



Cite this: *Food Funct.*, 2018, **9**, 6287

Maltol, a food flavor enhancer, attenuates diabetic peripheral neuropathy in streptozotocin-induced diabetic rats

Nan Guo,^{a,b} Caina Li,^{*a,b} Quan Liu,^{a,b} Shuainan Liu,^{a,b} Yi Huan,^{a,b} Xing Wang,^{a,b} Guoliang Bai,^{a,b} Miaomiao Yang,^{a,b} Sujuan Sun,^{a,b} Caimin Xu^c and Zhufang Shen^{id} ^{*a,b}

Scope: Maltol (3-hydroxy-2-methy-4-pyrone), a potent antioxidative agent, typically is used to enhance flavor and preserve food. This study evaluated its effects on preventing diabetic peripheral neuropathy (DPN) in streptozotocin (STZ)-induced diabetic rats and explored its mechanisms. **Methods and results:** We intraperitoneally injected Sprague-Dawley (SD) rats with STZ (65 mg kg⁻¹, ip) and treated the rats with different doses of maltol after 4 weeks of injection. During treatment, we evaluated motor nerve conduction velocity (MNCV) and thermal and mechanical hyperalgesia and assayed the oxidative stress, Na⁺-K⁺-ATPase activity, and apoptosis. Repeated treatment with maltol for 12 weeks significantly improved thermal and mechanical hyperalgesia, increased the MNCV, elevated the Na⁺-K⁺-ATPase activity, and ameliorated oxidative stress and apoptosis in STZ-induced diabetic rats. We cocultured RSC96 cells, a Schwann cell line, with maltol and hydrogen peroxide (H₂O₂, 0.6 mM). Evidently, maltol increased cell viability and inhibited apoptosis after injury by H₂O₂. **Conclusions:** Maltol was demonstrated to prevent DPN development and may provide a new alternative for the treatment of DPN.

Received 9th October 2018,
Accepted 16th October 2018

DOI: 10.1039/c8fo01964a

rsc.li/food-function

1. Introduction

Diabetic peripheral neuropathy (DPN) is a common complication of diabetes, which may be associated with age, duration of diabetes, glycemic control, blood lipid levels and blood pressure, but hyperglycemia is thought to be the most likely cause. A large amount of data demonstrated that the mechanisms of hyperglycemia causing diabetic complications, including DPN, mainly lay in the activation of the polyol pathway, formation of advanced glycation end products (AGEs), activation of protein kinase C isoforms, and increased hexosamine pathway flux, whereas the overproduction of superoxide by the mitochondrial electron-transport chain was a common element linking these four different pathogenic

mechanisms.¹ Despite therapeutic advances, no drugs have targeted any of these four mechanisms simultaneously. Antioxidants may be useful, such as α -lipoic acid (Nevralip Retard®, Medical Pharmaquality, Athens, Greece), a potent antioxidant. Through animal experiments and clinical trials, α -lipoic acid has been demonstrated to be effective in treating DPN,² and many other antioxidants, such as taurine, acetyl-L-carnitine, M40403, and β -carotene, also have been demonstrated to be effective in ameliorating the development of DPN.³

Maltol (3-hydroxy-2-methy-4-pyrone), a type of aromatic compound, exists in high concentrations in red ginseng, which is manufactured by steaming and drying fresh ginseng, a traditional herbal medicine.⁴ Currently, maltol is used extensively as a safe flavoring agent and food preservative, and it is always found in heat-processed food.⁵ As a well-known metal ion chelator, maltol is widely used in the field of catalysis, cosmetics, and medicine. Furthermore, studies have demonstrated that maltol was a strong antioxidant⁶ and it significantly inhibited the production of AGEs.⁷ Maltol also could defend against oxidation-induced damage to nerve cells, kidneys, and the liver by scavenging free radicals in the body,^{8,9} and effectively inhibiting the apoptosis induced by hydrogen peroxide (H₂O₂),¹⁰ which has attracted increasing attention in the research and development of drugs. Moreover, there are still no reports on the use of maltol to treat DPN.

^aState Key Laboratory of Bioactive Substances and Functions of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, China. E-mail: shenzhf@imm.ac.cn, leecaina@imm.ac.cn; Fax: +8610 83172669; Tel: +86 10 83172669

^bKey Laboratory of Polymorphic Drugs of Beijing, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, China

^cState Key Laboratory of Medical Molecular Biology, Department of Biochemistry and Molecular Biology, Institute of Basic Medical Sciences, Chinese Academy of Medical Sciences, School of Basic Medicine, Peking Union Medical College, Beijing 100005, China



Therefore, the current study prospectively investigated the effects of maltol on DPN following treatment for 12 weeks in streptozotocin (STZ)-induced diabetic rats and preliminarily explored the mechanisms *in vivo* and *in vitro*, to supply the experimental basis to extend its application.

2. Materials and methods

2.1. Animals

Male Sprague-Dawley (SD) rats (150–170 g; Beijing HFK Bioscience Co. Ltd, Beijing, China) were kept in a temperature- and humidity-controlled environment with a 12 h light/dark cycle and fed food and water *ad libitum*. The Experimental Animal Welfare Ethics Committee of the Institute of Materia Medica (Chinese Academy of Medical Sciences and Peking Union Medical College) under No. 00001042 approved all of the protocols for this research, and all of the rats were handled according to the commonly accepted “3R” and standards and guidelines for laboratory animals (GB14925-2001 and MOST 2006a) established by the People's Republic of China.

2.2. Cells

RSC96 cells, the rat Schwann cell line, were purchased from the Cell Resource Center of the Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences (Shanghai, China) and cultured in Dulbecco's modified Eagle's Medium (DMEM; Gibco Laboratories, Gaithersburg, MD, USA) plus 100 U mL⁻¹ penicillin (Sigma-Aldrich, St Louis, MO, USA), 100 mg mL⁻¹ streptomycin (Sigma-Aldrich) and 10% (v/v) fetal bovine serum (FBS; Gibco) at 37 °C under a 5% carbon dioxide-humidified atmosphere.

2.3. Experimental design and treatment

In vivo, after 3 days of acclimatisation, we intraperitoneally injected all of the rats with STZ (65 mg kg⁻¹, ip; Sigma-Aldrich) dissolved in citrate buffer (0.1 mol L⁻¹, pH 4.5) to induce hyperglycemia. We determined the fasting blood glucose (FBG) levels to verify the occurrence of diabetes 3 days later. Four weeks after the verification of diabetes, we determined the thermal and mechanical pain thresholds and the erythrocytic Na⁺-K⁺-ATPase activity of rats with blood glucose ≥ 11.1 mmol L⁻¹, and then we divided the rats into five groups ($n = 12$): the vehicle-treated group (Con), the maltol-treated groups (50, 100, and 200 mg kg⁻¹), and the α -lipoic acid-

treated group (100 mg kg⁻¹; see Table 1). We adopted normal age-matched SD rats as the healthy control (Nor, $n = 10$) and α -lipoic acid served as the positive reference. We treated the rats with maltol or α -lipoic acid dissolved in 0.5% carboxymethyl cellulose and sodium (CMC-Na) solution by gavage once daily for 12 weeks after division, and the Nor and Con rats were orally administered 0.5% CMC-Na solution. Insulin treatment was not given. We dynamically monitored the fasting blood glucose (FBG) levels and body weight (BW) during the treatment and evaluated other parameters, including motor nerve conduction velocity (MNCV), thermal and mechanical hyperalgesia, Na⁺-K⁺-ATPase activity, total antioxidant capacity (TAOC), malondialdehyde (MDA), glutathione (GSH), and superoxide dismutase (SOD) at the end of the experiment (Fig. 1A).

