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(–)-Cytisine: Access to a stereochemically defined and functionally flexible piperidine scaffold†

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N-Benzyl cytosine undergoes an efficient C(6)–N(7) cleavage via directed C(6) lithiation, borylation and oxidation to provide a “privileged” heterocyclic core unit comprising a highly functionalised, *cis*-3,5-disubstituted piperidine in enantiomerically pure form. The potential offered by this unit as a means to explore chemical space has been evaluated and methods have been defined (and illustrated) that allow for selective manipulation of N(1), C(3'), and the pyridone N. The pyridone core can also be diversified via bromination (at C(3'') and C(5'')) which is complementary to direct C–H activation based on Ir-catalyzed borylation to provide access to C(4''). The use of a boronate-based 1,2-migration as an alternative trigger to mediate C(6)–N(7) cleavage of cytosine was evaluated but failed. However, the stability of the intermediate boronate opens a new pathway for the elaboration of cytosine itself using both Matteson homologation and Zweifel olefination.

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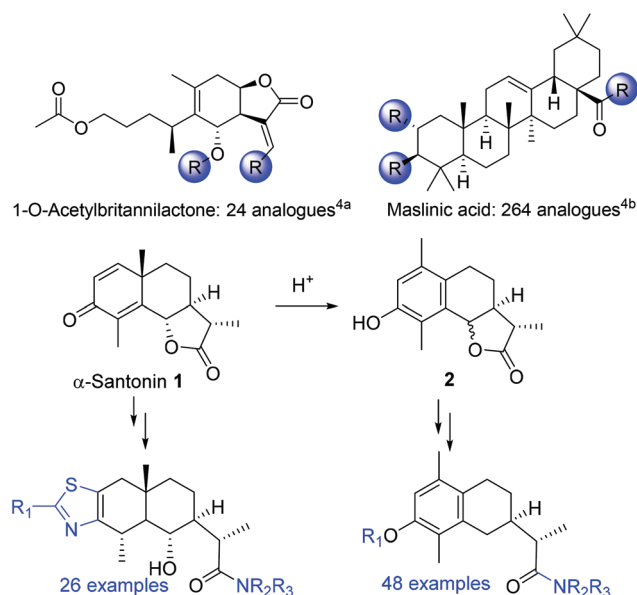
Introduction

Contemporary “small molecule” drug discovery relies on a variety of strategies to both identify and develop “hits” into leads and then drug candidates. Clearly, the ability to define and explore novel “chemical space” plays a key role in terms of generating (and then protecting to enable further exploitation) intellectual property.¹ For many years natural products provided a valued entry to enable structural variation but pharmaceutical industry interest in this area has, at points, waned in favor of, for example, combinatorial chemistry and related approaches as the preferred means by which to achieve molecular diversity.² However, the enormous structural variation associated with natural products has, regardless, continued to provide a source of inspiration (and a guide) that has led to the development of, for example, diversity-oriented and “chemical genetics” approaches to lead discovery.³

Natural products, which are often available in enantiomerically pure form and may contain multiply stereogenic centers, do provide one important starting point for a diversity-oriented approach to drug discovery. Several natural products or core components of natural products, such as 1-*O*-acetylbritannilactone

tone^{4a} and maslinic acid^{4b} have found application in this area (Scheme 1).

Taking this further, an ability to harness further the reactivity of a readily available natural product to widen the reach of the structural diversity available is well exemplified by Schwarz's work on α -santonin.⁵ Here, both the natural product **1** and a readily available variant **2** (available from **1** by acid-cat-



Scheme 1 Top: Example of natural products providing a functionalised core scaffold. Bottom: Derivatisation of α -santonin **1**.

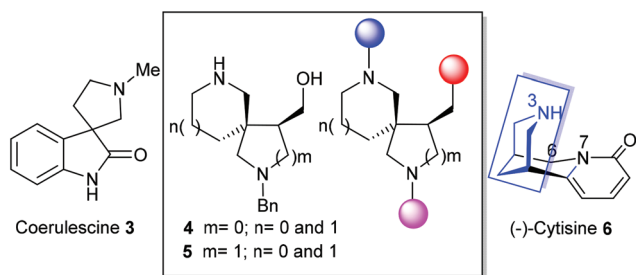
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alysed rearrangement) provided analogues of both of these related scaffolds that were explored as inhibitors of 5-lipoxygenase.

