwent diffusion-controlled growth. For the diffusion-controlled growth, the smaller particles grew more quickly than the larger ones. Consequently, the size distribution became narrower by growing particulate CaO_2 , and according to the PSD analysis, the span of distribution decreased to 0.333 (Table 3).

By increasing the rate of H_2O_2 addition to 0.88 ml min^{-1} , the PSD curve somewhat broadened. Since higher H_2O_2 concentration in the solution could result in a more extended nucleation stage by continuing the nucleation process during the growth process, it is concluded that additional nucleation caused an increment in the span of distribution. Eventually, the addition rate of 0.88 ml min^{-1} was considered as the optimal flow rate to precipitate the smallest particles (37 nm) with higher oxygen generation ability over a given time.

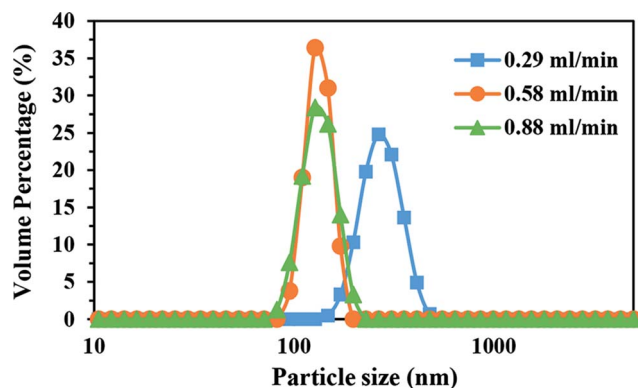


Fig. 5 Volume particle size distribution of synthesized CaO_2 with different H_2O_2 addition rates.

3.4. Characterization of PEG-coated calcium peroxide particles

In this study, TGA analysis and FTIR spectroscopy were used to conclusively prove the PEG coating on the surface of CaO_2 particles. The TGA thermograms of CP-PEG1, CP-PEG2, pure CaO_2 and pure PEG along with their first (DTG) and second derivatives (d2TG) are represented in Fig. 6. The TGA curve indicated that the weight loss processes of CP-PEG1 and CP-PEG2 occur in three steps in the temperature region of 50–550 °C. The first weight loss, which began at the onset of the recording, was due to the removal of water molecules and loss of water of hydration. The step height of this stage was about 4 and 4.8% for CP-PEG1 and CP-PEG2, respectively. In the case of the CP-PEG2 sample, there was a peak in the DTG curve (between 356 and 395 °C) before the sharp weight loss resulting from CaO_2 decomposition (Fig. 6b). Since the DTG curve of the pure CaO_2 sample (Fig. 6c) contained only one peak at 400 °C, which corresponds to CaO_2 decomposition, this peak could be attributed to the desorption and subsequent evaporation of the PEG molecular chains. The endpoint temperature for this step was defined by the corresponding zero crossover in the d2TG curve.⁴⁷ However, the DTG curve of the CP-PEG1 sample exhibited an inflection point instead of a distinct positive peak in the temperature range of 359 to 380 °C and then showed

Table 3 PSD analysis of synthesized CaO_2 with different addition rates of hydrogen peroxide

Addition rate of H_2O_2 (ml min^{-1})	PSD analysis			
	D10 (nm)	D50 (nm)	D90 (nm)	span
0.29	177	258	385	0.8031
0.58	106	123	147	0.3330
0.88	95	123	148	0.4363