In vitro, we incubated RSC96 cells with H₂O₂ (0.6 mM) to induce oxidative damage. We separately coincubated maltol at concentrations 0.1 and 0.5 mM, as well as α -lipoic acid at a

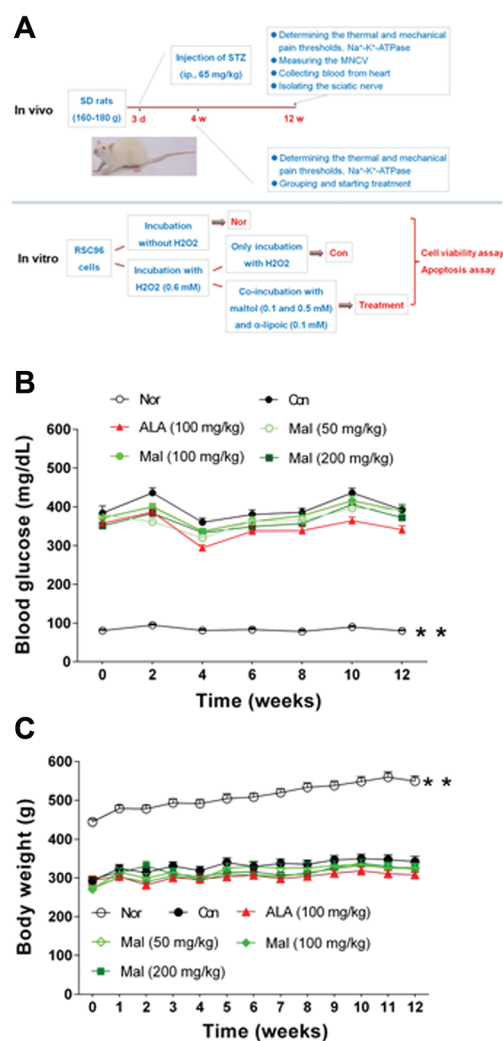


Fig. 1 Experimental scheme and the effects of treatment with maltol on fasting blood glucose and body weight in STZ-induced diabetic rats. (A) Experimental scheme, (B) blood glucose, and (C) body weight. Data expressed as mean \pm SEM, $n = 11$ –12, ** $P < 0.01$ vs. Con.

Table 1 Data for the grouping of STZ-induced diabetic rats

Group	Dose (mg kg ⁻¹)	Withdrawal latency (s)	Withdrawal threshold (g)	Na ⁺ -K ⁺ -ATPase (U mL ⁻¹)
Nor	—	15.8 \pm 0.8**	43.0 \pm 0.7**	7.4 \pm 0.3**
Con	—	10.0 \pm 0.5	27.4 \pm 1.1	5.1 \pm 0.5
ALA	100	10.0 \pm 0.5	27.4 \pm 1.1	5.0 \pm 0.7
Maltol	50	9.8 \pm 0.5	27.5 \pm 1.2	5.0 \pm 0.4
	100	10.0 \pm 0.5	27.4 \pm 1.2	4.9 \pm 0.5
	200	10.0 \pm 0.5	27.4 \pm 1.2	4.8 \pm 0.5

Data expressed as mean \pm SEM, $n = 10$ –12, ** $p < 0.01$ vs. Con.



concentration of 0.1 mM with H₂O₂ in RSC96 cells. We used cells without any treatment as the normal control (Nor). Then, we assayed cell viability by CCK8 and evaluated apoptosis-related proteins by western blot and real-time polymerase chain reaction (PCR; Fig. 1A).

2.4. Measurement of FBG and BW

To measure FBG, all of the rats were fasted for 4 h with water *ad libitum*, and then we determined the blood glucose levels using the glucose-oxidase method every 2 weeks. The BW was documented once weekly during treatment without a withdrawal of diet.

2.5. Measurement of MNCV

After treatment for 12 weeks, all of the rats were anesthetized with sodium pentobarbital (60 mg kg⁻¹, ip). We stripped the left sciatic nerve to receive a stimulation at the proximal end by square-wave pulses (duration: 0.01 ms, intensity: 1 V) delivered through bipolar recording electrodes, and the action potential at the distal end was recorded using the BL-420S bio-mechanical system (Chengdu Taimeng Technology Co., Ltd, Chengdu, China). We calculated MNCV (m s⁻¹) as follows: (the distance between the stimulating and recording electrode)/latency.¹¹

2.6. Measurement of the thermal and mechanical hyperalgesia

After treatment for 12 weeks, we monitored the thermal and mechanical hyperalgesia by a plantar test and recorded the latency or threshold of paws withdrawal after stimulation using Hargreaves Apparatus 37370 and Dynamic Plantar Aesthesiometer 37450 (Ugo Basile S.R.L., Comerio, Italy). In short, we placed the rats separately in six cages (17 × 11.5 × 14 cm) and stimulated the left hind paws with infrared light (cutoff period set at 25 s) or a mechanical needle (biting strength set at 50 g). The equipment automatically recorded paw withdrawal latency as soon as the paws withdrew.¹² We assessed each rat five times with a 5 min interval between trials and took the mean value as withdrawal threshold or latency.

2.7. Measurement of Na⁺-K⁺-ATPase activity

After treatment for 12 weeks, all of the rats were killed by drawing blood from the heart and the sciatic nerves were isolated. Then, we determined the Na⁺-K⁺-ATPase activities in erythrocytes and the sciatic nerves as follows. For the determination in erythrocytes, we collected 10 µl of whole blood and added it to 240 µl of distilled water to mix and immediately determine the Na⁺-K⁺-ATPase activity. For the determination in sciatic nerves, we first cut one side of the sciatic nerves into pieces with ophthalmic scissors in cold saline (100 mg:1800 µl) and homogenized the nerves with Beed Ruptor 24 Elite (OMNI International, Kennesaw, GA, USA). Then, we centrifuged the homogenate at 4 °C, 3500 rpm for 15 min and collected the supernatant to be used. The determination and calculation of Na⁺-K⁺-ATPase activities were pro-

cessed using commercial kits bought from Nanjing Jiancheng Bioengineering Institute (Nanjing, China).

