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Classic fungal natural products in the genomic age: the molecular legacy of Harold Raistrick†

Raissa Schor and Russell Cox *

Covering: 1893 to 2017

Harold Raistrick was involved in the discovery of many of the most important classes of fungal metabolites during the 20th century. This review focusses on how these discoveries led to developments in isotopic labelling, biomimetic chemistry and the discovery, analysis and exploitation of biosynthetic gene clusters for major classes of fungal metabolites including: alternariol; geodin and metabolites of the emodin pathway; maleidrides; citrinin and the azaphilones; dehydrocurvularin; mycophenolic acid; and the tropolones. Key recent advances in the molecular understanding of these important pathways, including the discovery of biosynthetic gene clusters, the investigation of the molecular and chemical aspects of key biosynthetic steps, and the reengineering of key components of the pathways are reviewed and compared. Finally, discussion of key relationships between metabolites and pathways and the most important recent advances and opportunities for future research directions are given.

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1. Harold Raistrick 1890–1971 and fungal natural products

In July 1929, at the age of 38, Harold Raistrick was appointed to his first academic post as University Chair of Biochemistry at the University of London.¹ By the end of 1934 he had published 42 scientific papers in the fledgling area of natural products chemistry and been elected to the Royal Society, publishing 20 papers during 1931 alone. This remarkable period of scientific productivity was built on an equally remarkable 8 year period when Raistrick was the head of biochemical research at the

Nobel Explosives Company based in Ardeer in Scotland. Raistrick's work at that time had focussed on the production of crucial feedstocks for the production of explosives by fermentation processes. However Raistrick realised that microorganisms, and fungi in particular, were adept at chemical transformations unavailable to industry at that time, and in the production of natural products with interesting and useful properties in other spheres of life. It was the results of these investigations which catalysed the start of his academic career in London, and which also helped to found today's science of fungal biotechnology and engineered biosynthesis (Fig. 1).

Raistrick described the discovery and investigation of compounds still of key interest today. These included kojic acid **1**,² citrinin **2** (ref. 3) the tropolones puberulic acid **3** (antimalarial), stipitatic acid **4** and sepedonin **5**, and the maleidride byssochlamic acid **6**. Compounds such as the xanthone ravenelin **7** were the first examples of compounds in their class. Others such as geodin **8** and griseofulvin **9** had unprecedented structures, and griseofulvin **9** and mycophenolic acid **10** are still clinically important compounds today. He focussed on the key orders of fungi which still make up the majority of the productive organisms exploited in industry and academia today including varied species of *Aspergillus*, *Penicillium* and *Fusarium*. Other compounds such as alternariol **11**, cyclophenin **12**, terrein **13** and patulin **14** remain of importance today as mycotoxins involved in crop and food spoilage. Raistrick also isolated and characterised numerous anthraquinones such as chrysophanol **15** which is a key intermediate in the biosynthesis of many anthraquinones and xanthenes, and curvularin **16**

Institut für Organische Chemie, BMWZ, Leibniz Universität Hannover, Germany.
E-mail: russell.cox@oci.uni-hannover.de

† Dedicated to professor T. J. Simpson to mark his retirement.



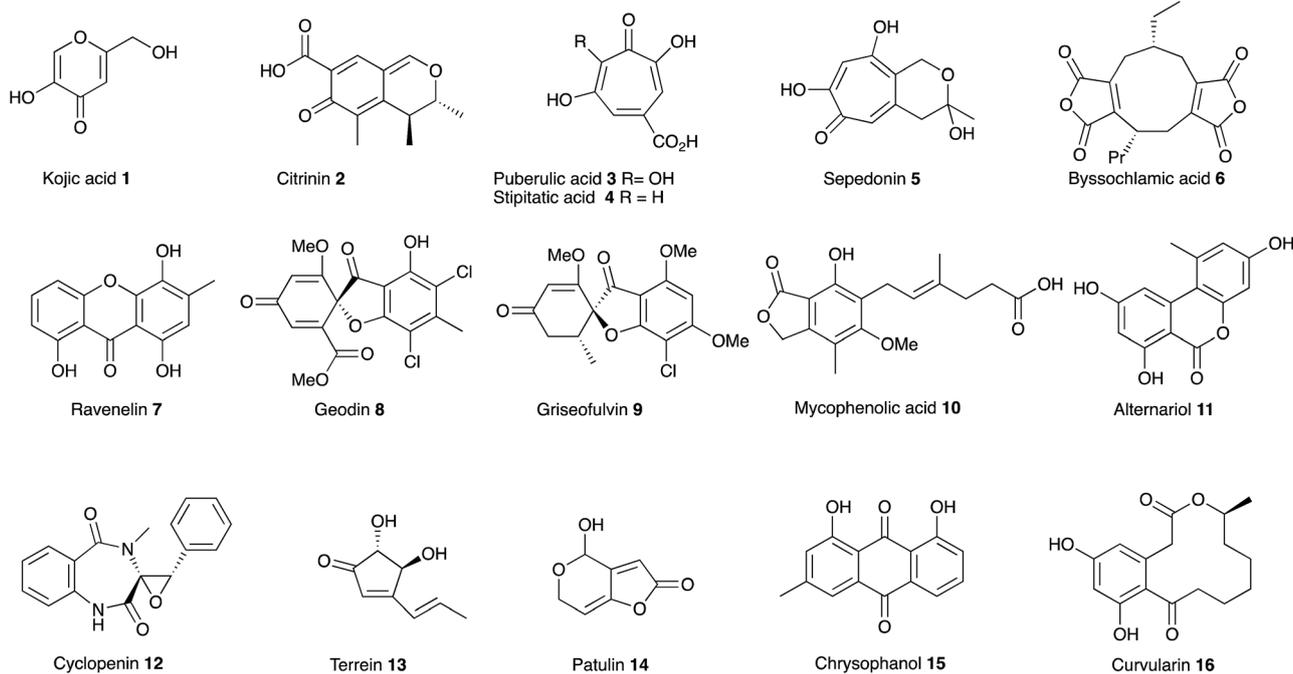


Fig. 1 Structures of key compounds.

which later had a role in development of ideas about the progressive nature of fungal polyketide biosynthesis. Raistrick also had a role in the penicillin story where he was an early collaborator of Fleming.⁴

In many cases these compounds were among the first examples of classes of compounds now taken for granted as natural products of fungi. Their structures became the

inspiration for generations of chemists involved in analysis, synthesis, biosynthesis, and more latterly genetic and genomic studies. Compounds such as citrinin 2 have served as key models at all stages of the development of the science of fungal natural products – from early isolation and structure elucidation (*e.g.* Cram and Whalley 1948),⁵ its first synthesis in 1949, through the earliest speculation and experiments on biosynthesis (for



Raissa Schor received a B. Sc. in Chemistry and Management from the University of Ulm in 2010 and a M. Sc. in Chemistry of the Julius-Maximilians-Universität Würzburg in 2013. She joined the Cox group as a PhD student in 2013, where she worked on the elucidation of the biosynthesis of fungal meroterpenoids.



Russell Cox was born in 1967 in the New Forest in the UK where he grew up. He studied chemistry at the University of Durham, and then worked with Prof. David O'Hagan at the same institution for his PhD, studying the biosynthesis of fungal metabolites. Post-doctoral periods with Professor John Vederas FRS in Edmonton Alberta, and Professors David Hopwood FRS and Tom Simpson FRS at Norwich and Bristol in the UK were followed by his appointment as a lecturer in the School of Chemistry at the University of Bristol where he rose to become full Professor of Organic and Biological Chemistry. He moved to become Professor of Microbiological Chemistry at the Leibniz Universität Hannover in Germany in 2013. He has served as an editorial board member of Natural Product Reports until 2012, is currently an editorial board member of RSC Advances, and has been past chair of the Directing Biosynthesis series of scientific conferences.



example Birch and coworkers in 1958),⁶ to the development of sophisticated isotopic labelling and NMR strategies (e.g. Staunton and coworkers⁷ and Sankawa and coworkers⁸ in the 1980s) to the more recent understanding of the biosynthetic gene cluster (e.g. Nihira and coworkers, 2007).⁹ Similarly compounds reported by Raistrick such as the tropolones (e.g. puberulic acid **3** in 1931) have stimulated important developments by others in the field of bonding theory (e.g. Dewar, 1945),¹⁰ and new ideas about synthesis (e.g. Todd) and biosynthesis (e.g. Robinson) from major figures in the field of organic chemistry. Other compounds such as the nonadrides (Section 4) also drove biosynthetic speculation, the development and application of new analytical methods to natural products chemistry, and also contributed to new ideas about biomimetic synthesis (e.g. Barton, Baldwin, Sutherland and coworkers). Raistrick's example also no-doubt contributed to the explosion in fungal natural product prospecting in the years after 1945.

Many of Raistrick's compounds have remained of key interest to microbiological chemists up to today because of their unusual structures, their useful biological properties, and the fascinating questions underpinning their biosynthesis. Modern investigations of many of the compounds have revealed the secrets of their biosynthesis, and illuminated the hidden relationships between the different classes of fungal natural products – particularly within the polyketides and meroterpenoids which form a rich vein of Raistrick's compounds. It seems highly likely that interest will continue in understanding and exploiting these compounds as the science moves towards rational manipulation of fungal biosynthetic pathways and production of new compounds. It seems timely, therefore, to review the most important work by briefly illustrating the historical context and most up-to-date discoveries for the main classes of Raistrick's compounds (Sections 2–9). Furthermore, despite the undoubted significant advances in the field, some key biosynthetic questions are still unanswered and some of the remaining challenges will be briefly explored in Section 10.

2. Alternariol

Raistrick and Thomas first reported alternariol **11** in 1953 as a crystalline solid isolated from *Alternaria tenuis* and its structure was determined by chemical means. Alternariol is a potent mycotoxin, and as many *Alternaria* species both produce **11** and infect food crops and stored feed, contamination with this compound presents a serious problem. Given its important ecological role, it is remarkable that relatively few studies of the biosynthesis of alternariol **11** have been reported. Early work by Thomas proved the compound was a typical non-reduced polyketide, biosynthesised in the expected way from [¹⁴C]-acetate.¹¹ More sophisticated experiments reported by Staunton, Abell, Garson and Leeper used [2-²H₃]-acetate and ²H NMR as part of a study designed to probe the enolization of intermediates prior to ring formation. The results showed that significant ²H wash-out was observed at position 4 (relative to position 6 and other deuterated positions) consistent with the idea that a 4,5-enol **18a** is a key intermediate, supporting the idea that a *cis*-intermediate must be formed prior to ring formation (Scheme 1).

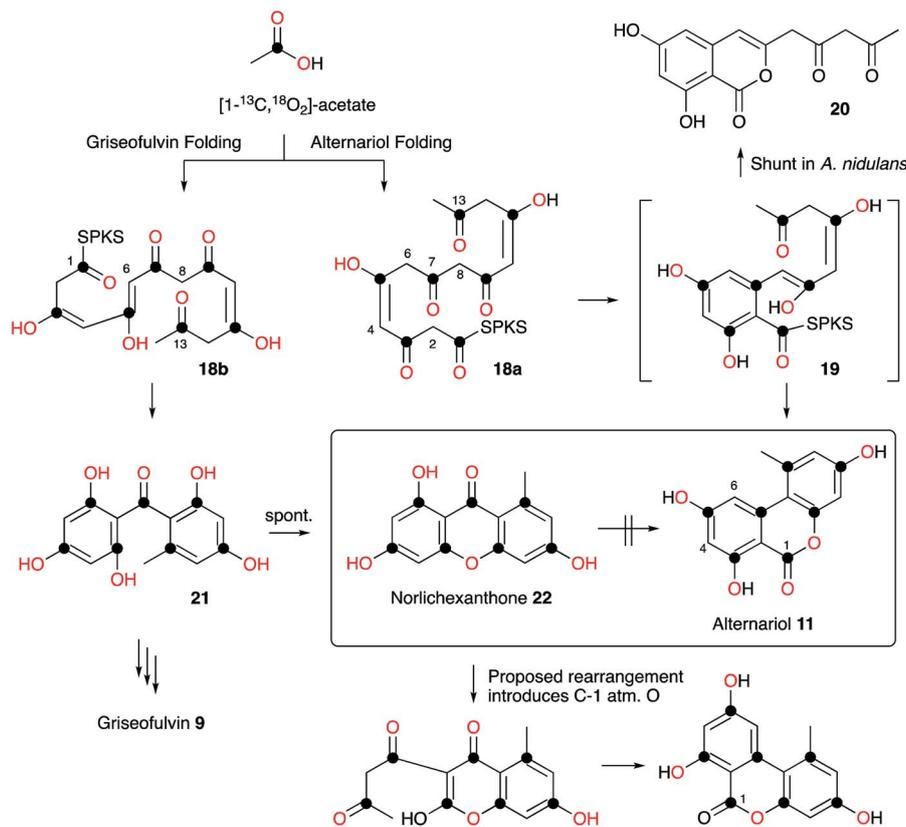
Wang, Oakley and coworkers reported a novel method for the discovery of the function of many of the cryptic non-reducing polyketide synthase (nrPKS) genes of *Aspergillus nidulans*. They systematically replaced the native promoters of nrPKS genes with constitutive promoters and isolated new compounds produced by the engineered strains.¹² In the case of the alternariol nrPKS, the lack of an obvious chain-releasing domain in the PKS itself meant that an adjacent gene encoding a β -lactamase-type thiolesterase also had to be induced. This led to production of alternariol **11** as well as the shunt dehydrocitreoisocoumarin **20**. Contemporaneous work by Fischer and coworkers identified a highly-reducing PKS (PKSJ) in *Alternaria alternata* as a potential alternariol synthase, but direct (e.g. knockout or heterologous expression) evidence for its involvement is lacking.¹³ More recently, Chooi and coworkers have isolated a true alternariol synthase gene (*SnPKS19*) from the plant pathogen *Parastagonospora nodorum* and proven its role by heterologous expression. *SnPKS19* encodes the expected nr-PKS. Intriguingly *SnPKS19* is a close homolog of the *Penicillium aethiopicum* norlichexanthone (NLX) synthase gene *gsfA* which is involved in the biosynthesis of griseofulvin (see Section 7): GfsA produces norlichexanthone **22** as a shunt metabolite. Previous labelling studies by Stinson and coworkers¹⁴ had suggested that norlichexanthone **22** was a precursor of alternariol, but labelling studies by Simpson using [1-¹³C, ¹⁸O₂]-acetate disproved this hypothesis (Scheme 1).¹⁵ The differences between the nrPKS *gsfA* and *SnPKS19* must control the regiochemistry of the first ring formation in each heptaketide which are 1–6 in norlichexanthone **22** and 2–7 in alternariol **11** biosynthesis.

Alternariol remains of key interest as a mycotoxin contaminant of food production, particularly soft fruits, berries and in viticulture. The current detailed level of understanding of its biosynthesis, coupled with the development of new CRISPR/Cas9 methods in the main producing host *A. alternata*¹⁶ may lead to methods for the production of *A. alternata* strains deficient in its biosynthesis, although the release of such strains to the wild would require significant regulatory work. The discovery of the close biochemical link between the alternariol and griseofulvin pathways suggests a possible common evolutionary history which may allow discovery of other related compounds and pathways, and future *in vitro* investigations could focus on the origins of the different ring-folding selectivities in the two closely related PKS, giving further details of how the intermediate polyketo intermediates are stabilised and selectively cyclised.

3. Geodin, ravenelin and the emodin pathway

The central role of the anthraquinone emodin **23** in fungal secondary metabolism is reflected in its very early discovery as a fungal metabolite and its structural proof by synthesis in 1924.¹⁷ Raistrick made many contributions to this area, notably through the first report of the xanthone ravenelin **7** (ref. 18) from *Helminthosporium ravenelii* and geodin **8** (ref. 19) from *Aspergillus terreus*, coincidentally both in 1936. It would not be





Scheme 1 Labelling studies and folding patterns in alternariol 11 and norlichexanthone 22.

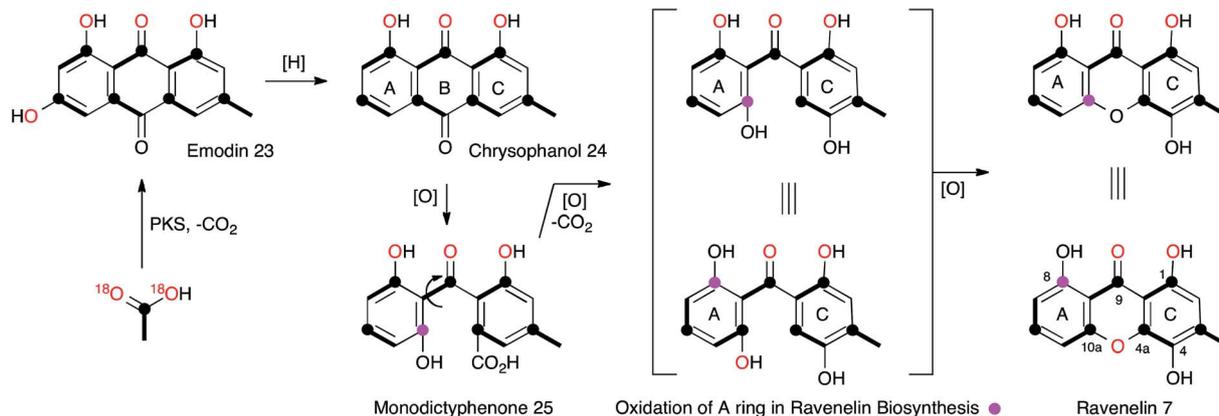
until many years later that the biosynthetic links between the seemingly very different anthraquinones, xanthenes and geodins would be elucidated, but these classes of compounds now form a key and widespread family of fungal metabolites, for which the molecular aspects of biosynthesis are only just becoming clear.

In the case of geodin 8, Raistrick proposed possible structures, with the assistance of Cedric Hassall, in 1947 based on extensive chemical degradation and derivatisation,²⁰ although it was not until 1958 when Ian Scott and Derek Barton provided the now-accepted structure, using IR spectroscopy as a key plank of evidence.²¹ The structure of ravenelin 7 was solved rapidly by Raistrick, and this proved to be the first fungal xanthone. Birch, Simpson and Westerman reported the results of the first important biosynthetic study of xanthenes in 1975. Feeding [1-¹³C]- and [1,2-¹³C₂]-acetate to *H. ravenelii* resulted in incorporation of intact acetates into the methylated aromatic C-ring, but scrambling of acetate in the A-ring, clearly indicating the intermediacy of a symmetrical benzophenone intermediate such as 25.²² Earlier work by Gatenbeck²³ and Hassall²⁴ had already shown that questin (1-*O*-methyl emodin) 26 is efficiently converted to the benzophenone sulochrin 27 (Scheme 3) in *Aspergillus terreus* and it seemed likely that a similar process in *H. ravenelii* could convert a simple non-reduced polyketide such as emodin 23 to a benzophenone, such as monodictyphenone 25, and then on to a xanthone such as ravenelin 7. However, further experiments would be needed to confirm this

hypothesis. Vederas and coworkers provided some of the required evidence through the use of [1-¹³C, ¹⁸O₂]-acetate feeding experiments (Scheme 2).²⁵ The results showed that the 4-oxygen and either the 8-oxygen or the 10a-oxygen are not derived from acetate, and are probably introduced from atmospheric oxygen. This supports an oxidative removal of the emodin 10-carbon, most likely by a biological Baeyer–Villiger-like reaction which would thereby introduce the atmospheric 8/10a oxygen prior to ring closure to the xanthone skeleton. The intact bond between ¹³C-10a and ¹⁸O-10a in 7 also confirms the mode of ring closure.

