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A versatile method for the determination of photochemical quantum yields *via* online UV-Vis spectroscopy†

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We have developed a simple method for determining the quantum yields of photo-induced reactions. Our setup features a fibre coupled UV-Vis spectrometer, LED irradiation sources, and a calibrated spectrophotometer for precise measurements of the LED photon flux. The initial slope in time-resolved absorbance profiles provides the quantum yield. We show the feasibility of our methodology for the kinetic analysis of photochemical reactions and quantum yield determination. The typical chemical actinometers, ferrioxalate and *ortho*-nitrobenzaldehyde, as well as riboflavin, a spiro-compound, phosphorus- and germanium-based photoinitiators for radical polymerizations and the frequently utilized photo-switch azobenzene serve as paradigms. The excellent agreement of our results with published data demonstrates the high potential of the proposed method as a convenient alternative to the time-consuming chemical actinometry.

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Introduction

Photo-induced reactions have been attracting substantial attention for many years. Recent developments in photocatalysis,^{1–3} molecular switches,^{4,5} photo-polymer chemistry^{6–9} and click-chemistry¹⁰ have pushed the field of photo-induced reactions to new frontiers. For assessing the efficiency of such systems, it is crucial to determine the quantum yields of the involved reactions. The quantum yield Φ indicates the efficiency for the formation of a certain product in a chemical reaction.^{11,12} Nevertheless, the accurate and facile measurement of quantum yields can become complicated, costly and cumbersome. The state-of-the-art procedures are based on time-consuming chemical actinometry for photon flux determination and the use of separation techniques (chromatography) combined with spectroscopic methods.^{13–15} However, these established methods suffer from several shortcomings. Typically, the analysis of concentrated solutions is necessary, requiring large sample quantities. In addition, the investigated compounds may absorb different amounts of light during the reaction due to the absorbing photoproducts, which mask the

starting materials (*i.e.* the “inner filter effect”¹²). Furthermore, quantitative analysis may require error prone dilution steps and long measurement times. Recently, alternative approaches for the determination of photo-dissociation quantum yields have been introduced, based on NMR techniques,¹⁶ laser-flash photolysis (LFP)¹⁷ and peak-height analysis of size exclusion chromatography (SEC) traces after pulsed-laser polymerization (PLP).¹⁸ However, the lack of a “standardized” procedure makes it difficult to compare the quantum-yield values obtained from different methods.

In this article, we wish to extend the scope of a recently presented setup based on online UV-Vis monitoring of photochemical reactions,^{19,20} to determine photochemical-reaction quantum yields. Irradiation is performed using light emitting diodes (LEDs). LEDs represent cost-efficient and flexible light sources and are readily available at various wavelengths with narrow bandwidths (*ca.* 20 nm). The photon flux can be easily varied by adjusting the forward current. We present the use of a calibrated spectrophotometer for photon flux determination replacing time-consuming photon flux determination *via* chemical actinometry. Additionally, as a reference, we use ferrioxalate actinometry with online detection.²⁰ We have evaluated our method by comparing the measured LED intensity values with the results of actinometric experiments using the standard actinometer *ortho*-nitrobenzaldehyde.²¹ We further illustrate the wide applicability of our procedure on the example of light-induced riboflavin degradation, photodissociation quantum yields of phosphorus-based and germanium-

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based radical photoinitiators (bisacylphosphane oxides and acylgermanes), as well as a kinetic study on a spirobenzopyran²² and the well-known azobenzene²³ isomerization.

Experimental

Chemicals

O-Nitrobenzaldehyde (1), spirobenzopyran 6 (1',3',3'-trimethyl-6-nitrospiro[2H-1-benzopyran-2,2'-indoline]), aceto-nitrile (purity >99%), methyl methacrylate (MMA, purity >99%), azobenzene (7) and 0.1 M H₂SO₄ were purchased from Sigma Aldrich and used as received. Ferrioxalate trihydrate (2) was purchased from Alfa Aesar. Riboflavin (vitamin B₂, 3) was obtained from Fluka. The bis(acyl)phosphane oxide (BAPO) photoinitiator Irgacure® 819 (4) was purchased from TCI. The bisacylgermane 5 was kindly provided by Ivoclar Vivadent (Schaan, Liechtenstein). Phosphate buffered saline tablets (Sigma Aldrich) were dissolved in deionized water.

Instrumentation

Azobenzene solutions were pre-irradiated with a mercury-xenon lamp (Hamamatsu LC4, *ca.* 3500 mW cm⁻²) to achieve a high enrichment of the Z isomer.

UV-Vis spectra were recorded on a UV-Vis spectrometer equipped with optical fibres and a 1024-pixel diode-array detector (J&M Analytik AG, Essingen, Germany). Standard fluorescence quartz cuvettes were used for all measurements. The power of the sample beam of the spectrometer was 0.08 mW, several orders of magnitude lower than the LEDs output that is typically between 5 and 50 mW.

Sample irradiation

We have modified the cuvette holder allowing sample illumination through an aperture ($d = 0.9$ cm) perpendicular to the spectrometer beam. The resulting optical paths are shown in Fig. 1. Accordingly, the path length is 1 cm for the irradiation and detection beam and independent of the fill height of the cuvette.¹⁹ It has to be justified that the LED light passing through the cuvette does not deteriorate the detection of the UV-Vis spectra.²⁴ Nevertheless some scattered light may reach the detector. However, spectra usually show no distortion at the LED peak wavelength. Placing the sample holder on a magnetic stirrer provides homogeneous illumination of the entire solution and provides reproducible reaction conditions. This setup allows the measurement of the absorbance spectra in regular sampling intervals while simultaneously irradiating and stirring the sample. To assess possible intensity losses of the irradiation beam in the sample solution, we have performed a control experiment, with the detection fibre of the spectrometer mounted opposite to the LED. The light intensity was measured in the absence of the cuvette, with an empty cuvette and with a cuvette filled with acetonitrile (see Fig. S3†). The detectable light intensity drops slightly upon insertion of the empty cuvette (~1%); however, the addition of acetonitrile increases the detected light intensity by 24%. This indicates

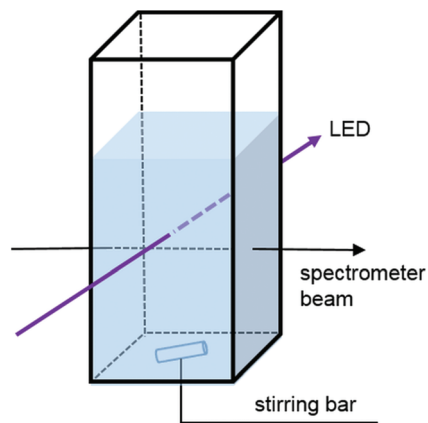


Fig. 1 Schematic representation of the experimental setup for the online monitoring of photochemical reactions. The solution is irradiated with an LED (purple arrow) perpendicular to the UV-Vis spectrometer beam (black arrow). The sample is stirred to provide a homogeneous solution.

that refraction at the glass/liquid interface occurs, leading to an essentially parallel light path.