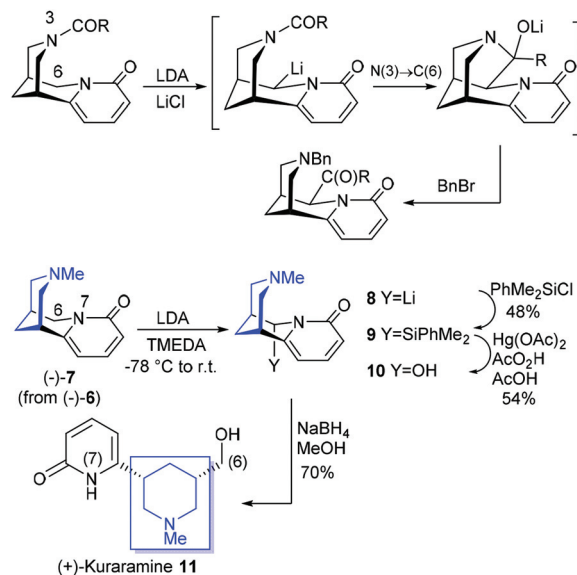
The role of “privileged scaffolds” serves as a further focus in this area, with many being based on or incorporating a heterocyclic core. Of these, piperidines are especially widely distributed and are found in a large number of bioactive and important pharmaceutical compounds.⁶ Our work has focussed on developing new and stereochemically-defined heterocyclic scaffolds based, in part, on nitrogen-based natural products that incorporate piperidine moieties.⁷ These include coerulescine **3**, which led to the development of a range of stereochemically-defined spirocyclic bis(azacycles), such as the piperidine and the pyrrolidine-containing units **4** and **5**.⁸ Each of these spirocyclic scaffolds (within which the individuals enantiomers are accessible) offers three discrete functionalisation sites: two secondary but readily differentiated amines (blue and purple), and a primary alcohol (red), all of which are distributed across a spatially distinctive spirocyclic core.



We have, more recently, aimed to extend this chemistry by evaluating the opportunity to “release” a stereochemically complex and functionally-flexible piperidine core unit from a readily accessible precursor; this is related to but distinctive from the concept outlined in Scheme 1. Our initial focus for this has been the development of the chemistry based on (-)-cytisine **6**, a readily accessible lupin alkaloid from *Laburnum anagyroides*,⁹ and the results of this work are summarised in this paper.

(-)-Cytisine, which is a partial agonist of the nicotinic acetylcholine receptor (the high affinity nicotine binding site in brain), is marketed (as Tabex®) within Eastern and Central Europe for smoking cessation.^{10,11} While that specific nicotinic profile may not be required (or desired), we recognised that cytisine **6** offered a readily available starting point from which to construct a series of structurally diverse libraries if, for example, the piperidine unit (coded blue) could be “released” from within the tricyclic heterocyclic core of **6**.

Cytisine **6** has already inspired a diversity-oriented synthesis approach to novel inhibitors of Bcl-2 and there has been extensive studies associated with the substitution of cytisine itself, driven largely by attempts to develop novel nicotinic ligands.¹² Of particular relevance to the work described here are the studies of Rouden and co-workers on the site-specific lithiation of *N*-acylated cytisines; this is illustrated by an *N* to *C* acyl transfer to functionalise at *C*(6) (Scheme 2).¹³ We had



Scheme 2 Top: *C*(6) lithiation triggering *N*(3) → *C*(6) acyl migration (Rouden *et al.*¹³). Bottom: *C*(6) oxidative functionalisation of *N*-methylcytisine **7**; a biomimetic synthesis of (+)-kuraramine **11** (Gallagher *et al.*¹⁴).

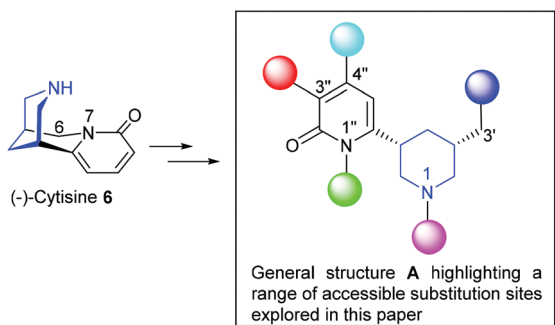
reasoned that using an appropriate electrophile, functionalisation of *C*(6) would allow for cleavage of the *C*(6)–*N*(7) link, and release of an intact piperidine unit. This was achieved as shown in Scheme 2 by *C*(6)-lithiation (*via* **8**) and (*in situ*) silylation of (-)-*N*-methylcytisine (caulophylline) **7** followed by *C*–Si → *C*–O oxidation followed by reduction. This sequence demonstrated the underlying concept by liberating the piperidine moiety (highlighted in blue) and provided a direct entry to the related lupin piperidine alkaloid (+)-kuraramine **11** in 18% overall yield from (-)-**7**.¹⁴

However, this initial solution involved the use of a silyl electrophile which necessitated a stoichiometric amount of a mercury-based oxidant to achieve the key Fleming–Tamao oxidative cleavage (**9** → hemiaminal **10**; overall 26% from **7**); alternatives to Hg(II) were evaluated but failed (see below). This issue alone, notwithstanding the modest yields, presents a clear block to any more general use of this approach to access a versatile heterocyclic (piperidine) scaffold or in a fragment-based approach to drug discovery. Given the goal of exploiting an ability to fragment cytisine, alongside the recognised importance of the piperidine scaffold within medicinal chemistry and a consequential ability to access efficiently differentiated and enantiomerically-pure piperidine-based libraries, this key synthetic obstacle had to be addressed.

