Anion transport by ortho-phenylene bis-ureas across cell and vesicle membranes†

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Ortho-phenylene bis-ureas serve as anionophores in cells expressing halide-sensitive yellow fluorescent protein, as well as in synthetic vesicles. Activities can reach high levels, and are strongly dependent on the deliverability of the transporters.

The development of synthetic anion transporters (anionophores) has become a major theme of supramolecular chemistry.1 An important motivation is the potential for anionophores to be applied therapeutically, for example in the treatment of cancer and channelopathies such as cystic fibrosis (CF).2 Much of the work focuses on using small molecule anion carriers, which can bind an anion on one side of the membrane, traverse the membrane and then release the anion on the other side.3,4

If anionophores are to be exploited in medicine, it is necessary to show that they can be effective in living cells, and not just in synthetic bilayer systems such as vesicles. We recently developed an assay for biological anion transport which employs Fischer rat thyroid cells expressing the iodide-sensing yellow fluorescent protein (YFP-FRT cells).5,6 The assay was applied to anionophores of the “1,5-diaxial” family3 with steroidal, trans-decalin or cyclohexane scaffolds (e.g. 1–3, see Fig. 1).6a All tested positive for chloride transport, with trans-decalin 2 showing especially high activity. Transporter 2 showed negligible toxicity to several cell lines, suggesting that anionophore activity is not necessarily damaging to cells and raising the hope of “channel replacement therapy” as a treatment for CF.

While the 1,5-diaxial anionophores are known to be exceptionally powerful,7 other families have also shown good activity in synthetic vesicle assays. In particular, ortho-phenylene bis-ureas (OPBUs) of general structure 4 have proven effective for Cl−/NO3−, Cl−/HCO3−, and Cl−/carboxylate antiport, in some cases at very low transporter:lipid ratios.8,9 These carriers are also especially easy to prepare, being accessible from commercially available starting materials in one or two steps. They thus provide a valuable opportunity for the study of structure-activity relationships. Here, we report that OPBUs are active in cells, at levels comparable to the 1,5-diaxial anionophores. We also explore the effects of substituents which modify lipophilicity and electronic characteristics, and the correlation between results in synthetic vesicles and data from the biological assay. The results highlight the importance of deliverability in the development of anionophores for biological applications.

The OPBU anionophores employed in this work are shown in Fig. 1. To ensure high activities (and thus ease of measurement), the urea units were uniformly terminated with 3,5-bis...
DMSO-d$_6$/0.5% H$_2$O with tetrabutylammonium chloride as function as anion receptors by performing NMR titrations in a

Table 1

<table>
<thead>
<tr>
<th>Compound</th>
<th>clog P</th>
<th>Binding to Bu$_4$N$^+$Cl$^-$</th>
<th>Transport in LUVs (initial slope $I$ of $F_0/F$)</th>
<th>Transport in YFP-FRT cells*</th>
</tr>
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<tbody>
<tr>
<td></td>
<td></td>
<td>$K_{a1}$ (M$^{-1}$)</td>
<td>$K_{a1,a2}$ (M$^{-1}$)</td>
<td>Preincorporated</td>
</tr>
<tr>
<td>4a</td>
<td>9.9</td>
<td>136 1</td>
<td>1</td>
<td>0.073</td>
</tr>
<tr>
<td>4b</td>
<td>10.4</td>
<td>146 1</td>
<td>1</td>
<td>0.073</td>
</tr>
<tr>
<td>4c</td>
<td>10.1</td>
<td>157 1</td>
<td>1</td>
<td>0.042</td>
</tr>
<tr>
<td>4d</td>
<td>8.8</td>
<td>166 3</td>
<td>1</td>
<td>0.0073</td>
</tr>
<tr>
<td>4e</td>
<td>9.2</td>
<td>146 2</td>
<td>1</td>
<td>0.011</td>
</tr>
<tr>
<td>4f</td>
<td>10.8</td>
<td>138 1</td>
<td>1</td>
<td>0.017</td>
</tr>
<tr>
<td>4g</td>
<td>11.6</td>
<td>134 1</td>
<td>1</td>
<td>0.019</td>
</tr>
</tbody>
</table>