The optical output power of the LEDs was measured using a calibrated spectrophotometer (GL Spectis 1.0, GL Optic Lichtmesstechnik GmbH, Weilheim, Germany) equipped with an integrating sphere. The LEDs were placed in front of the sphere aperture (diameter = 9 mm), resembling the geometry of the spectroscopic setup (see Fig. S2†). Low power LEDs (LED 385-33, XRL-400-50, 420-01, 450-06 and 490-06) were purchased from Roithner LaserTechnik GmbH (Vienna, Austria). A detailed description and test-measurement of LEDs emitting at different wavelengths is presented in the ESI.† The LEDs were operated using a custom-made power source (Fig. S1†), which was calibrated with a high precision voltmeter.

Photochemical processes involving free radicals and triplet-excited states are likely to be sensitive to dissolved oxygen. Therefore, we removed oxygen by saturating the solution with argon (5 min) before the measurement. Measuring single absorbance spectra before and after bubbling with argon allows correction of solvent evaporation during the deoxygenation procedure (see eqn (S1) in the ESI†).

Theoretical background

The quantum yield is defined as the number of events (reactions) that occur per absorbed photon. The quantitative relationship mirroring the amount of absorbed light by a photo-induced process is given by the Beer-Lambert law¹² (eqn (1)).

$$I_{\text{Abs.}} = I_0(1 - 10^{-A'}) \quad (1)$$

$I_{\text{Abs.}}$ represents the portion of the initial light intensity I_0 (LED light source) at a specific wavelength, which starts a photoreaction. The parameter A' is the absorbance at the specified wavelength. Accordingly, the knowledge of I_0 and A' yields $I_{\text{Abs.}}$. Moreover, I_0 is accessible *via* the photon flux of the LED emit-



ting at a specific wavelength. It can be conveniently determined with a spectrophotometer described in the Experimental section or by using a suitable chemical actinometer. Additionally, I_0 follows from radiant power values (P_{LED} , in W) accessible from technical data sheets. We tested four different LEDs emitting at 385, 420, 450 and 490 nm (Fig. S4†). Indeed, the radiant power of the LEDs shows a linear relationship with the forward current. The emitted light has a relatively narrow spectral distribution, with typical half widths in the range of 10 to 20 nm. We thus regard LEDs as monochromatic light sources and use eqn (2) to translate the radiant power into a photon flux I_0 in $\text{mol L}^{-1} \text{s}^{-1}$.

$$I_0 = \frac{\lambda P_{\text{LED}}}{hc N_A V} \quad (2)$$

Here, λ is the peak wavelength of the LED (in m), h is Planck's constant ($h = 6.63 \times 10^{-34} \text{ J s}$), c is the speed of light in a vacuum ($c = 299\,792\,458 \text{ m s}^{-1}$), N_A is Avogadro's constant ($N_A = 6.02 \times 10^{23} \text{ mol}^{-1}$) and V is the reaction volume (in L).

For the evaluation of the quantum yield, a connection between the absorbed light and the spectral changes based on the depletion of reactants must be established. The slope of the curve representing the time dependence of the absorbance is proportional to the amount of photons absorbed by the starting material ($I_{\text{Abs.}}^R$ in $\text{mol L}^{-1} \text{s}^{-1}$) multiplied by the quantum yield of the conversion (Φ). Hence, the time dependence reflecting the depletion of the reactant ($d[R]/dt$) in the photoreactor obeys eqn (3).

$$\frac{d[R]}{dt} = -I_{\text{Abs.}}^R \Phi \quad (3)$$

The molar absorption coefficients of R (ϵ_R) and the photoproduct (ϵ_P) and the optical path length d relate $[R]$ to the observed changes of the absorbance (A) at a certain wavelength:

$$\frac{dA}{dt} = -I_{\text{Abs.}}^R \Phi (\epsilon_R - \epsilon_P) d \quad (4)$$

In the case of $\epsilon_R > \epsilon_P$, the absorbance curve decays, whereas when $\epsilon_R < \epsilon_P$, an increase in absorbance is observed.

Eqn (4) can be simplified by considering the following: at the initial point of the reaction, the absorbance is given by $A^i = c_0 \epsilon_R d$, (c_0 is the starting concentration of R , $d = 1 \text{ cm}$). Accordingly, in the case of complete conversion, the residual absorbance (A^∞) is attributed to the photoproduct ($A^\infty = c_0 \epsilon_P d$). Thus, multiplying eqn (4) with c_0 yields eqn (5).

$$\frac{dA}{dt} c_0 = -I_{\text{Abs.}}^R \Phi (A^i - A^\infty) \quad (5)$$

The next step is quantifying the absorbed light by the reactant $I_{\text{Abs.}}^R$. After the reaction has started, the photoproduct may also absorb a fraction of the LED light. Therefore, it is justified to use eqn (5) only at the initial phase of the reaction, where

the light is exclusively absorbed by the parent species R ($I_{\text{Abs.}}^R = I_{\text{Abs.}}$).

$$\left(\frac{dA}{dt}\right)_{t=0} c_0 = -I_{\text{Abs.}} \Phi (A^i - A^\infty) \quad (6)$$

Accordingly, $I_{\text{Abs.}}$ is readily accessible either from the UV-Vis spectrum of the parent substrate or can be simply computed using established molar absorption coefficients (Beer-Lambert law). The initial slope is obtained by fitting an appropriate function to the experimental data and using its derivative at $t = 0$.¹³ We found that in case of weakly absorbing solutions ($A' < 0.1$), photoreactions virtually follow first order kinetics. Hence we use a mono-exponential function for data analysis. For strongly absorbing solutions ($A' > 2$), a linear function (reaction is apparent zero order) may be appropriate. Here it is crucial to analyse the resulting residuals, to ensure that the experimental data are well represented by the applied function. Errors due to this approximation can be expected to be lower than 2%. A detailed theoretical discussion and experimental data are given in the ESI.† Fitting the data with a mono-exponential function and taking the derivative at $t = 0$, yields

$$\left(\frac{dA}{dt}\right)_{t=0} = -k_{\text{fit}} (A^i - A^\infty). \quad (7)$$

Combining eqn (6) and (7) eliminates the factor ($A^i - A^\infty$). Together with the Beer-Lambert law (eqn (1)) an easy-to-use expression follows, with the time constant k_{fit} and the initial absorbance at the LED's peak wavelength A'_i as parameters, leading to the quantum yield Φ of the photo-induced conversion.

$$\Phi = \frac{k_{\text{fit}} c_0}{I_0 (1 - 10^{-A'_i})} \quad (8)$$

To obtain Φ with this method, the initial absorbance at the irradiation wavelength (A'_i) and the concentration of the substrate is required. We measured A'_i directly before starting the photo-induced reaction and calculated c_0 from the concentrations of the stock solutions.