In this paper, we describe a much more user-friendly solution to the cleavage of the *C*(6)–*N*(7) bond of cytisine shown in Scheme 2, which also offers access to the piperidine core with flexibility around the secondary amine protecting group. Further, the chemistry we have now developed avoids any heavy metals and is significantly more efficient and scalable. We have not explicitly focussed on a specific medicinal chemistry target, rather we have aimed to exemplify how the piper-

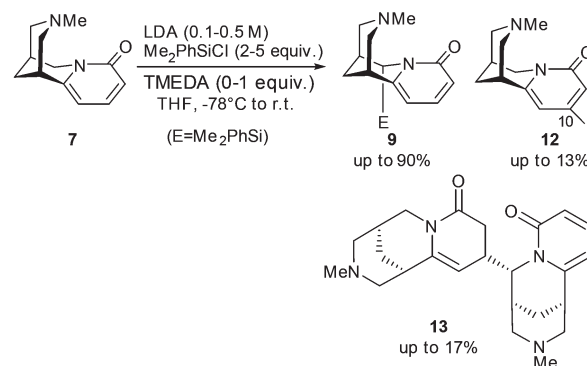
idine core can be both accessed and manipulated at various sites to provide access to a wide range of functional variants. The scope of this chemistry is outlined, in terms of the sites that become accessible, by general structure **A** and the issues around accessing each of these sites serves as the focus of this paper. The positions immediately accessible for derivatisation include two differentiated nitrogen centres (N(1) and N(1'')) in the piperidine and the pyridone moieties respectively); an ability to manipulate the C(3') methylene unit; further functionalisation of the pyridone moiety *via* either electrophilic bromination (which occurs predominantly at C(3'')) or, and of more utility and novelty, Ir-catalysed C–H activation which selectively targets C(4'').

This program has been driven in part by an increased awareness of the power and potential within medicinal chemistry associated with cost effective *in silico* assessment of ligands against protein targets of interest. Indeed, the goal would be to enable a full evaluation of the “effectiveness” of a ligand or family of ligands prior to embarking on an expensive program of chemical synthesis and evaluation by biological assay.¹⁵ As the dependability of these *in silico* methods increases so will their value as a reliable predictive tool. This would then enable the routine application of *in silico* prioritization of putative ligands as an early step in lead selection. It is likely that this, in turn, will lead to a rebalancing of the current reliance on existing library collections (*i.e.* physical samples) towards evaluating synthetic methodologies that offer opportunities to access as-yet-unexplored ligands.¹⁶ Those methodologies must, necessarily, encompass versatile, drug-like scaffolds such as **A**.



Results and discussion

To improve the efficiency of the fragmentation of cytosine, we have explored the chemistry illustrated in Scheme 2 in a number of ways: (i) the conditions for C(6) lithiation must avoid metalation at other sites, see below. (ii) The variation of the N(3) substituent that is possible; this was *N*-methyl earlier but this is less attractive for downstream synthetic utility. (iii) The choice of electrophile (ideally employed *in situ* during the deprotonation step) used to trap the C(6) lithiated species (*i.e.* **8**) in order facilitate C(6)–N(7) cleavage. (iv) The conditions needed to achieve that critical transformation that also avoid heavy metals such as Hg(II).



Scheme 3 Optimization of C(6) lithiation. Variables evaluated were substrate concentration, equivalents of LDA and Me₂PhSiCl, and presence of TMEDA (key to a high yield of **9**) Details are available in the ESI.†

Revisiting C(6) lithiation of *N*-substituted cytosines

The original conditions used to lithiate *N*-methylcytosine **7** (which was trapped with Me₂PhSiCl to give **9**) had proceeded in 48% yield. Relatively straightforward variation of some of the basic reaction parameters allowed us to optimise this basic transformation to 83% in favour of C(6) lithiation and trapping.

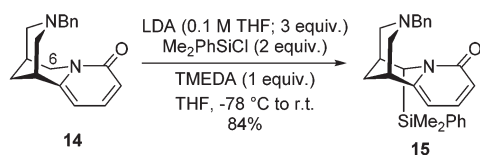
Although we did observe yields of **9** of up to 90%, 83% reflected the yield obtained when competing lithiation (and trapping) at C(10) was suppressed. The latter led to a C(10) silylation adduct **12** (and this was exacerbated in the absence of TMEDA) and we also observed a (known)^{13b} dimerization product **13** (Scheme 3).