*Calculated log P, an estimate of lipophilicity where P is the partition coefficient between n-octanol and water. Values were calculated using TorchLite (available as freeware from http://www.cresset-group.com). + Determined by $^1$H NMR titration in DMSO-d$_6$/0.5% H$_2$O at 298 K. Data were analysed using a 1 : 1 + 1 : 2 (host : guest) binding model. † Values are proportional to rates of Cl$^-$/NO$_3^-$ exchange. Experiments were conducted in 200 nm POPC/cholesterol [7 : 3] LUVs with transporter: lipid = 1 : 25 000. I was obtained by fitting $F_0/F$ versus time (0–500 s) to a double exponential decay function, for further details see ref. 7 and ESI. ‡ Ratio of I from experiments employing external addition/preincorporation of transporter (see discussion in text). § Cl$^-$/Cl$^-$ exchange mediated by the transporter (50 μM) in YFP-FRT cells. Obtained by fitting the decay in YFP fluorescence to a first order exponential function and subtracting fluoresence quenching of the vehicle (0.5–1% v/v DMSO).
(25 mM). The decay in lucigenin fluorescence emission corresponds to an increase in intravesicular chloride concentration. To obtain values for comparison, the data were expressed as \( \frac{F_0}{F} \) (which is proportional to intravesicular chloride concentrations) then initial slopes \( I \) were obtained using a curve fitting procedure.\(^7\) Decay curves \( \frac{F_0}{F} \) for a series of experiments with transporter : lipid = 1 : 25 000 are shown in Fig. 3, and corresponding values for \( I \) are listed in Table 1.

The experiments revealed that all compounds in this series possess high intrinsic anionophore activity, clearly measurable at a very low transporter loading (0.004 mol%). The most powerful were bis-ureas \( 4a \) and \( 4b \), based on halogenated central scaffolds. In the case of \( 4a \), a dose–response study was performed and showed activity down to transporter : lipid = 1 : 1000k (Fig. S38†).

When comparing the activities of esters \( 4d-g \), there is clearly a positive correlation between initial rate and the length of the ester alkyl chain/lipophilicity (Table 1 and Fig. 4, red circles). In principle this effect could arise due to different amounts of transporter in the membrane; less lipophilic esters (shorter chains) might be present in lower concentrations due to leaching from the membrane into the aqueous phase. To examine this possibility, we conducted a leaching study\(^10c,12\) on methyl ester \( 4d \), the least lipophilic member in this series and thus the most likely to partition into water. Briefly, LUVs with \( 4d \) preincorporated in the membrane were prepared as previously described. The vesicles were then successively diluted with aqueous NaNO\(_3\) which should result in a loss of transporter from the lipid bilayers if it were capable of leaching. However, these studies indicated that \( 4d \) resides exclusively within the membrane and is not lost to the aqueous phase (see ESI† for further details). It thus seems that the longer alkyl chains promote intrinsic transport activity, possibly by positioning the transporter or complex more favourably within the membrane.\(^10c,13,14\)

After measuring the intrinsic activities of OPBUs \( 4a-g \) we next sought to establish how readily they could be delivered by external addition to membranes (the second component of practical effectiveness). LUVs were prepared in a manner analogous to that previously described, except that the anionophore was not added to the initial lipid mixture. Instead a solution of the anionophore in acetone was externally added to rapidly stirred preformed vesicles to give a notional transporter to lipid loading of 1 : 25 000. After 5 minutes, NaCl (25 mM) was added and the decay in fluorescence analysed as before (Table 1 and ESI†). Deliverability (\( D \)) was calculated by dividing the initial rate of transport from this experiment to that observed for preincorporated anionophore. If a transporter is perfectly deliverable, values for \( D \) are found to be \( \geq 1 \) and this was observed for esters \( 4d \) and \( 4e \).\(^15\) However, all other OPBUs tested displayed inferior activity when externally added, indicating that they are not perfectly deliverable (\( D < 1 \)). Within the ester series it is evident that deliverability decreases as a function of lipophilicity (Table 1 and Fig. 4). This has a substantial effect on the activity of \( 4g \); whereas it was the most powerful transporter in the ester series when preincorporated inside vesicles, it is the least active when added externally. Octyl ester \( 4g \) would thus be an ideal candidate for further investigation in coiled-coil driven membrane fusion studies. As demon-