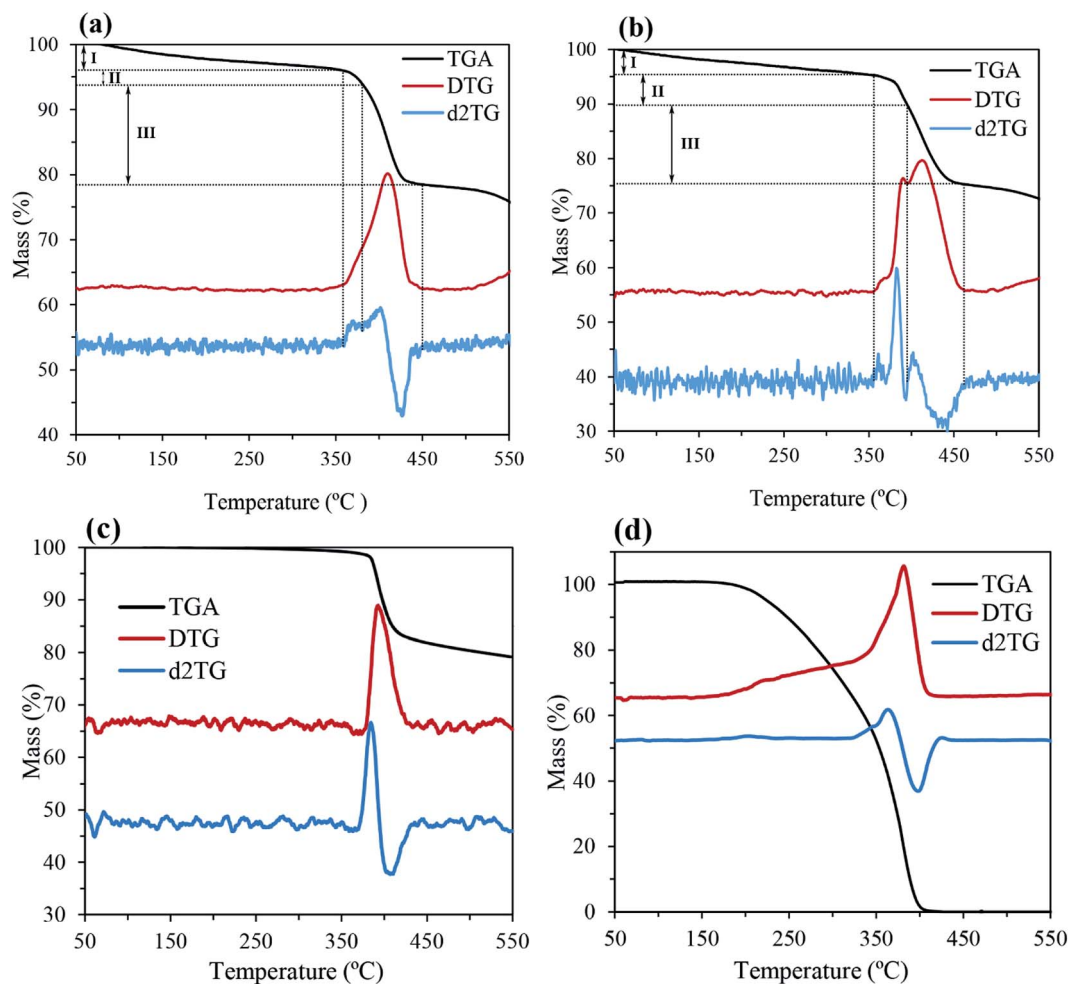
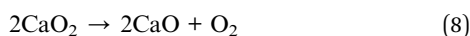


Fig. 6 TGA, DTG, and d2TG curves of (a) CP-PEG1, (b) CP-PEG2, (c) pure CaO₂ and (d) pure PEG.

a minimum in the second derivative of the TGA curve; this minimum is used to determine the endpoint of this step⁴⁷ (Fig. 6a). The weight losses for two samples of CaO₂ are indicated in Table 4. The content of adsorbed PEG on the surface of the CP-PEG2 sample (higher molar ratio of PEG/calcium) was about 3% higher than that on the CP-PEG1 surface. According to the results, a molar ratio of 0.34 was necessary for the adsorption of PEG on the CaO₂ surface.

Furthermore, one common peak between 380 and 460 °C in the differential of the TGA traces of CP-PEG1 and CP-PEG2 samples was observed (Fig. 6a and b). Since the CaO₂ decomposition begins at 375 °C,⁹ these peaks can be assigned to the intensive decomposition of CaO₂ and liberation of oxygen, so that the CaO₂ content can be obtained from the stoichiometry of the following decomposition reaction [eqn (8)]:



The percentage of CaO₂ in each sample, CP, is calculated using eqn (9):

$$\text{CP (\%)} = \frac{\Delta m}{m_0} \times \frac{72.08}{0.5 \times 32} \times 100 \quad (9)$$

where Δm is the weight loss (g), m_0 is the initial (anhydrous) mass of the sample (g), the ratio of 72.08 : 32 is the molar mass of CaO₂ to the eliminated oxygen, and 0.5 is the number of moles of oxygen liberated for each mole of the sample.