While LDA provided cleanly lithiation at C(6), Rouden observed C(10) (as well as competing C(9)) metalation and (*in situ*) silylation of *N*-methylcytosine **7** when a particularly bulky base (LiTMP) was used. We did see some level of C(10) lithiation with LDA and an analogous metalation (*cf.* **12**) within a simple pyridone (*i.e.* at C(4), pyridone numbering) has been reported by Katritzky.¹⁷ We evaluated *N*-Boc cytosine as a substrate for C(6) metalation but analysis of the crude reaction mixture (following reaction with LDA and silylation) indicated substitution at C(2) and/or C(4) had occurred which is associated with Boc-directed metalation.¹⁸

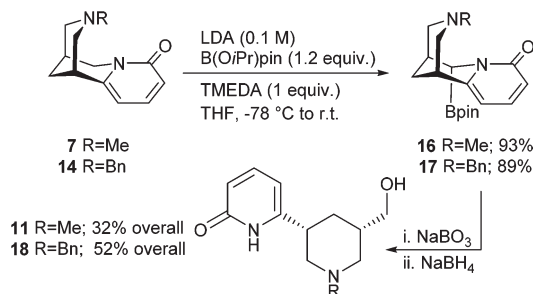
Use of *N*-benzyl cytosine **14** was more attractive both for downstream manipulations and because we did not observe competitive metalation (as **12**) or dimer formation (as **13**). Although Rouden has examined lithiation of **14**, in our hands and using a more synthetically useful aryl-based chlorosilane (Me₂PhSiCl), TMEDA (1 equiv.) did have an impact in terms of reaction efficiency and in particular suppressed a small amount of competing lithiation at C(10); **15** was isolated in 84% yield free of competing side products, with the mass balance accounted for by recovered **14** (Scheme 4).

Options around electrophilic component to facilitate C(6)–N(7) cleavage

Our original conditions for fragmentation of the cytosine tri-cycle to generate the piperidine unit associated with kurar-



Scheme 4 C(6) lithiation and silylation of *N*-benzylcytisine **14**.



Scheme 5 C(6) borylation and ring cleavage.†

amine **11** involved silylation and Fleming–Tamao oxidation (using peracid and stoichiometric $\text{Hg}(\text{OAc})_2$) followed by a reductive ring opening of hemiaminal **10** (Scheme 2). Closer inspection of this process indicated two competing reactions: desilylation of **9** to regenerate **7** and (ii) possible competing *N*-oxidation of the tertiary ($\text{N}(3)$) amine was suggested by MS analysis. These oxidation conditions were modified to give a small increase in yield of **11** (from 38% to 47% from **8**) by using a combination of H_2O_2 and peracetic acid (rather than peracetic acid alone) but this transformation still required $\text{Hg}(\text{OAc})_2$. All attempts, however, to achieve the conversion of **9** to **10** by other means *e.g.* KBr , AcOOH/AcOH ; HBF_4 , then H_2O_2 ; KH , $t\text{BuOOH}$ then TBAF , thereby avoiding mercury salts, failed. This forced consideration of an electrophile that would offer an alternative cleavage mechanism. Lithiation followed by halogenation at C(6) was evaluated using a variety of halogen sources (Br_2 , I_2 , *N*-bromosuccinimide, $\text{BrCCl}_2\text{CCl}_2\text{Br}$, CBr_4) proved unproductive. However, diphenyl disulfide (not shown, but see ESI†) and boron-based electrophiles were effective *in situ* traps for the C(6) lithio intermediate derived from **7** (Scheme 5). In the boron series, further optimisation using both *N*-methyl and *N*-benzylcytisine **7** and **14** respectively led to (OiPr)Bpin as the electrophile of choice, and adducts **16** and **17** were obtained in 93 and 89% yields respectively. While boronate esters **16** and **17** were relatively tolerant of chromatography, further purification was unnecessary. Direct oxidation of the crude boronates (using NaBO_3) followed by reduction (NaBH_4) gave the piperidine core structures, (+)-kuraramine **11** and the synthetically more flexible *N*-benzyl variant **18** in 32% and 52% overall yields (from **7** and **14**) respectively.

† When we applied a more conventional oxidation protocol (NaOH , H_2O_2) to the *N*-methyl boronate ester **16**, we only observed decomposition.

In summary, cytosine **6** is easily derivatised to the *N*-methyl and *N*-benzyl variants **7** and **14**, each of which provides efficient access to the corresponding enantiomerically pure *cis*-3,5-disubstituted piperidines **11** and **18** respectively. The results reported here serve to improve significantly the synthesis of (+)-kuraramine **11** from *N*-methylcytosine and avoid heavy metal oxidants to mediate the key C(6)–N(7) bond cleavage. Use of a boron-based electrophile, and specifically (OiPr)Bpin to trap the C(6) lithiated intermediate, was applicable to both the *N*-methyl and *N*-benzyl series.

Elaboration of the piperidine scaffold; identification of protecting group arrays that enable selective site manipulation