2.8. Measurement of TAOC, MDA, GSH and SOD

At the end of the experiment, we isolated the serum by centrifugation at 4 °C, 3000 rpm for 10 min. We then determined the content of TAOC, MDA, GSH, and SOD in serum and sciatic nerve homogenates with commercial kits bought from Nanjing Jiancheng Bioengineering Institute (Nanjing, China) according to the manufacturer's instructions.

2.9. Measurement of caspase-3 activity

After treatment for 12 weeks, we also collected the dorsal root ganglia (DRG) under a stereoscope to measure the caspase-3 activities with a caspase-3 colorimetric assay kit (Applygen Technologies Inc., Beijing, China). First, we cut the DRG into pieces with ophthalmic scissors in a lysis buffer (3–10 mg:100 µl) and homogenized with Beed Ruptor 24 Elite (OMNI International). Then, we collected the supernatant by centrifugation at 4 °C, 12 000g for 10 min. We quantified the protein in the supernatant following the Bradford method (Applygen Technologies Inc.). We calculated the caspase-3 activities with protein correction.

2.10. CCK-8 assay

We planted RSC96 cells into a 96-well plate (15 000 cells per well) and cultured the cells overnight in DMEM plus 100 U mL⁻¹ penicillin, 100 mg mL⁻¹ streptomycin and 10% (v/v) FBS. Six hours after treatment, as described in Section 2.3, we added 10 µl of the CCK-8 assay reagent (Dojindo Molecular Technologies, Inc., Kumamoto, Japan) and incubated the cells for another 3 h. Then, we measured the absorbance at 450 nm and used the optical density (OD) values on the y-axis to express the cell viability.

2.11. Western blot

We evaluated the apoptosis-related proteins, including Bcl-XL, BAX, and caspase-3 in sciatic nerves of STZ-induced diabetic rats after treatment for 12 weeks, and evaluated Bcl-XL in RSC96 cells cultured and treated as noted in Section 2.10, with western blot. First, we extracted total protein from one side of the sciatic nerve and RSC96 cells planted in 6-well plates (200 000 cells per well) with RIPA lysate plus inhibitors of protease and protein phosphatase (Applygen Technologies Inc.), followed by protein quantification using a BCA assay kit (Applygen Technologies Inc.). We electrophoretically separated equal amounts of protein by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and transferred the protein to a polyvinylidene difluoride (PVDF) membrane (Millipore, Darmstadt, Germany). After blocking for 1 h with 5% nonfat milk, we separately incubated the PVDF membrane with primary antibodies, including Bcl-XL, BAX, caspase-3, and β-actin (1:1000, Cell Signaling Technology, Danvers, MA, USA) at 4 °C overnight, followed by incubation with mouse anti-rabbit secondary antibody conjugated with horseradish peroxidase (1:1000, Applygen Technologies Inc.) after washing.



We visualized the protein bands through an enhanced chemiluminescence detection system (ChemiScope 2850; Clinx Science Instruments, Shanghai, China) and quantified the relative band densities using β -actin as the internal standard.

2.12. Quantitative real-time PCR

We also evaluated the apoptosis-related genes, including BAG4, BAX, and caspase-3 in the sciatic nerve and evaluated BAX and caspase-3 in RSC96 cells with real-time PCR. First, we extracted total RNA from one side of the sciatic nerve and RSC96 cells planted and treated as noted in Section 2.11 using TRIzol reagent (Invitrogen, Carlsbad, CA, USA). Then, we synthesized the cDNA using a VigoScript first strand cDNA synthesis kit (Vigorous Biotechnology Beijing Co., Ltd, Beijing, China). We conducted quantitative real-time PCR with SYBR Green Master Mix (Takara, Dalian, China) using the 7900HT fast real-time PCR system (Applied Biosystems, Foster City, CA, USA) according to the following protocol: 1 cycle at 95 °C for 30 s, then 40 cycles at 95 °C for 5 s, and 60 °C for 31 s. Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) served as the internal standard. We used the following primer sequences: BAG4, forward 5'-GCGTTTCCTCACTCGTGACA-3' and reverse 5'-AAGGGGGTTAGGACGGC TAC-3', BAX forward 5'-TT GCTACAGGGTTTCATCCAGG-3' and reverse 5'-CACTCGCT CAGCTTCTTGGT-3', caspase-3 forward 5'-GGAGCTTGGA ACGCAAGAA-3' and reverse 5'-ACACAAGCCCATTTCAGGGT-3', and GAPDH forward 5'-CCTGTTGCTGTAGCCGTATTCA-3' and reverse 5'-CCAGGTT GTCTCCTGCGACTT-3'.

2.13. Statistical analysis

All of the results were presented as mean \pm SEM. We analyzed data through one-way analysis of variance with Bonferroni's correction and Student's *t*-test. We defined statistical significance as $P < 0.05$.

3. Results and discussion

3.1. Effects of maltol on FBG and BW in STZ-induced diabetic rats

In comparison with nondiabetic rats (Nor), STZ-induced diabetic rats displayed about 4.5-fold increase in FBG levels and 1.6-fold decrease in BW during the treatment, suggesting the occurrence of diabetes. In comparison with STZ-induced diabetic rats (Con), repeated treatment with maltol ranging from 50 mg kg⁻¹ to 200 mg kg⁻¹ for 12 weeks produced some changes in the FBG level but with fluctuations (Fig. 1B). Repeated treatment with maltol had no effect on BW during the experiment (Fig. 1C). These results suggested that maltol had no significant influences on controlling blood glucose levels and body weight.

STZ-induced diabetic rats are a commonly used model to explore the mechanisms of neuropathy and to evaluate the effects of potential therapeutics. One study reported that DPN developed within 6 weeks after the induction of diabetes.¹³ Researchers have opted to start drug intervention at 4 or

8 weeks after the induction of diabetes.^{3,14} We divided STZ-induced diabetic rats according to the thermal and mechanical pain thresholds and the erythrocytic Na⁺-K⁺-ATPase activity 4 weeks after the induction of diabetes, when all three markers were significantly decreased in comparison with nondiabetic rats (Table 1), suggesting pain hyperalgesia, accumulation of intra-axonal Na⁺, and nodal axonal swelling.¹⁵

Uncontrolled chronic hyperglycemia in STZ-induced diabetic rats was the source of DPN, and keeping the blood glucose level in a suitable range without a large fluctuation significantly decreased the rate of DPN. Thus, in the present study, we first investigated the influences of maltol on FBG and BW. We found that maltol (200 mg kg⁻¹) produced some changes in the FBG level but with fluctuations during the treatment, which we thought may have resulted from the states of the animals before the measurement was taken, such as food intake, water consumption, and β -cell function. Repeated treatment with maltol did not influence the BW of STZ-induced diabetic rats.