The CaO₂ content for CP-PEG1 and CP-PEG2, calculated by the above method, was 73.19 and 70.24 wt%, respectively. The purity percentage of the as-prepared CaO₂ coincides with that of commercial CaO₂ since the purity of these products such as 466271 Aldrich and IXPEN® 60C is around 60–80% CaO₂. The weight loss continuing after the endpoint of CaO₂ decomposition was attributed to CaCO₃ decomposition, because CaCO₃ decomposes in the temperature range of 500 to 700 °C.⁴⁸ As indicated in the XRD results (Fig. 2), calcite is the only calcium compound other than CaO₂ that was precipitated over the synthesis, and no other characteristic peak was identified in the XRD pattern. The main reason for calcite production was possibly the carbonation of Ca(OH)₂, which is formed as a result of hydrolysis of the precipitated calcium peroxide, during the synthesis. Hence, the main impurity was CaCO₃ at about 24.81 and 24.76 wt% for the CP-PEG1 and CP-PEG2 samples, respectively.



Table 4 Weight losses (%) and endpoint temperature of the three steps for CaO₂ samples using thermogravimetric analysis

Event	Weight loss in the TGA curve (%)		Endpoint temperature (°C)	
	CP-PEG1	CP-PEG2	CP-PEG1	CP-PEG2
The removal of physical adsorptive water	4	4.8	359	356
Degradation of PEG	2.2	5.2	380	395
Decomposition of CaO ₂	15.2	14.8	450	464

Fig. 7 shows the comparison of DTG curves of pure PEG, pure CaO₂, CP-PEG1, and CP-PEG2. It can be seen that the DTG curves of the PEG-coated CaO₂ particles showed a definite shift to higher values compared to those of pure PEG and CaO₂. In the case of the CP-PEG2 sample, it was worth mentioning that the degradation temperature of PEG shifted from 364 to 390 °C. Furthermore, the temperature of CaO₂ decomposition shifted toward a higher temperature, from 400 to 412 °C.

Fig. 8 depicts the FTIR spectra of the precipitated particles at molar ratio of PEG/calcium of 0.34 and the spectra of the polymer alone in the range 400–4000 cm⁻¹. One of the common absorption peaks between PEG alone and CP-PEG2 was a broadband peak, which is attributed to the stretching mode of the -OH group of the PEG and that of the lattice water molecules of the synthesized calcium peroxide in the spectral region of 3000–3700 cm⁻¹. Moreover, two peaks at 2857 and 2929 cm⁻¹ (-CH₂- band),⁴⁹ 1635 cm⁻¹ band of the H-O-H bending of residual free water,⁵⁰ and C-O-C band at 1076 cm⁻¹ (ref. 23) were common. Consequently, the presence of the bands assigned to PEG in the CP-PEG2 sample and a slight reduction in mass by 5% resulting from PEG degradation in the TGA curve of CP-PEG2 confirmed the adsorption of the polymer onto the particle surface.

Moreover, the existence of CaO₂ and calcite, which XRD and TGA results identified as an impurity of CaO₂ synthesis, was also confirmed by FTIR analysis. For example, the absorption peak at 875 cm⁻¹ was attributed to the O-O bridge of CaO₂ (ref. 51) and the very weak absorption at 711 cm⁻¹ was due to the vibration of the carbonate ion in calcite (Fig. 8).⁵² Furthermore, a broad peak in the interval of 1600–1200 cm⁻¹ was caused from

the superposition of O-Ca-O of CaO₂ and the CO₃²⁻ group of calcite. Therefore, the smoothed second derivative spectra were calculated to determine the position of the overlapping peaks (Fig. 9). As represented in Fig. 9b, several peaks at 1490, 1440, 1413, and 1384 cm⁻¹ were observed. In fact, those at 1413 and 1490 cm⁻¹ could be associated with the bending vibration mode of O-Ca-O⁵³ and the two peaks at 1440 and 1384 cm⁻¹ corresponded to the CO₃²⁻ group.^{53–55}

In this study, the effective factors on the variation of the measured oxygen concentration in distilled water over a period of 4 days for two inorganic peroxides were determined. Fig. 10a shows the oxygen release kinetics and pH changes of the surrounding medium for the CP-PEG1 and CP-PEG2 particles, as well as for the control sample during the initial minutes. The release profile of PEG-coated CaO₂ particles showed an initial burst release of oxygen, in which the dissolved oxygen (DO) of CP-PEG1 and CP-PEG2 could reach nearly 2- and 1.7-fold of dissolved oxygen equilibrium concentration in distilled water within 90 minutes, respectively. This significant burst release was due to the hydrolysis reaction on the surface of the particles with high surface-to-volume ratio and the lack of a diffusional barrier initially. Interestingly, the oxygen saturation level of the CP-PEG1 sample was higher than the CP-PEG2 sample because, according to the TGA results, the calcium peroxide content or available oxygen of CP-PEG1 was higher than that of CP-PEG2.