Meanwhile a direct link between emodin 23 and geodin 8 was proven by Fujimoto and coworkers who fed [UL-¹⁴C]-emodin to surface cultures of *A. terreus* and showed incorporation into geodin 8 and dihydrogeodin 28.²⁶ This led to a different strategy to understand the biosynthetic pathway by Japanese groups who purified active enzymes from cell-free extracts of *A. terreus*. The first success came in the isolation of an emodin 1-*O*-methyl transferase (e.g. a questin 26 synthase) by Sankawa, Ebizuka and Fujii.²⁷ Next came evidence for the final step of the pathway when dihydrogeodin oxidase (DHGO) was purified from *A. terreus* and shown to effectively stereoselectively convert dihydrogeodin 28 to geodin 8 itself.²⁸ The enzyme is an unusual blue-copper protein, and quite different in sequence to the P450 enzyme which catalyses the parallel step in griseofulvin biosynthesis (Section 7). The DHGO gene was cloned through a process involving expression of an *A.*





Scheme 2 Labelling pattern in ravenelin 7 revealed from isotopic feeding experiments.

terreus cDNA library and probing it with a DHGO antibody. This led to identification of a partial DHGO encoding gene which could be used to capture a full-length genomic clone.²⁹

The enzymatic system which catalyses the key anthraquinone ring cleavage was also purified from WT *A. terreus* by the Sankawa group. The purified protein effectively converts questin 26 to desmethylsulochrin 29 *in vitro*, presumably via a Baeyer–Villiger/hydrolysis sequence, and requires O₂ and NADPH.³⁰ Fractionation of this preparation into two less active fractions which regained full activity on recombination suggested the active system consists of an oxygenase and an associated electron transfer protein. Another oxidase, putatively involved in the route, was isolated from *A. terreus* by the same group.³¹ Emodin anthrone oxygenase is a non-heme iron protein which converts emodin anthrone 30 to emodin 23. Finally, the sulochrin PKS was identified by degenerate PCR³² using primers specific for non-reducing PKS which identified a gene fragment which could be used for targeted knockout.³³

Availability of the *A. terreus* genome, combined with the sequence information obtained from the experiments described above, allowed Mortensen, Larsen and coworkers to identify the geodin BGC (*ged*) and reconstitute it in *A. nidulans*.³⁴ Thirteen open reading frames were transferred from *A. terreus* to *A. nidulans*, along with their native promoters and terminators and this converted *A. nidulans* to a geodin 8 producer. These experiments also unambiguously identified the required chlorinase protein (GedL), but did not dissect the precise timings of all steps.

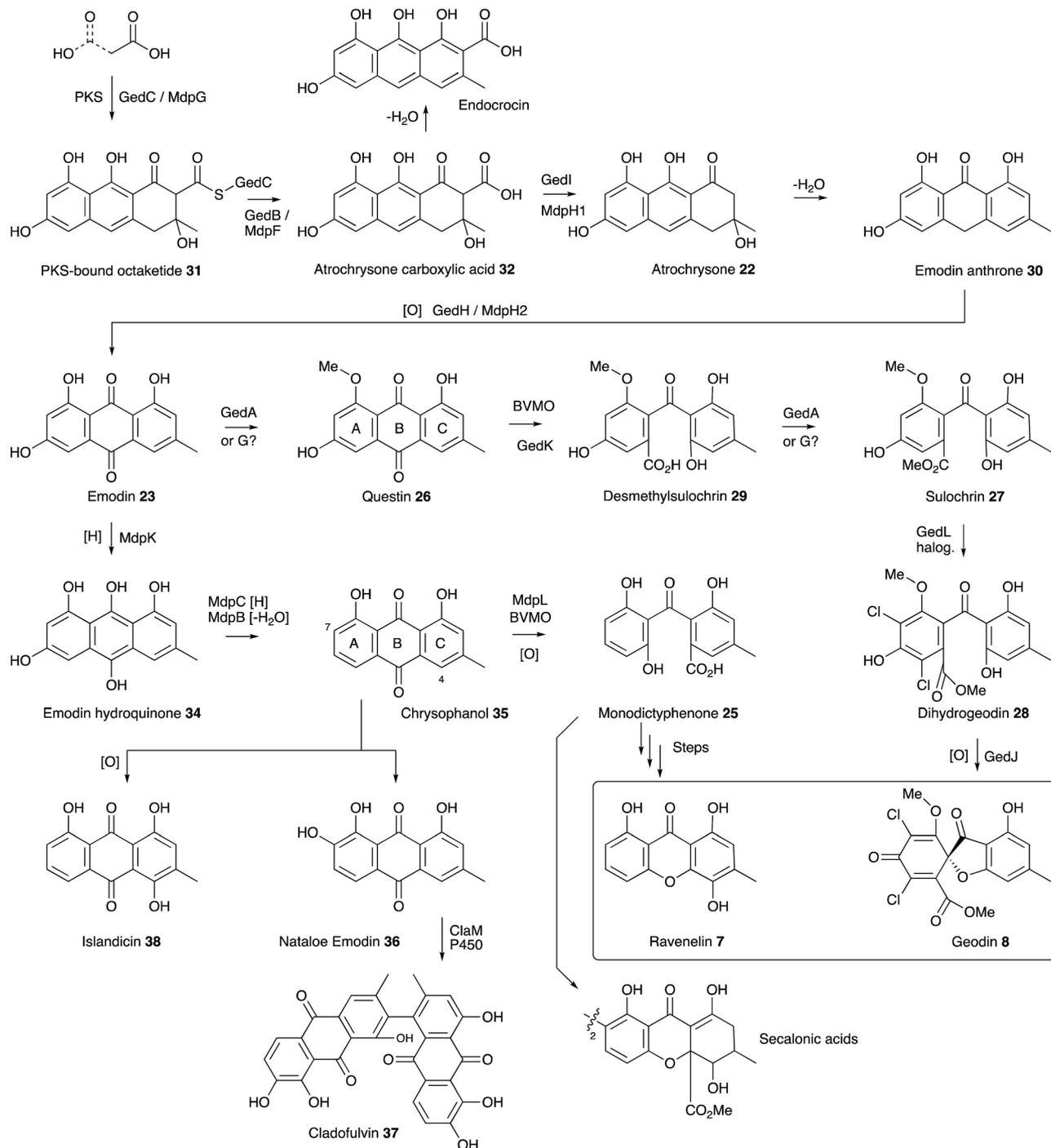
The ubiquity of fungal xanthenes has stimulated very many biosynthetic studies over the past half century, and there is no space here for a comprehensive discussion. However recent molecular advances, particularly into the biosynthesis of monodictyphenone 25 (*mdp*),³⁵ the prenylated xanthenes,³⁶ and the secalonic acids (otherwise known as the ergochromes)³⁷ have shed significant light onto the early steps of ravenelin 7 and geodin 8 biosynthesis and highlighted the key intermediate as emodin 23. It is now certain that the non-reducing PKS MdpG (homologous to GedC) produces a non-reduced PKS-bound octaketide 31 which is hydrolytically released by MdpF (GedB) to produce atrochrynone carboxylic acid 32, which in turn gives

atrochrynone 33 and emodin anthrone 30 after decarboxylation and hydrolysis (GedB/MdpF). Conversion to emodin 23 is catalysed by MdpH2 (GedH), although Müller and coworkers have convincingly argued that emodin 23 is probably present as the corresponding reduced emodin hydroquinone 34 *in vivo*.³⁸ In *in vitro* experiments MdpC and MdpB respectively provide the key intermediate chrysophanol 35 (Raistrick, 1950)³⁹ via a reduction–elimination sequence. Simpson has convincingly argued that chrysophanol 35 is subjected to a Baeyer–Villiger oxidation catalysed by MdpL to give monodictyphenone 25 which would be a precursor of ravenelin 7 and the secalonic acids.⁴⁰ Information concerning these later molecular steps is so far missing. Chrysophanol 35 is also the substrate for hydroxylation, *e.g.* at C-7 to provide nataloe emodin 36, the precursor of cladofulvin 37,⁴¹ or at C-4 to provide the isomeric islandicin 38 (Raistrick 1949).⁴²

The geodin pathway branches from emodin 23 and proceeds *via* questin 26 to desmethylsulochrin 29. Labelling studies show that the regioselectivity of this BVMO must be opposite to that operating in the *mdp* pathway, *i.e.* C-ring oxidation. Finally, methyl ester formation, chlorination and oxidative ring closure give geodin 8. Although the geodin pathway is now well understood, key questions regarding the later-stage steps and mechanisms of many other anthraquinone and xanthenone pathways, such as those to the secalonic acids, remain to be solved. One recent highlight has been the discovery of the P450 system responsible for the oxidative dimerisation steps which form the bis-anthraquinones such as cladofulvin 37.⁴⁰

Genomic studies are beginning to show that the *mdp* pathway is very widespread in fungi and it appears that early evolution of the steps to emodin 23 have formed the foundations for later diversification to many different bioactive structures involved in key processes of pathogenicity, particularly with respect to plants and other fungi. It seems likely that genomic investigations of these pathways using an evolutionary rationale will probably lead to discoveries of many other related compounds and pathways. Of the pathways which are now known to flow from emodin 23, for example to the large class of secalonic acids, key molecular questions remain regarding the (often) oxidative rearrangements leading to the complex final structures.





Scheme 3 Summary of the emodin pathway to geodin 8, ravenelin 7 and related compounds.

4. The maleidrides

Gluconic acid 39 (*Penicillium purpurogenum*), reported by Nadine Wijkman in 1931,⁴³ was the first member of a family of carbocyclic maleic anhydrides which have become known as maleidrides. Raistrick and Smith reported byssochlamic acid 6 from *Byssochlamys fulva* soon after in 1933.⁴⁴ As in the cases of many other compounds isolated and purified during this period, the structure elucidation proved very difficult, and it was

not until the early 1960s that definitive structures were elucidated by Baldwin, Barton, Sutherland and coworkers (chemical degradation)⁴⁵ and Sim and Robertson (Crystallography)⁴⁶ for both compounds. The related gluconic acid 40 (Fig. 2) is isomeric to byssochlamic acid 6 and Barton and Sutherland speculated that such compounds could be derived by dimerisation of C₉ monomers – coining the term nonadride for this class⁴⁷ – with different regiochemical dimerisations explaining the structural differences. Heaveadride 41 (*Helminthosporium*

heveae) isolated by MacMillan and coworkers in the early 1970s falls into the same C₉-derived family,⁴⁸ but other compounds such as rubratoxin B **42** (ref. 49) and the structurally more complex phomoidrides **43** (ref. 50) must be derived from larger precursors. Most family members contain two maleic anhydride moieties, but the selective herbicide cornexistin **44** (ref. 51) contains only one. Despite the increasing numbers of larger compounds discovered the name nonadrides later became associated with the central 9-membered ring. Later related *octadrides* such as zopfiellin **45** (ref. 52) and viburspiran **46**,⁵³ and *heptadrides* such as agnestadride A **47** (ref. 54) have been discovered with 8- and 7-membered central rings respectively, leading to the general term maleidrides (Fig. 2).

As soon as the structures of **6** and **39** were elucidated their biosynthetic origins were of interest. Sutherland and coworkers reported the results of [1-¹⁴C]- and [2-¹⁴C]-acetate feeds in 1968 which showed the incorporation of three acetates head-to-tail in each C₉ monomer during biosynthesis of glauconic acid **39** in *Penicillium purpurogenum*. Scrambling of label from acetate at the other three carbons suggested the use of a Krebs cycle intermediate for the remaining C₃ unit and this was confirmed by feeding [2,3-¹⁴C₂]-succinate which suggested that oxaloacetate was a more likely precursor than pyruvate.⁵⁵ The results supported a previous suggestion that a polyketide or fatty acid is condensed with oxaloacetate to make a citrate-like intermediate which dehydrates to the observed maleic anhydride **48** (Scheme 4).⁴⁶ The symmetrical incorporation of the labels strongly supported the idea that dimerisation of such an intermediate formed the nine-membered ring in **39** *etc.* This hypothesis was confirmed by feeding 2-(1-³H)-*n*-butyl-3-methylmaleic anhydride (*i.e.* dihydro-**48**) to *P. purpurogenum* and observation of symmetrical tritium incorporation into **39**, albeit at low levels of incorporation (<0.5%). Later results showed that the unsaturated anhydride **48** is incorporated much more efficiently (*ca.* 50%).⁵⁶

These results encouraged Sutherland and coworkers to attempt a biomimetic synthesis of glauconic acid **39**, reported in 1972. Treatment of the anhydride precursor **48** with either NaH or Et₃N produced a low yield (2–4%) of the nonadride isoglaucanic acid **49**. This observation supported the proposed mechanism of formation of the nonadrides as either a concerted 6π + 4π cyclodimerisation, or a stepwise polar process (Scheme 4A).⁵⁵ Baldwin and coworkers extensively reinvestigated this biomimetic process using an *n*-pentenyl maleic anhydride **50** (rather than the *n*-butenyl analogue **48** used by Sutherland) during the late 1990s. Although still low, optimisation of reaction conditions resulted in yield improvements (up to 8.5% for **51**), and the isolation of isomeric compounds with six- (**53**) and seven-membered rings (**52**) in similar amounts.⁵⁷ The structural relationships between these compounds suggest an initial step in which an anion attacks at C-4 to form the common intermediate **54** (Scheme 4B). Three different modes of cyclisation are then possible to afford the observed 9-, 7- and 6-membered products, and this favours the stepwise mechanism over the concerted 6π + 4π addition. The use of polar solvents improved overall yields through stabilisation of the required anionic starting material, and the addition of Mg²⁺ ions favoured formation of cyclised products over polymers, presumably via coordination and activation of the maleic anhydride electrophile.

Further progress in understanding the biosynthesis of maleidrides did not come for another 15 years and had to await the development of full genome sequencing and effective molecular methods in fungi. A thorough reinvestigation of the secondary metabolite profile of the original *B. fulva* strain investigated by Raistrick in the 1930s by Cox, Simpson and coworkers revealed the presence of several related compounds.⁵⁴ Two heptadrides (**47** and its dehydration product) were isolated and these appear to derive from the same pathway as byssochlamic acid **6** itself.

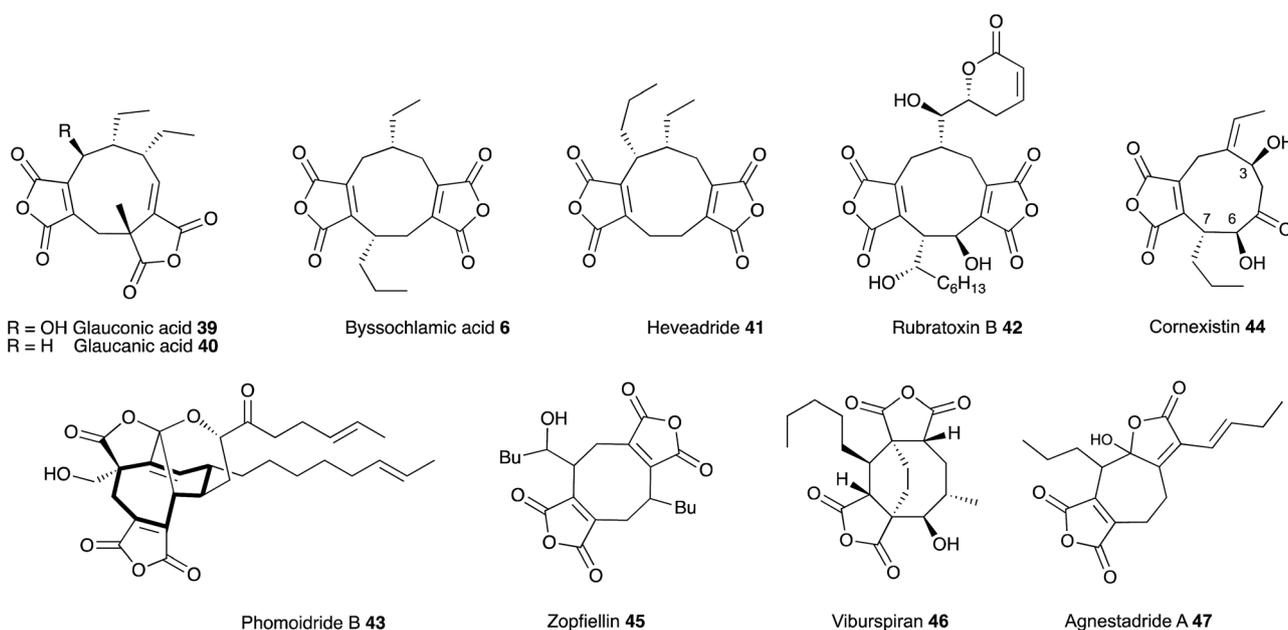
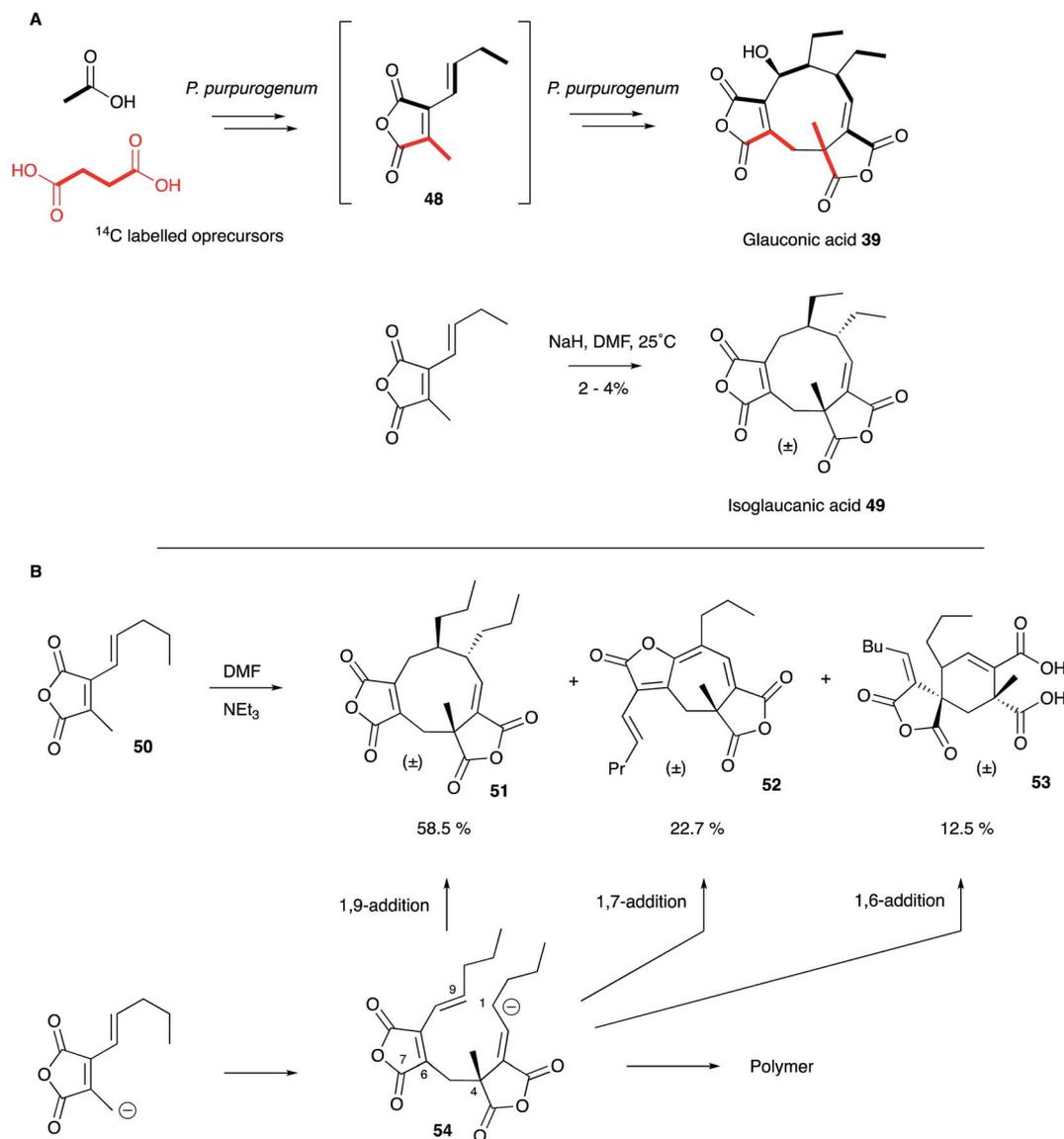


Fig. 2 Structures of key maleidrides.





