



Cite this: *Med. Chem. Commun.*,
2018, 9, 1311

Ammonium [¹¹C]thiocyanate: revised preparation and reactivity studies of a versatile nucleophile for carbon-11 radiolabelling†

Tom Haywood,^a Sara Cesarec,^{id}^a Steven Kealey,^a
Christophe Plisson^b and Philip W. Miller^{id}^{*a}

Received 18th August 2017,
Accepted 11th May 2018

DOI: 10.1039/c7md00425g

rsc.li/medchemcomm

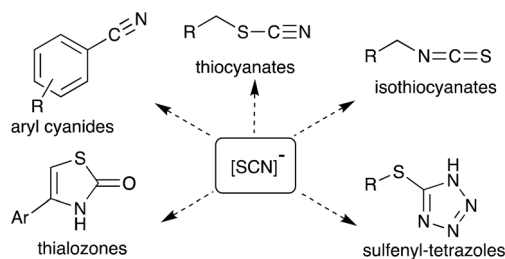
Herein we report the preparation of ammonium [¹¹C]thiocyanate *via* the reaction of [¹¹C]CS₂ with ammonia. The [¹¹C]SCN⁻ ion is demonstrated as a potent nucleophile that can be used to readily generate a range of ¹¹C-labelled thiocyanate molecules in high conversions. Furthermore, novel ¹¹C-labelled thiazolone molecules can be easily prepared from the intermediate α -thiocyanatophenones *via* an acid mediated cyclisation reaction.

Positron emission tomography (PET) is now a widely used clinical and research imaging technique for the study and diagnosis of a range of neurological, oncological and cardiac conditions.^{1–5} The expansion of PET imaging centres worldwide over the past decade has led to an accompanying increase in demand for both new and existing PET tracers. The development of PET tracers presents a number of key chemical and biological challenges due to the short half-lives of positron-emitting nuclides, the peculiarity of tracer level chemical reactions, the complexity of biological interactions, and the strict working environment required for safe handling of radioactive substances. The synthetic chemistry hurdles are particularly apparent considering that the half-lives of the two most common PET nuclides, carbon-11 and fluorine-18, are only 20.4 min and 109.7 minutes, respectively. For example, a clinical radiopharmaceutical carbon-11 tracer production involving radiosynthesis, purification and formulation, typically needs to be complete within a 30–45 min timeframe. The development of new tracers is further confounded by the limited number of available PET precursors and methods for labelling, meaning that there are only a certain number of reactions that can be used to introduce the radionuclide.⁶ This limitation clearly restricts access to what can ultimately be radiolabelled and where on the target molecule the radionuclide can be introduced.

Despite these challenges, an array of novel fluorine-18 and carbon-11 labelling methods have been reported in recent years to try overcome these challenges.^{7–12} Carbon-11 is typi-

cally incorporated into tracers *via* well-established methylation protocols using [¹¹C]CH₃I or the more reactive [¹¹C]CH₃SO₃CF₃.^{6,13} Carbon-11 methylation is, however, unsuitable for labelling many interesting biological molecules, either due to the absence of a methyl group on the target molecule or due to unfavourable metabolism of the labelled methyl group *in vivo*. A range of other reactive small molecule carbon-11 reagents have therefore been developed to label in different positions. For example [¹¹C]CO₂,¹⁴ [¹¹C]CO,¹⁵ [¹¹C]HCN,¹¹ [¹¹C]CH₂O¹⁶ and [¹¹C]COCl₂¹⁷ have been used to radiolabel carbonyl and cyano functional groups. Recently, our group reported the production of a novel carbon-11 labelling reagent, [¹¹C]carbon disulfide, and described its initial reactivity studies for the efficient labelling of organosulfur compounds such as dithiocarbamates, thioureas and other biological molecules.^{18–20} Herein, we report the facile production of ammonium [¹¹C]thiocyanate, prepared *via* the reaction of [¹¹C]CS₂ with ammonia, and describe preliminary radiolabelling reactions of this versatile nucleophilic reagent.