3.2. Effects of maltol on MNCV in STZ-induced diabetic rats

In comparison with nondiabetic rats (Nor), STZ-induced diabetic rats (Con) showed decreased MNCV, suggesting the occurrence of peripheral neuropathy. In comparison with STZ-induced diabetic rats (Con), repeated administration of maltol at doses of 100 and 200 mg kg⁻¹ for 12 weeks improved the MNCV in a dose-dependent manner by 34.0% ($p < 0.05$) and 58.2% ($p < 0.01$), respectively, and the MNCV of rats in the maltol-treated group at a dose of 200 mg kg⁻¹ showed no difference from that of nondiabetic rats (Nor), suggesting the improvement in peripheral nerve function (Fig. 2).

The slowing of a large fiber nerve conduction velocity (NCV) has been a gold standard for defining neuropathy in patients with type 2 diabetes mellitus and for assessing the efficacy of potential therapeutics. The slowing of MNCV and sensory NCV (SNCV) also was an early feature of diabetic mice and may be detected within weeks of the onset of diabetes.¹⁶ In our study, the MNCV of STZ-induced diabetic rats decreased by 47.9% compared with nondiabetic rats after 16 weeks of untreated diabetes. The increase in MNCV following maltol treatment at a dose of 200 mg kg⁻¹ to a level that was not statistically

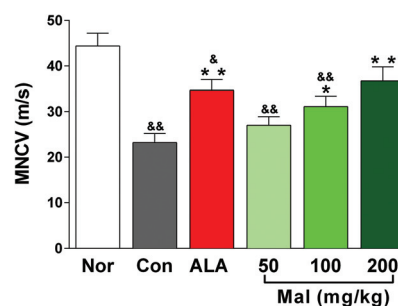


Fig. 2 Effect of treatment with maltol on the MNCV in STZ-induced diabetic rats. Data expressed as mean \pm SEM, $n = 10-12$, [§] $P < 0.05$, ^{§§} $P < 0.01$ vs. Nor; * $P < 0.05$, ** $P < 0.01$ vs. Con.



different from that of nondiabetic rats indicated its benefits in treating DPN. In addition, as a potent antioxidant, α -lipoic acid has been a useful clinical option for patients with DPN, which also significantly reduced the oxidative-stress-induced injury and improved the NCV and blood flow.^{2,17}

3.3. Effects of maltol on thermal and mechanical hyperalgesia in STZ-induced diabetic rats

In comparison with nondiabetic rats (Nor), STZ-induced diabetic rats exhibited decreased withdrawal threshold and latency in response to mechanical and thermal stimulations, indicating evident pain hyperalgesia. In comparison with STZ-induced diabetic rats (Con), repeated administration of maltol at doses of 50, 100, and 200 mg kg⁻¹ for 12 weeks significantly extended the withdrawal threshold and latency following mechanical stimulation by 18.1%, 24.0%, and 27.7%, and thermal stimulation by 47.2%, 87.9%, and 100.0%, respectively. Furthermore, the withdrawal thresholds and latency of rats in the groups of maltol at doses of 100 and 200 mg kg⁻¹ was equivalent to that of nondiabetic rats (Nor), suggesting amelioration of the pain hyperalgesia (Fig. 3).

DPN, a major cause of foot ulceration and limb loss for patients with diabetes, has been associated with sensory loss or abnormal sensations, such as hyperalgesia, allodynia and spontaneous pain. Thermal and mechanical hyperalgesia have been a clinical phenomenon in early DPN, whereas hypoalgesia has been found in advanced DPN;¹⁸ however, the mechanisms of abnormal nociception and the time for turnover of hyperalgesia to hypoalgesia in diabetes have not been well understood. In our study, we found that the thermal and mechanical pain thresholds of STZ-induced diabetic rats significantly decreased by 51.2% and 24.9% after 16 weeks of untreated diabetes compared with nondiabetic rats, suggesting hyperalgesia. These results were consistent with previous studies, wherein hyperalgesia has been reported after 6, 8, and 12 weeks of hyperglycemia.^{19–21} Many studies, however, have still reported hypoalgesia. For example, Davidson *et al.* found that STZ-induced diabetic rats displayed thermal hypoalgesia after 16 weeks of untreated diabetes, and the high-fat-fed/low-

dose STZ-induced diabetic rats showed thermal hypoalgesia after 8 weeks of a high fat diet and 16 weeks of hyperglycemia induced by STZ,^{22,23} whereas Stavnichuk *et al.* reported that STZ-induced diabetic C57Bl6/J mice exhibited thermal hypoalgesia but tactile allodynia.¹⁴

Behavioral responses to mechanical touch may incorporate the function of large myelinated sensory fibers that terminated in the dermis, whereas the responses to thermal stimulation were transduced by small sensory C fibers that terminated in the epidermis. Calcutt *et al.* reported that diabetic rodents developed increased sensitivity to a series of manual von Frey filaments within 2–4 weeks of hyperglycemia and an initial paw thermal hyperalgesia that progressed to hypoalgesia without insulin therapy.^{24,25} Kambiz *et al.* found that STZ-induced WAG/RijHsd diabetic rats did not display significantly different responses to mechanical stimulation after 4 weeks of diabetes. These rats, however, developed evident hyposensitivity after 6 and 8 weeks of diabetes and showed remarkable heat hypersensitivity in the hot plate test at 4, 6, and 8 weeks after the induction of diabetes.²⁶ Thus, we deemed the behavioral responses to thermal and mechanical stimulation of rodents to be complicated, which may have varied as a result of the duration of diabetes, blood glucose level, and animal species *etc.* among different labs.

3.4. Effects of maltol on Na⁺-K⁺-ATPase activities in STZ-induced diabetic rats

We considered the Na⁺-K⁺-ATPase activity to be an index of microvascular blood supply and peripheral nerve injury. In comparison with nondiabetic rats (Nor), STZ-induced diabetic rats (Con) displayed decreased Na⁺-K⁺-ATPase activities in erythrocytes and sciatic nerves. In comparison with STZ-induced diabetic rats (Con), repeated treatment with maltol at doses of 50, 100, and 200 mg kg⁻¹ for 12 weeks increased the erythrocytic Na⁺-K⁺-ATPase activity in a dose-dependent manner by 19.2%, 24.8%, and 28.6%, respectively ($p < 0.01$). Meanwhile, maltol at doses of 100 and 200 mg kg⁻¹ also evidently increased the Na⁺-K⁺-ATPase activity in sciatic nerves by 13.9% ($p < 0.05$) and 16.8% ($p < 0.01$), respectively.