Moreover, most of the oxygen was released on the first day, and oxygen generation decreased subsequently (Fig. 10b). Two main reasons of oxygen release reduction can be related to:

- According to the Fig. 10a, the pH value in the medium surrounding the particles increased from neutral to 11 within

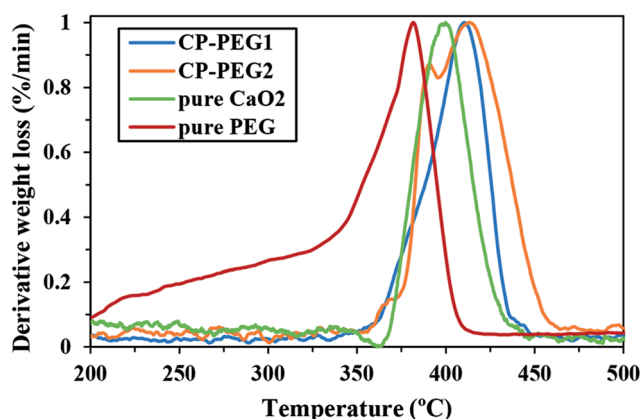
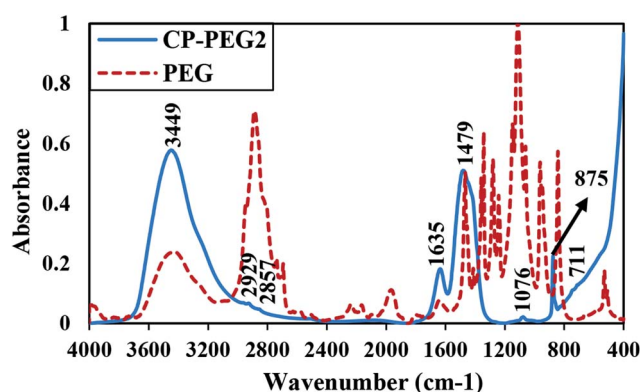
Fig. 7 DTG curves of CP-PEG1, CP-PEG2, pure CaO₂, and pure PEG.

Fig. 8 FTIR spectra of PEG alone and PEG stabilized particles (CP-PEG2).



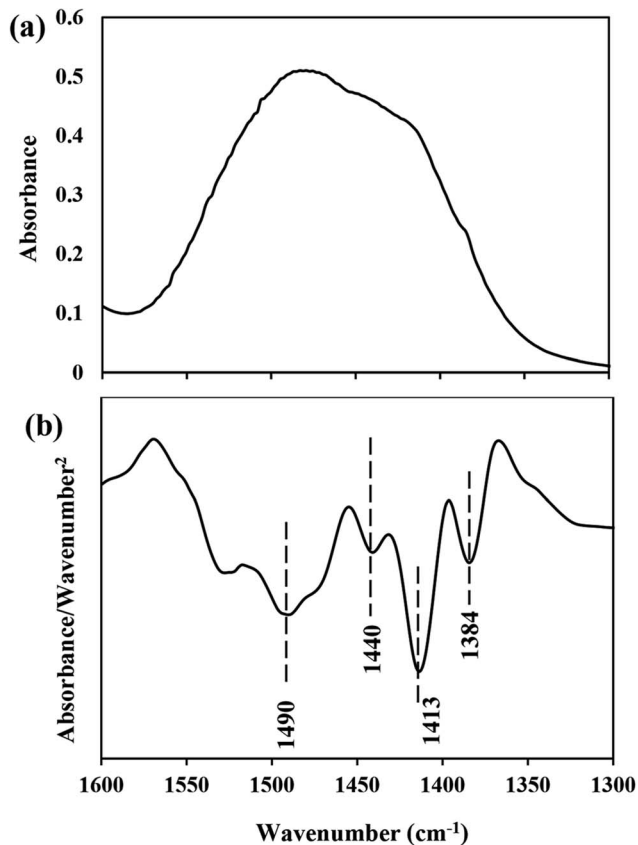


Fig. 9 (a) FTIR spectra and (b) second derivative of CP-PEG2.

90 minutes. The pH variation was attributed to the calcium hydroxide generation (a secondary product of the CaO_2 hydrolysis). An increase in the pH level of the medium increases the O_2 yield, however acts as an inhibitor of the oxygen release process and lower dissolution rate of calcium peroxide.¹¹ Therefore, an increase in the pH value of the exterior medium of two samples caused a noticeable decrease in the oxygen levels.