We investigated the scope and limitations of eqn (8) for the determination of quantum yields using various model compounds and reactions. The light-driven conversions range from intramolecular rearrangements over the reduction of metal-organic complexes, biologically relevant decompositions and photo-induced bond cleavage to "switching" reactions such as *E-Z*-isomerization. They are presented in the following section.

Results and discussion

ortho-Nitrobenzaldehyde (1) as a chemical actinometer

The conversion of *o*-nitrobenzaldehyde (1) to *o*-nitrosobenzoic acid (**P1**, see Fig. 2) was followed with online UV-Vis spectroscopy. Due to its absorbance in the UV region, solutions or crystals of compound 1 serve as an actinometric system for



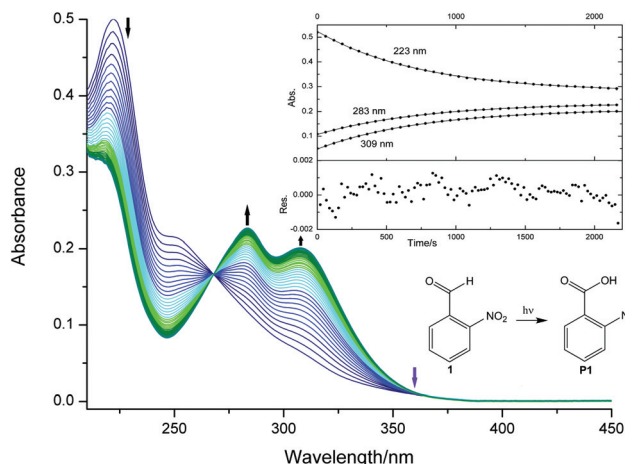


Fig. 2 Time-dependent spectral changes observed upon the illumination of **1** in acetonitrile. The inset shows absorbance–time curves monitored at 3 wavelengths (223 nm, 283 nm, and 309 nm). Sample details: 1.9 mL 44 μ M **1** in acetonitrile, illumination with a 385 nm LED operated at 20 mA, time steps between each spectrum: 1 minute. The irradiation wavelength (385 nm) is indicated with a purple arrow.

determining the intensity of UV light in different fields of photo-chemistry.^{13,25–27} The spectral changes of a degassed solution of **1** in acetonitrile (44 μ M) upon irradiation with a 385 nm LED operated at 20 mA for 35 minutes are shown in Fig. 2. The absorbance at 223 nm decays, whereas new absorbance bands appear at 283 and 309 nm. The sharp isosbestic point and similar time profiles for the maxima at 223, 283 and 309 nm reveal that only **1** and **P1** contribute to the spectra. Obviously, the molar absorption coefficient of **1** at the electronic absorbance maximum (223 nm) exceeds that of its photoproduct **P1** ($\epsilon_R > \epsilon_P$). The absorbance decays exponentially ($k_{\text{fit}} = 0.013 \text{ s}^{-1}$, see the inset in Fig. 2), whereas an increase is observed for the bands at 283 and 309 nm with irradiation time ($\epsilon_R < \epsilon_P$). However, as it is more intuitive, we chose to follow the reaction at 223 nm.

Ideally, the quantum yield Φ is independent of the LED light intensity. Higher light intensities (I_0) increase k_{fit} , thus leaving the quantum yield constant. Therefore, we repeated the experiment with different LED forward currents (5, 10, 15, 20, 25 and 30 mA) while leaving the concentration of **1** constant. In this case, Φ is determined from the slope of a plot according to eqn (8) ($k_{\text{fit}}c_0$ vs. $I_0(1-10^{-A_1})$) as depicted in Fig. 3. The inset shows the acceleration of the reaction as the light intensity increases due to higher forward currents (5 to 35 mA). Randomly distributed residuals (Fig. S10†) and R^2 values >0.9998 demonstrate that the applied model is appropriate.

The slope in Fig. 3 yields a quantum yield of 0.43 ± 0.02 , well in line with the reported values also obtained in acetonitrile ($\Phi = 0.43 \pm 0.03$).²⁷ Control experiments referenced to ferrioxalate^{14,28} (compound **2**, see the ESI†) virtually gave the same results. This underpins the validity of 0.43 instead of the previously reported value of 0.5 (ref. 29) in acetonitrile (see Table 1).

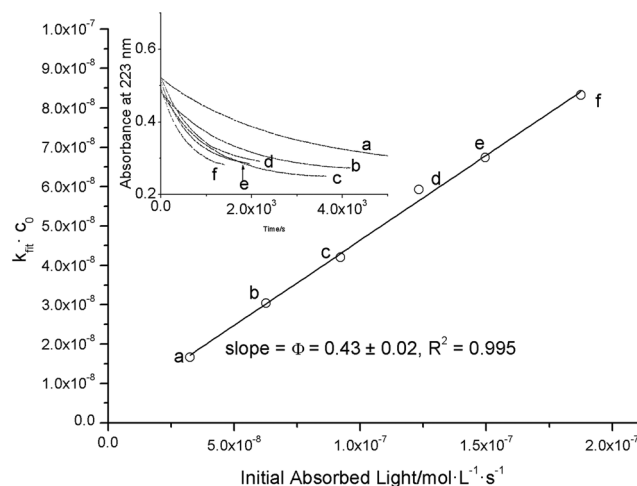


Fig. 3 Plot of the parameter $k_{\text{fit}}c_0$ vs. the denominator in eqn (8) for different solutions of **1** in acetonitrile. The inset shows the absorbance changes monitored at 223 nm that were used to obtain k_{fit} . The letters a, b, c, d, e and f indicate LED forward currents 5, 10, 15, 20, 25 and 30 mA, respectively. The displayed error (± 0.02) results from the linear fit of 6 independent measurements. Residuals are shown in Fig. S10†.

Table 1 Summary of quantum-yield experiments: studied compounds, LEDs, observation wavelengths λ , values for quantum yields from our work and published data. The displayed uncertainty is given as the standard deviation from the mean of three independent experiments

| Compound | LED/nm | λ/nm | Φ_{measured} | $\Phi_{\text{published}}$ |
|----------------------|--------|---------------------|--------------------------|---------------------------|
| 1 | 385 nm | 223 | 0.43 ± 0.02 | $0.43^{27-0.5^{29}}$ |
| 3 | 420 nm | 446 | 0.08 ± 0.008^a | — |
| 4 | 385 nm | 370 | 0.60 ± 0.03 | $0.6^{36,55}$ |
| 5 | 385 nm | 294 | 0.80 ± 0.04 | 0.85^{37} |
| 6 | 385 nm | 569 ^b | $\sim 0.08-0.12^c$ | $0.22^{d,56}$ |
| 7^e | 400 nm | 436 | 464 ± 5^e | $509^{f,53,54}$ |

^a Measured in argon-saturated solution, uncertainty was set to 10% of the obtained value. ^b Absorbance maximum of **P6** in acetone, precise molar absorption coefficient not available. ^c Quantum yield for color formation (using 3.5×10^4 and 5.2×10^4 for ϵ_{P6} (569 nm), respectively). ^d Relative value determined by laser flash photolysis. ^e Pseudototal quantum yield, for explanation see the text. ^f Literature value calculated from the reported quantum yields and extinction coefficients at 405 nm.