The thiocyanate ion is a ‘pseudohalide’ and potent nucleophile that can react either *via* the nitrogen atom or



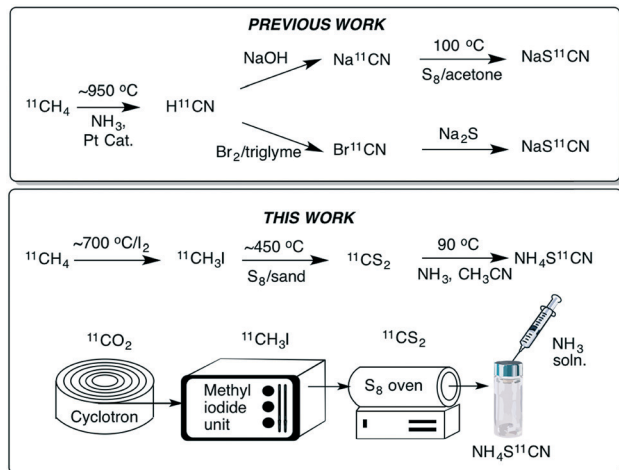
Scheme 1 A selection of organic molecules that can be prepared using the thiocyanate ion.

^a Department of Chemistry, Imperial College London, South Kensington, SW7 2AZ, London, UK. E-mail: philip.miller@imperial.ac.uk; Tel: +44 (0)2875942847

^b Imanova Limited, Imperial College London, Hammersmith Hospital, Burlington Danes Building, Du Cane Road, London, W12 0NN, UK

† Electronic supplementary information (ESI) available: Experimental details and HPLC traces. See DOI: 10.1039/c7md00425g

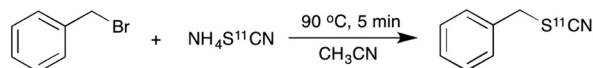




Scheme 2 Previously reported routes to $[^{11}\text{C}]\text{SCN}^-$ relying on generating $[^{11}\text{C}]\text{HCN}$ first (top); this work, which reports the synthesis of $[^{11}\text{C}]\text{SCN}^-$ via $[^{11}\text{C}]\text{CS}_2$ (bottom).

sulfur atom. It has been widely used to prepare organic thiocyanates and a range of other sulfur containing compounds (Scheme 1).²¹ The thiocyanate ion is attractive from a radiolabelling perspective because of its reactivity and potential to form a wide range of organosulfur derivatives. $[^{11}\text{C}]\text{Thiocyanate}$ has been previously reported, however, its synthesis relies on the production of $[^{11}\text{C}]\text{HCN}$, which is not readily accessible in most PET centres, thus limiting its potential wider use. Typically, $[^{11}\text{C}]\text{SCN}^-$ is prepared by first producing $[^{11}\text{C}]\text{HCN}$, followed by conversion to a salt and then reaction with a sulfur source. The first reported method describes a two-step process where $[^{11}\text{C}]\text{HCN}$ is passed through a phosphorous pentoxide trap into a solution of sodium or potassium hydroxide to generate $[^{11}\text{C}]\text{CN}^-$, which is then reacted with elemental sulfur to give $[^{11}\text{C}]\text{SCN}^-$ (Scheme 2, top).^{22,23} In an alternative approach, $[^{11}\text{C}]\text{HCN}$ can be converted to $[^{11}\text{C}]\text{NaSCN}$ via bromination in triglyme to give $[^{11}\text{C}]\text{BrCN}$ followed by distillation over antimony and reaction with sodium sulfide (Scheme 2, top).²⁴ In an effort to prepare a range of sulfur based labelled compounds we sought to explore alternative methods to synthesise $[^{11}\text{C}]\text{SCN}^-$ that circumvent the need for specialised $[^{11}\text{C}]\text{HCN}$ synthesis equipment (Scheme 2).