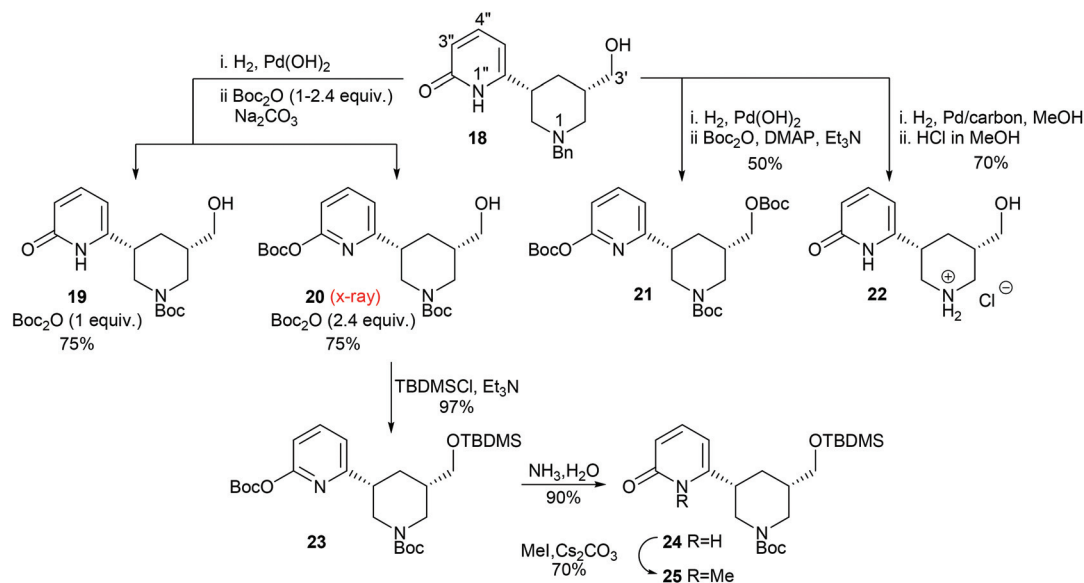
Heterocyclic scaffolds, such as **11** and **18**, offer significant potential with the *N*-Bn residue in particular cleavable under a variety of different reaction conditions. Consequently, the majority of the remainder of this paper is focused on developing the potential of the *N*-benzyl piperidine **18** as the basis of a flexible heterocyclic core structure. Our aim here is to demonstrate how the periphery of **18** can be functionalised in ways that would make this unit well suited to a general fragment-based drug discovery strategy, for the reasons articulated above. We have explored chemistry around four specific sites illustrated in general scaffold **A**: *via* the piperidine nitrogen ($\text{N}(1)$), at C(3') (in **18**, a primary alcohol), and in three areas of the pyridone moiety, at C(3''), C(4'') and across the lactam unit ($\text{N}(1'')$ and $\text{C}=\text{O}$). The latter is especially important to protect (deactivate) when pursuing functionalization of C(3') (see below). Given the array of functional groups present within **18**, we have determined methods for differentiating at these different sites. One consequence is that a series of protecting group arrays have been assessed and this is illustrated in Scheme 6.

Selective *N*- and *O*-Boc protection in combination with *O*-silylation provided a comprehensive set of options, which has allowed us to generate a number of differentially protected variants **19**–**24**; this includes the fully deprotected variant **22**. Additionally, the structure of the *N,O*-bis-Boc protected alcohol **20** was solved by X-ray crystallographic analysis (see ESI†). The pyridone lactam underwent *O*-acylation in the presence of an excess of Boc_2O , a protecting group array that is then cleavable under basic/nucleophilic conditions, see below. Further, the ability to manipulate selectively the pyridone nitrogen $\text{N}(1'')$ is illustrated by *N*-methylation of **24** to give adduct **25**.

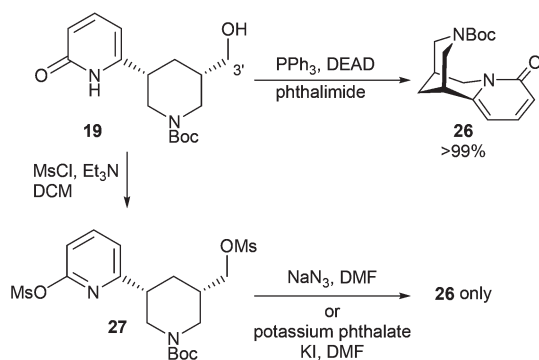
Elaboration at C(3'); variation of C(3') oxidation level

That there is a requirement to protect/deprotect easily the pyridone moiety is exemplified attempts to derivatise the C(3') primary alcohol (Scheme 7). Reaction of *N*-Boc piperidine **19** with phthalimide under Mitsunobu conditions in an attempt to access the 3' amino variant led cleanly (and in essentially quantitative yield) to *N*-Boc cytosine **26**.

Of course, *cis*-disubstituted piperidine **19** is well set up to undergo intramolecular alkylation, a possibility that was both recognized and exploited elegantly in one of the very early syntheses of cytosine carried out 60 years ago by van Tamelen and Baran,^{19a,b} and subsequently used in related contexts by others.^{19c–e}



Scheme 6 Selective protection of piperidine, hydroxyl and pyridone moieties associated with **18**.