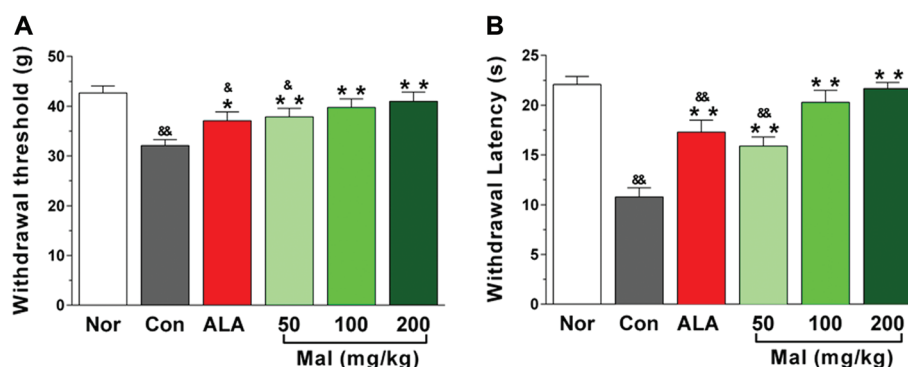


Fig. 3 Effects of treatment with maltol on the sensitivity of plantar to mechanical and thermal stimulations in STZ-induced diabetic rats. (A) Plantar withdrawal threshold to mechanical stimulations and (B) plantar withdrawal latency to thermal stimulations. Data expressed as mean \pm SEM, $n = 11–12$, ^a $P < 0.05$, ^{ab} $P < 0.01$ vs. Nor; * $P < 0.05$, ** $P < 0.01$ vs. Con.



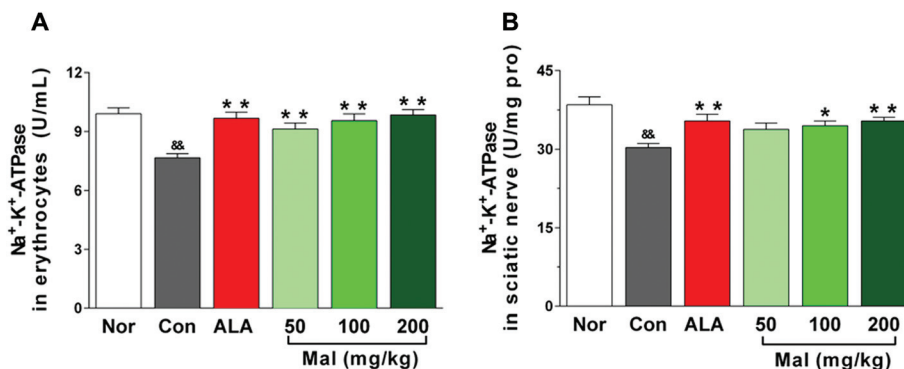


Fig. 4 Effects of treatment with maltol on the Na⁺-K⁺-ATPase activities in erythrocytes and the sciatic nerve of STZ-induced diabetic rats. (A) Na⁺-K⁺-ATPase activity in erythrocytes and (B) Na⁺-K⁺-ATPase activity in the sciatic nerve. Data expressed as mean \pm SEM, $n = 6$, ^{ab} $P < 0.01$ vs. Nor; * $P < 0.05$, ** $P < 0.01$ vs. Con.

Furthermore, the Na⁺-K⁺-ATPase activity in both erythrocytes and sciatic nerves after treatment with maltol were not different from that of nondiabetic rats (Nor) (Fig. 4).

Na⁺-K⁺-ATPase is a ubiquitous membrane-associated enzyme that is expressed in most erythrocytes. Early studies have shown that diabetes induced a parallel decrease of Na⁺-K⁺-ATPase activity in erythrocytes and sciatic nerve, and monitoring the Na⁺-K⁺-ATPase activity in erythrocytes seemed to reflect the activity in the peripheral nerve.²⁷ Furthermore, decreased Na⁺-K⁺-ATPase activity in erythrocytes may be responsible for the decreased cell deformability and NCV and impaired nerve histology.²⁸ Therefore, the erythrocytic Na⁺-K⁺-ATPase activity could be a potential marker for a predisposition to diabetic neuropathy.²⁹ Our study found that long-term treatment with maltol significantly increased the Na⁺-K⁺-ATPase activity in both erythrocytes and sciatic nerve of STZ-induced diabetic rats, correlating with the improvement of sciatic NCV and thermal and mechanical hyperalgesia, which suggested that maltol may treat DPN by elevating the decreased Na⁺-K⁺-ATPase activity in diabetic rats.

3.5. Effects of maltol on oxidative stress markers in STZ-induced diabetic rats

In comparison with nondiabetic rats (Nor), STZ-induced diabetic rats (Con) showed a significant imbalance between oxidation and antioxidation with increased serum MDA levels and decreased TAOC, GSH, and SOD content in serum and sciatic nerves. In comparison with STZ-induced diabetic rats (Con), repeated administration of maltol for 12 weeks significantly increased the serum TAOC, GSH, and SOD levels and decreased the serum MDA level in a dose-dependent manner (Fig. 5A–D). Furthermore, maltol also increased the SOD activity ($p < 0.01$) and GSH content ($p < 0.05$) in sciatic nerves, especially at a dose of 200 mg kg⁻¹ (Fig. 5E–F). Note that the antioxidative markers, TAOC, SOD, and GSH, both in serum and the sciatic nerves of maltol-treated (100 and 200 mg kg⁻¹) rats were not different from those of nondiabetic rats (Nor), but the MDA level evidently was still higher than that of Nor, suggesting that an improvement in the imbalance between oxi-

dation and antioxidation by maltol mainly were attributed to an increase in antioxidative ability.

Accumulating evidence has supported the finding that oxidative stress caused by chronic hyperglycemia was a key mechanism of DPN and seeking excellent antioxidative therapies was essential for the prevention of neuropathy in patients with diabetes.³⁰ Oxidative stress occurred in a cellular system when the generation of free radicals exceeded their rate of utilization.³¹ It established a linkage between the physiological and metabolic initiators involved in the progressive nerve fiber injury, dysfunction, and loss during the development of DPN.³² GSH, the most sensitive index of ongoing oxidative stress, was a first line of defense against free radicals and the most potent endogenous antioxidant in most mammalian cells. Depletion of GSH significantly impaired the mitochondrial function of motor neurons and rendered them susceptible to oxidative damage and death,³³ while sparing GSH by antioxidants was highly effective in protecting neuronal cells from damage.³⁴ As a potent antioxidant, repeated treatment with maltol significantly increased the antioxidative capacity of diabetic rats by elevating the serum TAOC, SOD, and GSH levels and decreasing the serum MDA level, and increasing the SOD and GSH levels in the sciatic nerve, which suggested that maltol may treat DPN by protecting the sciatic nerve from oxidative damage in STZ-induced diabetic rats.

3.6. Effects of maltol on caspase-3 activities in STZ-induced diabetic rats

In comparison with nondiabetic rats (Nor), STZ-induced diabetic rats (Con) exhibited increased caspase-3 activity in DRG ($p < 0.01$), suggesting the occurrence of apoptosis. In comparison with STZ-induced diabetic rats (Con), repeated treatment with maltol for 12 weeks had no significant influence on the activity of caspase-3, but it separately produced a decrease by 14.6%, 15.4%, and 23.3% at doses of 50, 100, and 200 mg kg⁻¹. The caspase-3 activities of rats in maltol-treated groups were higher than those of nondiabetic rats (Nor) (Fig. 6). These results suggested that treatment with maltol may inhibit cell apoptosis in DRG.