• Waite *et al.*¹⁰ stated that oxygen release rate is controlled by diffusion through the $\text{Ca}(\text{OH})_2(\text{s})$ product layer that precipitates during the CaO_2 dissolution. However, according to the solubility of $\text{Ca}(\text{OH})_2$ in water (0.16^{20} g/100 g water⁵⁶) and 1 : 1 mole ratio between $\text{Ca}(\text{OH})_2$ and CaO_2 in the hydrolysis reaction, the amount of ions added in this study was lower than $\text{Ca}(\text{OH})_2$ solubility. Alternatively, the declining pH level of the medium after the first day (Fig. 10b) confirmed the gradual decrease of hydroxide ion content due to carbonation. Thus, the rate of oxygen release was controlled by diffusion through the CaCO_3 precipitated layer as a result of carbonation of the $\text{Ca}(\text{OH})_2$ over time.

As shown in Fig. 10, the oxygen release pattern of CaO_2 nanoparticles was biphasic: an initial burst phase for the first 24 hours followed by a descending release rate for three days. The initial burst oxygen release was affected by the nanoparticle characteristics including particle size, particle size distribution, purity of CaO_2 , and amount of adsorbed PEG. However, the oxygen release rate in the second release phase was slightly affected by the nanoparticle properties; the second phase was

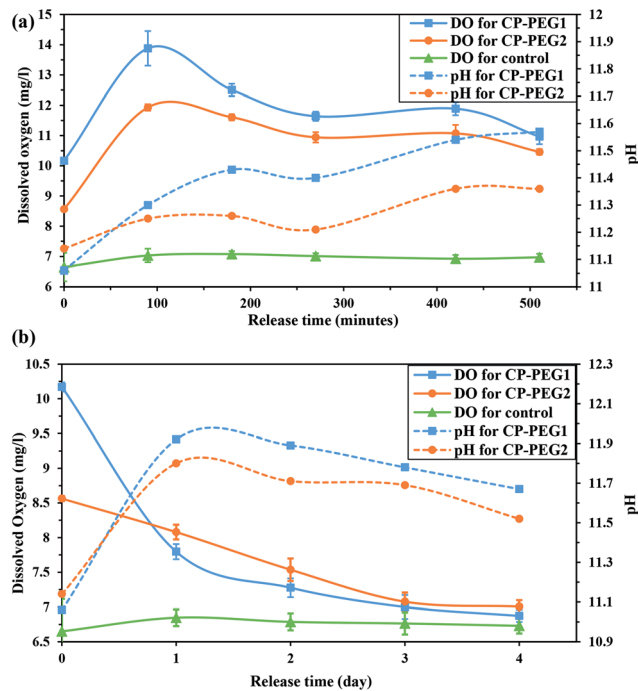


Fig. 10 pH variation and release of O_2 from two CaO_2 samples (a) within 510 minutes and (b) within 4 days.

mainly controlled by high pH value of the exterior medium and diffusion through the precipitated product layer on the surface of the particles (as discussed previously). Moreover, the precipitation of the product layer and the increase in the pH level of the medium as a result of the significant oxygen production during the initial hours induced a descending oxygen release rate. In order to compare oxygen production by CP-PEG1 and by CP-PEG2 during the two phases, the area under the curve [AUC_(0–510 min)] was calculated by the linear trapezoidal rule.⁵⁷ The level of total oxygen production by the CP-PEG1 particles (28.84 mg l^{-1}) over 510 minutes was higher than that by the CP-PEG2 particles (22.91 mg l^{-1}) possibly due to the higher purity of CP-PEG1. However, the oxygen released by CP-PEG2 (1.83 mg l^{-1}) and CP-PEG1 (1.28 mg l^{-1}) particles substantially declined over the last three days. Decreasing the oxygen generation by CP-PEG1 and CP-PEG2 samples in the second stage confirmed the significant impact of oxygen release controlling factors after burst oxygen release. Therefore, the oxygen release rate of CP-PEG1 and CP-PEG2 samples due to impacts of oxygen release controlling factors was almost the same after the first day.

The statistical comparison between the released oxygen amount of CaO_2 particles and the dissolved oxygen in the control sample exhibited a significant O_2 level introduced by the CP-PEG2 sample throughout the analysis ($p < 0.05$), although there was no meaningful difference among the CP-PEG1 and control samples on the third and fourth days (Fig. 11). It is possibly due to the higher adsorption of PEG on the surface of CP-PEG2 particles, which roughly inhibited the agglomeration of particles and thereby boosted the oxygen production capability by virtue of exposing a higher surface area for O_2 release.