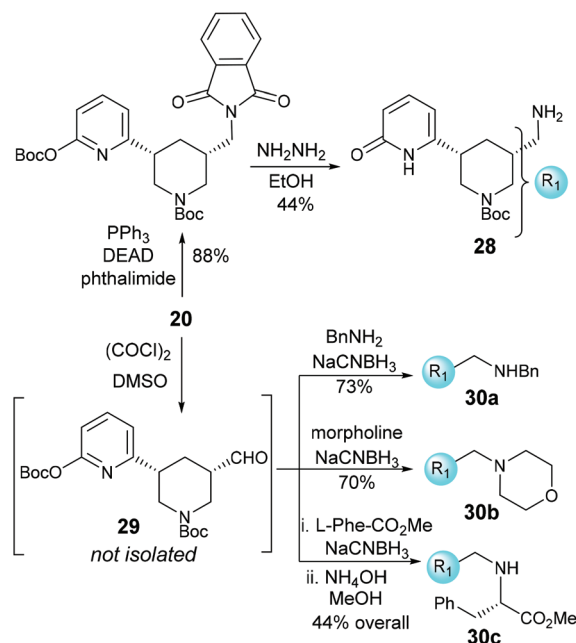


Scheme 7

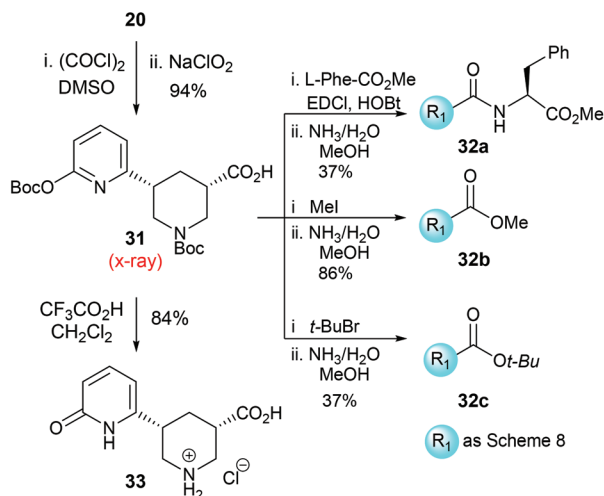
We attempted to circumvent this issue *via* double mesylation of **19** to generate intermediate **27** (not isolated, but assignment of a double mesylate was based on MS). However, cyclisation still occurred with **26** being the only product observed on exposure of **27** to azide or phthalate. Clearly, the *O*-mesylate is labile in the presence of a good nucleophile but *O*-Boc protection solves this issue. Using intermediate **20**, activation and phthalimide displacement at C(3') followed by imide cleavage provided the C(3') amino derivative **28** (Scheme 8). Here, hydrazine also cleaves the *O*-Boc residue liberating the pyridone unit, but this is presumed to occur after nucleophilic displacement at C(3').

In addition, the scope of the C(3') amino substituent can be extended using aldehyde **29** derived from **20** using Swern oxidation.

Aldehyde **29** was not routinely isolated (to minimize any risk of epimerization) but used directly for reductive amination or further oxidation (see below). Reductive amination using nucleophilic amines was generally accompanied by *O*-Boc cleavage under the reaction conditions (see **30a** and **30b**), and where (as in the case of an α -amino ester) this does not occur, *O*-Boc cleavage is then readily achieved using aqueous ammonia (leading to **30c**). The latter is a generally applicable deprotection method²⁰ and is illustrated further below. Alcohol **20** is also readily converted to the corresponding carboxylic acid **31** in excellent overall yield and acid **31** offers access to a representative library of C(3') derivatives (Scheme 9).



Scheme 8 Successful displacement at C(3'). Yields of **30a–c** are from alcohol **20**.



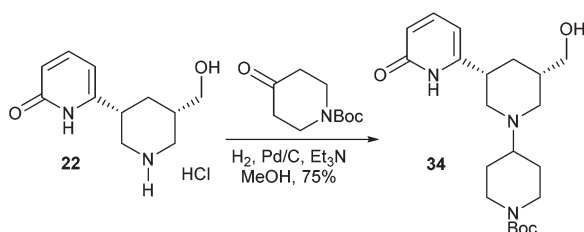
Scheme 9 Ester/amide library based on carboxylic acid **31**; global deprotection to provide acid **33**.

Again, the pyridone unit is unmasked by *O*-Boc cleavage under nucleophilic conditions without disruption of other functionality to provide pyridones **32a–c**. In addition, global deprotection of **31** is readily achieved to give the heterocyclic β -amino acid **33**.

Acid **33** is related to (*S*)-nipecotic acid (the parent piperidine-based β -amino acid), where 5-aryl substituted nipecotic acid derivatives have attracted medicinal chemistry interest.²¹ We had recognized a risk associated with C(3) epimerization of *e.g.* aldehyde **29**. For that reason we secured the X-ray crystal structure of acid **31** (see ESI†), which confirmed both the (intact) *cis*-configuration and the presence of the *O*-Boc protected pyridine moiety.

Elaboration at N(1)

The final target site for substitution on this piperidine scaffold was at N(1). This can be carried out in a number of ways, and we have simply illustrated the accessibility of this key position by *N*-debenzylation of **18** to give **22** (Scheme 6) as an isolable and versatile intermediate, followed by a representative reductive amination leading to the bis(piperidine) **34** (Scheme 10). This does, however, serve to show that protection elsewhere is not necessary in order to access N(1).



Scheme 10 Reductive amination at N(1).