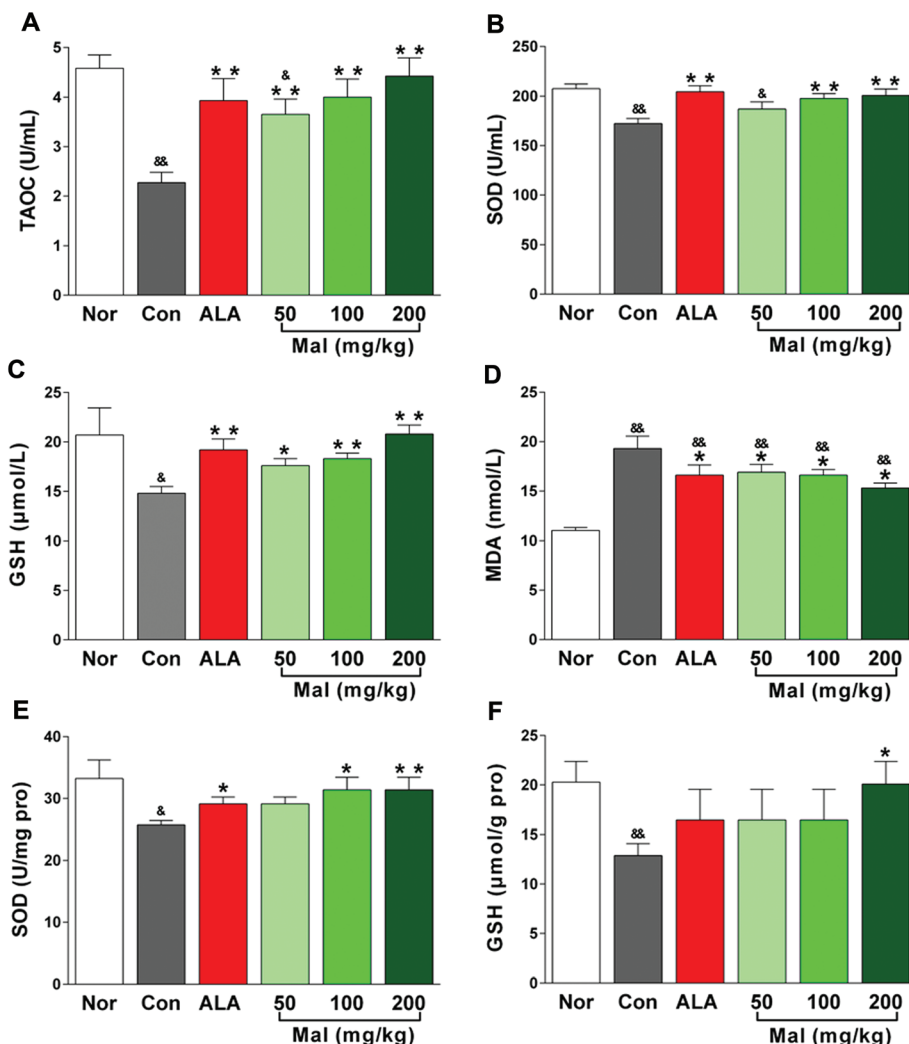


Fig. 5 Effects of treatment with maltol on the oxidative markers in STZ-induced diabetic rats. (A) TAOC in serum, (B) SOD in serum, (C) GSH in serum, (D) MDA in serum, (E) SOD in the sciatic nerve, and (F) GSH in the sciatic nerve. Data expressed as mean \pm SEM, $n = 6-8$ for (A) and (C), $n = 9-11$ for (B), $n = 7-10$ for (D) and (F), $n = 7-11$ for (E), $^{\circ} P < 0.05$, $^{\circ\circ} P < 0.01$ vs. Nor; $^* P < 0.05$, $^{**} P < 0.01$ vs. Con.

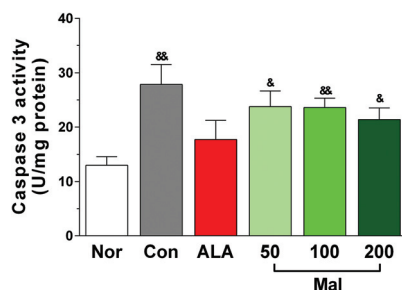


Fig. 6 Effect of treatment with maltol on the caspase-3 activity in DRG of STZ-induced diabetic rats. Data expressed as mean \pm SEM, $n = 8-12$, $^{\circ} P < 0.05$, $^{\circ\circ} P < 0.01$ vs. Nor.

The axons of DRG neurons were afferents that transmitted sensory information to the central nervous system. Due to the exuberant metabolism and super sensitivity to hyperglycemia, DRG neurons were more easily injured and subjected to apop-

tosis during the progression of diabetes, ultimately leading to numbness and pain in patients.³⁵ In addition, there were also close connections between SCs and DRG neurons that SCs played important roles in the neurite outgrowth of DRG neurons and a decrease of the nerve growth factor level secreted by SCs might cause a defect in axonal regeneration.³⁶ Thus, DRG neurons were considered to be an important experimental object of DPN. Large amounts of research have reported that elevated glucose levels could produce oxidative damage to DRG neurons and push them to apoptosis, with the mitochondria serving as an important target.^{37,38} Caspase-3 belongs to the cysteine-aspartic acid protease family and is an important executor of cell apoptosis. To fully assess the effects of maltol treatment on apoptosis, we also collected the DRG neurons to determine the pro-apoptotic caspase-3 activities. We found that long-term treatment with maltol produced a decrease by at least 14.6%, although without statistical significance, which was consistent with the expression of caspase-3