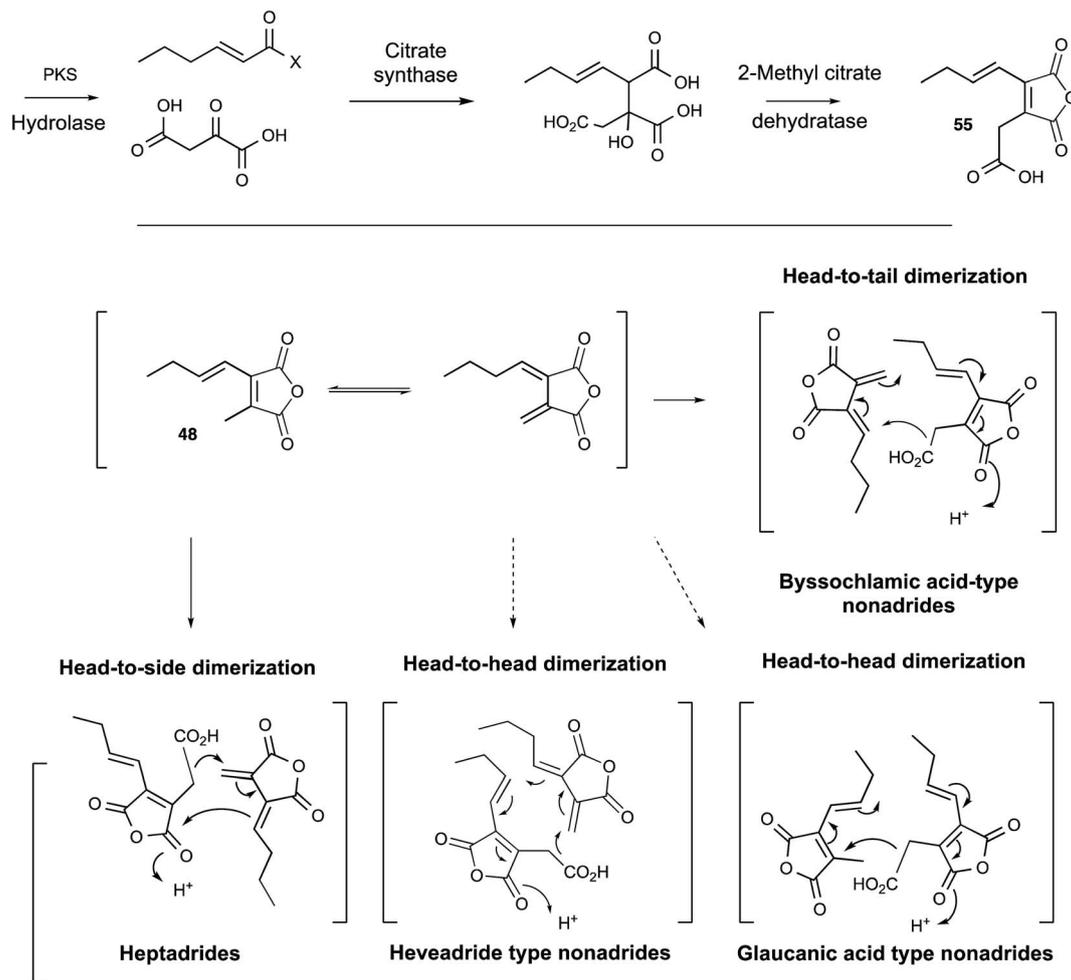
Scheme 4 Biosynthetic and Biomimetic routes to maleidrides: (A) Sutherland's work; (B) Baldwin's work.

The carboxymethyl maleic anhydride **55** was also isolated from a fungus for the first time, and was shown to be unstable to spontaneous decarboxylation in solution to give the methyl maleic anhydride **48** shown by Sutherland and Baldwin to be an *in vitro* precursor of the nonadrides. This compound appears to be derived from condensation of a polyketide and oxaloacetate in a citrate synthase-like process in agreement with the original suggestions of Sutherland and others. Full genome sequencing of *B. fulva* was then achieved. A search for a biosynthetic gene cluster (BGC) encoding a fungal highly reducing polyketide synthase (hr-PKS) and a citrate synthase (CS) enzyme revealed a candidate cluster which also encoded a hydrolase, a 2-methylcitrate dehydratase (2MCDH) and several proteins of unknown function.⁵⁸ Transcriptomic analysis linked activity of the cluster with metabolite production and defined possible limits of the BGC. Knockout of the PKS gene then gave definitive evidence for its involvement in the biosynthesis of the maleic

anhydride **55** and both the nonadride **6** and heptadride **47** components (Scheme 5).

Stepwise reconstruction of the gene cluster in the fungal host *Aspergillus oryzae* was then used to probe the function of the biosynthetic components. Coexpression of the PKS, hydrolase, CS and 2MCDH encoding genes led to production of the carboxymethyl maleic anhydride **55**, as expected, but not the nonadrides or heptadrides.⁵⁷ Parallel work by the group of Oikawa, focussing on the biosynthesis of the phomoidrides **43**, came to a similar conclusion that a hr-PKS, CS and 2MCDH are involved in the production of the monomer.⁵⁹ The byssochlamic acid BGC contains several genes of unknown function. Two of these encode proteins with homology to ketosteroid isomerases (KSI), and two more show homology to phosphatidylethanolamine binding proteins (PEBP). Coexpression of the core genes encoding the biosynthesis of **55** with the two KSI-encoding genes increases the titre of **55** and produces byssochlamic





Scheme 5 Biosynthesis of maleidrides.

acid **6**. Addition of the two PEBP genes increases the titre again, giving significantly more **6** and also produces the heptadride **47**.

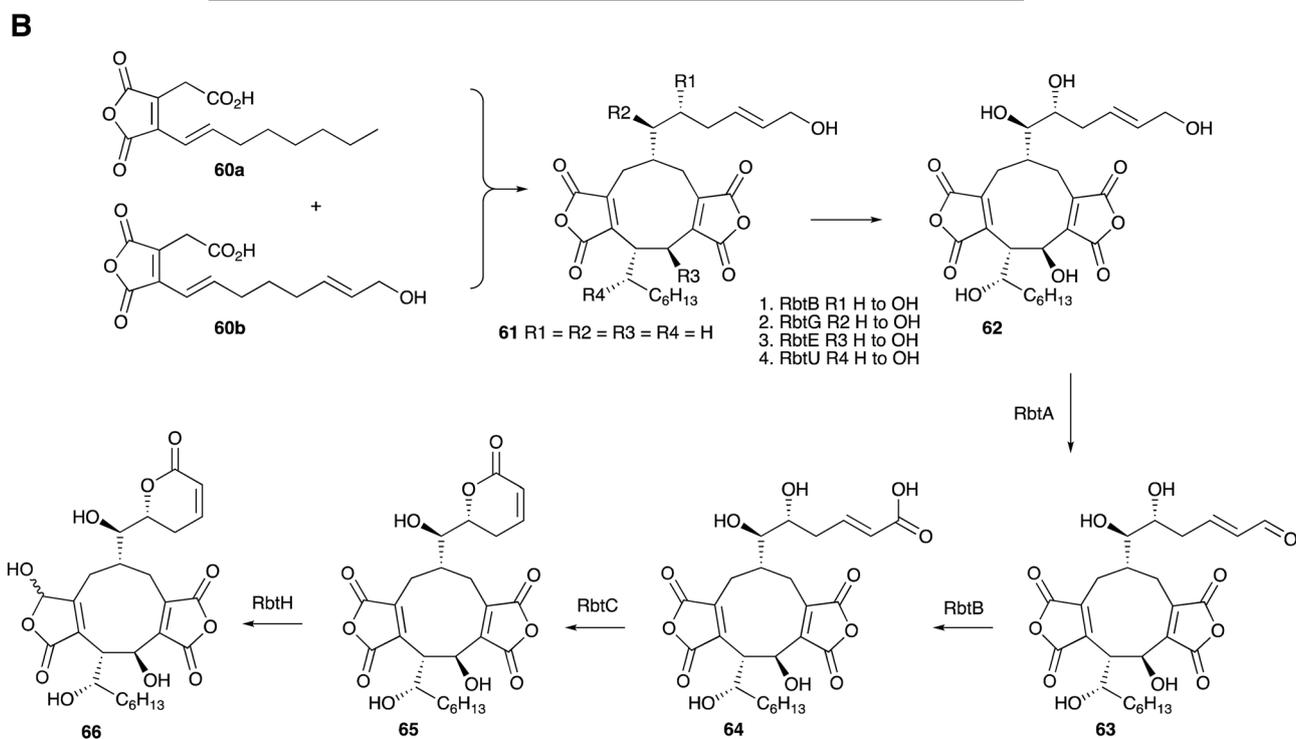
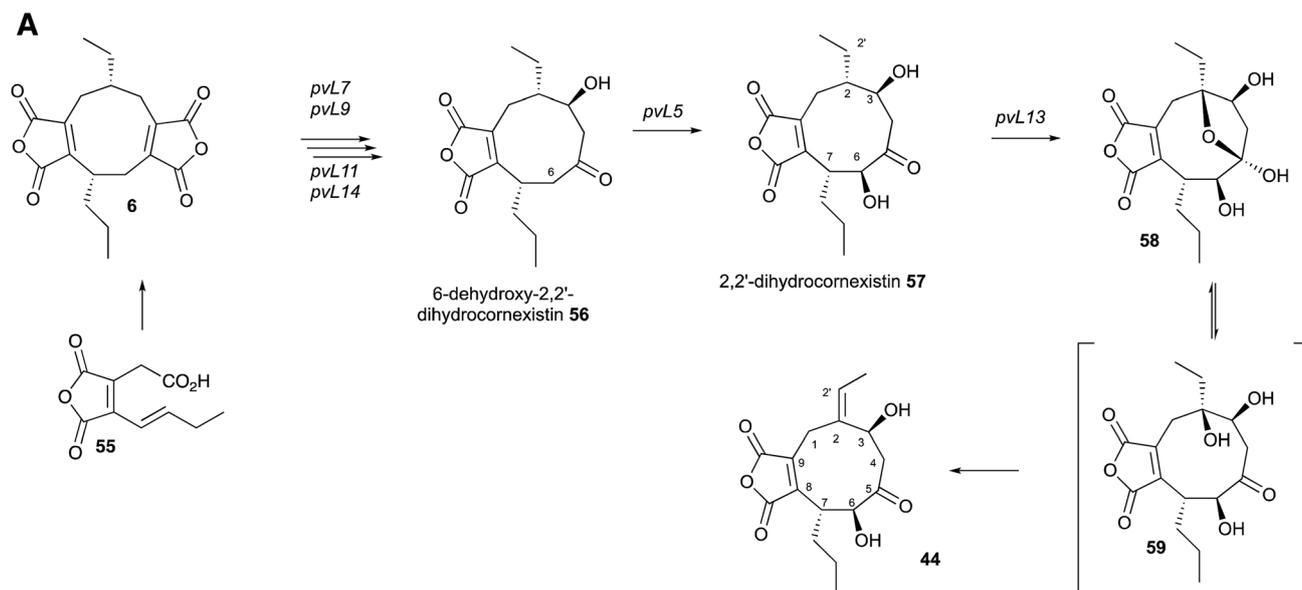
Two recent studies have revealed the complex series of oxidative steps which convert the nonadride skeleton into more complex and biologically more interesting compounds (Scheme 6A). Genome sequencing strategies have revealed the BGC responsible for the biosynthesis of cornexistin **44** which is a potent and selective herbicide.⁶⁰ Once again the core genes encode an hr-PKS, hydrolase, CS and 2MCDH. However the cornexistin BGC differs from that responsible for byssochlamic acid **6** biosynthesis in including only single copies of the KSI and PEBP genes. Unlike the BGC for **6**, the cornexistin cluster encodes numerous oxidases and other genes of unknown function. While the oxidases were shown to be involved in late-stage decoration of the cornexistin skeleton, the processes involved in removal of one of the maleic anhydrides could not be elucidated by knockout experiments and remain cryptic.

In the case of the biosynthesis of rubratoxin B **65**, a potent and specific inhibitor of protein phosphatase 2, Yu, Tang, Hu and coworkers revealed the BGC by full genome sequencing of *Penicillium dangeardii* and genome mining using the byssochlamic acid biosynthetic genes as targets.⁶¹ This revealed

a BGC containing the expected PKS, CS, 2MCDH, KI and PEBP genes, as well as genes encoding seven further redox enzymes. Knockout experiments supported by re-feeding, and *in vitro* assays of isolated recombinant enzymes elucidated the oxidative steps, and also implicated an interesting possibility of an early hydroxylation of one maleic anhydride precursor **60** and an asymmetric dimerisation to form the initial nonadride **61** (Scheme 6B). The pathway also features a dual functional non-heme iron dependent oxygenase (RbtB) which appears to act at two unrelated points in the pathway. Finally, an interesting new reductase (RbtH) which can convert one of the maleic anhydrides to the corresponding γ -hydroxybutenolide of rubratoxin A **66** was discovered.

Thus the early predictions of Sutherland and others regarding the likely biosynthesis have been proven correct with combination of polyketide and citrate-processing steps. The use of PKS, hydrolase and CS also features in the biosynthesis of other fungal metabolites such as the squales-tatins⁶² and these genes form a functional motif which appears to be widely distributed in fungi. Once again it seems that evolution of a particular biosynthetic pattern leads to an intermediate structure which can be diversified to many other





Scheme 6 Biosynthesis of cornexistin 44 and rubratoxin B 66.

bio-active compounds by addition of genes encoding new functions.

It seems clear that the KSI component of the synthetic machinery can form the nine-membered ring by a dimerisation process, but the precise chemical steps remain to be elucidated – for example it is not yet clear if the decarboxylation of the initial maleic anhydride (*e.g.* 55) is coupled to the dimerisation or not. In the case of byssochlamic acid 6 biosynthesis the KSI and PEBP proteins can produce both 9- and 7-membered rings, but as yet no explanation has been

found for the biosynthesis of the related 8-membered octadrides such as viburspiran 46 and zopfiellin 45. Baldwin's biomimetic work shows that the chemistry to form the various regioisomers is feasible, even without enzyme catalysis, but selectivity-controlling features of the biological processes remain elusive. There are also numerous examples of linear family members such as cordyanhydrides⁶³ A and B 67 and 68 (Fig. 3) and the biosynthesis of these has not yet been investigated. It seems likely that future advances will focus on *in vitro* studies of these mysterious enzymes to provide a better



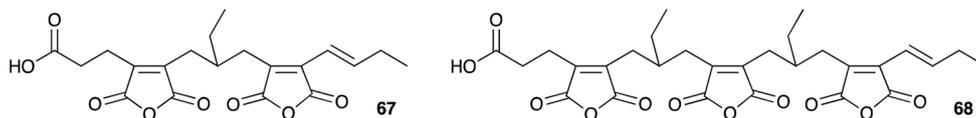


Fig. 3 Cordyanhydrides.

level of understanding, particularly of their mechanism and selectivity.

5. Citrinin and the azaphilones

Citrinin **2** was first reported by Raistrick and Hetherington in 1931 as a yellow component of the organic extract of *Penicillium citrinum*.⁶⁴ Lack of useful spectroscopic methods meant that Raistrick and Robert Robinson's early studies to elucidate its structure relied exclusively on chemical and physical methods.⁶⁵ The deduced structures were unsatisfactory and many groups struggled with this problem through the 1940s during which a lively series of papers disputed each other's claims.⁶⁶ Further efforts by Cram refined, but did not solve the structure⁶⁷ and it was not until 1948 that Whalley and coworkers elucidated the now accepted quinomethide structure.⁶⁸ Early researches focussed on the potential antibacterial properties of citrinin, no doubt stimulated by the emerging work with penicillin from another *Penicillium* species, but it was soon clear that citrinin was too toxic for medical use.

Schwenk and coworkers⁶⁹ and Birch and coworkers⁷⁰ published almost contemporaneous reports of the incorporation of ¹⁴C-labelled acetate into citrinin which confirmed its polyketide origin. The results of Birch showed that the three branching carbons derived from the C₁ pool (formic acid), but Schwenk showed that the *S*-methyl of methionine is incorporated more efficiently and these were the first demonstrations of the incorporation of methyl branches into fungal polyketides, confirming an earlier idea of R. B. Woodward.⁷¹ Staunton and coworkers concluded that these branching groups must be added prior to aromatisation because ¹⁴C-sclerotinin **69** was weakly incorporated into citrinin by *P. citrinum*, but its desmethyl analogue **70** was not (Scheme 7A).⁷²

Investigations of citrinin biosynthesis served as the basis for the development of a number of innovative isotopic labelling experiments by the Staunton group using stable isotopes during the late 1970s and early 1980s. Development of procedures for the synthesis of specifically isotopically labelled late-stage precursors were developed by Staunton, and independently by Scolastico and coworkers, to show that the ketoaldehyde **71** is the likely first enzyme-free intermediate (Scheme 7B);⁷³ and inverse feeding experiments using [1,2-¹³C₂-2,2,2-¹H₃]-acetate in media made from D₂O provided a sensitive way to trace acetate protons during biosynthesis. For example, observation of ¹H at C-4 of **2** after feeding [1,2-¹³C₂, ¹H₃]-acetate to *P. citrinum* grown in D₂O showed that **73** cannot be a direct precursor (Scheme 7C).⁷⁴ The previously observed conversion of **69** to **2** in low yield (Scheme 7A) therefore probably arises from adventitious reactions in *P. citrinum*. Together, results from these experiments

showed that the polyketide is most likely methylated as the chain is assembled (processive biosynthesis) to give an enzyme-bound intermediate such as **74A**, which then cyclises and aromatises to **74B** before reductive release to give **71** (Scheme 7D). Subsequent complex tailoring reactions must then give **2**.