In cases where a calibrated photometer is not available, the conversion of **1** or a concentrated solution of **2** can be used to determine the photon flux into the UV-Vis cuvette. A simple rearrangement of eqn (8) (see eqn (S8)†) and the use of $\Phi = 0.43$ allows calculating the optical output power from the kinetic traces. As for the quantum yield measurement, each experiment gives a value for the applied LED intensity. Fig. S11† compares the results of the spectrophotometric with the actinometric measurements and the light intensities specified in the LED data sheets (11 mW at 20 mA). The intensities at all forward currents are in very good agreement. The above example demonstrates that this method is applicable for the conversion of **1** to **P1**, involving only one single reactant and photoproduct.



Photolysis of riboflavin (3)

As a more complex example, we have studied the photo-decomposition of the biologically relevant compound riboflavin (vitamin B₂, **3**, structure shown in Fig. S18†). When exposed to light, riboflavin can decompose *via* several pathways that have been extensively studied.^{30,31} Kinetic studies revealed the role of singlet and triplet excited states, oxygen, pH, irradiation wavelength, light intensity and different buffer systems. However, these studies were usually based on high intensity irradiation of large sample quantities, *e.g.* with mercury vapour lamps and taking samples at regular intervals for separate steps for analysis.^{32,33} Therefore, the online monitoring of the photo-degradation offers the possibility of a straightforward screening of many different reaction conditions. As an example, we irradiated 8 μM aqueous riboflavin solutions (pH 7.4, 0.01 M phosphate buffer containing 0.0027 M potassium chloride and 0.137 sodium chloride) with a 420 nm LED. Spectral changes for aerobic conditions along with the decay of the absorbance at 446 nm are shown in Fig. 4. To assess the efficiency of degassing, we have bubbled the solutions with argon for 1, 2, 3, 4 and 5 min, respectively.

The time traces for the degassed solutions (Fig. S19†) clearly show that the removal of oxygen accelerates the reaction drastically and the effect of degassing levels off after 3 min. Oxygen can quench singlet and triplet excited states and oxidize intermediates *e.g.* leucodeutero-flavin to formylmethyl-flavin that in turn forms lumichrome when exposed to irradiation. After the complete conversion (no further absorbance change), all solutions possess similar spectra that are well in line with the absorbance spectrum of lumichrome, the main photoproduct under these conditions.³⁴ The kinetic curves of the degassed solutions are not well described by a single step photoreaction (see residuals in Fig. S19†), although

the absorbance at the irradiation wavelength was adjusted to small values (~ 0.07 , see Table S5†). This may be due to the involvement of a short-lived intermediate in the formation of lumichrome as suggested by Remucal *et al.*³⁵ Another interesting feature in Fig. 4 is the region with “negative” absorbance above 500 nm. This is due to the strong green fluorescence of riboflavin, reaching the detector. Switching the spectrometer beam off and repeating the experiment with the spectrometer in the emission mode allows one to record time resolved fluorescence spectra using a 385 nm LED light as the excitation source (see Fig. S20†). Fluorescence light may be reabsorbed by riboflavin and in turn lead to its decomposition. However, the photon flux and the absorbance of the solution in this spectral region (450–540 nm) are small compared to the excitation source (420 nm). Applying eqn (8) to the time trace obtained under anaerobic conditions yields an apparent quantum yield of 8% that is reduced to 0.4% in the aerobic solution.

Cleavage of photoinitiators (4 and 5)

In addition to the decomposition of riboflavin, we further indicate the scope of our methodology for extended systems involving several photoproducts. In this respect, photoinitiators for radical polymerizations are of high interest. Irradiation at a suitable wavelength cleaves the photoinitiator and the formed radical species may initiate polymerizations. The photoinitiator fragments usually possess less extended chromophores and therefore do not absorb in the visible or near UV region.

This process known as “photobleaching” leads to the decay of UV-absorption bands of the parent compound upon irradiation. Applications like dental filling composites require photoinitiators that cleave efficiently even upon irradiation with moderate light intensities. The α -cleavage of the photoinitiator and the corresponding quantum yield of decomposition is, therefore, a key parameter for classifying the efficiency of such systems.^{17,36,37} As paradigms, we have investigated phosphorus- and germanium-based photoinitiators (**4** and **5**, Scheme 1) with respect to their decomposition quantum yields.^{7,9,36–41} Upon photocleavage, these initiators produce a benzoyl-type radical together with a phosphorus- or

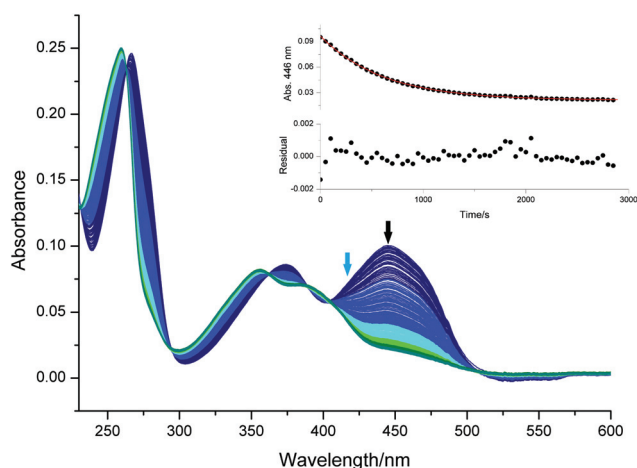
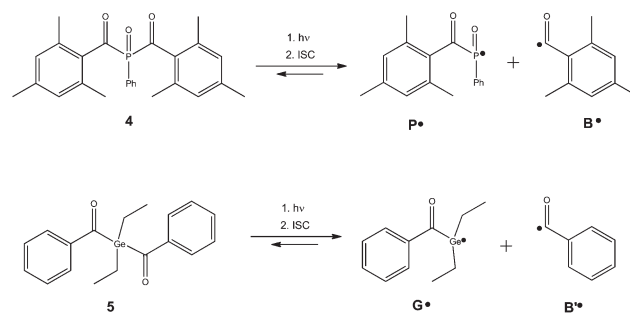


Fig. 4 Irradiating 7.7 μM riboflavin (**3**) in 0.01 M phosphate buffer with a 420 nm LED. Each spectrum = 10 s. The insert shows the absorbance decay at 446 nm under aerobic conditions, together with a mono-exponential fit and the corresponding residuals used to determine the reaction rate and the quantum yield.