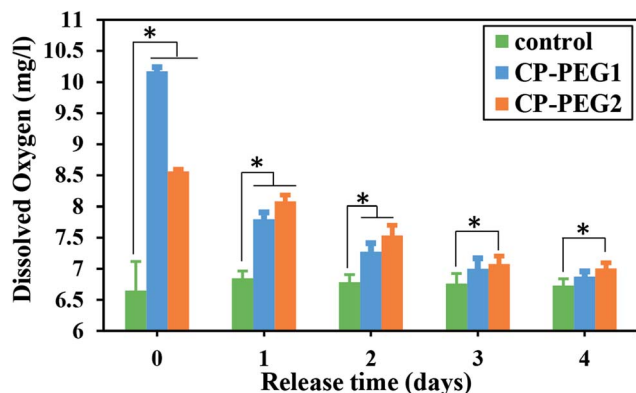


Fig. 11 Histogram of the released oxygen from two CaO_2 samples in distilled water per day. Asterisk represents the significant difference between each other, $n = 3$ for each group, $p < 0.05$.

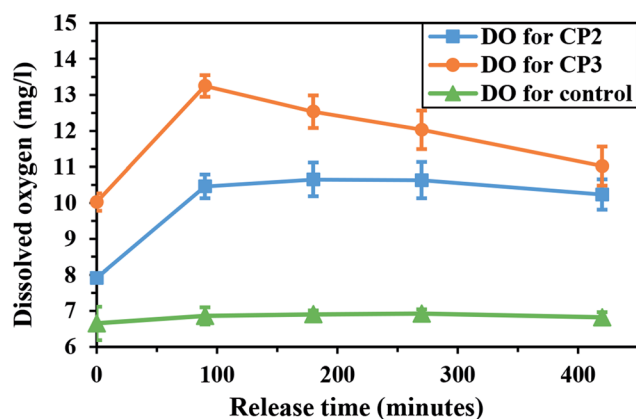


Fig. 12 Release of O_2 from synthesized CaO_2 with different H_2O_2 addition rates within 420 minutes.

Therefore, a higher amount of adsorbed PEG led to a more sustained oxygen release from the particles.

The synthesis of the calcium peroxide particles with smaller size can lead to an increase in oxygen generation ability over a particular time interval (as discussed before). In order to confirm the effect of the CaO_2 crystallite size on the oxygen release, the oxygen release behavior of calcium peroxide synthesized with different reactant addition rates of 0.58 (CP2) and 0.88 ml min^{-1} (CP3) was examined (Fig. 12). The area under the curve [AUC_(0–420 min)] was used to evaluate oxygen generation ability within 420 minutes. The level of total oxygen production for CP3 particles (24.02 mg l^{-1}) over 420 minutes was higher compared to that for CP2 particles (15.76 mg l^{-1}) with larger crystallite size. Therefore, the initial oxygen production increased with an increment in the H_2O_2 addition rate. The oxygen released during the initial hours was probably due to hydrolysis reaction on the surface of the particles. Consequently, smaller crystallites with a higher surface-to-volume ratio as a result of increasing the rate of H_2O_2 addition led to higher oxygen production over a specific period.

4. Conclusion

In this study, calcium peroxide particles with oxygen release capability and orthorhombic structure were synthesized *via* the precipitation reaction of an ammonia calcium chloride solution with hydrogen peroxide. The effect of hydrogen peroxide addition rate as an effective factor on the purity, crystallite size, and particle size distribution of the synthesized CaO_2 was inspected. It was found that the H_2O_2 addition rate affects the purity of the synthesized CaO_2 . Moreover, smaller crystallites were produced from the solutions with a faster addition rate, so there could be an indirect relationship between the crystallite size and reactant addition rate. The results showed that the slow addition rate could increase the span of distribution due to the Ostwald ripening process; however, the higher local supersaturation could induce diffusion controlled growth and lead to narrower PSD. Additionally, PEG-coated CaO_2 particles were achieved by the synthesis of particles directly in PEG medium. The presence of PEG on the surface of CaO_2 particles with a higher molar ratio of PEG to calcium was proved by the TGA analysis and FTIR spectroscopy. The PEG-coated particles exhibited an initial burst release of oxygen due to the high surface-to-volume ratio; however, the rate of oxygen release was controlled by diffusion through the precipitated layer over time. Moreover, a more sustained oxygen release from the particles having more amount of adsorbed PEG was identified.

Conflicts of interest

There are no conflicts to declare.