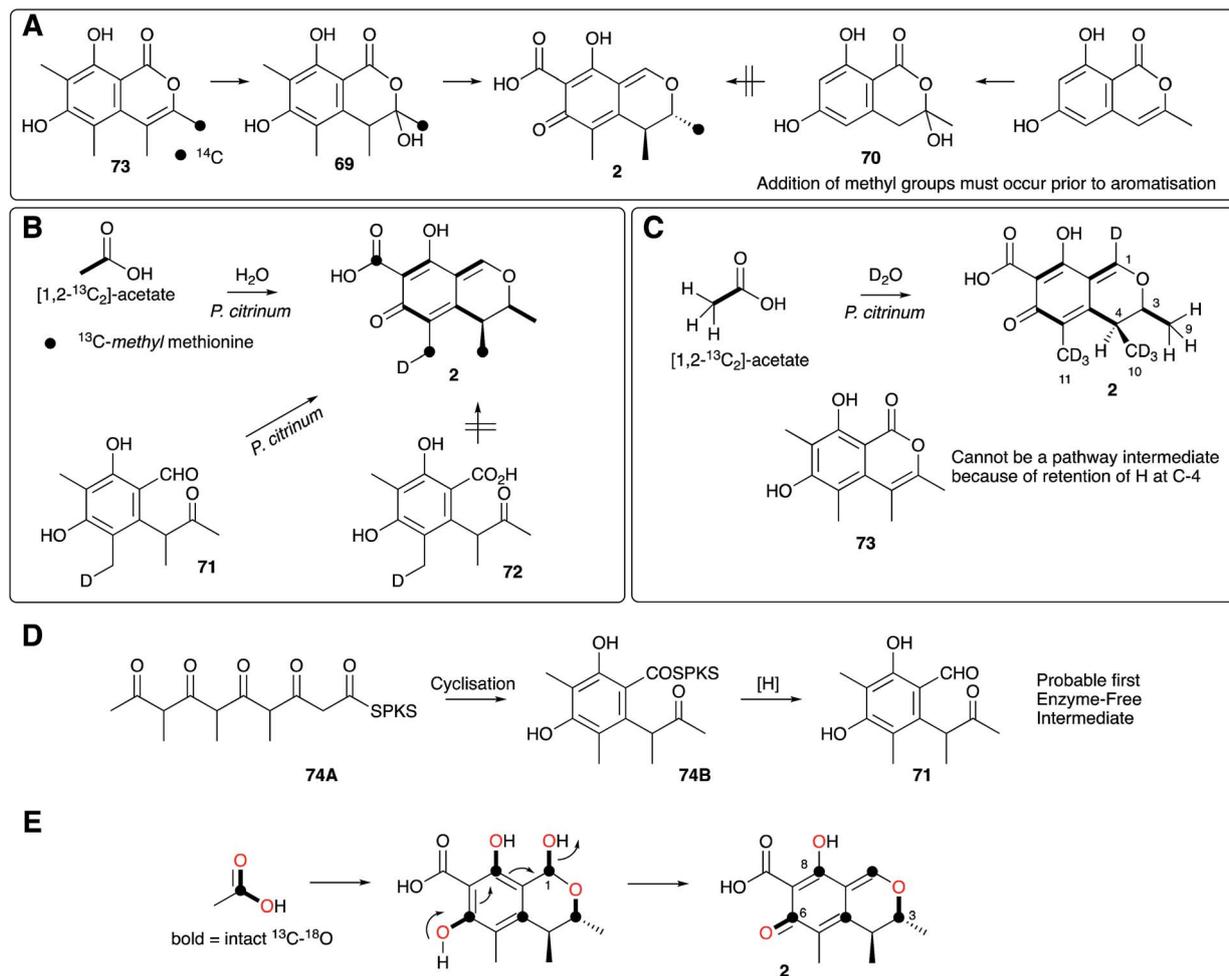
Meanwhile, Sankawa and coworkers used citrinin as a test-bed for early experiments using ¹⁷O- and ¹⁸O-labelled acetates, dual labelled with ¹³C in *Aspergillus terreus*.⁷⁵ Their work clearly showed that both ¹⁸O and ¹⁷O can be effective tracers in biosynthetic studies. ¹⁸O is most conveniently detected by its isotope shift on attached ¹³C nuclei in ¹³C NMR, while ¹⁷O is NMR active itself and can be directly observed. Retention of intact ¹³C-¹⁸O bonds from [1-¹³C, ¹⁸O₂]-acetate at C-6 suggested that the quinomethide is formed by a simple mechanism involving elimination of water from a hemiacetal at C-1 (Scheme 7E).

Despite the extensive work of many groups on the topic of citrinin biosynthesis reported in the literature over 30 years, other poorly interpreted work has also appeared and been highly cited. In particular the work of Francois, Hajjaj and coworkers into the biosynthesis of citrinin in *Monascus ruber* is widely referenced despite an incorrect interpretation of the labelling pattern from [1,2-¹³C₂]-acetate leading to a biosynthetic hypothesis which is starkly different from the previously deduced pathways in *A. terreus* and *P. patulum*.⁷⁶ This route is proposed to involve the cleavage of an acetate despite the labelling pattern from their own data clearly showing that all acetates are, in fact, incorporated intact. This report has misled several subsequent researchers,⁷⁷ and is unfortunately not the only misinterpretation of data in the citrinin story.

A gene encoding the citrinin 2 PKS (*citS*, also known as *pkscI*) was first discovered in *Monascus purpureus* by Nihira and coworkers, reported in 2005, by using a PCR screening approach.⁷⁸ This led to the gradual discovery of a larger genomic fragment containing the complete citrinin BGC and an activator gene *ctnA*.⁷⁹ In a key experiment Nihira and coworkers transferred the entire genomic fragment from *M. purpureus* to the fungal host *Aspergillus oryzae* with the aim of heterologously producing citrinin **2**. Citrinin **2** could be detected in this experiment, but in vanishingly low titre. Titres could be increased to ca. 1.5 mg L⁻¹ by over-expression of *ctnA*.⁸⁰ However no significant analysis of the clustered genes, or accompanying biochemical investigations were reported.

Significant progress has been made more recently in understanding citrinin biosynthesis at the molecular level. Cox and He reported a series of experiments in which the citrinin BGC from *Monascus ruber* was systematically reconstructed in *A. oryzae* by using strong promoters for each structural gene.⁸¹ The citrinin synthase gene *citS* encodes an iterative non-reducing





Scheme 7 Summary of key isotopic labelling experiments which elucidated various steps of citrinin 2 biosynthesis.

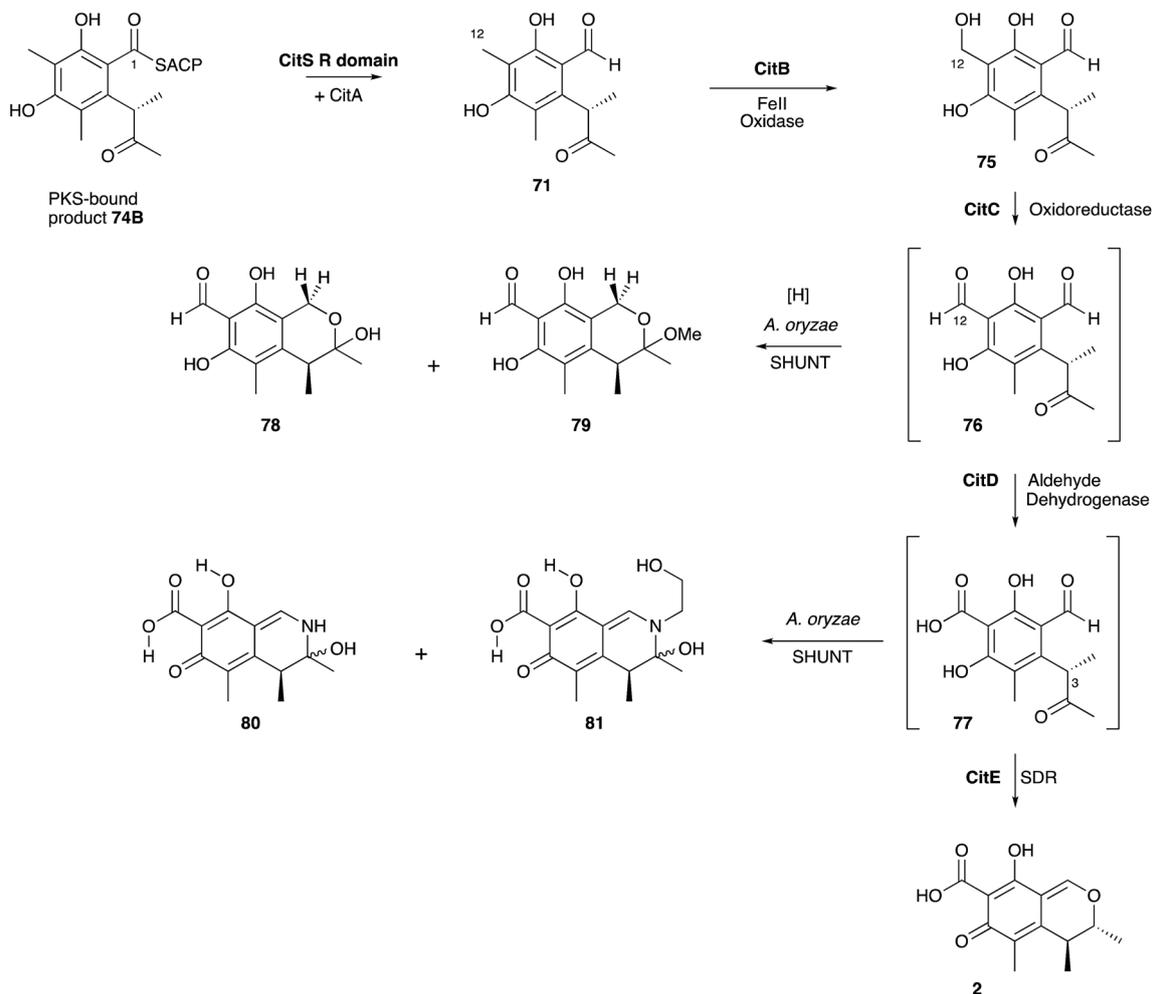
PKS. Expression of *citS* alone led to the production of the keto-aldehyde 71 in agreement with the earlier hypotheses of Staunton and others. This is evidently released from the PKS by its terminal reductive release domain, so the PKS requires no other release mechanism. However, if *citA* encoding a serine hydrolase is coexpressed with *citS* a much higher titre of 71 was observed. Townsend and coworkers recently showed that the hydrolase appears to act as an ‘unblocking’ system, releasing incorrectly synthesised intermediates from the PKS and restoring its processivity.⁸² Systematic co-expression with each of the remaining citrinin BGC components then elucidated the post-PKS pathway in full. The first step is oxidation of C-12 by a non-heme iron dioxygenase encoded by *citB* to form the 12-alcohol 75. This is then oxidised to the C-12 aldehyde 76 by *citC*, a nicotinamide dependent oxidoreductase, and then again to the 12-carboxylic acid 77 by *citD* which is a typical aldehyde dehydrogenase. The final step, catalysed by *citE*, is stereo-selective reduction of the 3-ketone of 77, closure of the dihydropyran ring and formation of the quinomethide 2 by dehydration. The pathway is in full agreement with the previous labelling studies, and negates the pathways suggested by Hajjaj and others. A further advantage of the *A. oryzae* expression

system described here is the high titres obtained-up to 50 mg L⁻¹. The pathway proceeds *via* unstable aldehyde intermediates 76 and 77 and these give rise to numerous shunt pathways in which probable spontaneous reactions diversify the citrinin skeleton to 78–81 (Scheme 8).

Citrinin is closely related to the azaphilone class of fungal metabolites. These compounds are so-named because of their propensity to react with amines non-enzymatically. This is illustrated by the fact that aldehyde intermediates such as 76 and 77 in the citrinin pathway are rapidly shunted to chromanes and dihydroisoquinolines such as 78–79 and 80–81 respectively. In *A. oryzae* ethanolamine appears to act as the nucleophilic amine. Citrinin itself is resistant to amination. The biosynthesis of the closely related azaphilones is now well understood in the cases of chaetoviridin and related compounds such as the *Monascus* pigments and proceeds *via* a pathway parallel to the citrinin route in which reductive release of a non-reduced polyketide is followed by successive oxidations.⁸³

Full understanding of the citrinin pathway at the molecular level allows other parallels to be drawn with known fungal pathways. For example *citB*, a non-heme Fe-II oxygenase, is





Scheme 8 Biosynthetic pathway to citrinin 2 and related shunts.

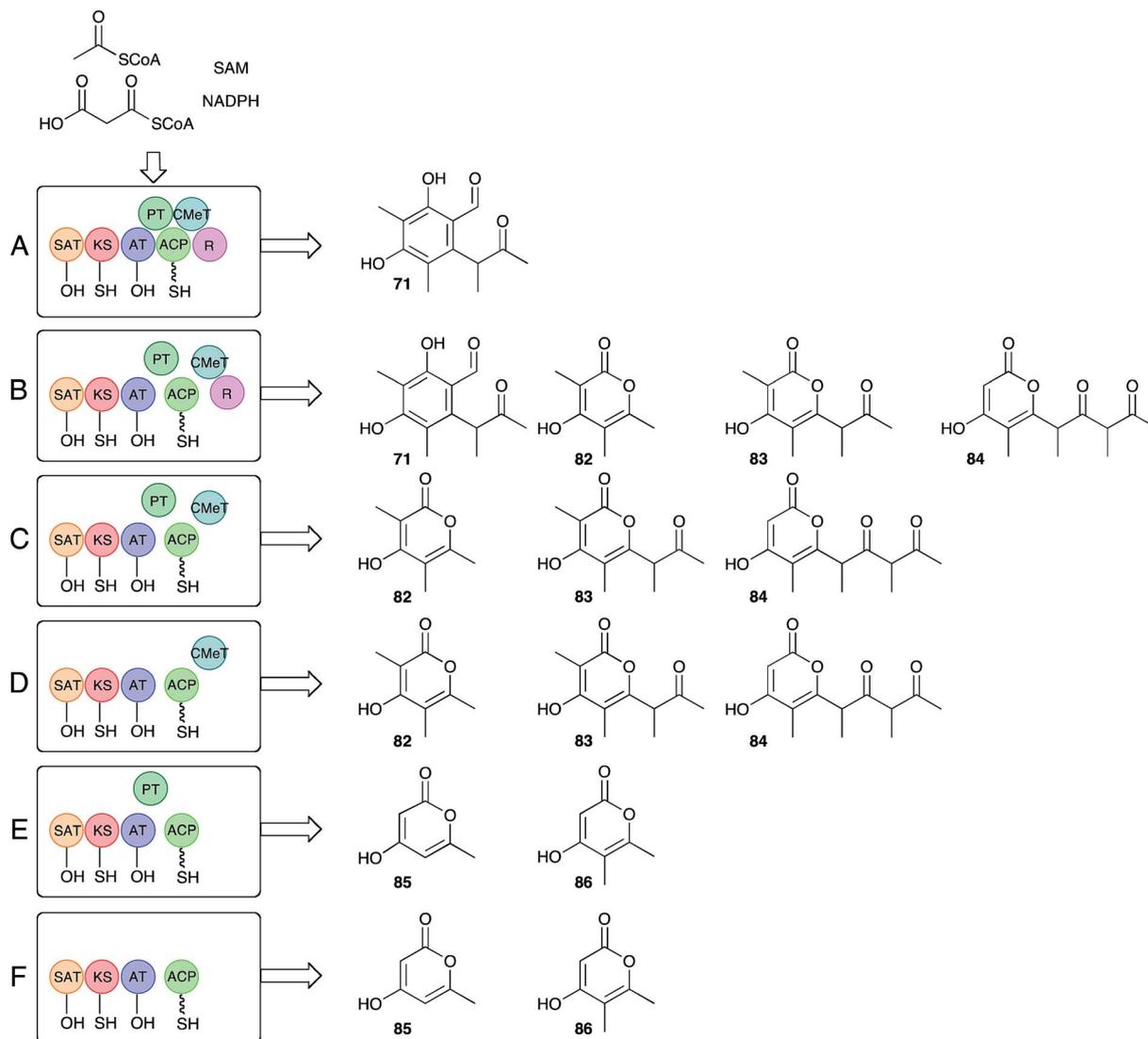
functionally equivalent to TropC in the tropolone pathway (Section 9). However during tropolone biosynthesis the aromatic ring has been oxidised (by the FAD-dependent TropB) and this allows oxidative ring expansion to form the tropolone nucleus. In the case of citrinin 2 the more reduced aromatic nucleus is stable and does not rearrange. Compounds such as sepedonin 5,⁸⁴ another of the Raistraick compounds, feature structural motifs common to both the tropolones and citrinin and it seems likely that the sepedonin BGC will feature both a *citB*-type and a TropC-type oxidase.

Recent work by Townsend and coworkers has set about answering long-standing questions regarding the precise function of the citrinin PKS itself.⁸⁵ Bioinformatic analysis of the citrinin PKS shows it belongs to the non-reducing class of iterative fungal PKS, and consists of (N to C) starter unit acyl transferase (SAT), ketosynthase (KS), acyl transferase (AT), product template (PT), acyl carrier protein (ACP), C-methyltransferase (C-MeT) and reductive release (R) catalytic domains. Much is already known about this class of PKS. For example the SAT domain loads the starter unit (acetate in this case) from CoA to KS, while AT loads a malonyl unit to the ACP. KS catalyses chain extension to create acetoacetyl ACP. In the

absence of other reactions these processes are iterative (4 extensions in the case of citrinin), with the growing β -ketoacyl chain being stabilised and cyclised by the PT domain prior to reductive release giving 2 (Scheme 9A). In the case of *citS*, however, the CMeT domain shows a programmed role and methylates β -carbons after extensions 1, 2 and 3, but not after extension 4.

The approach of Townsend and coworkers was to take a 'deconstruction-reconstruction' approach. They expressed fragments of *citS* in *E. coli* to obtain separate SAT-KS-AT, PT, ACP, CMeT and R domains which were recombined *in vitro* in the presence of substrates and cofactors. Remarkably the dissected and reassembled synthase retains the ability to synthesise 71, although pyrones 82–84, indicative of spontaneous release, were also produced (Scheme 9B). In the absence of the reductive release domain a range of pyrones were also made (Scheme 9C), and further removal of the PT domain gave much the same result (Scheme 9D). Removal of the CMeT domain provided only triketides showing that methylation must occur before chain extension (Scheme 9E and F). Observation of very low levels of 86 were rationalised by copurification of methyl-acetoacetyl CoA which could act as a starter unit. These results





Scheme 9 Townsend's deconstruct/reconstruct experiments with *citS*.

clearly show that the interplay between the C-MeT and KS selectivities is crucial for correct product production in this type of PKS, and support other recent observations of kinetic competition between catalytic domains for ACP-bound substrates as a likely key factor controlling the programming of iterative fungal PKS more generally.^{86–88}

While citrinin itself is a mycotoxin, other azaphilones and related compounds are of major interest as safe food colourants and preservatives, especially in oriental cuisine. Many efforts to understand and exploit their biosynthesis derive from attempts to produce the dyestuffs in higher titre and more selectively. The recent successes in using full and partial pathway expression in the case of the monascus pigments have begun to link the molecular and chemical features required for the production of particular colours of pigments and it seems likely that this work will be extended to include rational efforts to produce new compounds to-order. Such engineering efforts could obviously focus on both the PKS itself, building on the knowledge

gained by Townsend and coworkers of the citrinin PKS, but parallel work by Molnar (see Section 6 below) shows that rational domain-swaps can also be deployed. Increased knowledge of the activities and selectivities of the numerous tailoring redox transformations in varied pathways should also lead to productive mix and match experiments in which recombinant PKS and tailoring genes can be combined.

6. Curvularin and the dihydroxyphenylacetic acid lactones

Raistrick and Rice discovered curvularin 16, a metabolite of *Penicillium gilmanii*, in 1952. The paper communicating its discovery was not published until 1971 shortly after Raistrick's death.⁸⁹ This was the first example of a widespread class of fungal metabolites now known as the dihydroxyphenylacetic acid lactones (DAL), which are in-turn related to the resorcylic acid lactones (RAL). Musgrave, coincidentally working at



Raistrick's old laboratories at the ICI Nobel division at Ardeer, made the first reports of curvularin in 1956,⁹⁰ and the first biosynthetic studies and structure were reported by Arthur Birch, Herchel Smith and coworkers in 1959.⁹¹ The structure was one of the first solved using Birch's acetate hypothesis. Dehydrocurvularin **87** was reported by Musgrave in 1967,⁹² and this compound has proved to be an important testbed for developing ideas about the biosynthesis of highly reduced polyketides in fungi.