Our recent work on the development of $[^{11}\text{C}]\text{CS}_2$ led us to investigate this reagent as an alternative approach to prepare



Scheme 3 Substitution reaction of benzyl bromide with $[^{11}\text{C}]\text{NH}_4\text{SCN}$ to generate benzyl $[^{11}\text{C}]\text{thiocyanate}$.

$[^{11}\text{C}]\text{SCN}^-$. Carbon disulfide is known to react with ammonia to give ammonium dithiocarbamate. On heating ammonium dithiocarbamate to 90 °C, ammonium thiocyanate is generated along with one equivalent of H_2S . We previously reported a fast and easily accessible route to $[^{11}\text{C}]\text{CS}_2$ via the reaction of sulfur vapour with $[^{11}\text{C}]\text{CH}_3\text{I}$. Unlike $[^{11}\text{C}]\text{HCN}$, $[^{11}\text{C}]\text{CH}_3\text{I}$ is a much more commonly used carbon-11 labelling reagent that is prevalent in many PET centres. In our initial experiments to prepare $[^{11}\text{C}]\text{SCN}^-$, a stream of gaseous $[^{11}\text{C}]\text{CS}_2$ was passed into an ammonia solution (0.1 M in MeCN) and heated to 90 °C for 5 min. Analysis of the crude reaction mixture confirmed $[^{11}\text{C}]\text{NH}_4\text{SCN}$ formation, albeit in low radiochemical yields (RCY) (<10%, Table 1). We hypothesised that passing the $[^{11}\text{C}]\text{CS}_2$ gas stream directly through the ammonia solution resulted in significant loss of the volatile ammonia, hence greatly reducing its concentration and therefore limiting the reaction. To circumvent this, the order of addition was changed to avoid ammonia loss; first trapping the $[^{11}\text{C}]\text{CS}_2$ in acetonitrile (1 mL), then adding the ammonia solution (50 μL , 2 M NH_3 in CH_3OH) and heating to 90 °C for 5 min. Pleasingly, this resulted in near quantitative radiochemical conversion of $[^{11}\text{C}]\text{CS}_2$ to $[^{11}\text{C}]\text{NH}_4\text{SCN}$ as observed by radio-HPLC (Table 1).

Given its ease of synthesis and potential utility as a nucleophile, we aimed to explore the reactivity of the $[^{11}\text{C}]\text{thiocyanate}$ ion for substitution reactions. A simple model reaction of $[^{11}\text{C}]\text{NH}_4\text{SCN}$ with benzyl bromide was initially investigated. In a typical labelling experiment, a solution of benzyl bromide was injected into the reaction vial containing *in situ* generated $[^{11}\text{C}]\text{NH}_4\text{SCN}$, and the mixture heated for 5 min at 90 °C (Scheme 3).

The crude product mixture was analysed by radio-HPLC and the product identity confirmed by comparison of the radioactive sample with unlabelled reference material. $[^{11}\text{C}]\text{NH}_4\text{SCN}$ was found to react cleanly and quickly with benzyl bromide showing virtually quantitative conversion to benzyl $[^{11}\text{C}]\text{thiocyanate}$ (Chart 1). Encouraged by this result, we proceeded to explore this reaction with a range of α -ketobromides to generate their respective ^{11}C -labelled

Table 1 Conversion of $[^{11}\text{C}]\text{CS}_2$ to $[^{11}\text{C}]\text{NH}_4\text{SCN}$ via reaction with ammonia solution

$^{11}\text{CS}_2 + 2\text{NH}_3 \rightarrow \text{NH}_4\text{S}^{11}\text{CN} + \text{H}_2\text{S}$		
Synthesis method ^a	RCY ^b $[^{11}\text{C}]\text{CS}_2$ (%)	RCY ^b $[^{11}\text{C}]\text{NH}_4\text{SCN}$ (%)
A	90	10
B	1	99

^a Method A: $[^{11}\text{C}]\text{CS}_2$ gas stream bubbled through ammonia solution (50 μL , 2 M NH_3 in CH_3OH , acetonitrile 1 mL) followed by heating to 90 °C for 5 min. Method B: addition of ammonia solution (50 μL , 2 M NH_3 in CH_3OH) to $[^{11}\text{C}]\text{CS}_2$ in acetonitrile (1 mL) followed by heating to 90 °C for 5 min. ^b Non-isolated radiochemical yield (RCY) determined by analytical radio HPLC of the crude product based on conversion from $[^{11}\text{C}]\text{CH}_3\text{I}$. Average of minimum $n = 3$.