Scheme 1 Photoinduced α -cleavage of initiators **4** and **5** to produce a phosphanyl or germynyl radical (P^\bullet or G^\bullet) and a benzoyl-type radical (B^\bullet or B''^\bullet).



germanium-centered radical (Scheme 1). Both radicals add to the double bonds of monomers, initiating radical polymerizations.^{9,36,40–42} We have determined the quantum yields for the α -cleavage of **4** and **5** in acetonitrile with the monomer methyl methacrylate (MMA) (1/1 v/v ratio). The samples were prepared to achieve low absorbance at the irradiation wavelength ($A_{385} < 0.1$), and the photon flux of the 385 nm LED, operated at 20 mA into 3 mL of the solution was determined with three reference measurements with ferrioxalate (photon flux = $(1.53 \pm 0.03) \times 10^{-5} \text{ mol L}^{-1} \text{ s}^{-1}$, Table S4†).

Fig. 5 presents the UV-Vis spectra recorded upon the photolysis of bis(acyl)germane **5** at 385 nm. The kinetic trace shown in the inset of Fig. 5 is monitored at 294 nm (black arrow). Upon irradiation, this characteristic π - π^* band decays. As described before, the parameter k_{fit} is extracted by the exponential fitting of the absorbance/time traces obtained from three independent measurements (Table S7, Fig. S23†).

The corresponding data for compound **4**, together with fitting residuals are presented in the ESI (Fig. S21, Fig. S22, Table S6†). The obtained dissociation quantum yields are in excellent agreement with previously published values and listed in Table 1. In accordance with the literature, bisacylgermane **5** exhibits a higher quantum yield for decomposition than the bisacylphosphane oxide **4**.^{9,37,40}

Color formation of a spirobenzopyran (**6**)

Up to this point, we applied our method to irreversible photo-induced reactions. However, it is also well suited for kinetic studies on more complex reversible systems, *e.g.* the formation of an intense colored isomer (color formation) of spirobenzopyrans upon exposure to UV light. Spirobenzopyran **6** (Fig. 6) undergoes a cascade of excited state reactions,^{22,43–45} eventually leading to the formation of an open form, **P6** (Scheme

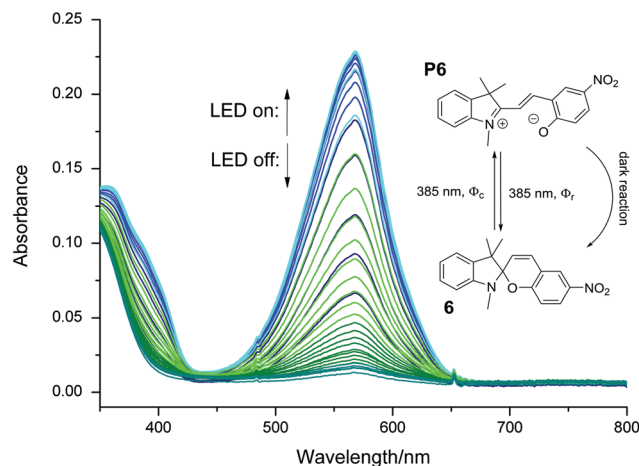


Fig. 6 Color formation of **6** in acetone under irradiation with UV light (385 nm). One spectrum = 20 s. The insert shows structure and the three reactions linking **6** with **P6**. The corresponding time profile of the absorbance at 569 nm is shown in Fig. S24.†

shown in Fig. S25†). This intensely purple colored species exhibits an absorption maximum at 569 nm in acetone.

Viewed at a slow timescale, the system can be described by two components (**6** and **P6**) that are linked by three reactions:⁴⁶ two photo-isomerizations interconverting **6** and **P6**, characterized by the quantum yields Φ_c and Φ_r . An additional first order, dark-reaction, described by k_r , regenerates **6**. Fig. 6 displays the spectral changes of a 12 μM solution of **6** in acetone (argon-saturated) irradiated with a 385 nm LED.

Upon irradiation, the absorbance at 569 nm increases, until an equilibrium between the color formation and the thermal back reaction is established (*i.e.* photostationary state, PSS, see Fig. S24†). Stopping the irradiation results in a recovery of **6** with the rate constant k_r .

It is useful to evaluate different PSSs reached with different light intensities, *i.e.* LED forward currents. Fig. 7 shows the spectra of different PSSs established by irradiating the sample with varying intensities. At low photon fluxes, the absorbance at 569 nm in the PSS follows the light intensity linearly; for higher optical powers, we observe saturation behavior. In the linear region, the photochemical reverse reaction is negligible, since k_r ($= 8.2 \times 10^{-3} \text{ s}^{-1}$) exceeds the photochemical back reaction ($I_0 = 8.8 \times 10^{-7} \text{ mol L}^{-1} \text{ s}^{-1}$). In principle, data from a photostationary state in this linear region can be used to determine the quantum yield of color formation Φ_c . However, due to the lack of a precise molar absorption coefficient of **P6** at 569 nm, this value should be taken as an estimate. Taking data from the PSS reached with the lowest light intensity (denoted as “a” in Fig. 7), the quantum yield ranges from 12% to 8% using the published values of 3.5×10^4 and $5.2 \times 10^4 \text{ L mol}^{-1} \text{ cm}^{-1}$ for the molar absorption coefficient ϵ_{P6} (569 nm),⁴⁴ respectively.

Isomerization of azobenzene (**7**)

As an example of a molecular photo-switch, we have investigated the photochemical *E/Z*-isomerization of azobenzene (**7**).

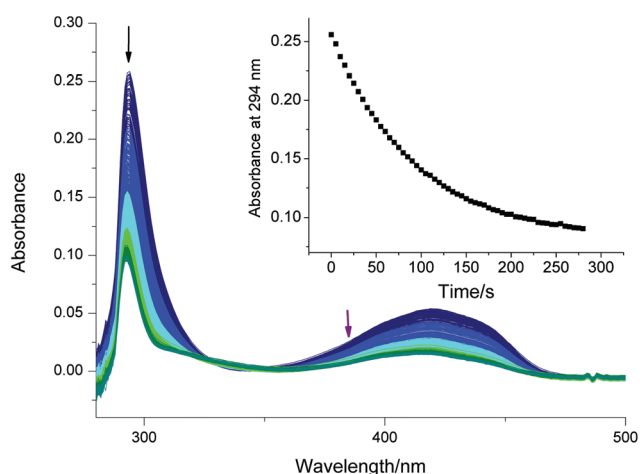


Fig. 5 Spectral changes observed upon irradiation of **5** with 385 nm (purple arrow). Time steps between each spectrum: 0.5 s. The insert shows the decay of the absorbance at 294 nm (black arrow) vs. the irradiation time. (Sample: 3 mL of 0.17 mM **5** in acetonitrile/MMA (1/1 v/v), degassed by argon bubbling.)