References

- M. Gholipourmalekabadi, S. Zhao, B. S. Harrison, M. Mozafari and A. M. Seifalian, *Trends Biotechnol.*, 2016, **34**, 1010–1021.
- T. L. Schenck, M. N. Chávez, A. P. Condurache, U. Hopfner, F. Rezaeian, H.-G. Machen and J. T. Egaña, *J. Visualized Exp.*, 2014, 51428.
- H. Steg, A. T. Buizer, W. Woudstra, A. G. Veldhuizen, S. K. Bulstra, D. W. Grijpma and R. Kuijjer, *J. Mater. Sci.: Mater. Med.*, 2015, **26**, 1–4.
- J. Folkman, P. Hahnfeldt and L. Hlatky, *Nat. Rev. Mol. Cell Biol.*, 2000, **1**, 76–79.
- G. Camci-Unal, N. Alemdar, N. Annabi and A. Khademhosseini, *Polym. Int.*, 2013, **62**, 843–848.
- E. Pedraza, M. M. Coronel, C. A. Fraker, C. Ricordi and C. L. Stabler, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 4245–4250.
- L. Yang, L. Zhu, W. Dong, Y. Cao and Z. Rong, *Bone*, 2013, **57**, 322–323.
- A. L. Farris, A. N. Rindone and W. L. Grayson, *J. Mater. Chem. B*, 2016, **4**, 3422–3432.
- I. I. Volnov, *Peroxides, Superoxides, and Ozonides of Alkali and Alkaline Earth Metals*, Plenum Press, New York, 1966.
- A. J. Waite, J. S. Bonner and R. Autenrieth, *Environ. Eng. Sci.*, 1999, **16**, 187–199.