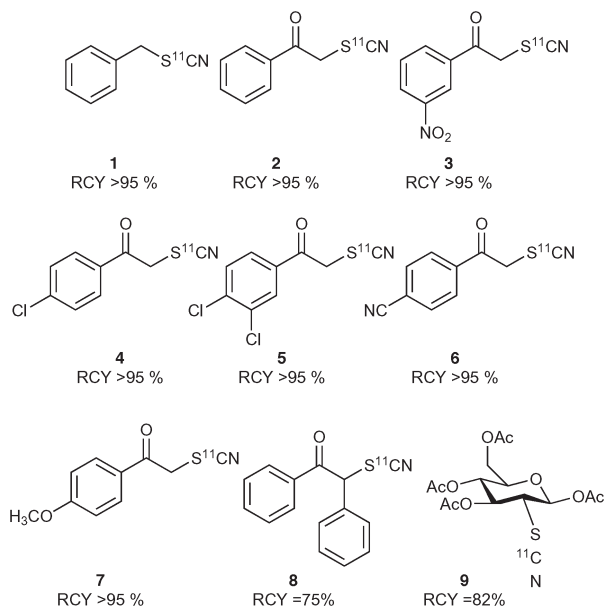
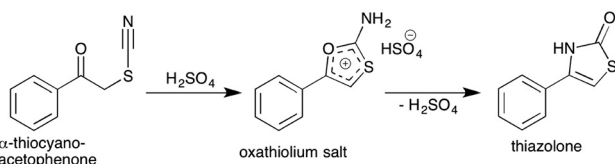
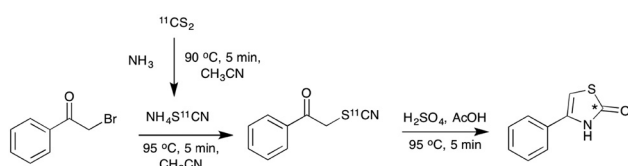


Chart 1 Radiolabelled thiocyanate molecules. Average of minimum $n = 2$.



Scheme 4 Acid mediated cyclisation of α -thiocyanatophenone which is hypothesised to proceed via the intermediate oxathiolium salt prior to thiazolone formation and elimination of sulfuric acid.



Scheme 5 One pot radiolabelling of 4-phenylthiazol-2-[^{11}C]one (10) via acid mediated cyclisation of the intermediate 1-phenyl-2-[^{11}C]thiocyanatoethanone (2), (*) denotes the ^{11}C labelling position.

α -thiocyanatophenones. We discovered that excellent conversions could be achieved for the C-11 thiocyanation of all seven α -ketobromides we tested, within a 5 min reaction time (Chart 1). Lower conversions were observed for the reaction of the diphenyl substrate (Chart 1, entry 8), most likely due to the greater steric hindrance of this substrate inhibiting nucleophilic attack of the thiocyanate ion. [^{11}C] NH_4SCN was also found to react cleanly with mannose triflate, the precursor used for 2-([^{18}F]fluoro)-2-deoxy-D-glucose synthesis, to give the labelled sugar thiocyanate derivative (9).