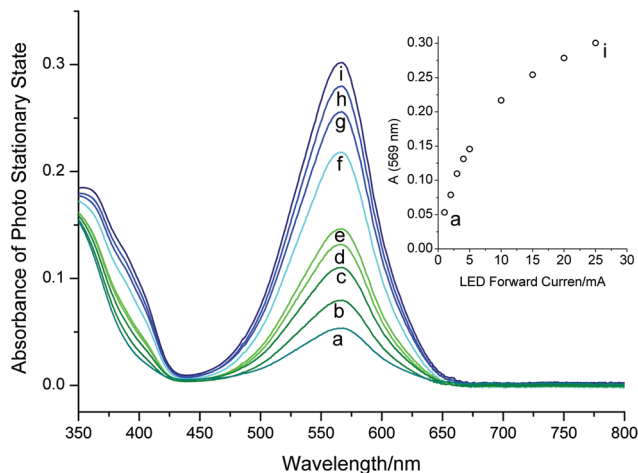


Fig. 7 UV-Vis spectra of different photostationary states of **6** in acetone reached by LED forward currents of 1, 2, 3, 4, 5, 10, 15, 20 and 25 mA (a, b, c, etc.). The spectra were recorded while the LED was on. The insert correlates the forward voltage with the absorbance in the PSS at 569 nm.

In contrast to compound **6**, the thermal reaction rate of **7** is extremely slow at room temperature, making compound **7** an archetype of a light-driven photo-switch. Its characteristic behavior has been exploited in numerous applications^{23,47–49} and the photochemical properties have been studied extensively.^{50–52}

Upon irradiation at low wavelengths (<360 nm), an excess of the Z-isomer (Z-7) is formed. Irradiation of the n- π^* band leads to the regeneration of E-7. Very recently, the molar absorption coefficients and photo-isomerization quantum yields in methanol have been re-determined and reviewed.^{53,54} Here we examine the conversion of a pre-irradiated solution containing predominantly Z-7 with a 400 nm LED in a kinetic approach. In the case of azobenzene, the change in absorbance dA/dt induced by the continuous irradiation I_0 is given by:⁵⁰

$$\frac{dA}{dt} = -dQI_0F(A - A^\infty). \quad (9)$$

The “pseudo total quantum yield” (Q) is independent of the light intensity and concentration and the current isomer ratio (defined as $Q = \epsilon_Z\Phi_Z + \epsilon_E\Phi_E$), d is the path length of irradiation ($d = 1$ cm) and F is the photokinetic factor ($F = (1 - 10^{-A'})/A'$) with A' being the absorbance at the irradiation wavelength. When a PSS is reached, the absorbance is constant and hence, $A = A^\infty$. Again, the data are analyzed at the very beginning of the reaction. Using the same methodology as above (eqn (7)), the following expression is obtained:

$$\frac{k_{fit}}{dI_0F} = Q. \quad (10)$$

An excess of Z-7 was formed by irradiating the cuvette with a high intensity mercury-UV-lamp for 60 s. The cuvette was allowed to cool, and was irradiated with a 400 nm LED while the absorbance at 436 nm was monitored. The experiment was

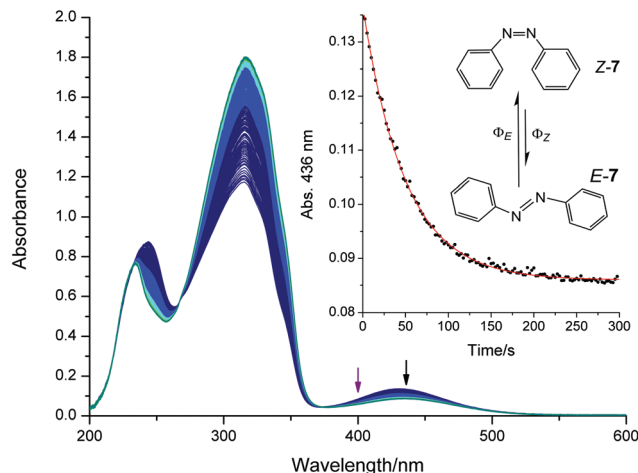


Fig. 8 UV-Vis spectra showing the isomerization of a 0.12 mM solution of E/Z-7 in methanol to the thermodynamically more stable E-7 upon irradiation with a 400 nm LED (purple arrow). The insert shows the exponentially fitted traces (solid line) of the absorbance at 436 nm (black arrow). Single spectrum = 0.5 s, overall time = 280 s.

repeated three times with fresh solutions. UV-Vis spectra and the corresponding time trace are shown in Fig. 8. The photon flux of the LED was measured three times with ferrioxalate (see Table S8†) yielding a photon flux of $(2.11 \pm 0.06) \times 10^{-5}$ mol L⁻¹ s⁻¹. We evaluated the factor F^i by measuring single absorbance spectra before the kinetic experiment ($n = 3$, $A'_i \sim 0.08$). Using this photon flux, these three measurements yield a pseudo total quantum yield of 464 ± 6 , lower (9%) than the reported value for 405 nm ($Q = 509$ (ref. 53 and 54) see Table 1). However, lower molar absorption coefficients of both isomers at 400 nm compared with 405 nm can account for this difference. Since the LED emits light in a wavelength range (FWHM = 13 nm), this value is an average in the region around 400 nm.

Conclusions

In this article, we present a versatile approach for the straightforward determination of quantum yields of photo-induced reactions. Our experimental setup features a simple modification of a cuvette holder used with a fibre coupled diode array spectrometer. This allows online monitoring of spectral changes induced upon photoreactions.¹⁹ Weakly absorbing solutions (<0.1 at the irradiation wavelength) follow apparent first order kinetics in good approximation. We have shown that for these cases, a simple relationship between the initial slope, the absorbed LED light and the quantum yield can be established and used to calculate the quantum yield for the observed process. This approximation offers the advantage that neither kinetic modeling nor numeric solving of differential equations is required.⁵⁴

Using ferrioxalate with online detection as a reference, we could reproduce quantum yields of several model systems. Our