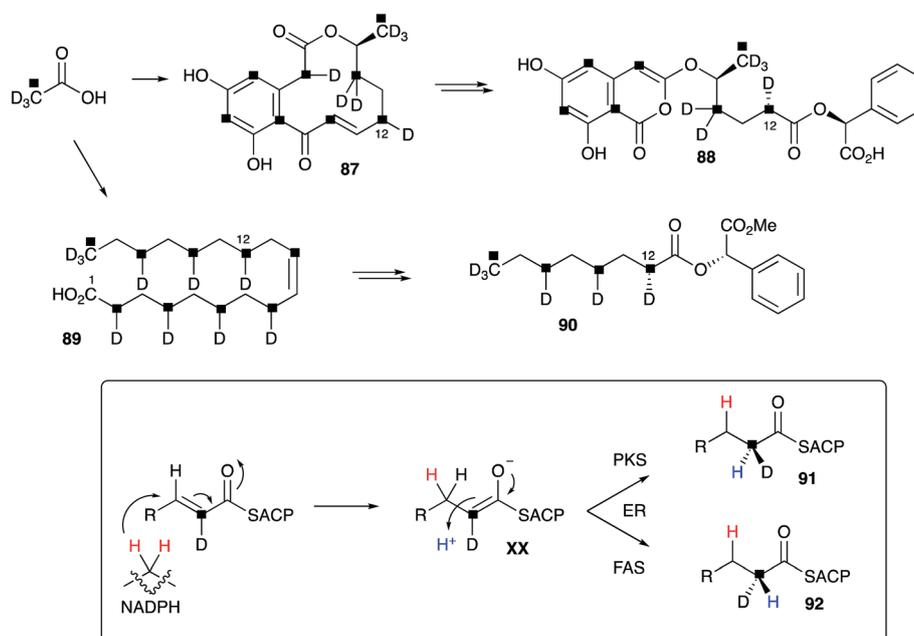
Vederas fed *Alternaria cinerariae*, a producer of **87**, with a wide range of ¹³C, ²H and ¹⁸O-labelled acetates and used ¹³C-NMR to determine the origin of all atoms and bonds in **87**. In particular [2-¹³C, ²H₃]-acetate labelled intact C-D units (Scheme 10), and chemical conversion to **88** provided a system in which the stereochemistry at C-12 could be assessed by NMR. Oleic acid **89** was isolated from the same fermentations, and degradation to **90** also allowed stereochemical assignment. The results of these studies showed clearly that the remaining ²H, at C-2-derived carbons, is incorporated with opposite stereoselectivity in the polyketide and fatty acid (Scheme 10). This indicates that fungal fatty acid synthase (FAS) and the dehydrocurvularin hrPKS must possess enoyl reductase (ER) components with opposite stereoselectivity for the reprotonation step, forming **91** and **92** respectively as intermediates.⁹³

A key early question regarding the mechanism of fungal hrPKS was whether β -processing occurs processively (after each chain extension reaction) or after construction of a complete poly- β -keto carbon skeleton such as occurs during the construction of non-reduced polyketides. Dehydrocurvularin **87** is an interesting case in which the initial tetraketide appears highly reduced, while the final tetraketide appears non-reduced. Vederas and coworkers neatly addressed this point by synthesising and feeding isotopically labelled di-, tri- and tetra-ketides **93**–**98** to *A. cinerariae* as their *N*-acetyl cysteamine thioesters

(SNAC). To prevent degradation of these advanced precursors they also supplemented β -oxidation inhibitors. In addition, to detect incorporation of intact precursors, the compounds were labelled with ¹³C₂ units placed between polyketide acetate positions. ¹³C NMR of isolated **87** showed intact incorporation of **93**, **94** and **97**, strongly supporting the processive mechanism of highly reduced polyketide formation.⁹⁴ The tetraketide **97** was incorporated with surprisingly high efficiency (*ca.* 70%) (Scheme 11).

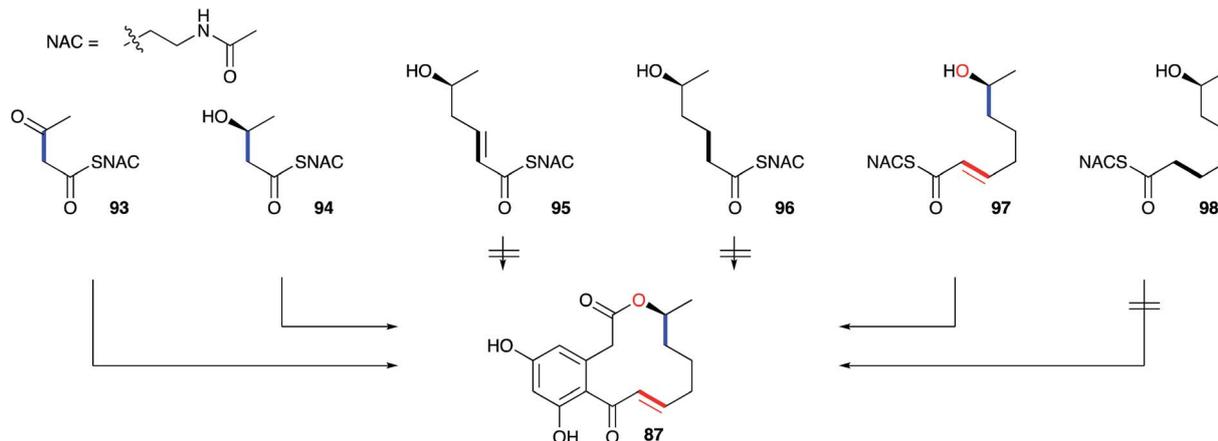
Molecular information was supplied by Molnar's group who isolated the dehydrocurvularin BGC from *Aspergillus terreus*. Two PKS are required for synthesis, a hrPKS (AtCURS1) and a nrPKS (AtCURS2), and this was proven by heterologous expression of these two components in yeast. The hrPKS provides a tetraketide starter unit for the nrPKS, and experiments in yeast in which only the nrPKS was expressed allowed DAL formation when synthetic tetraketides (*i.e.* **97**) were fed as SNAC thioesters (Scheme 12A).⁹⁵ This neatly explains the fact that the tetraketide **97** is so efficiently incorporated into **87** – it is the starter unit for the nrPKS. The nrPKS extends **97** by four more units and catalyses the aromatic cyclisation. Final off-loading by the nrPKS TE domain forms the macrocycle. The use of two PKS, where a hrPKS supplies the starter unit for a subsequent nrPKS is the same as observed to be involved in the biosynthesis of resorcylic acid lactones (RAL), which are another large class of fungal metabolites.

The Molnar group also reported significant progress in understanding and engineering the cyclisation events in the nrPKS component.⁹⁶ During biosynthesis of the DAL class of metabolites nrPKS ring formation occurs at the 3–8 positions (Scheme 12A), while in the RAL class, for example monocillin **99**, the ring formation is 2–7 (Scheme 12B). Ring formation is catalysed by the product template (PT) domain of the nrPKS, and this evidently also controls the regiochemistry since



Scheme 10 Isotopic feeding experiments in *Alternaria cinerariae*.





Scheme 11 Incorporation of advanced labelled precursors into dehydrocurvularin **87** in *Alternaria cinerariae*.

swapping the PT domains led to the formation of the reprogrammed compounds (Scheme 12C). Homology modelling of the PT domains themselves led to the identification of key residues involved in folding and cyclisation and a triple mutation of the AtCURS2 PT domain led to clean conversion from a synthase which made **87** to one which makes **100b**.

The AtCURS2 nrPKS releases its normal product, **87**, using its thioesterase (TE) domain. Similarly the monocillin nrPKS CcRADS2 releases **99**. Molnar and coworkers used a similar domain swap strategy to probe the intrinsic selectivities of these TE domains.⁹⁷ All 8 permutations of TE (AtCURS2 or CcRADS2), aromatic substitution pattern (3–8 or 2–7) and side chain structure (tetraketide or pentaketide) were assessed by building chimeric PKS systems *in vivo*. For example, to assess a pentaketide starter unit with a 3,8 cyclisation pattern and the dehydrocurvularin TE, CcRADS1 was paired with CcRADS2 in which the PT and TE domains were swapped-in from ArCURS2. This provided the new compound radilarin **102** (Scheme 12D). Further work to increase the range of functional DAL/RAL hybrids showed that combinations of the hrPKS and nrPKS can be made with PKS systems which make saturated and unsaturated starter units and nrPKS which make only three extensions.⁹⁸ Methylated compounds such as **104** can be created if the hrPKS component has a CMeT domain, such as AfoG, the starter unit synthase for preasperfuranone **103** which is an early intermediate in azaphilone biosynthesis.⁹⁹ In this case the AtCURS2 PKS had to be engineered to contain the AfoE SAT domain (Scheme 13). These experiments clearly show the opportunities for rational production of defined compounds within the DAL/RAL family and are among the most advanced to date in the sphere of natural product biosynthetic engineering in general.

Thus, significant progress has been made in the field of understanding and engineering RAL and DAL biosynthesis since the first discovery of **16** in the 1950s. Enough is now known to engineer the basic carbon skeletons and cyclisation patterns at-will in a process which is competitive with synthetic chemistry. Future advances will have to address the question of titre, and will no-doubt focus on expanding the range of post-

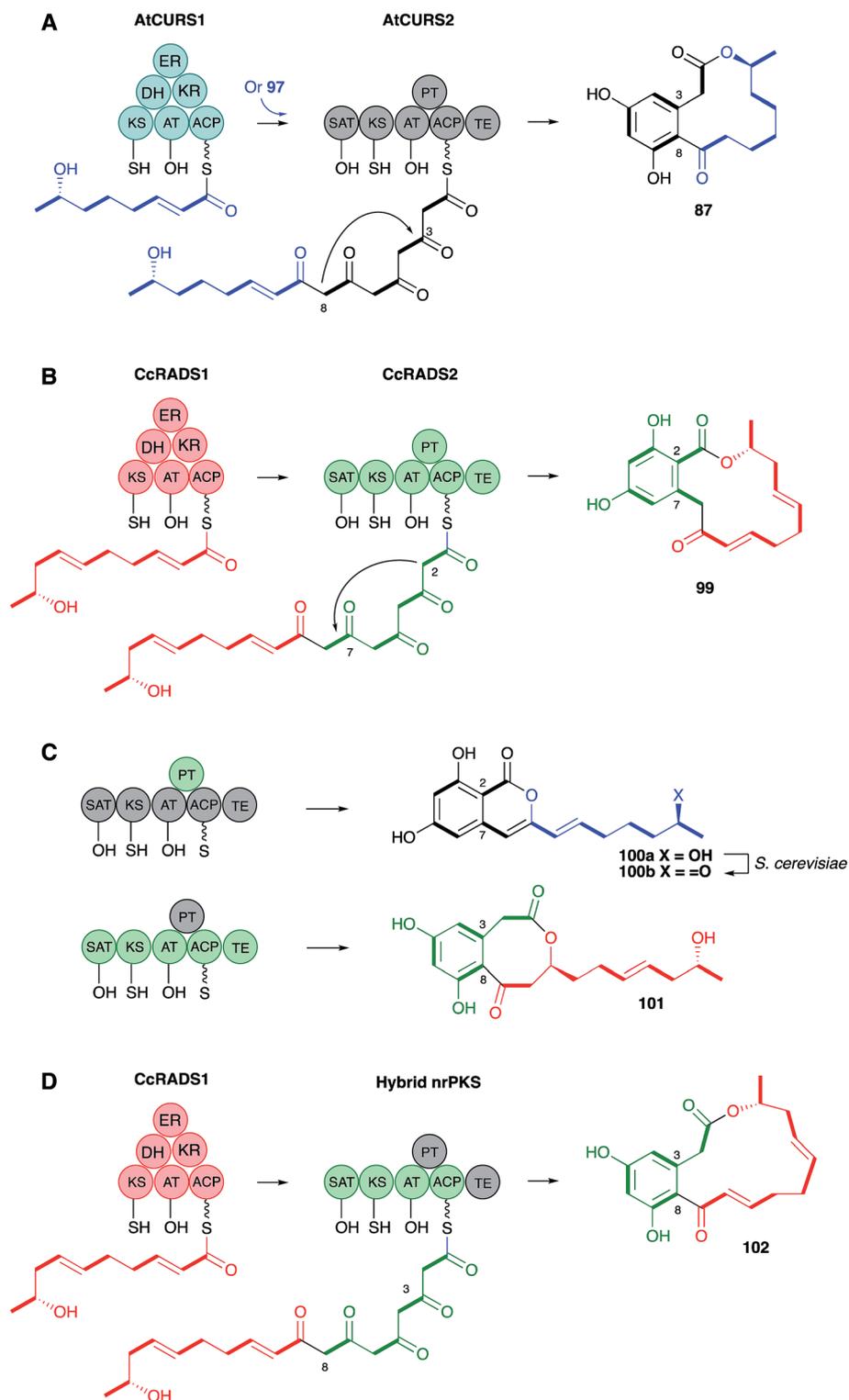
PKS tailoring steps which can be rationally incorporated into the synthetic pathways. New developments in the creation of modular and controllable expression systems in fungi, such as those reported by Lazarus and coworkers,¹⁰⁰ will underpin efforts to move synthesis of this class compounds out of the synthetic chemistry laboratory, and into the sphere of molecular and microbiology.

7. Griseofulvin

Griseofulvin **9** is a useful orally available antifungal agent used in human medicine, and is included in the World Health Organisation's list of essential drugs. Raistrick and coworkers first reported **9** as an unusual chlorinated metabolite of *Penicillium griseofulvum* in early 1939, after around 10 years of work.¹⁰¹ Chemical analysis was able to provide a preliminary structure, and capture the 6,5,6 ring system, but it was not until 1952 when Grove, MacMillan¹⁰² and coworkers at ICI determined the distinctive spirocyclic structure.¹⁰³ Earlier work by Grove had already identified the antifungal properties of **9** when he showed that griseofulvin and the so called 'curling factor' isolated from *P. janczewskii* were identical. Macmillan also reported one of the very first examples of precursor-directed biosynthesis when he fed chloride-starved cultures of *P. griseofulvum* with KBr and isolated bromogriseofulvin.¹⁰⁴ ICI and Glaxo developed and commercialised griseofulvin in a joint venture during the 1950s and 1960s.¹⁰⁵

Birch reported the first studies of the biosynthesis of griseofulvin **9** using [$1-^{14}\text{C}$]-acetate, which were important for confirming the acetate hypothesis of polyketide biosynthesis. Feeding studies with *P. griseofulvum* showed that **9** is constructed from a linear chain of seven acetates.¹⁰⁶ Rhodes and coworkers at Glaxo experimented with fermentation conditions which led to the production of putative precursors of **9**. For example limiting chloride concentration in the media led to the production of various griseophenones **105–106** and **111**, while inhibiting C_1 metabolism gave different methylation isomers **105–108**.¹⁰⁷ Combination of this work with radio-labelling (including the use of ^{36}Cl) then gave various labelled



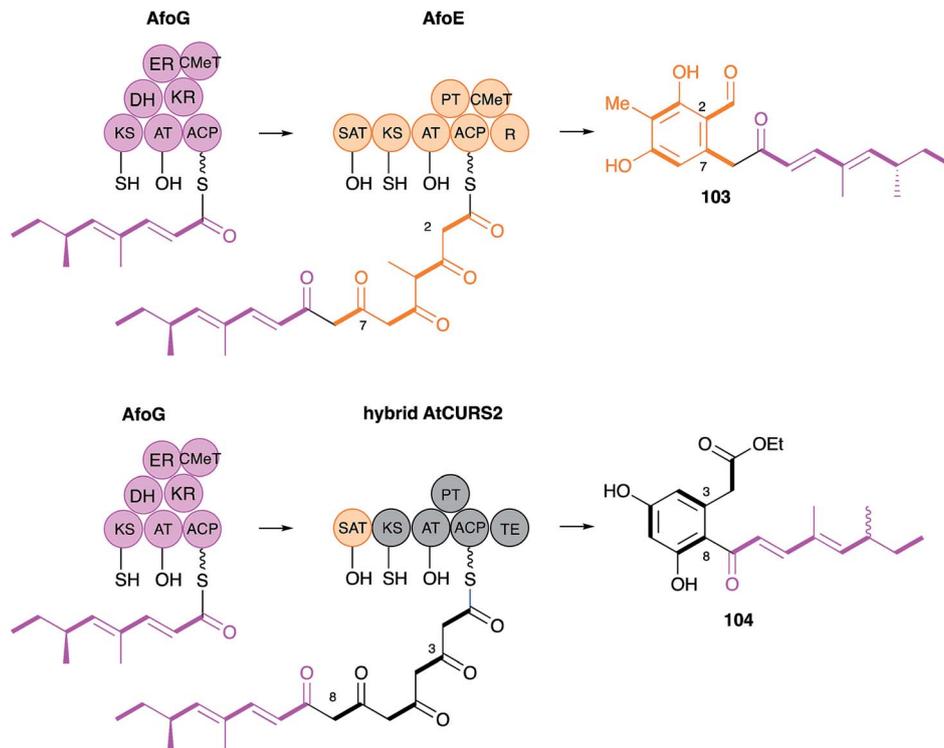


Scheme 12 Engineering of DAL and RAL systems to control ring cyclisation and TE regioselectivity.

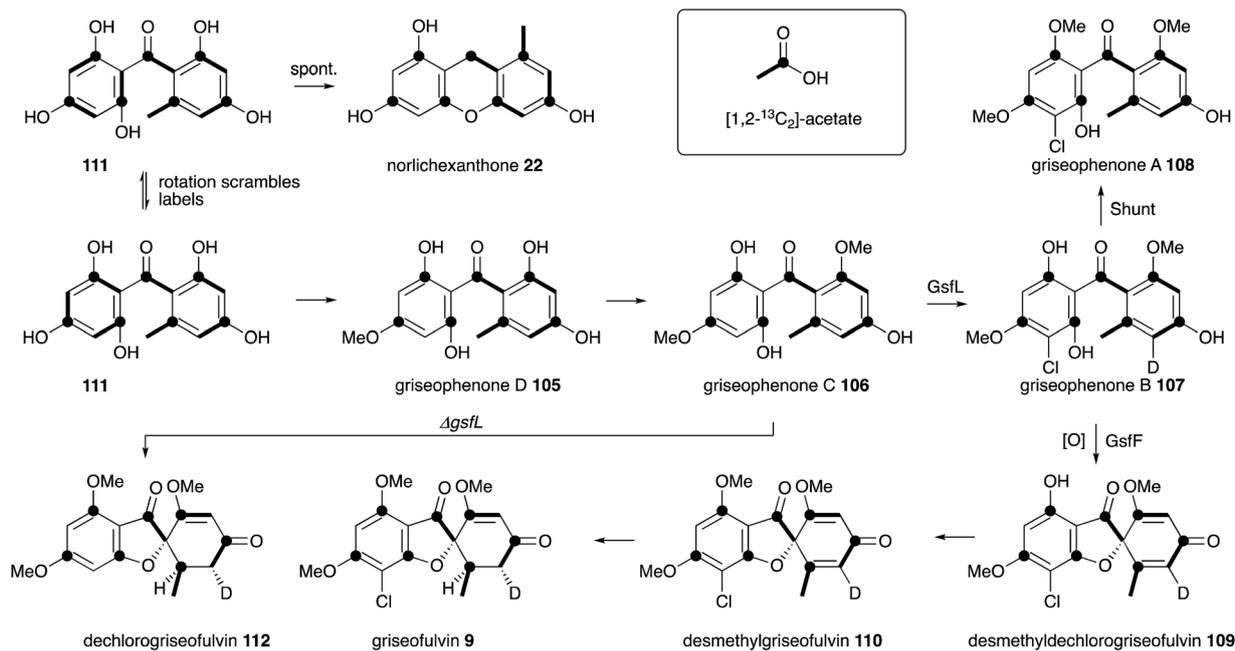
intermediates which were supplemented back to producing cultures. These experiments showed that griseophenone C **106** is converted to griseophenone B **107** which is in-turn a precursor of **9** itself and the shunt metabolite griseophenone A **108**.¹⁰⁸ Harris and coworkers considerably expanded the known pathway by feeding tritium labelled synthetic

intermediates, and this revealed the fuller pathway shown in Scheme 14 in which the unmethylated pentahydroxybenzophenone **111** is the likely first enzyme free intermediate. Feeding of [1,2-¹³C₂]-acetate by Simpson and Holker was used to elucidate the acetate folding pattern,¹⁰⁹ while feeding of [5'-²H]-griseophenone B **107** led to the elucidation of the cryptic





Scheme 13 Engineered hybrid azaphilone and DAL pathways.