Regioselective functionalisation of the pyridone moiety; electrophilic bromination and Ir-catalyzed borylation

The remaining region of the piperidine scaffold that was of interest is associated with the pyridone unit. Electrophilic halogenation of pyridones is well-established and leads predominantly to 3-substitution, with the 5-halo isomer as (usually) the minor product. Using *N*-bromosuccinimide, this pattern was observed with pyridone **19** (Scheme 11).

Bromides **35** and **36** are readily separated (and easily differentiated by ¹H NMR) and provide an obvious and very useful handle at C(3'') and C(5'').‡ Access to C(4'') is also achievable using Ir-catalyzed borylation, as has been reported recently.²² In our hands, however, NH pyridones (*e.g.* **19**) are not viable substrates for this C–H activation method. *N*-Alkyl pyridones (*e.g.* **7**) are, though, generally reactive and with *O*-Boc protected pyridines (*i.e.* **23**) highly selective borylation at C(4'') is observed.

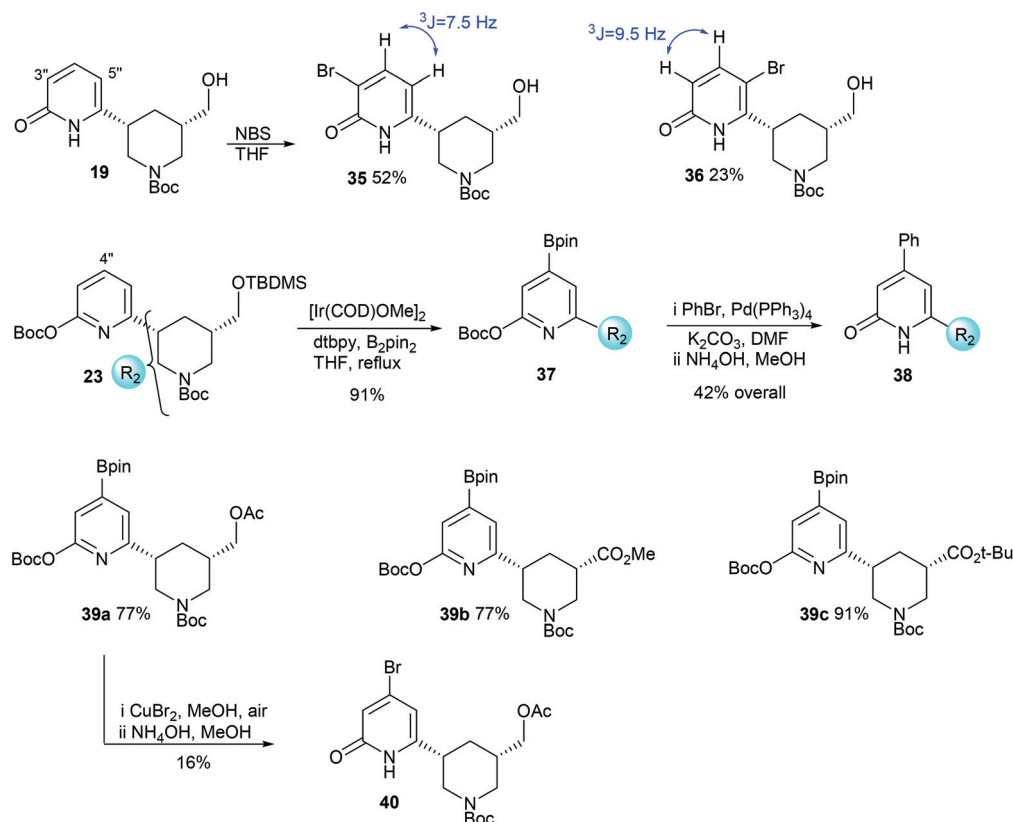
The scope and potential of this chemistry is illustrated in Scheme 11, and this includes two representative transformations of the intermediate boronate esters: Suzuki coupling of adduct **37**, followed by *O*-Boc cleavage (using basic ammonia) gave **38**; Cu-mediated bromination and deprotection of adduct **39a** gave **40**, offering a reactant that complements the corresponding Bpin esters. It is useful to note that the low yield of **40** reflects the instability in solution of the intermediate bromide *prior to O*-Boc cleavage but this transformation has not been optimized.

The chemistry outlined above serves to demonstrate the ability to manipulate the piperidine scaffold aligned to general structure **A**. This scaffold can be manipulated in a variety of distinct locations, using different transformations, and the only major requirement is the engagement of a suitable protecting group array that is primarily associated with moderating the reactivity of the pyridone unit. That moderation, largely based on *N*- and *O*-(Boc) protection, is readily achieved. Thus, we believe that the ready availability of (–)-cytisine and the efficiency with which the piperidine core unit can be revealed and then differentially manipulated makes this a potentially attractive starting point for a fragment-based discovery approach.