method utilizes cost-efficient low power LEDs for irradiation, and requires low amounts of sample, while allowing quick determination of quantum yields of photochemical reactions. We could demonstrate that the photon flow can be measured with a spectrophotometer or using the conversion of nitrobenzaldehyde in acetonitrile and ferrioxalate combined with online spectrophotometric detection. Our method is feasible for the light-induced decomposition of riboflavin, two widely used phosphorus- and germanium-based photoinitiators. It is furthermore suitable for studying complex reaction kinetics like the photo-isomerization of a spirobenzopyran and photo-switches like azobenzene. Small required sample amounts, simple analysis and short conversion times make this technique ideally suited to evaluate photocatalyzed reactions. We are currently exploring the scope of the method for further photochemical systems like light-driven spin switches.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 J. Kou, C. Lu, J. Wang, Y. Chen, Z. Xu and R. S. Varma, Selectivity Enhancement in Heterogeneous Photocatalytic Transformations, *Chem. Rev.*, 2017, **117**, 1445–1514.
- 2 X. Li, J. Yu and M. Jaroniec, Hierarchical photocatalysts, *Chem. Soc. Rev.*, 2016, **45**, 2603–2636.
- 3 K. Wenderich and G. Mul, Methods, Mechanism, and Applications of Photodeposition in Photocatalysis: A Review, *Chem. Rev.*, 2016, **116**, 14587–14619.
- 4 M. M. Lerch, M. J. Hansen, W. A. Velema, W. Szymanski and B. L. Feringa, Orthogonal photoswitching in a multi-functional molecular system, *Nat. Commun.*, 2016, **7**, 12054.
- 5 M. M. Lerch, S. J. Wezenberg, W. Szymanski and B. L. Feringa, Unraveling the Photoswitching Mechanism in Donor–Acceptor Stenhouse Adducts, *J. Am. Chem. Soc.*, 2016, **138**, 6344–6347.
- 6 J.-F. Xing, M.-L. Zheng and X.-M. Duan, Two-photon polymerization microfabrication of hydrogels: an advanced 3D printing technology for tissue engineering and drug delivery, *Chem. Soc. Rev.*, 2015, **44**, 5031–5039.
- 7 J. Wang, G. Siqueira, G. Müller, D. Rentsch, A. Huch, P. Tingaut, J. Levalois-Grützmaier and H. Grützmaier, Synthesis of new bis(acyl)phosphane oxide photoinitiators for the surface functionalization of cellulose nanocrystals, *Chem. Commun.*, 2016, **52**, 2823–2826.
- 8 M. Tromayer, P. Gruber, M. Markovic, A. Rosspeintner, E. Vauthey, H. Redl, A. Ovsianikov and R. Liska, A biocompatible macromolecular two-photon initiator based on hyaluronan, *Polym. Chem.*, 2017, **8**, 451–460.
- 9 J. Radebner, A. Eibel, M. Leybold, C. Gorsche, L. Schuh, R. Fischer, A. Torvisco, D. Neshchadin, R. Geier, N. Moszner, R. Liska, G. Gescheidt, M. Haas and H. Stueger, Tetraacylgermanes: Highly Efficient Photoinitiators for Visible-Light-Induced Free-Radical Polymerization, *Angew. Chem., Int. Ed.*, 2017, **56**, 3103–3107.
- 10 N. Corrigan, S. Shanmugam, J. Xu and C. Boyer, Photocatalysis in organic and polymer synthesis, *Chem. Soc. Rev.*, 2016, **45**, 6165–6212.
- 11 A. D. McNaught and A. Wilkinson, in *IUPAC Compendium of Chemical Terminology*, IUPAC, Research Triangle Park, NC, 1997.
- 12 S. E. Braslavsky, Glossary of terms used in photochemistry, 3rd edition (IUPAC Recommendations 2006), *Pure Appl. Chem.*, 2007, **79**, 293–465.
- 13 K. L. Willett and R. A. Hites, Chemical Actinometry: Using o-Nitrobenzaldehyde to Measure Lamp Intensity in Photochemical Experiments, *J. Chem. Educ.*, 2000, **77**, 900.
- 14 H. J. Kuhn, S. E. Braslavsky and R. Schmidt, Chemical actinometry (IUPAC Technical Report), *Pure Appl. Chem.*, 2004, **76**, 2105–2146.
- 15 M. Montalti, A. Credi, L. Prodi and M. Teresa Gandolfi, *Handbook of Photochemistry*, 3rd edn, 2006.
- 16 N. D. Dolinski, Z. A. Page, F. Eisenreich, J. Niu, S. Hecht, J. Read de Alaniz and C. J. Hawker, A Versatile Approach for In Situ Monitoring of Photoswitches and Photopolymerizations, *ChemPhotoChem*, 2017, **1**, 125–131.
- 17 J. Lalevée, X. Allonas and J. P. Fouassier, Acylgermanes: Excited state processes and reactivity, *Chem. Phys. Lett.*, 2009, **469**, 298–303.
- 18 Y. W. Marien, P. H. M. Van Steenberge, K. B. Kockler, C. Barner-Kowollik, M.-F. Reyniers, G. B. Marin and D. R. D'hooge, Estimating the photodissociation quantum yield from PLP-SEC peak heights, *Polym. Chem.*, 2017, **8**, 3124–3128.
- 19 M. Gombár, É. Józsa, M. Braun and K. Ósz, Construction of a photochemical reactor combining a CCD spectrophotometer and a LED radiation source, *Photochem. Photobiol. Sci.*, 2012, **11**, 1592.
- 20 T. Lehoczki, É. Józsa and K. Osz, Ferrioxalate actinometry with online spectrophotometric detection, *J. Photochem. Photobiol., A*, 2013, **251**, 63–68.
- 21 R. Smith and K. Cantrell, Modeling the Effect of Polychromatic Light in Quantitative Absorbance Spectroscopy, *J. Chem. Educ.*, 2007, **84**, 1021.
- 22 H. Görner, Photochromism of nitrospiropyrans: effects of structure, solvent and temperature, *Phys. Chem. Chem. Phys.*, 2001, **3**, 416–423.
- 23 H. M. D. Bandara and S. C. Burdette, Photoisomerization in different classes of azobenzene, *Chem. Soc. Rev.*, 2012, **41**, 1809–1825.