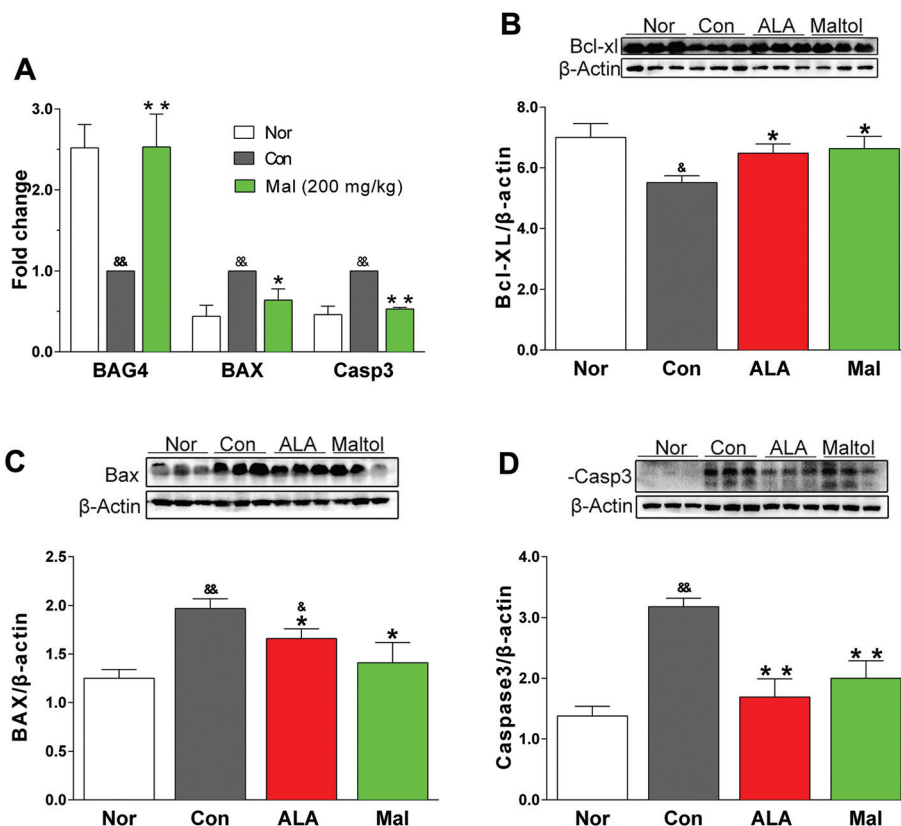


Fig. 7 Effects of maltol on the expression of genes and proteins related to apoptosis in sciatic nerves of STZ-induced diabetic rats. (A) The expression of BAG4, BAX, and caspase-3 genes, (B) the expression of Bcl-XL protein, (C) the expression of BAX protein, and (D) the expression of caspase-3 protein. Data expressed as mean \pm SEM, $n = 5-6$ for (A), and $n = 3$ for (B), (C), and (D), [§] $P < 0.05$, ^{§§} $P < 0.01$ vs. Nor; * $P < 0.05$, ** $P < 0.01$ vs. Con.

in sciatic nerves shown in Fig. 7, suggesting that maltol may inhibit cell apoptosis to improve the peripheral nerve function.

3.7. Effects of maltol on the expression of apoptosis-related proteins in STZ-induced diabetic rats

BAG4 and Bcl-XL were two major negative regulators of apoptotic pathways, whereas BAX and caspase-3 were important pro-apoptotic proteins. In comparison with nondiabetic rats (Nor), STZ-induced diabetic rats (Con) displayed increased expressions of BAX and caspase-3 and decreased expressions of BAG4 and Bcl-XL, suggesting activation of the apoptotic pathway. In comparison with STZ-induced diabetic rats (Con), repeated treatment with maltol for 12 weeks significantly lowered the expression of BAX and caspase-3 both in gene and protein levels, and it remarkably elevated the expression of BAG4 gene and Bcl-XL protein. Furthermore, the expressions of BAX, caspase-3, BAG4, and Bcl-XL in the maltol-treated group were nearly equivalent to the expressions of nondiabetic rats (Nor) (Fig. 7).

Oxidative stress was closely associated with apoptosis in many cells by increasing the peroxidation of membrane lipids, impairing the functional proteins and signal transduction pathways, and changing the cellular redox potential.³⁹ The pathogenesis of DPN involved demyelination, axonal atrophy,

and degeneration, indicating that apoptosis participated in its occurrence and development.³⁹ The sciatic nerve is the longest and largest peripheral nerve in the body, and we analyzed the expressions of apoptosis-associated proteins in the sciatic nerves of diabetic rats to further investigate how maltol treated DPN. Bcl-XL and BAX were two important proteins in cell survival, whereas Bcl-XL belonged to the pro-survival Bcl-2 family and could bind to the pro-apoptotic protein BAX to suppress the induction of apoptosis.⁴⁰ Our study showed that repeated administration of maltol significantly decreased the expression of BAX and increased the expression of Bcl-XL of sciatic nerves in STZ-induced diabetic rats, and evidently decreased the expression of caspase-3. The results suggested that maltol may improve the nerve function by inhibiting cell apoptosis.

3.8. Effects of maltol on the expression of apoptosis-related proteins in RSC96 cells injured by H₂O₂

Schwann cells (SCs) accounted for most of the peripheral nervous system and RSC96 cells belonged to the rat Schwann cell line. We used RSC96 cells to evaluate the effect of maltol in the present study. H₂O₂ is a commonly used oxidant to induce cell apoptosis through the activation of oxidative stress. In comparison with the cells without H₂O₂ treatment (Nor),



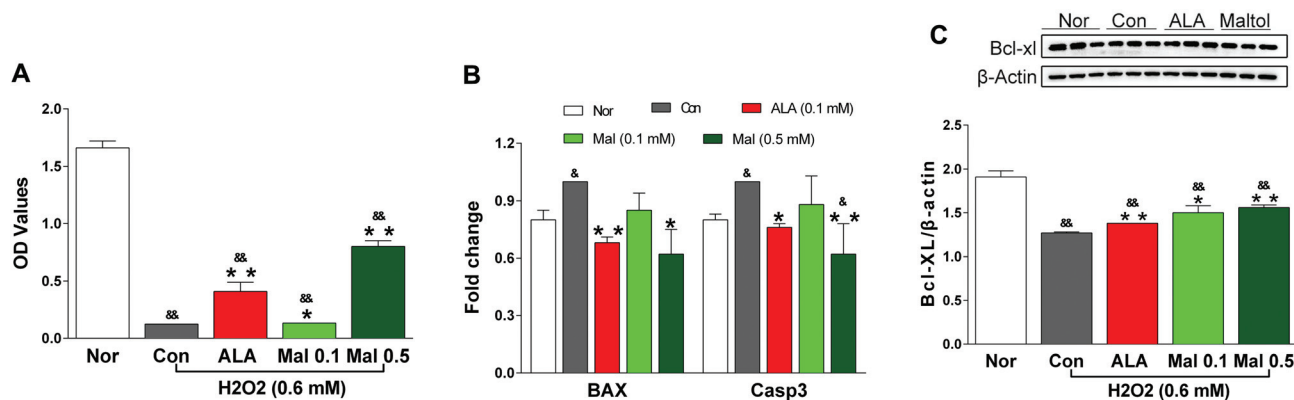


Fig. 8 Effects of maltol on the viability of RSC96 cells injured by H_2O_2 and the expression of genes and proteins related to apoptosis. (A) Cell viability, (B) the expression of BAX and caspase-3 gene, (C) the expression of Bcl-XL protein. Data expressed as mean \pm SEM, $n = 5$ for (A), $n = 3-6$ for (B) and (C), $^{\circ} P < 0.05$, $^{\circ\circ} P < 0.01$ vs. Nor; $^* P < 0.05$, $^{**} P < 0.01$ vs. Con.