Thiocyanate compounds are interesting intermediates because they can be converted into other organosulfur and

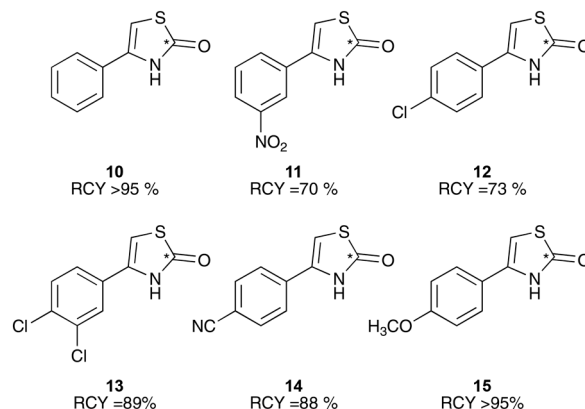


Chart 2 ^{11}C -Radiolabelled thiazolone molecules. Average of minimum $n = 2$.

heterocyclic structures.²⁵⁻²⁷ Thiazole structures are prevalent in a number of important naturally occurring compounds, for example thiamine and penicillin, and in a wide range of bioactive small molecules such as benzothiazoles, while the related thiazolones have been investigated for anti-cancer activity.²⁸ Carbon-11 radiolabelling within the ring of such sulfur based heterocyclic structures is challenging, and to the best of our knowledge has not been achieved until now. Since thiazolones can be accessed in high yield from α -thiocyanatophenones via an acid mediated cyclisation reaction (Scheme 4), we sought to explore this pathway using our *in situ* generated α -[^{11}C]thiocyanatophenones.²⁹

To test this in the radiochemistry laboratory, a solution of 1-phenyl-2-[^{11}C]thiocyanatoethanone (2) was added to a mixture of sulfuric and acetic acid, and heated to 90°C for 5 min. This resulted in complete conversion of 2 to the cyclised thiazolone, 4-phenylthiazol-2-[^{11}C]one (10) (Scheme 5, Chart 2). This cyclisation reaction was then performed with the remaining phenyl-substituted α -[^{11}C]thiocyanatophenones, producing the corresponding 4-phenylthiazol-2-[^{11}C]ones in high radiochemical purity (70–95%, Chart 2).

In conclusion, we have developed a new synthetic route to ammonium [^{11}C]thiocyanate via the reaction of ammonia with [^{11}C]carbon disulfide. The speed and efficiency of this synthesis has enabled us to exploit the [^{11}C]thiocyanate ion as a potent nucleophilic species for the preparation of a range of radiolabelled thiocyanate compounds in high radiochemical yields. Furthermore, we have been able to rapidly conduct cyclisation reactions with these α -[^{11}C]thiocyanatophenones to generate unprecedented ^{11}C -thiazolone molecules. We anticipate that this labelling route will find further applications for the preparation of carbon-11 based organosulfur tracers. Given that [^{11}C] CS_2 can be easily prepared from the widely available carbon-11 precursor [^{11}C] CH_3I we also expect that this method could be easily adopted by other PET centres with carbon-11 facilities.

We are grateful for financial support from the EPSRC (grant no. EP/L025140/1) and the Royal Society (grant no. RG110449).



Conflicts of interest

The authors declare no competing interest.