Scheme 14 Biosynthesis of griseofulvin 9 deduced by classical labelling and feeding experiments.

stereochemistry of the griseofulvin methylene by ^2H NMR during the 1970s.¹¹⁰

Once again a period of more than 30 years had to elapse before further progress could be made in understanding the molecular steps of biosynthesis. Tang and coworkers sequenced

the genome of *Penicillium aethiopicum*, a known griseofulvin 9 producer, using 454 methodology to generate 1522 contigs with N50 of 149 Kb. Out of a total of 30 PKS genes, 7 encoded non-reducing PKS, and one of these genes was embedded in a cluster also encoding a halogenase and three



methyltransferases.¹¹¹ This BGC was thus designated as the most likely to encode the biosynthesis of **9**. Knockout experiments targeting the core pks, *gfsA*, proved this hypothesis correct. The GfsA PKS does not encode a classic C-terminal thioesterase (TE) domain (e.g. see Section 6) and so it likely releases the heptaketide product **111** by internal Claisen reaction (Scheme 15A). The three methyltransferases (GsfB, GsfC and GsfD) encoded by the cluster and the halogenase (GsfL) must then provide key intermediate griseophenone B **107**. Knockout of the halogenase gene *gsfL* resulted in accumulation of dechlorogriseofulvin **112** in *P. aethiopicum*. It was expected that a copper-dependent enzyme would be involved in formation of the spirocycle, in analogy to known chemistry in the geodin pathway (Section 3). The cluster does not encode such a protein; however, a cytochrome P450 enzyme encoded by *gsfF* was speculated to fulfil this role.

Tang and coworkers have more recently proven all of the steps of biosynthesis by reconstructing the complete pathway *in vitro* using purified proteins, malonyl CoA and cofactors.¹¹² This confirms the order of steps, and the remarkable oxidative formation of the grisan system **109** by the cytochrome P450 enzyme GsfF. The mechanism of this step has been probed computationally and the results indicate that hydrogen atom abstraction from a phenolic OH of **107** to give **114**, followed by direct spiran ring formation to **115** before collapse to **109** is the most likely mechanism. The alternative possibility of initial epoxidation of the toluene ring is of too high energy to be competitive (Scheme 15B).¹¹³

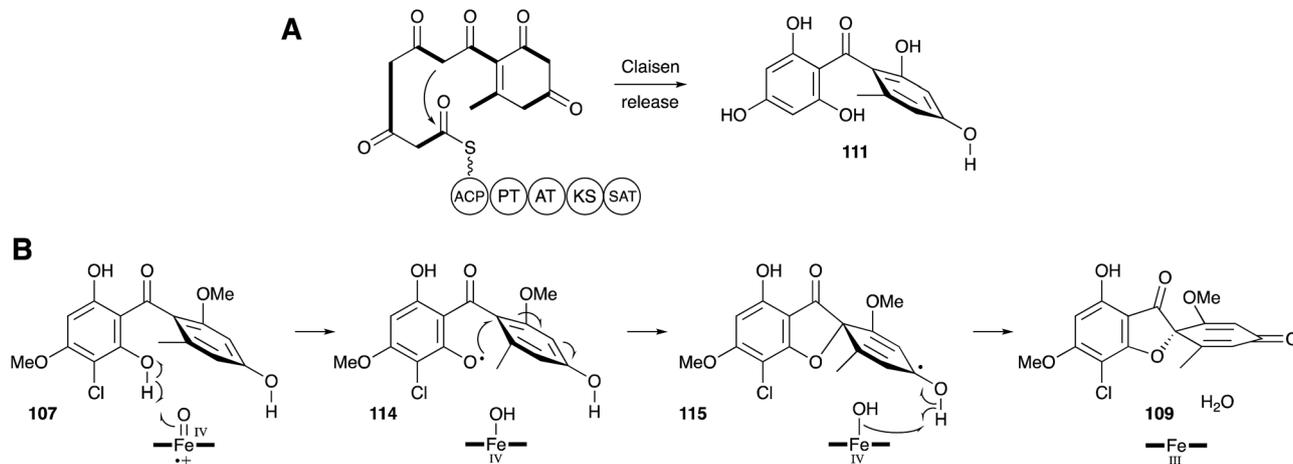
Once again the early work of Raistrick has reached a high level of maturity and a near complete understanding of the pathway and key metabolic steps has been achieved. Again the key advances have come after full pathway reconstruction in a heterologous host – a technique which is increasingly shown to give unambiguous answers to questions regarding the order and selectivity of the chemical steps. Comparison of the griseofulvin and geodin pathways also illustrate that nature can find more than one route for the synthesis of compounds with potent bioactivities (Scheme 16).

8. Mycophenolic acid

Mycophenolic acid **10** was first described by Gosio in the 1890's as a metabolite of *Penicillium brevicompactum* isolated from maize.¹¹⁴ Gosio had already discovered that an extract of *P. brevicompactum* showed antibiotic activity against *Bacillus anthrax*, and as such mycophenolic acid was the first antibacterial compound purified from any source.¹¹⁵ Over the following century **10** was reported to display antifungal,¹¹⁶ antiviral,^{117,118} antitumor,^{119,120} antipsoriasis¹²¹ and immunosuppressing^{122,123} activity, and it is now clinically important. However, **10** was only named mycophenolic acid by Alsberg & Black who purified it from another maize colonising fungus, *Penicillium stoloniferum* in 1913.¹²⁴ It was Raistrick who recognized that the described compounds are identical, and, after studying the secondary metabolite production of 15 related fungal strains, 12 were found to produce **10**.^{125,126} Although Alsberg & Black had identified the correct chemical formula of C₁₇H₂₀O₆, the successful structure elucidation of **10** by extensive chemical reaction studies took Raistrick and co-workers until 1952.^{127–129}

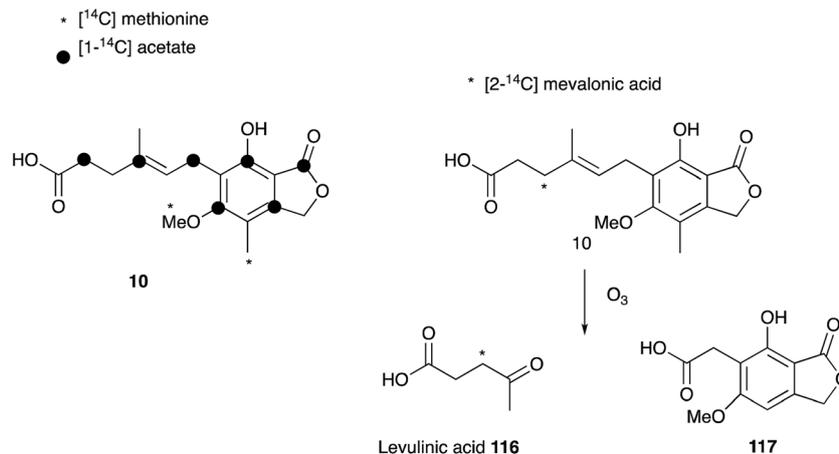
Shortly after structure elucidation, the biosynthesis of **10** became of interest to Arthur Birch, Herchel Smith and coworkers. The origin of the aromatic methyl and the methoxy group was confirmed by feeding [¹⁴C]-methionine.¹³⁰ Further labelling studies with [1-¹⁴C]-acetate confirmed equal integration of acetate into the aromatic nucleus and side chain, which was consistent with polyketide biosynthesis, but also a possible terpene origin. Feeding of [2-¹⁴C]-mevalonic acid resulted in labelled **10**, and label was located in the side-chain by degradation to levulinic acid **116**. This led to the proposal of a geranyl precursor as terpene moiety.¹³¹

However, later feeding experiments with advanced precursors containing C₁₅ or C₁₀ side chains showed that C₁₅ is the preferred terpene chain length to be incorporated into **10**.¹³² Two mechanisms were proposed and proven equally important for the degradation of the C₁₅ farnesyl chain to the observed C₇ side chain in **10**.¹³³ Either stepwise oxidation of the terminal alkene^{134–136} of **116** to the C₁₂ intermediate **113** (Scheme 17A), or direct oxidation of the central alkene¹³⁷ of **116** (Scheme 17B), can occur.

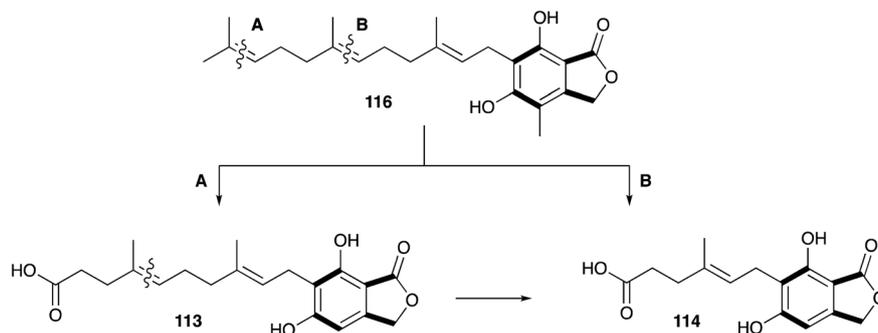


Scheme 15 Key griseofulvin **9** biosynthetic steps revealed by Tang, Houk and coworkers.





Scheme 16 Early isotopic feeding experiments to deduce the origin of mycophenolic acid.



Scheme 17 Oxidative processing of the C₁₅ side chain towards mycophenolic acid.

In 2011 the BGC encoding **10** biosynthesis was first identified in *P. brevicompactum* by Nielsen and coworkers. The strategy employed knowledge about its bioactivity.¹³⁸ Mycophenolic acid inhibits inosine monophosphate dehydrogenase (IMPDH) and thus *de novo* purine biosynthesis. The genome of *P. brevicompactum* IBT23078 was mined for a gene encoding an IMPDH which confers resistance to **10**. In this way Nielsen and coworkers identified BGC of approx. 25 Kb which contains 7 open reading frames (ORF), encoding a NR-PKS (MpaC), a chimeric P450-hydrolase (known as MpaDE), a prenyltransferase (MpaA), an oxidative cleavage enzyme (MapH), an *O*-methyltransferase (MapG), an IMPDH (MapF) and a protein of unknown function (MpaB). Homologous clusters were subsequently identified in one additional strain of *P. brevicompactum*¹³⁹ and in *Penicillium roqueforti*,¹⁴⁰ both known producers of **10**.

The first three steps were determined by heterologous expression and knockout experiments.^{133,141,142} The NR-PKS synthesises the tetraketide **119**, which is hydroxylated at position 6 by the P450 domain of the chimeric MpaDE to give **120**. The hydrolase domain of MpaDE then acts as a lactone synthase yielding **121**.¹³⁷ The intermediate steps, prenylation of **121** to **116** and oxidative cleavage of the C₁₅ to C₇ side chain are proposed to be catalysed by MpaA (prenyltransferase) and MpaH (oxidative cleavage enzyme) but need further experimental confirmation. These early steps of meroterpenoid

biosynthesis are similar to many others (*e.g.* see the comprehensive review of Abe and Matsuda)¹⁴³ in that an electron-rich aromatic nucleus serves as the prenyl acceptor. However, unusually in the case of mycophenolic acid (and related compounds such as yanuthone D)¹⁴⁴ oxidative cyclisation of the terpene does not occur (Scheme 18).

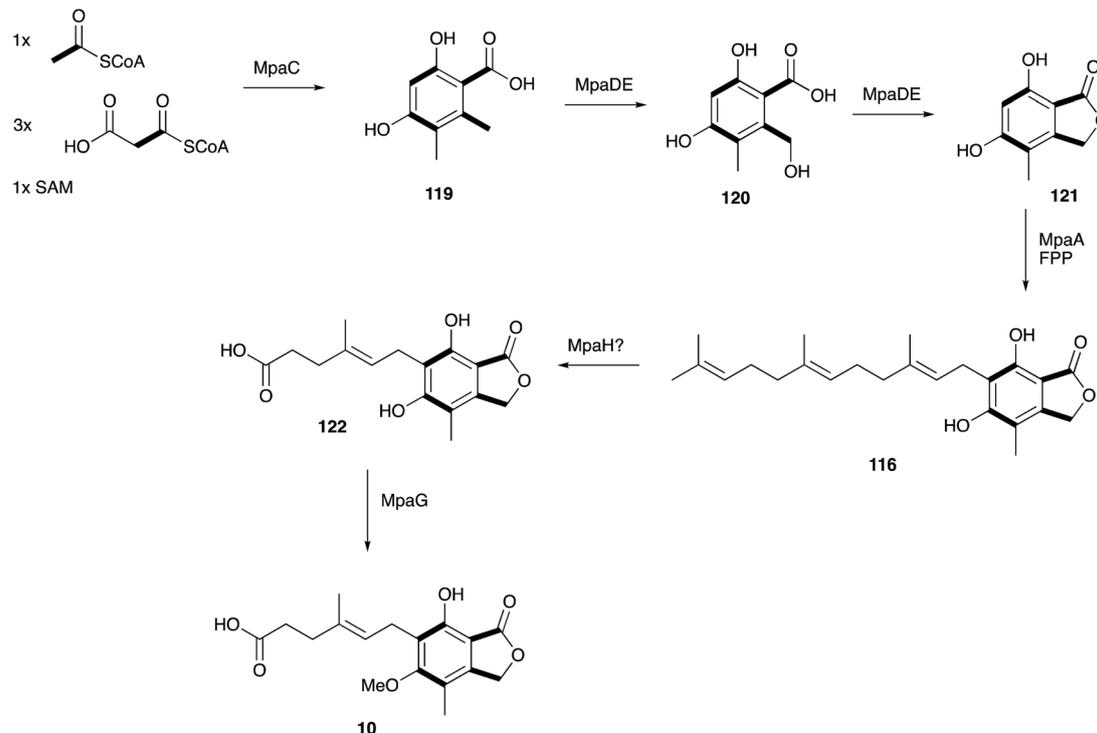
A SAM-dependent *O*-methyltransferase was purified from *P. stoloniferum* and shown to catalyse the methylation of 4-OH in **122** in 1975.¹⁴⁵ Due to technical limitations at the time no DNA sequence is available, but it is most likely to be MpaG. Further *in vitro* assays of recombinant MpaG' (from *P. brevicompactum* NRRL 864) in 2015 confirmed these initial findings and showed a relatively broad substrate selectivity, which could be useful for generation of analogues of **10**.¹³⁴

Thus while impressive advances have been made, a full molecular description of mycophenolic acid biosynthesis is still lacking. It is likely, as in other cases discussed here, that full pathway expression will lead to an understanding of all the steps of biosynthesis and conclusively link these to each gene of the BGC. Once this is complete, then engineering efforts can commence.

9. Tropolones

Raistrick, Birkenshaw and Chambers first reported the isolation of stiptatic acid **4**, isolated from *Penicillium stiptatum* (later





Scheme 18 Overall pathway to mycophenolic acid 10.

renamed as *Talaromyces stipitatus* in 1942. Extensive chemical derivatisation determined the compound to be aromatic with the formula C₈H₆O₅, but a combination of wartime conditions and a lack of a theoretical framework underpinning concepts of aromaticity meant that the structure could not be solved. However, chemical similarities to other fungal compounds such as puberulic acid 3 (first reported in 1932),^{146,147} which also defied structural elucidation, led Raistrick to conclude that these compounds formed a new class of fungal metabolites. The structure was solved in 1945 when Michael Dewar realised that 4 must be a new type of aromatic system consisting of a 7-membered ring he named a tropolone. In a parallel paper Dewar also solved the long mystery of the structure of colchicine which he also showed to contain a tropolone,¹⁴⁸ masked as its methyl ether.

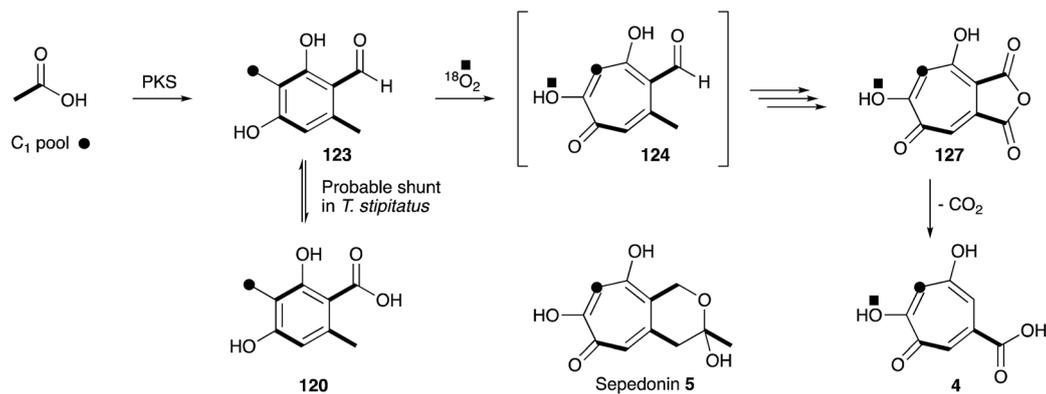
After 1945 the study of tropolone synthesis and biosynthesis became highly topical. For example, Alexander Todd and coworkers reported the first synthesis of 4 in 1950,¹⁴⁹ and Robert Robinson speculated on the biosynthetic origin from “the condensation of polyhydric phenols with formaldehyde or its biological equivalent”.¹⁵⁰ The first experimental investigations into fungal tropolone biosynthesis came from Bentley and coworkers during the early 1960s, with the development of systematic methods for purification¹⁵¹ and controlled degradation of 4.¹⁵² These methods allowed an investigation using ¹⁴C labelled precursors, and confirmed a pathway utilising acetate, malonate and a C₁ source (Scheme 19).¹⁵³

The next concrete result came from the group of Ian Scott who showed that ¹⁴C-labelled 3-methylorsellinic acid 120 was incorporated into stipitonic acid 127,¹⁵⁴ which Bentley had

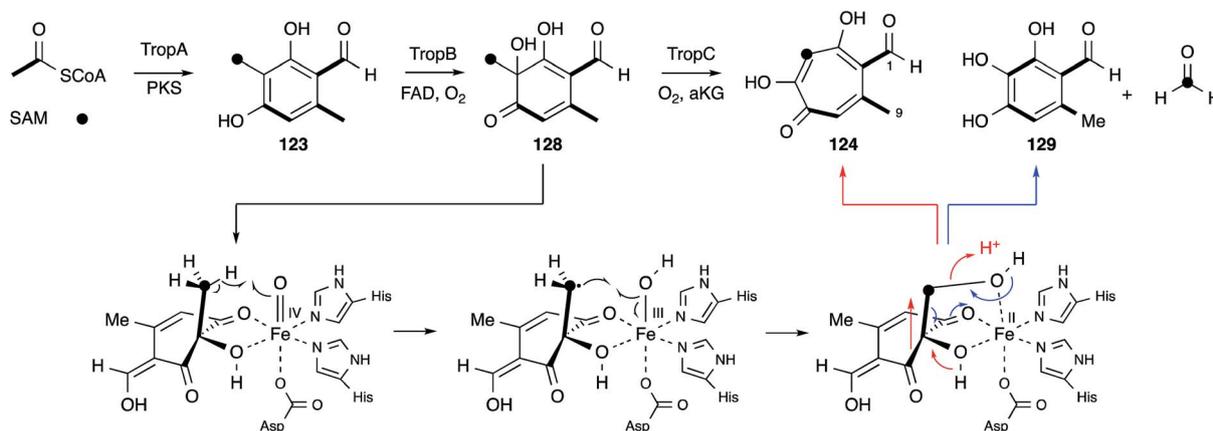
already shown is decarboxylated to 4 (Scheme 19).¹⁵⁵ Later work by Bryant and Light showed that methylorsinaldehyde 123 is incorporated much more efficiently.¹⁵⁶ Vining and coworkers were the first to use ¹³C labels and NMR in the field of tropolone biosynthesis when they showed that sepedonin 5 is derived from acetate and formate in only the second reported use of ¹³C NMR for the direct elucidation of a biosynthetic pathway.¹⁵⁷ The results clearly show that the C₁-derived carbon is inserted between carbons derived from acetate. Scott and coworkers then used ¹⁸O₂ incubations, contrasted with dual ¹⁴C-¹⁸O-labelled acetate feeding experiments, to differentiate mechanisms involving ring expansion by a migration or by a ring-opening/ring-closing sequence.¹⁵⁸ Analysis of the produced stipitonic acid 127 by mass spectrometry showed an enhancement of the M²⁺ peak, and no evidence of an M⁴⁺ peak, fully in agreement with the proposed migration mechanism.