Enhancing the complexity of (–)-cytisine via C(6) lithiation; Matteson homologation and Zweifel olefination

A key aspect of a fragment-based approach must be an ability to increase rapidly molecular complexity. We have also explored additional avenues to address that issue where the goal has been to identify methods to introduce additional (and novel) complexity to the cytisine-based (tricyclic) precursor prior to any fragmentation and further functionalisation. While these reactions do not contribute to extending the scope of the piperidine scaffold discussed earlier, they do provide access to novel cytisine variants that can be considered as alternative scaffold configurations and progenitors.

‡ It is noteworthy that the corresponding *O*-Boc protected variant **23** was unreactive under these same conditions.



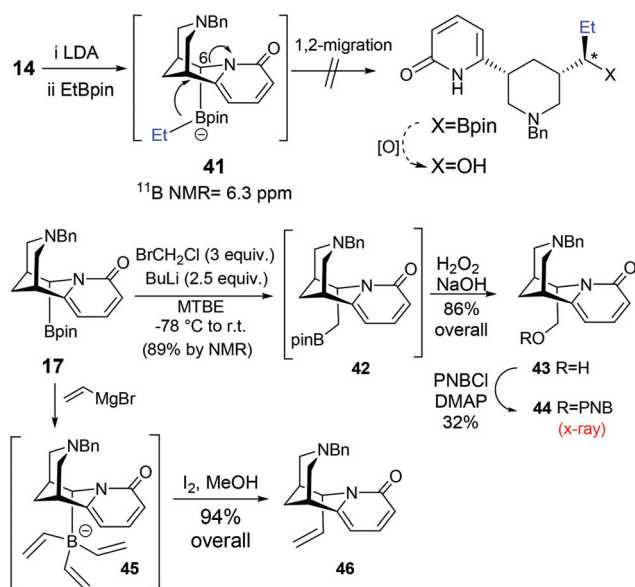
Scheme 11 Pyridone functionalisation.

Of particular relevance here was the potential associated with trapping a C(6) lithiated cytosine (see Scheme 2, intermediate **8**) with a secondary boronate ester (e.g. EtBpin). Our goal here had been to trigger a 1,2-migration sequence that would also serve to fragment the tricycle of cytosine and simultaneously create an addition stereocenter(*) at a key position on the periphery. The concept, which draws on previous work done within the Aggarwal group,²³ is illustrated in Scheme 12.

Using *N*-benzyl cytosine **14**, lithiation and trapping with EtBpin appeared to generate the requisite borate intermediate **41** based on ¹¹B NMR which showed a signal at 6.3 ppm. However, all attempts to achieve 1,2-migration using a range of Lewis acids failed. Given that Bpin esters are sterically demanding, we also evaluated Et₃B as a less demanding electrophile. Again, “ate” formation (analogous to **41**) occurred (as judged by ¹¹B NMR) but no migration products could be detected.

The most plausible explanation for these outcomes is that the pyridone N is not a sufficiently good leaving group to drive the migration step. However, an appreciation of the stability of the pyridone moiety with respect to a 1,2-migration provided an opportunity to assess to other “ate”-based chemistries available using boronate ester **17**.

Boronate **17** undergoes efficient Matteson homologation²⁴ to give boronate **42** in 89% yield. Oxidation of **42** provided primary alcohol **43** and the structure of the corresponding 4-nitrobenzoate **44** confirmed the configuration at C(6) (see

Scheme 12 Top: Attempt to couple 1,2-migration with tricycle cleavage. Bottom: C(6) alkenylation of *N*-benzyl cytosine **14**.

ESI⁺). A detailed ¹H NMR analysis of **44** was also carried (see ESI⁺) which aided in the assignment of adduct **46** (see below). Zweifel olefination²⁵ also involves generation of a stable borate intermediate and using boronate **17** proceeds very efficiently.

Using a modified procedure developed in the Aggarwal group,^{25c} exposure of **17** to vinyl Grignard (and this likely generates the triethenyl borate intermediate **45**), followed by exposure to I₂ in methanol gave the C(6) ethenyl adduct **46** in 94% yield (Scheme 12). The configuration at C(6) was confirmed by NOESY experiments (using **44** for comparison) and details of these studies are also available in the ESI.†

Conclusions

In summary, C(6) metalation of *N*-alkyl cytosine derivatives provides a direct and versatile method of functionalization that allows for ring fragmentation to provide a synthetically flexible, enantiomerically pure piperidine based heterocyclic scaffold. Given the privileged nature of the piperidine ring within medicinal chemistry, we suggest that this is an attractive unit for the development of more complex heterocyclic ligands within the context of a fragment-based approach to drug discovery. The potential of this functionalised piperidine scaffold has been exemplified using the *N*-benzylated variant **18**. Critically it is possible to differentiate all key elements and, using C–H activation, to provide an additional functional handle within the pyridone unit. Attempts to induce a B-based 1,2-migration as a means to increase the molecular complexity within the piperidone scaffold failed, however, the relative stability of the key intermediate (identified by a lack of reactivity) can be exploited. Derivatives such as **43** and **46** that arise from this offer new opportunities for modification of the established nicotinic ligand profile of cytosine and studies in this area are underway.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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Notes and references

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