Notes and references

- 1 T. Jones and E. A. Rabiner, *J. Cereb. Blood Flow Metab.*, 2012, **32**, 1426–1454.
- 2 A. Nordberg, J. O. Rinne, A. Kadir and B. Långström, *Nat. Rev. Neurol.*, 2010, **6**, 78–87.
- 3 D. J. Brooks, *J. Nucl. Med.*, 2010, **51**, 596–609.
- 4 E. M. Rohren, T. G. Turkington and R. E. Coleman, *Radiology*, 2004, **231**, 305–332.
- 5 L. W. Dobrucki and A. J. Sinusas, *Nat. Rev. Cardiol.*, 2010, **7**, 38–47.
- 6 P. W. Miller, N. J. Long, R. Vilar and A. D. Gee, *Angew. Chem., Int. Ed.*, 2008, **47**, 8998–9033.
- 7 S. Preshlock, M. Tredwell and V. Gouverneur, *Chem. Rev.*, 2016, **116**, 719–766.
- 8 T. W. Price, J. Greenman and G. J. Stasiuk, *Dalton Trans.*, 2016, **45**, 15702–15724.
- 9 V. Bernard-Gauthier, J. J. Bailey, Z. Liu, B. Waengler, C. Waengler, K. Jurkschat, D. M. Perrin and R. Schirrmacher, *Bioconjugate Chem.*, 2016, **27**, 267–279.
- 10 J.-P. Meyer, P. Adumeau, J. S. Lewis and B. M. Zeglis, *Bioconjugate Chem.*, 2016, **27**, 2791–2807.
- 11 B. H. Rotstein, S. H. Liang, M. S. Placzek, J. M. Hooker, A. D. Gee, F. Dollé, A. A. Wilson and N. Vasdev, *Chem. Soc. Rev.*, 2016, **45**, 4708–4726.
- 12 M. B. Haskali and V. W. Pike, *Chem. – Eur. J.*, 2017, **1003**, 8156–8160.
- 13 G. Antoni, *J. Labelled Compd. Radiopharm.*, 2015, **58**, 65–72.
- 14 B. H. Rotstein, S. H. Liang, J. P. Holland, T. L. Collier, J. M. Hooker, A. A. Wilson and N. Vasdev, *Chem. Commun.*, 2013, **49**, 5621–5629.
- 15 S. Kealey, A. Gee and P. W. Miller, *J. Labelled Compd. Radiopharm.*, 2014, **57**, 195–201.
- 16 J. M. Hooker, M. Schoenberger, H. Schieferstein and J. S. Fowler, *Angew. Chem., Int. Ed.*, 2008, **47**, 5989–5992.
- 17 D. Roeda and F. Dolle, *Curr. Top. Med. Chem.*, 2010, **10**, 1680–1700.
- 18 P. W. Miller and D. Bender, *Chem. – Eur. J.*, 2012, **18**, 433–436.
- 19 T. Haywood, S. Kealey, S. Sanchez-Cabezas, J. J. Hall, L. Allott, G. Smith, C. Plisson and P. W. Miller, *Chem. – Eur. J.*, 2015, **21**, 9034–9038.
- 20 S. Merchant, L. Allott, L. Carroll, V. Tittrea, S. Kealey, T. H. Witney, P. W. Miller, G. Smith and E. O. Aboagye, *RSC Adv.*, 2016, **6**, S7569–S7579.
- 21 T. Castanheiro, J. Suffert, M. Donnard and M. Gulea, *Chem. Soc. Rev.*, 2016, **45**, 494–505.
- 22 S. Stone-Elander, P. Roland, C. Halldin, M. Hassan and R. Seitz, *Nucl. Med. Biol.*, 1989, **16**, 741–746.
- 23 P. Johnstrom, L. Bergman, K. Varnas, J. Malmquist, C. Halldin and L. Farde, *Nucl. Med. Biol.*, 2015, **42**, 555–560.
- 24 G. Westerberg and B. Langstrom, *J. Labelled Compd. Radiopharm.*, 1994, **34**, 545–548.
- 25 F. R. Bisogno, A. Cuetos, I. Lavandera and V. Gotor, *Green Chem.*, 2009, **11**, 452.
- 26 A. K. Yadav and L. D. S. Yadav, *Green Chem.*, 2015, **17**, 3515–3520.
- 27 J. Rudolph, H. Theis, R. Hanke, R. Endermann, L. Johannsen and F. Geschke, *J. Med. Chem.*, 2001, **44**, 619–626.
- 28 D. Havrylyuk, B. Zimenkovsky, O. Vasylenko, L. Zaprutko, A. Gzella and R. Lesyk, *Eur. J. Med. Chem.*, 2009, **44**, 1396–1404.
- 29 K. Pihlaja, V. Ovcharenko, E. Kolehmainen, K. Laihia, W. M. F. Fabian, H. Dehne, A. Perjéssy, M. Kleist, J. Teller and Z. Šusteková, *J. Chem. Soc., Perkin Trans. 2*, 2002, **2**, 329–336.