No further original information was available for around 40 years until genomic sequence data for *Talaromyces stipitatus* became publically available. The fact that 3-methylorsinaldehyde 123 is likely to be the first enzyme free intermediate stimulated a search of the genome using the sequence of the PKS methylorsinaldehyde synthase (MOS) from *Acremonium strictum*.¹⁵⁹ This highlighted a BGC encoding a homologous PKS (TropA), an FAD-dependent oxygenase (TropB) and a non-heme iron dependent oxygenase (TropC).¹⁶⁰ *In vitro* assay of purified TropB and TropC showed that the first step is oxidative dearomatization of the polyketide to form the dienone 128 (Scheme 20). TropC then attempts to hydroxylate the adjacent methyl, but pinacol-type rearrangement gives the tropolone nucleus of 124. Competing deformylation also affords the





Scheme 19 Early labelling results obtained during the study of the biosynthesis of stipitatic acid 4.



Scheme 20 Likely molecular steps during the oxidative ring expansion to form tropolones in fungi.

trihydroxybenzene **129** at least *in vitro*. Knockout experiments of all the other genes in the BGC then illustrated how the pathway involves sequential oxidation of the C-1 and C-9 positions, followed by late anhydride hydrolysis and decarboxylation.¹⁶¹

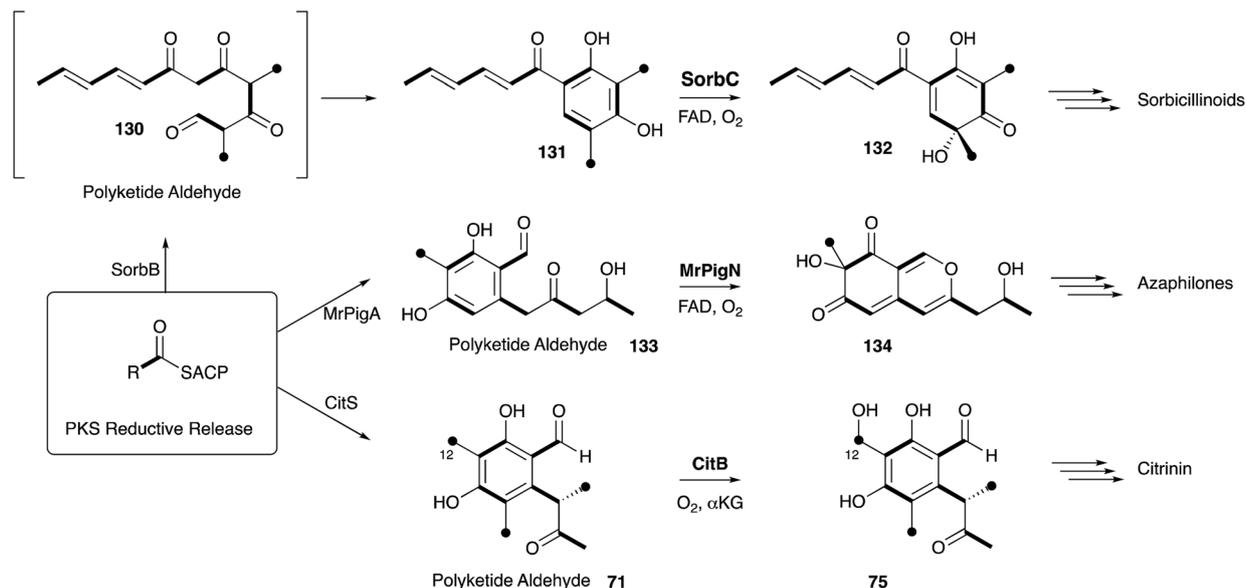
Elucidation of the early steps of the stipitatic acid **4** pathway also shed light on other related processes in fungal secondary metabolite biosynthesis. For example, TropB, responsible for the oxidative dearomatisation step, has homologues in the sorbicillinoid (SorBC)¹⁶² and azaphilone (MrPigN)⁸¹ pathways where it is also involved in the oxidative dearomatisation of polyketide aldehydes – a key step in the activation of the otherwise poorly reactive aromatic system. A TropC analogue, which in the stipitatic acid **4** pathway catalyses the ring expansion, is involved in hydroxylation of an aromatic methyl group in the citrinin pathway (Section 5).⁸⁰ Meanwhile the nrPKS for all three pathways are closely related, producing aromatic polyketide aldehydes, differing in chain-length and methylation pattern, but all with a distinctive reductive release. These chemical observations thus point strongly to the evolution and close relationships of the biosynthetic pathways leading to these different classes of polyketides. Thus the continuing study of the classic Raistrick compounds such as **4** inevitably leads to insights and understanding of many other fungal pathways. Several other classes of fungal metabolites are

known to feature tropolones in their structures, or as putative precursors or intermediates. Such compounds include Xenoulene A,¹⁶³ the epolones,¹⁶⁴ pycnidione¹⁶⁵ and the eupenifidins.¹⁶⁶ However no sequence or molecular information has yet been revealed in these cases, and while it is tempting to speculate that their tropolone moieties may be derived as discussed above, other modes of tropolone biosynthesis are known in plants and bacteria.¹⁶⁷

10. Other compounds and outstanding problems

Many of the classic Raistrick compounds have continued to yield fascinating insights into fungal secondary metabolism through the past decade, as new genomic and molecular methods have built on the foundations formed from isotopic feeding experiments. Much of the work reported in the literature has focussed on the more biologically (*e.g.* mycophenolic acid **10**, griseofulvin **9**) significant or structurally interesting (*e.g.* maleidrides **6**, tropolones **3**) metabolites where much new chemistry has been revealed. In many cases understanding the biosynthesis at the genomic and molecular levels is revealing hitherto unsuspected relationships, such as between geodin **8**





Scheme 21 Chemical relationships between the biosynthesis of sorbicillinoids, azaphilones and citrinin.

and ravenelin 7 (Section 3) or between citrinin 2, the tropolones 3, the sorbicillins and the azaphilones (Sections 3 and 9). Such relationships will no doubt help to uncover evolutionary relationships between pathways, and this understanding will also aid in the future rational redesign of individual biosynthetic steps (e.g. as exemplified in the DALs, Section 6) and new pathways in their entirety. It seems likely that the Raistrick compounds will continue to be of interest in this field because of the significant depth of knowledge accrued over the past 80–90 years.

Other compounds which might be considered as of more minor structural or biological significance, such as the fruit spoilage mycotoxin patulin 14 produced by *Penicillium* species and the mycotoxin terrein 13 from *Aspergillus* species remain to be fully investigated. For example, in the case of patulin 14 a biosynthetic gene cluster has been reported,¹⁶⁸ and some P450 enzymes have been isolated,¹⁶⁹ but no detailed pathway investigations have yet been made. The pathway should include interesting ring contracting steps. Likewise the BGC for terrein biosynthesis has also been revealed in *A. terreus*,¹⁷⁰ but no detailed pathway has yet been reported, and again fascinating oxidative ring contractions are likely to be revealed. In the case of kojic acid 1 the genes have been identified,^{171,172} and biotechnological processes are already being engineered to increase the production of this valuable metabolite through a rather straight-forward pathway from glucose.¹⁷³ Significant progress has been made in elucidating the early steps of the emodin pathway to chrysophanol, but later steps to many of the ergochromes and reduced xanthenes remain cryptic. Likewise, recent progress in understanding the molecular mechanisms of maleidride biosynthesis have revealed the new proteins involved, but not yet the chemical mechanisms and selectivity factors which must be at play, or how these systems could be extended to explain the biosynthesis of the octadrides or linear family members (Scheme 21).

Thus it seems certain that interest in the Raistrick compounds, and their use as models for understanding and exploiting fungal biosynthetic pathways will continue into the future.

11. Conflicts of interest

There are no conflicts to declare.

12. Acknowledgements

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13. References

- 1 J. H. Birkinshaw, *Biogr. Mem. Fellows R. Soc.*, 1972, **18**, 488–509.
- 2 R. Bentley, *Nat. Prod. Rep.*, 2006, **23**, 1046–1062.
- 3 B. R. Clark, R. J. Capon, E. Lacey, S. Tennant and J. H. Gill, *Org. Biomol. Chem.*, 2006, **4**, 1520.
- 4 P. W. Clutterbuck, R. Lovell and H. Raistrick, *Biochem. J.*, 1932, **26**, 1907–1918.
- 5 D. J. Cram, *J. Am. Chem. Soc.*, 1948, **70**, 440–441; J. P. Brown, N. J. Cartwright, A. Robertson and W. B. Whalley, *Nature*, 1948, **162**, 72–73.
- 6 A. J. Birch, P. Fitton, E. Pride, A. Ryan, H. Smith and W. Whalley, *J. Chem. Soc.*, 1958, 4576–4581.
- 7 J. Barber and J. Staunton, *J. Chem. Soc., Chem. Commun.*, 1979, 1098–1099.
- 8 U. Sankawa, Y. Ebizuka, H. Noguchi, Y. Isikawa, S. Kitagawa, Y. Yamamoto, T. Kobayashi and Y. Iitak, *Tetrahedron*, 1983, **39**, 3583–3591.



- 9 T. Shimizu, H. Kinoshita and T. Nihira, *Appl. Environ. Microbiol.*, 2007, **73**, 5097–5103.
- 10 M. Dewar, *Nature*, 1945, **155**, 50–51.
- 11 R. Thomas, *Biochem. J.*, 1961, **78**, 748–758.
- 12 M. Ahuja, Y.-M. Chiang, S.-L. Chang, M. B. Praseuth, R. Entwistle, J. F. Sanchez, H.-C. Lo, H.-H. Yeh, B. R. Oakley and C. C. C. Wang, *J. Am. Chem. Soc.*, 2012, **134**, 8212–8221.
- 13 D. Saha, R. Fetzner, B. Burkhardt, J. Podlech, M. Metzler, H. Dang, C. Lawrence and R. Fischer, *PLoS One*, 2012, **7**, e40564.
- 14 E. E. Stinson, W. B. Wise, R. A. Moreau, A. J. Jurewicz and P. E. Pfeffer, *Can. J. Chem.*, 1986, **64**, 1590–1594.
- 15 J. Dasenbrock and T. J. Simpson, *J. Chem. Soc., Chem. Commun.*, 1987, 1235–1242.
- 16 M. Wenderoth, C. Pinecker, B. Voß and R. Fischer, *Fungal Genet. Biol.*, 2017, **101**, 55–60.
- 17 R. A. Jacobson and R. Adams, *J. Am. Chem. Soc.*, 1924, **46**, 1312–1316.
- 18 H. Raistrick, R. Robinson and D. E. White, *Biochem. J.*, 1936, **30**, 1303–1314.
- 19 H. Raistrick and G. Smith, *Biochem. J.*, 1936, **30**, 1315–1322.
- 20 C. T. Calam, P. W. Clutterbuck, A. E. Oxford and H. Raistrick, *Biochem. J.*, 1947, **41**, 458–463.
- 21 D. H. R. Barton and A. I. Scott, *J. Chem. Soc.*, 1958, 1767–1776.
- 22 A. J. Birch, T. J. Simpson and P. W. Westerman, *Tetrahedron Lett.*, 1975, **16**, 4173–4177.
- 23 S. Gatenbeck and L. Malmström, *Acta Chem. Scand.*, 1969, **23**, 3493–3497.
- 24 R. F. Curtis, C. H. Hassall and D. R. Parry, *J. Chem. Soc., Perkin Trans. 1*, 1972, 240–245.
- 25 J. G. Hill, T. T. Nakashima and J. C. Vederas, *J. Am. Chem. Soc.*, 1982, **104**, 1745–1748.
- 26 H. Fujimoto, H. Flasch and B. Franck, *Chem. Ber.*, 1975, **108**, 1224–1228.
- 27 I. Fujii, Y. Ebizuka and U. Sankawa, *Chem. Pharm. Bull.*, 1982, **30**, 2283–2286.
- 28 I. Fujii, H. Iijima, Y. Ebizuka and U. Sankawa, *Chem. Pharm. Bull.*, 1983, **31**, 337–340.
- 29 K. X. Huang, I. Fujii, Y. Ebizuka, K. Gomi and U. Sankawa, *J. Biol. Chem.*, 1995, **270**, 21495–21502.
- 30 I. Fujii, Y. Ebizuka and U. Sankawa, *J. Biochem.*, 1988, **103**, 878–883.
- 31 Z. G. Chen, I. Fujii, Y. Ebizuka and U. Sankawa, *Phytochemistry*, 1995, **38**, 299–305.
- 32 T. P. Nicholson, B. A. Rudd, M. Dawson, C. M. Lazarus, T. J. Simpson and R. J. Cox, *Chem. Biol.*, 2000, **8**, 157–178.
- 33 R. D. Couch and G. M. Gaucher, *J. Biotechnol.*, 2004, **108**, 171–177.
- 34 M. T. Nielsen, J. B. Nielsen, D. C. Anyaogu, D. K. Holm, K. F. Nielsen, T. O. Larsen and U. H. Mortensen, *PLoS One*, 2013, **8**, e72871–10.
- 35 Y.-M. Chiang, E. Szewczyk, A. D. Davidson, R. Entwistle, N. P. Keller, C. C. C. Wang and B. R. Oakley, *Appl. Environ. Microbiol.*, 2010, **76**, 2067–2074.
- 36 J. F. Sanchez, R. Entwistle, J.-H. Hung, J. Yaegashi, S. Jain, Y.-M. Chiang, C. C. C. Wang and B. R. Oakley, *J. Am. Chem. Soc.*, 2011, **133**, 4010–4017.
- 37 L. Neubauer, J. Dopstadt, H.-U. Humpf and P. Tudzynski, *Fungal Biol. Biotechnol.*, 2016, **3**(2), 1–14.
- 38 M. A. Schätzle, S. M. Husain, S. Ferlaino and M. Müller, *J. Am. Chem. Soc.*, 2012, **134**, 14742–14745.
- 39 B. H. Howard and H. Raistrick, *Biochem. J.*, 1950, **46**, 49–53.
- 40 T. J. Simpson, *ChemBioChem*, 2012, **13**, 1680–1688.
- 41 S. Griffiths, C. H. Mesarich, B. Saccomanno, A. Vaisberg, P. J. G. M. de Wit, R. J. Cox and J. Collemare, *Proc. Natl. Acad. Sci. U.S.A.*, 2016, **113**, 6851–6856.
- 42 B. H. Howard and H. Raistrick, *Biochem. J.*, 1949, **44**, 227–233.
- 43 N. Wijkman, *Justus Liebigs Ann. Chem.*, 1931, **485**, 61–73.
- 44 H. Raistrick and G. Smith, *Biochem. J.*, 1933, **27**, 1814–1819.
- 45 J. E. Baldwin, D. H. R. Barton, J. L. Bloomer, L. M. Jackman, L. Rodriguez-Hahn and J. K. Sutherland, *Experientia*, 1962, **18**, 345.
- 46 T. Hamor, I. Paul, J. Monteath Robertson and G. Sim, *Experientia*, 1962, **18**, 352–354.
- 47 D. H. R. Barton and J. K. Sutherland, *J. Chem. Soc.*, 1965, 1769–1774.
- 48 R. I. Crane, P. Hedden, J. MacMillan and W. B. Turner, *J. Chem. Soc., Perkin Trans. 1*, 1973, 194–197.
- 49 G. Büchi, K. M. Snader, J. D. White, J. Z. Gougoutas and S. Singh, *J. Am. Chem. Soc.*, 1970, **92**, 6638–6641.
- 50 G. A. Sulikowski, F. Agnelli, P. Spencer, J. M. Koomen and D. H. Russell, *Org. Lett.*, 2002, **4**, 1447–1450.
- 51 M. Nakajima, K. Itoi, Y. Takamatsu, S. Sato, Y. Furukawa, K. Furuya, T. Honma, J. Kadotani, M. Kozasa and T. Haneishi, *J. Antibiot.*, 1991, **44**, 1065–1072.
- 52 M. Futagawa, D. E. Wedge and F. E. Dayan, *Pestic. Biochem. Physiol.*, 2002, **73**, 87–93.
- 53 M. Saleem, H. Hussain, I. Ahmed, S. Draeger, B. Schulz, K. Meier, M. Steinert, G. Pescitelli, T. Kurtán, U. Flörke and K. Krohn, *Eur. J. Org. Chem.*, 2010, **2011**, 808–812.
- 54 A. J. Szwalbe, K. Williams, D. E. O'Flynn, A. M. Bailey, N. P. Mulholland, J. L. Vincent, C. L. Willis, R. J. Cox and T. J. Simpson, *Chem. Commun.*, 2015, **51**, 17088–17091.
- 55 J. L. Bloomer, C. E. Moppett and J. K. Sutherland, *J. Chem. Soc. C*, 1968, 588–591.
- 56 R. K. Huff, C. E. Moppett and J. K. Sutherland, *J. Chem. Soc., Perkin Trans. 1*, 1972, 2584.
- 57 J. E. Baldwin, A. Beyeler, C. Keats, G. J. Pritchard, R. M. Adlington and D. J. Watkin, *Tetrahedron*, 1999, **55**, 7363–7374.
- 58 N. P. Mulholland, J. L. Vincent, A. M. Bailey, C. L. Willis and R. J. Cox, *Angew. Chem., Int. Ed.*, 2016, **55**, 6784–6788.
- 59 R. Fujii, A. Minami, S. Nagamine, I. Takeuchi and H. Oikawa, *Org. Lett.*, 2015, **17**, 5658–5661.
- 60 K. Williams, A. J. Szwalbe, C. Dickson, T. R. Desson, N. P. Mulholland, J. L. Vincent, J. M. Clough, A. M. Bailey, C. P. Butts, C. L. Willis, T. J. Simpson and R. J. Cox, *Chem. Commun.*, 2017, **53**, 7965–7968.
- 61 J. Bai, D. Yan, T. Zhang, Y. Guo, Y. Liu, Y. Zou, M. Tang, B. Liu, Q. Wu, S. Yu, Y. Tang and Y. Hu, *Angew. Chem.*, 2017, **129**, 4860–4864.
- 62 B. Bonsch, V. Belt, C. Bartel, N. Duensing, M. Koziol, C. M. Lazarus, A. M. Bailey, T. J. Simpson and R. J. Cox, *Chem. Commun.*, 2016, **52**, 6777–6780.