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**Fig. 3** Chloride transport mediated by OPBUs in 200 nm POPC/cholesterol (7:3) LUVs as measured using the lucigenin method. The anionophore was preincorporated in the membrane at a transporter : lipid molar ratio of 1 : 25 000 in all cases.
strated previously, this is a highly efficient method of delivering lipophilic anionophores to membranes.

Following characterisation of their transport properties in synthetic membranes, OPBUs 4a–g were tested in cells. For these studies, we used Fischer rat thyroid cells expressing the halide sensor YFP-H148Q/I152L (YFP-FRT cells). YFP-FRT cells were seeded in a 96-well plate and cultured for 3 to 4 days until 90% confluence. The cells were washed twice with phosphate buffered saline (PBS) solution before incubating with a PBS solution containing the test anionophore (50 μM) for 10 min. The experiment was commenced by the addition of NaI (100 mM) and the rate of chloride/iodide exchange measured by fitting the decay in YFP fluorescence to a first order exponential function (see ESI† for full experimental details).

All compounds were found to mediate chloride/iodide exchange in cells leading to a decay in YFP fluorescence (Fig. 5, Fig. S45† and Table 1). The most active by some margin was bis-urea 4a, based on the difluoroaromatic scaffold. Reassuringly, transporter 4a was also the most active when externally delivered to LUVs, and joint first when preincorporated. This adds to our confidence that the YFP-FRT assay does indeed measure anion transport, rather than some other phenomenon. Anionophore 4a is somewhat more active than 2, which gave |df/dt| = 0.013 s⁻¹ in the present assay. The overall relationship between activity in cells and vesicles is shown in Fig. 6. Provided the vesicle measurements involve external addition of anionophore, a rough correlation emerges (Fig. 6a). There is, however, considerable scatter; the poor performance of dichloride 4b in the cells, despite its good activity in LUVs, is especially noticeable. This is difficult to explain, but may result from a specific interaction between 4b and the natural membrane. The results imply that, while experiments in LUVs can give some guidance, these synthetic assemblies are not ideal surrogates for cells and cannot substitute for biological testing. When results in cells are compared to those in vesicles with preincorporated OPBU, the correlation is very poor (Fig. 6b). This highlights the importance of considering deliverability as well as intrinsic activity when developing biologically active anionophores.

In conclusion, we have shown that a series of ortho-phenylene bis-urea (OPBU) anionophores are active in cells as well as synthetic vesicles. The most effective in all assays is bis-urea 4a, based on a difluorinated central scaffold. Activities depend both on electronic factors and lipophilicity. In particular, the sequence 4d–g illustrates the role of lipophilicity in promoting intrinsic activity, but the contrary effect on deliverability and thus on effectiveness in cells. These bis-ureas are readily prepared, so that many other variations are potentially accessible. They therefore hold promise as tools for obtaining further insight into anionophore biological activity, and perhaps for the development of medical applications.

Conflicts of interest

There are no conflicts to declare.
Acknowledgements

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Notes and references


11 For modelling and crystallography of OPBU-Cl− complexes, see ref. 8 and 9 respectively. Both methods support the participation of all 4 NH groups as H-bond donors.


15 With some anionophores, external addition to LUVs gives higher rates than preincorporation, resulting in values of $D > 1$. We presume that this observation is due to the actual concentration of lipid being lower than the notional 0.4 mM. Vesicle preparation involves some loss of lipid, so that subsequent addition of anionophore (external addition) results in a transporter to lipid loading that is higher than intended. In contrast, when the transporter is preincorporated in the membrane, the lipid and transporter are lost together during vesicle preparation so that the intended transporter to lipid ratio is maintained.