- 11 A. Northup and D. Cassidy, *J. Hazard. Mater.*, 2008, **152**, 1164–1170.
- 12 D. P. Cassidy and R. L. Irvine, *J. Hazard. Mater.*, 1999, **69**, 25–39.
- 13 P. Ehrlich, *Handbook of preparative inorganic chemistry*, G. Brauer, Academic Press, New York, 2nd edn, 1963, ch. 17, vol. 1, pp. 887–949.
- 14 W. H. Philipp and P. A. Kraft, *Annual Meeting of the American Institute of Chemical Engineers*, San Francisco, CA, 1989.
- 15 J. Khodaveisi, H. Banejad, A. Afkhami, E. Olyaie, S. Lashgari and R. Dashti, *J. Hazard. Mater.*, 2011, **192**, 1437–1440.
- 16 S. H. Oh, C. L. Ward, A. Atala, J. J. Yoo and B. S. Harrison, *Biomaterials*, 2009, **30**, 757–762.
- 17 B. S. Harrison, D. Eberli, S. J. Lee, A. Atala and J. J. Yoo, *Biomaterials*, 2007, **28**, 4628–4634.
- 18 C. L. Ross, B. Bonfert, L. Smith, S. Y. Jeong and B. S. Harrison, *J. Sci. Appl. Biomed.*, 2015, **3**, 1–4.
- 19 J. Wang, Y. Zhu, H. K. Bawa, G. Ng, Y. Wu, M. Libera, H. C. Van Der Mei, H. J. Busscher and X. Yu, *ACS Appl. Mater. Interfaces*, 2011, **3**, 67–73.
- 20 H. Steg, A. T. Buizer, W. Woudstra, A. G. Veldhuizen, S. K. Bulstra, D. W. Grijpma and R. Kuijter, *Polym. Adv. Technol.*, 2016, **28**, 1252–1257.
- 21 N. G. Rim, C. S. Shin and H. Shin, *Biomed. Mater.*, 2013, **8**, 014102.
- 22 H. W. Kim, H. H. Lee and J. C. Knowles, *J. Biomed. Mater. Res., Part A*, 2006, **79**, 643–649.
- 23 S. Garcia-Jimeno and J. Estelrich, *Colloids Surf., A*, 2013, **420**, 74–81.
- 24 H. Pardoe, W. Chua-anusorn, T. G. St. Pierre and J. Dobson, *J. Magn. Magn. Mater.*, 2001, **225**, 41–46.
- 25 B. R. Jarrett, M. Frendo, J. Vogan and A. Y. Louie, *Nanotechnology*, 2007, **18**, 035603.
- 26 S. A. Simakov and Y. Tsur, *J. Nanopart. Res.*, 2007, **9**, 403–417.
- 27 A. S. Karakoti, S. Das, S. Thevuthasan and S. Seal, *Angew. Chem., Int. Ed.*, 2011, **50**, 1980–1994.
- 28 R. A. Sperling and W. J. Parak, *Philos. Trans. R. Soc., A*, 2010, **368**, 1333–1383.
- 29 P. Kaewdee, N. Chandet, G. Rujijjanagul and C. Randorn, *Catal. Commun.*, 2016, **84**, 151–154.
- 30 Z. Xuefei, Z. Alec, Q. Yajie, Z. Jing, C. Jiabin and H. Haiping, *CN Pat.*, CN103601155 A, 2014.
- 31 Z. Xuefei, Z. Alec, Z. Jing, Q. Yajie and H. Haiping, *CN Pat.*, CN103964395 A, 2014.
- 32 M. Kaszuba, D. McKnight, M. T. Connah, F. K. McNeil-Watson and U. Nobbmann, *J. Nanopart. Res.*, 2008, **10**, 823–829.
- 33 Malvern Instruments Ltd., *Zetasizer nano series user manual*, 2013.
- 34 X. Zhao, M. C. Nguyen, C.-Z. Wang and K.-M. Ho, *RSC Adv.*, 2013, **3**, 22135–22139.
- 35 C. Suryanarayana and M. Grant Norton, *X-Ray Diffraction A practical Approach*, Springer, New York, 1998.
- 36 B. D. Cullity, *Elements of X Ray Diffraction*, Addison-Wesley Publishing Company, Inc., 2nd edn, 1978, pp. 281–285.
- 37 V. Mote, Y. Purushotham and B. Dole, *J. Theor. Appl. Phys.*, 2012, **6**, 1–8.
- 38 M. A. Larson and J. Garside, *Chem. Eng. Sci.*, 1986, **41**, 1285–1289.
- 39 E. Kramer, J. Podurgiel and M. Wei, *Mater. Lett.*, 2014, **131**, 145–147.
- 40 L. Suber, I. Sondi, E. Matijević and D. V. Goia, *J. Colloid Interface Sci.*, 2005, **288**, 489–495.
- 41 K. Lu, *Nanoparticulate Materials: Synthesis, Characterization, and Processing*, Wiley, Hoboken, 2012.
- 42 D. Chicea, E. Indrea and C. M. Cretu, *J. Optoelectron. Adv. Mater.*, 2012, **14**, 460–466.
- 43 M. V. Sujitha and S. Kannan, *Spectrochim. Acta, Part A*, 2013, **102**, 15–23.
- 44 G. K. Soon and T. Hyeon, *Acc. Chem. Res.*, 2008, **41**, 1696–1709.
- 45 J. Luan, J. Zhang, X. Yao, F. Li and B. Jia, *Ceram. Int.*, 2016, **42**, 16262–16265.
- 46 G. M. Gurgel, L. X. Lovisa, O. L. A. Conceição, M. S. Li, E. Longo and C. A. Paskocimas, *J. Mater. Sci.*, 2017, **52**, 4608–4620.
- 47 Reproducible Step Evaluation for TGA Curves, http://www.mt.com/es/en/home/supportive_content/matchar_apps/MatChar_HB415.html, accessed Jan 2017.
- 48 T. Kim and J. Olek, *Transp. Res. Rec.*, 2012, **2290**, 10–18.
- 49 D. L. Pavia, G. M. Lampman, G. S. Kriz and J. R. Vyvyan, *Introduction to Spectroscopy*, Cengage Learning, 5th edn, 2014.
- 50 L. Tortet, J. R. Gavarri, G. Nihoul and A. J. Dianoux, *J. Solid State Chem.*, 1997, **132**, 6–16.
- 51 J. Żegliński, G. P. Piotrowski and R. Piękoś, *J. Mol. Struct.*, 2006, **794**, 83–91.
- 52 F. A. Andersen and L. Brecevic, *Acta Chem. Scand.*, 1991, **45**, 1018–1024.
- 53 S. Yan, M. Kim, S. O. Salley and K. S. Ng, *Appl. Catal., A*, 2009, **360**, 163–170.
- 54 Z. Y. Zhou, N. Tian, Y. J. Chen, S. P. Chen and S. G. Sun, *J. Electroanal. Chem.*, 2004, **573**, 111–119.
- 55 M. Derrick, D. Stulik and J. Landry, *Infrared spectroscopy in conservation science*, Getty Publications, Los Angeles, 2000.
- 56 *CRC Handbook of Chemistry and Physics*, ed. D. R. Lide, CRC Press, Boca Raton, 84th edn, 2003.
- 57 J. C. Pruessner, C. Kirschbaum, G. Meinschmid and D. H. Hellhammer, *Psychoneuroendocrinology*, 2003, **28**, 916–931.