- 63 M. Isaka, M. Tanticharoen and Y. Thebtaranonth, *Tetrahedron Lett.*, 2000, **41**, 1657–1660.
- 64 A. C. Hetherington and H. Raistrick, *Philos. Trans. R. Soc., B*, 1931, **220**, 269–295.
- 65 F. P. Coyne, H. Raistrick and R. Robinson, *Philos. Trans. R. Soc., B*, 1931, **220**, 297–300.
- 66 T. S. Gore, T. B. Panse and K. Venkataraman, *Nature*, 1946, **157**, 333–333.
- 67 D. J. Cram, *J. Am. Chem. Soc.*, 1948, **70**, 440–441.
- 68 J. P. Brown, N. J. Cartwright, A. Robertson and W. B. Whalley, *Nature*, 1948, **162**, 72–73.
- 69 E. Schwenk, G. J. Alexander, A. M. Gold and D. F. Stevens, *J. Biol. Chem.*, 1958, **233**, 1211–1213.
- 70 A. J. Birch, P. Fitton, E. Pride, A. J. Ryan, H. Smith and W. B. Whalley, *J. Chem. Soc.*, 1958, 4576–6.
- 71 R. B. Woodward, *Angew. Chem.*, 1956, **68**, 13–20.
- 72 R. Carter, M. Garson and J. Staunton, *J. Chem. Soc., Chem. Commun.*, 1979, 1097–1098.
- 73 J. Barber, R. H. Carter, M. J. Garson and J. Staunton, *J. Chem. Soc., Perkin Trans. 1*, 1981, 2577–2583; L. Colombo, C. Gennari, D. Potenza, C. Scolastico, F. Aragozzini and C. Merendi, *J. Chem. Soc., Perkin Trans. 1*, 1981, 2594–2597.
- 74 J. Barber and J. Staunton, *J. Chem. Soc., Chem. Commun.*, 1979, 1098–1099; J. Barber and J. Staunton, *J. Chem. Soc., Perkin Trans. 1*, 1980, 2244–2248.
- 75 U. Sankawa, Y. Ebizuka, H. Noguchi, Y. Isikawa, S. Kitagawa, Y. Yamamoto, T. Kobayashi and Y. Iitak, *Tetrahedron*, 1983, **39**, 3583–3591.
- 76 H. Hajjaj, A. Klabe, M. Loret, G. Goma, P. Blanc and J. Francois, *Appl. Environ. Microbiol.*, 1999, **65**, 311–314.
- 77 P. C. Y. Woo, C.-W. Lam, E. W. T. Tam, K.-C. Lee, K. K. Y. Yung, C. K. F. Leung, K.-H. Sze, S. K. P. Lau and K.-Y. Yuen, *Sci. Rep.*, 2014, **4**, 6728–8.
- 78 L. E. Bingle, T. J. Simpson and C. M. Lazarus, *Fungal Genet. Biol.*, 1999, **26**, 209–223; T. P. Nicholson, B. A. Rudd, M. Dawson, C. M. Lazarus, T. J. Simpson and R. J. Cox, *Chem. Biol.*, 2000, **8**, 157–178.
- 79 T. Shimizu, H. Kinoshita and T. Nihira, *Appl. Environ. Microbiol.*, 2007, **73**, 5097–5103.
- 80 K. Sakai, H. Kinoshita, T. Shimizu and T. Nihira, *J. Biosci. Bioeng.*, 2008, **106**, 466–472.
- 81 Y. He and R. J. Cox, *Chem. Sci.*, 2016, **7**, 2119–2127.
- 82 P. A. Storm and C. A. Townsend, *Chem. Commun.*, 2017, **54**, 50–53.
- 83 W. Chen, R. Chen, Q. Liu, Y. He, K. He, X. Ding, L. Kang, X. Guo, N. Xie, Y. Zhou, Y. Lu, R. J. Cox, I. Molnár, M. Li, Y. Shao and F. Chen, *Chem. Sci.*, 2017, **8**, 4917–4925.
- 84 P. V. Divekar, H. Raistrick, T. A. Dobson and L. C. Vining, *Can. J. Chem.*, 1965, **43**, 1835–1848.
- 85 P. A. Storm, D. A. Herbst, T. Maier and C. A. Townsend, *Cell Chem. Biol.*, 2017, **24**, 1–11.
- 86 R. A. Cacho, J. Thuss, W. Xu, R. Sanichar, Z. Gao, A. Nguyen, J. C. Vederas and Y. Tang, *J. Am. Chem. Soc.*, 2015, **137**, 15688–15691.
- 87 D. M. Roberts, C. Bartel, A. Scott, D. Ivison, T. J. Simpson and R. J. Cox, *Chem. Sci.*, 2017, **8**, 1116–1126.
- 88 E. Liddle, A. Scott, L.-C. Han, D. Ivison, T. J. Simpson, C. L. Willis and R. J. Cox, *Chem. Commun.*, 2017, **53**, 1727–1730.
- 89 H. Raistrick and F. A. H. Rice, *J. Chem. Soc. C*, 1971, 3069–3072.
- 90 O. C. Musgrave, *J. Chem. Soc.*, 1956, 4301–4305.
- 91 A. J. Birch, O. C. Musgrave, R. W. Rickards and H. Smith, *J. Chem. Soc.*, 1959, 3146–3147.
- 92 H. D. Munro, O. C. Musgrave and R. Templeton, *J. Chem. Soc. C*, 1967, 947–948.
- 93 K. Arai, B. Rawlings, Y. Yoshizawa and J. Vederas, *J. Am. Chem. Soc.*, 1989, **111**, 3391–3399.
- 94 Y. Yoshizawa, Z. Li, P. Reese and J. Vederas, *J. Am. Chem. Soc.*, 1990, **112**, 3212–3213.
- 95 Y. Xu, P. Espinosa-Artiles, V. Schubert, Y. M. Xu, W. Zhang, M. Lin, A. A. L. Gunatilaka, R. Sussmuth and I. Molnar, *Appl. Environ. Microbiol.*, 2013, **79**, 2038–2047.
- 96 Y. Xu, T. Zhou, Z. Zhou, S. Su, S. A. Roberts, W. R. Montfort, J. Zeng, M. Chen, W. Zhang, M. Lin, J. Zhan and I. Molnár, *Proc. Natl. Acad. Sci. U.S.A.*, 2013, **110**, 5398–5403.
- 97 Y. Xu, T. Zhou, S. Zhang, L.-J. Xuan, J. Zhan and I. Molnár, *J. Am. Chem. Soc.*, 2013, **135**, 10783–10791.
- 98 Y. Xu, T. Zhou, S. Zhang, P. Espinosa-Artiles, L. Wang, W. Zhang, M. Lin, A. A. L. Gunatilaka, J. Zhan and I. Molnár, *Proc. Natl. Acad. Sci. U.S.A.*, 2014, **111**, 12354–12359.
- 99 J. Bai, Y. Lu, Y.-M. Xu, W. Zhang, M. Chen, M. Lin, A. A. L. Gunatilaka, Y. Xu and I. Molnár, *Org. Lett.*, 2016, **18**, 1262–1265.
- 100 K. A. K. Pahirulzaman, K. Williams and C. M. Lazarus, *Meth. Enzymol.*, 2012, **517**, 241–260.
- 101 A. E. Oxford, H. Raistrick and P. Simonart, *Biochem. J.*, 1939, **33**, 240–248.
- 102 J. MacMillan, *Annu. Rev. Plant Physiol.*, 1996, **47**, 1–21.
- 103 J. F. Grove, J. MacMillan, T. P. C. Mulholland and M. A. T. Rogers, *J. Chem. Soc.*, 1952, 3977–3987.
- 104 J. MacMillan, *J. Chem. Soc.*, 1954, 2585–2587.
- 105 *Glaxo: A History to 1962*, ed. R. P. T. Davenport-Hines and J. Slinn, Cambridge University Press, 1992.
- 106 A. J. Birch, R. A. Massy-Westropp, R. W. Rickards and H. Smith, *J. Chem. Soc.*, 1958, 360–365.
- 107 A. Rhodes, B. Boothroyd, P. McGonagle and G. A. Somerfield, *Biochem. J.*, 1961, **81**, 28–37.
- 108 A. Rhodes, G. A. Somerfield and M. P. McGonagle, *Biochem. J.*, 1963, **88**, 349–357.
- 109 T. J. Simpson and J. Holker, *Phytochemistry*, 1977, **16**, 229–233.
- 110 Y. Sato, T. Oda and H. Saitô, *J. Chem. Soc., Chem. Commun.*, 1978, 135–136.
- 111 Y.-H. Chooi, R. Cacho and Y. Tang, *Chem. Biol.*, 2010, **17**, 483–494.
- 112 R. A. Cacho, Y.-H. Chooi, H. Zhou and Y. Tang, *ACS Chem. Biol.*, 2013, **8**, 2322–2330.
- 113 J. M. Grandner, R. A. Cacho, Y. Tang and K. N. Houk, *ACS Catal.*, 2016, **6**, 4506–4511.
- 114 B. Gosio, *G. Accad. Med. Torino*, 1893, **61**, 484.
- 115 R. Bentley, *Chem. Rev.*, 2000, **100**, 3801–3825.



- 116 R. Nicoletti, M. De Stefano, S. De Stefano, A. Trincone and F. Marziano, *Mycopathologia*, 2004, **158**, 465.
- 117 M. S. Diamond, M. Zachariah and E. Harris, *Virology*, 2002, **304**, 211.
- 118 K. Borroto-Esoda, F. Myrick, J. Feng, J. Jeffrey and P. Furman, *Antimicrob. Agents Chemother.*, 2004, **48**, 4387.
- 119 F. Kavanagh, *J. Bacteriol.*, 1947, **54**, 761.
- 120 R. D. Torrenegra, J. E. Baquero and J. S. Calderon, *Rev. Latinoam. Quim.*, 2005, **33**, 76.
- 121 W. Epinette, C. Parker, E. Jones and M. Greist, *J. Am. Acad. Dermatol.*, 1987, **17**, 962.
- 122 J. C. Cline, J. D. Nelson, K. Gerzon, R. H. Williams and D. C. DeLong, *Appl. Microbiol.*, 1969, **18**, 14.
- 123 C. M. Smith, L. J. Fontenelle, H. Muzik, H. Unger, L. W. Brox and J. F. Henderson, *Biochem. Pharmacol.*, 1974, **23**, 2727.
- 124 C. L. Alsberg and O. F. Black, U.S. Department of Agriculture, Bureau of Plant Industry, Bulletin No. 270, 1913.
- 125 W. P. Clutterbuck, A. E. Oxford and H. Raistrick, *Biochem. J.*, 1932, **26**, 1441–1458.
- 126 A. E. Oxford and H. Raistrick, *Biochem. J.*, 1933, **27**, 634–653.
- 127 H. Birkinshaw, H. Raistrick and D. J. Ross, *Biochem. J.*, 1952, **50**, 630.
- 128 J. Birkinshaw, A. Bracken, E. N. Morgan and H. Raistrick, *Biochem. J.*, 1948, **43**, 216.
- 129 W. P. Clutterbuck and H. Raistrick, *Biochem. J.*, 1933, **27**, 654.
- 130 A. J. Birch, R. J. English, R. A. Massy-Westropp, M. Slaytor and H. Smith, *Proc. Chem. Soc., London*, 1957, 204–204.
- 131 A. J. Birch, R. J. English, R. A. Massy-Westropp and H. Smith, *Proc. Chem. Soc., London*, 1957, 233–234.
- 132 L. Canonica, W. Kroszczyński, B. M. Ranzi, B. Rindone, E. Santaniello and C. Scolastico, *J. Chem. Soc., Perkin Trans. 1*, 1972, 2639.
- 133 L. Colombo, C. Gennari, D. Potenza and C. Scolastico, *J. Chem. Soc., Perkin Trans. 1*, 1980, 365.
- 134 C. P. Nulton and I. M. Campbell, *Can. J. Microbiol.*, 1978, **24**, 199–201.
- 135 L. Colombo, C. Gennari, D. Potenza and C. Scolastico, *J. Chem. Soc., Chem. Commun.*, 1979, 1021–1022.
- 136 D. L. Doerfler, L. A. Ernst and I. M. Campbell, *J. Chem. Soc., Chem. Commun.*, 1980, 329–330.
- 137 L. Colombo, C. Gennari and C. Scolastico, *J. Chem. Soc., Chem. Commun.*, 1978, 434–434.
- 138 T. B. Regueira, K. R. Kildegaard, B. G. Hansen, U. H. Mortensen, C. Hertweck and J. Nielsen, *Appl. Environ. Microbiol.*, 2011, **77**, 3035.
- 139 W. Zhang, S. Cao, L. Qiu, F. Qi, Z. Li, Y. Yang, S. Huang, F. Bai, C. Liu, X. Wan and S. Li, *ChemBioChem*, 2015, **16**, 565.
- 140 A. Del-Cid, C. Gil-Durán, I. Vaca, J. F. Rojas-Aedo, R. O. García-Rico, G. Levicán and R. Chávez, *PLoS One*, 2016, **11**, e0147047.
- 141 B. G. Hansen, B. Salomonsen, M. T. Nielsen, J. B. Nielsen, N. B. Hansen, K. F. Nielsen, T. B. Regueira, J. Nielsen, K. R. Patil and U. H. Mortensen, *Appl. Environ. Microbiol.*, 2011, **77**, 3044–3051.
- 142 B. G. Hansen, E. Mních, K. F. Nielsen, J. B. Nielsen, M. T. Nielsen, U. H. Mortensen, T. O. Larsen and K. R. Patil, *Appl. Environ. Microbiol.*, 2012, **78**, 4908–4913.
- 143 Y. Matsuda and I. Abe, *Nat. Prod. Rep.*, 2015, **33**, 26–53.
- 144 D. K. Holm, L. M. Petersen, A. Klitgaard, P. B. Knudsen, Z. D. Jarczyńska, K. F. Nielsen, C. H. Gotfredsen and T. O. Larsen, *Chem. Biol.*, 2014, **21**, 519–529.
- 145 W. L. Muth and C. H. Nash, *Antimicrob. Agents Chemother.*, 1975, **8**, 321.
- 146 J. H. Birkinshaw and H. Raistrick, *Biochem. J.*, 1932, **26**, 441–453.
- 147 M. Iwatsuki, S. Takada, M. Mori, A. Ishiyama, M. Namatame, A. Nishihara-Tsukashima, K. Nonaka, R. Masuma, K. Otoguro, K. Shiomi and S. Omura, *J. Antibiot.*, 2011, **64**, 183–188.
- 148 M. Dewar, *Nature*, 1945, **155**(3927), 141–142.
- 149 R. Corbett, A. Johnson and A. Todd, *J. Chem. Soc.*, 1950, 147–149.
- 150 R. Robinson, *Chem. Ind.*, 1951, 12.
- 151 R. Bentley and C. Thiessen, *J. Biol. Chem.*, 1963, **238**, 1880–1888.
- 152 R. Bentley, *J. Biol. Chem.*, 1963, **238**, 1889–1894.
- 153 R. Bentley, *J. Biol. Chem.*, 1963, **238**, 1895–1902.
- 154 A. I. Scott, H. Guilford and E. Lee, *J. Am. Chem. Soc.*, 1971, **93**, 3534–3536.
- 155 R. Bentley and C. P. Thiessen, *J. Biol. Chem.*, 1963, **238**, 3811–3816.
- 156 R. Bryant and R. Light, *Biochemistry*, 1974, **13**, 1516–1522.
- 157 A. McInnes, D. Smith and L. Vining, *J. Chem. Soc., Chem. Commun.*, 1971, 325–326.
- 158 A. I. Scott and K. J. Wiesner, *J. Chem. Soc., Chem. Commun.*, 1972, 1075–1077.
- 159 A. M. Bailey, R. J. Cox, K. Harley, C. M. Lazarus, T. J. Simpson and E. Skellam, *Chem. Commun.*, 2007, 4053–4055.
- 160 J. Davison, A. Al Fahad, M. Cai, Z. Song, S. Y. Yehia, C. M. Lazarus, A. M. Bailey, T. J. Simpson and R. J. Cox, *Proc. Natl. Acad. Sci. U.S.A.*, 2012, **109**, 7642–7647.
- 161 A. Al Fahad, A. Abood, T. J. Simpson and R. J. Cox, *Angew. Chem., Int. Ed.*, 2014, **53**, 7519–7523.
- 162 A. A. Fahad, A. Abood, K. M. Fisch, A. Osipow, J. Davison, M. Avramović, C. P. Butts, J. Piel, T. J. Simpson and R. J. Cox, *Chem. Sci.*, 2014, **5**, 523–527.
- 163 M. E. Raggatt, T. J. Simpson and M. I. ChicarelliRobinson, *Chem. Commun.*, 1997, 2245–2246.
- 164 P. Cai, D. Smith, B. Cunningham, S. Brown-Shimer, B. Katz, C. Pearce, D. Venables and D. Houck, *J. Nat. Prod.*, 1998, **61**, 791–795.
- 165 G. H. Harris, K. Hoogsteen, K. C. Silverman, S. L. Raghoobar, G. F. Bills, R. B. Lingham, J. L. Smith, H. W. Dougherty, C. Cascales and F. Peláez, *Tetrahedron*, 1993, **49**, 2139–2144.
- 166 F. Mayerl, Q. Gao, S. Huang, S. Klohr, J. Matson, D. Gustavson, D. Pirnik, R. Berry, C. Fairchild and W. Rose, *J. Antibiot.*, 1993, **46**, 1082–1088.



- 167 R. J. Cox and A. Al Fahad, *Curr. Opin. Chem. Biol.*, 2013, **17**, 532–536.
- 168 A.-R. Ballester, M. Marcet-Houben, E. Levin, N. Sela, C. Selma-Lázaro, L. Carmona, M. Wisniewski, S. Droby, L. González-Candelas and T. Gabaldón, *Mol. Plant-Microbe Interact.*, 2015, **28**, 232–248.
- 169 M. P. Artigot, N. Loiseau, J. Laffitte, L. Mas-Reguieg, S. Tadrist, I. P. Oswald and O. Puel, *Microbiology*, 2009, **155**, 1738–1747.
- 170 C. Zaehle, M. Gressler, E. Shelest, E. Geib, C. Hertweck and M. Brock, *Chem. Biol.*, 2014, **21**, 719–731.
- 171 Y. Terabayashi, M. Sano, N. Yamane, J. Marui, K. Tamano, J. Sagara, M. Dohmoto, K. Oda, E. Ohshima, K. Tachibana, Y. Higa, S. Ohashi, H. Koike and M. Machida, *Fungal Genet. Biol.*, 2010, **47**, 953–961.
- 172 Y. Terabayashi, M. Sano, N. Yamane, J. Marui, K. Tamano, J. Sagara, M. Dohmoto, K. Oda, E. Ohshima, K. Tachibana, Y. Higa, S. Ohashi, H. Koike and M. Machida, *Fungal Genet. Biol.*, 2010, **47**, 953–961.
- 173 R. Yamada, T. Yoshie, S. Wakai, N. Asai-Nakashima, F. Okazaki, C. Ogino, H. Hisada, H. Tsutsumi, Y. Hata and A. Kondo, *Microb. Cell Fact.*, 2014, **13**, 71–78.