- 24 T. Ditrói, J. Kalmár, J. A. Pino-Chamorro, Z. Erdei, G. Lente and I. Fábián, Construction of a multipurpose photochemical reactor with on-line spectrophotometric detection, *Photochem. Photobiol. Sci.*, 2016, **15**, 589–594.
- 25 H. J. Kuhn, S. E. Braslavsky and R. Schmidt, Chemical actinometry (IUPAC Technical Report), *Pure Appl. Chem.*, 2004, **76**, 2105–2146.
- 26 N. De la Cruz, V. Romero, R. F. Dantas, P. Marco, B. Bayarri, J. Giménez and S. Esplugas, o-Nitrobenzaldehyde actinometry in the presence of suspended TiO₂ for photocatalytic reactors, *Catal. Today*, 2013, **209**, 209–214.
- 27 E. S. Galbavy, K. Ram and C. Anastasio, 2-Nitrobenzaldehyde as a chemical actinometer for solution and ice photochemistry, *J. Photochem. Photobiol. A*, 2010, **209**, 186–192.
- 28 C. G. Hatchard and C. A. Parker, A New Sensitive Chemical Actinometer. II. Potassium Ferrioxalate as a Standard Chemical Actinometer, *Proc. R. Soc. A*, 1956, **235**, 518–536.
- 29 M. V. George and J. C. Scaiano, Photochemistry of o-nitrobenzaldehyde and related studies, *J. Phys. Chem.*, 1980, **84**, 492–495.
- 30 I. Ahmad and F. H. M. Vaid, in *Comprehensive Series in Photochemical & Photobiological Sciences*, 2006, vol. 6, pp. 13–40.
- 31 M. A. Sheraz, S. H. Kazi, S. Ahmed, Z. Anwar and I. Ahmad, Photo, thermal and chemical degradation of riboflavin, *Beilstein J. Org. Chem.*, 2014, **10**, 1999–2012.
- 32 I. Ahmad and H. D. C. Rapson, Multicomponent spectrophotometric assay of riboflavine and photoproducts, *J. Pharm. Biomed. Anal.*, 1990, **8**, 217–223.
- 33 M. A. Sheraz, S. H. Kazi, S. Ahmed, T. Mirza, I. Ahmad and M. P. Evstigneev, Effect of phosphate buffer on the complexation and photochemical interaction of riboflavin and caffeine in aqueous solution: A kinetic study, *J. Photochem. Photobiol. A*, 2014, **273**, 17–22.
- 34 I. Ahmad, Q. Fasihullah, A. Noor, I. A. Ansari and Q. N. M. Ali, Photolysis of riboflavin in aqueous solution: a kinetic study, *Int. J. Pharm.*, 2004, **280**, 199–208.
- 35 C. K. Remucal and K. McNeill, Photosensitized Amino Acid Degradation in the Presence of Riboflavin and Its Derivatives, *Environ. Sci. Technol.*, 2011, **45**, 5230–5237.
- 36 S. Jockusch and N. J. Turro, Phosphinoyl radicals: Structure and reactivity. A laser flash photolysis and time-resolved ESR investigation, *J. Am. Chem. Soc.*, 1998, **120**, 11773–11777.
- 37 B. Ganster, U. K. Fischer, N. Moszner and R. Liska, New Photocleavable Structures. Diacylgermane-Based Photoinitiators for Visible Light Curing, *Macromolecules*, 2008, **41**, 2394–2400.
- 38 K. Dietliker, T. Jung, J. Benkhoff, H. Kura, A. Matsumoto, H. Oka, D. Hristova, G. Gescheidt and G. Rist, New developments in photoinitiators, *Macromol. Symp.*, 2004, **217**, 77–97.
- 39 N. Moszner, F. Zeuner, I. Lamparth and U. K. Fischer, Benzoylgermanium derivatives as novel visible-light photoinitiators for dental composites, *Macromol. Mater. Eng.*, 2009, **294**, 877–886.
- 40 D. Neshchadin, A. Rosspeintner, M. Griesser, B. Lang, S. Mosquera-Vazquez, E. Vauthey, V. Gorelik, R. Liska, C. Hametner, B. Ganster, R. Saf, N. Moszner and G. Gescheidt, Acylgermanes: Photoinitiators and sources for Ge-centered radicals. insights into their reactivity, *J. Am. Chem. Soc.*, 2013, **135**, 17314–17321.
- 41 A. Eibel, M. Schmallegger, M. Zalibera, A. Huber, Y. Bürkl, H. Grützmacher and G. Gescheidt, Extending the Scope of Bis(acyl)phosphane Oxides: Additional Derivatives, *Eur. J. Inorg. Chem.*, 2017, **2017**, 2469–2478.
- 42 J. Radebner, M. Leybold, A. Eibel, J. Maier, L. Schuh, A. Torvisco, R. Fischer, N. Moszner, G. Gescheidt, H. Stueger and M. Haas, Synthesis, Spectroscopic Behavior, and Photoinduced Reactivity of Tetraacylgermanes, *Organometallics*, 2017, **36**, 3624–3632.
- 43 A. K. Chibisov and H. Görner, Complexes of spiropyran-derived merocyanines with metal ions: relaxation kinetics, photochemistry and solvent effects, *Chem. Phys.*, 1998, **237**, 425–442.
- 44 A. K. Chibisov and H. Görner, Photoprocesses in Spiropyran-Derived Merocyanines, *J. Phys. Chem. A*, 1997, **101**, 4305–4312.
- 45 H. Görner, Photoprocesses in spiropyrans and their merocyanine isomers: Effects of temperature and viscosity, *Chem. Phys.*, 1997, **222**, 315–329.
- 46 V. Pimienta, D. Lavabre, G. Levy, A. Samat, R. Guglielmetti and J. C. Micheau, Kinetic Analysis of Photochromic Systems under Continuous Irradiation. Application to Spiropyrans, *J. Phys. Chem.*, 1996, **100**, 4485–4490.
- 47 S. Venkataramani, U. Jana, M. Dommaschk, F. D. Sönnichsen, F. Tuczek and R. Herges, Magnetic bistability of molecules in homogeneous solution at room temperature, *Science*, 2011, **331**, 445–448.
- 48 J. Kind, L. Kaltschnee, M. Leyendecker and C. M. Thiele, Distinction of trans-cis photoisomers with comparable optical properties in multiple-state photochromic systems – examining a molecule with three azobenzenes *via in situ* irradiation NMR spectroscopy, *Chem. Commun.*, 2016, **52**, 12506–12509.
- 49 C. Slavov, C. Yang, L. Schweighauser, C. Boumrifak, A. Dreuw, H. A. Wegner and J. Wachtveitl, Connectivity matters - ultrafast isomerization dynamics of bisazobenzene photoswitches, *Phys. Chem. Chem. Phys.*, 2016, **18**, 14795–14804.
- 50 G. Gauglitz, Azobenzene as a convenient actinometer for the determination of quantum yields of photoreactions, *J. Photochem.*, 1976, **5**, 41–47.
- 51 G. Zimmerman, U. Paik and L.-Y. Chow, The Photochemical Isomerization of Azobenzene, *J. Am. Chem. Soc.*, 1958, **80**, 3528–3531.
- 52 G. Gauglitz and S. Hubig, Chemical actinometry in the UV by azobenzene in concentrated solution: A convenient method, *J. Photochem.*, 1985, **30**, 121–125.



- 53 E. Vetráková, V. Ladányi, J. Al Anshori, P. Dvořák, J. Wirz and D. Heger, The absorption spectrum of *cis*-azobenzene, *Photochem. Photobiol. Sci.*, 2017, **16**, 1749–1756.
- 54 V. Ladányi, P. Dvořák, J. Al Anshori, E. Vetráková, J. Wirz and D. Heger, Azobenzene photoisomerization quantum yields in methanol redetermined, *Photochem. Photobiol. Sci.*, 2017, **16**, 1757–1761.
- 55 S. Jockusch, I. V. Kopyug, P. F. McGarry, G. W. Sluggett, N. J. Turro and D. M. Watkins, A steady-state and picosecond pump-probe investigation of the photophysics of an acyl and a Bis(acyl)phosphine oxide, *J. Am. Chem. Soc.*, 1997, **119**, 11495–11501.
- 56 H. Görner, Photochemical ring opening in nitrospiropyrans: triplet pathway and the role of singlet molecular oxygen, *Chem. Phys. Lett.*, 1998, **288**, 589.