H_2O_2 -treated (0.6 mM) cells (Con) showed a significant decrease in viability and anti-apoptotic protein Bcl-XL expression as well as an evident increase in pro-apoptotic protein BAX and caspase-3 expression, suggesting the apoptosis of RSC96 cells. In comparison with the H_2O_2 -treated cells (Con), maltol at concentrations of 0.1 and 0.5 mM significantly increased the viability of RSC96 cells after coincubation with H_2O_2 (Fig. 8A), evidently elevated the expression of Bcl-XL protein in a dose-dependent manner (Fig. 8B), and decreased the expression of BAX and caspase-3 genes (Fig. 8C), although the cell viability and expression of Bcl-XL remained lower than that of the cells without H_2O_2 treatment (Nor). The results suggested that maltol may inhibit the H_2O_2 -induced oxidative injury to RSC96 cells.

SCs, the only glial cells in the peripheral nervous system, generated myelin and supported the structural and functional integrity of nerves.⁴¹ The disruption to SCs' metabolism triggered by hyperglycemia resulted in an accumulation of neurotoxic intermediates, which contributed to axonal degeneration, endothelial dysfunction, and diabetic neuropathy.⁴² Furthermore, SCs also imposed cellular specializations that allowed for the fast conduction of action potentials.⁴³ Reduced NVC in DPN was largely due to the impaired function of SCs, which generated and maintained a multilamellar insulating myelin sheath around an associated axon and provided a permissive environment for peripheral nerve regeneration.⁴⁴ Previous studies showed that the exposure of SCs to H_2O_2 *in vitro* significantly caused oxidative stress and induced apoptosis *via* the mitochondrial pathway.⁴⁵ In the present study, we incubated RSC96 cells with maltol and H_2O_2 (0.6 mM) to investigate the effect of maltol on the apoptosis induced by oxidative stress. A previous study revealed that maltol could help SCs recover from H_2O_2 -induced oxidative injury and protect SCs from apoptosis *in vitro*, suggesting it may be a potential neuroprotective therapeutic agent for oxidative-stress-induced injury.⁴⁶ Our study found that maltol significantly increased the viability of RSC96 cells after incubation with H_2O_2 , remarkably decreased the expressions of pro-apop-

totic BAX and caspase-3, and elevated the expression of anti-apoptotic Bcl-XL in a dose-dependent manner.

Studies have reported, however, that maltol (1.2 mM) induced an apoptotic cell death together with iron(II) sulfate (FeSO_4 ; 0.4 mM) in HL60 cells, which may have resulted from the generation of reactive oxygen species by redox cycling, but maltol or FeSO_4 alone did not affect the cells.⁴⁷ Hironishi *et al.* found that maltol induced apoptosis of neuroblastoma cell lines and primary murine fetal hippocampal neurons early in 1996,⁴⁸ and recent studies showed that the cytotoxicity of maltol was not the result of the apoptotic pathway in some human tumor cell lines.⁴⁹ In our study, we found that maltol at concentrations of 0.1 and 0.5 mM protected RSC96 cells from H_2O_2 -induced apoptosis. Thus, we thought that the influence of maltol on cells, (*i.e.*, benefit or toxicity) may lay in its concentration in the cells and the cell types.

4. Conclusion

As a common flavor-enhancing agent and metal ion chelator, maltol also possesses a strong antioxidative activity. The present study first found that repeated treatment with maltol at doses ranging from 50 mg kg^{-1} to 200 mg kg^{-1} , selected according to previous research,^{7,8,50} in STZ-induced diabetic rats produced the following results: (1) significantly enhanced the conduction velocity of the sciatic nerve, (2) improved the thermal and mechanical hyperalgesia, (3) increased Na^+/K^+ -ATPase activity, (4) elevated the antioxidative ability, (5) inhibited the apoptosis of the sciatic nerve in diabetic rats and apoptosis induced by H_2O_2 in RSC96 cells, and (6) decreased the caspase-3 activities in DRG, but (7) had no significant influences on the FBG level and BW. These results suggested that the benefits of maltol on DPN were not attributed to the control of blood glucose but rather to improvements in nerve function through antioxidative stress and apoptosis.

We found maltol to be effective in slowing the progression of DPN without affecting hyperglycemia in STZ-induced dia-



betic rats. These benefits may be associated with an increase in Na⁺-K⁺-ATPase activity, amelioration of oxidative stress, and inhibition of apoptosis. Thus, maltol may provide a new alternative for the treatment of DPN.

Author contributions

N. G. and C. L. designed the entire study, performed most of the experiments and prepared the manuscript. Q. L. and S. L. participated in the measurement of thermal and mechanical thresholds. Y. H. and X. W. took part in monitoring the MNCV. G. B. and M. Y. determined the oxidative stress-related index. S. S. was in charge of the dosages in the animal experiment. C. X. supplied the maltol. Z. S. supplied academic and technical support. Both Z. S. and C. L. are the corresponding authors.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

This work was supported by the CAMS Initiative for Innovative Medicine (CAMS-I2 M) under No. 2016-I2M-2-006 and No. 2017-I2M-1-010. We thank LetPub for its linguistic assistance during the preparation of this manuscript.

References

- 1 M. Brownlee, Biochemistry and molecular cell biology of diabetic complications, *Nature*, 2001, **414**, 813–820.
- 2 E. Agathos, A. Tentolouris, I. Eleftheriadou, P. Katsaouni, I. Nemtzas, A. Petrou, C. Papanikolaou and N. Tentolouris, Effect of α -lipoic acid on symptoms and quality of life in patients with painful diabetic neuropathy. Journal of international medical research, *J. Int. Med. Res.*, 2018, **46**, 1779–1790.
- 3 S. G. Sayyed, A. Kumar and S. S. Sharma, Effects of U83836E on nerve functions, hyperalgesia and oxidative stress in experimental diabetic neuropathy, *Life Sci.*, 2006, **79**, 777–783.
- 4 H. C. Jeong, H. D. Hong, Y. C. Kim, Y. K. Rhee, S. Y. Choi, K. T. Kim, S. S. Kim, Y. C. Lee and C. W. Cho, Quantification of maltol in Korean ginseng (*Panax ginseng*) products by high-performance liquid chromatography-diode array detector, *Pharmacogn. Mag.*, 2015, **11**, 657–664.
- 5 K. G. Lee and T. Shibamoto, Antioxidant properties of aroma compounds isolated from soybeans and mung beans, *J. Agric. Food Chem.*, 2000, **48**, 4290–4293.
- 6 K. S. Kang, H. Y. Kim, J. S. Pyo and T. Yokozawa, Increase in the free radical scavenging activity of ginseng by heat-processing, *Biol. Pharm. Bull.*, 2006, **29**, 750–754.
- 7 K. S. Kang, N. Yamabe, H. Y. Kim and T. Yokozawa, Role of maltol in advanced glycation end products and free radicals: *in vitro* and *in vivo* studies, *J. Pharm. Pharmacol.*, 2008, **60**, 445–452.
- 8 Y. Han, Q. Xu, J. N. Hu, X. Y. Han, W. Li and L. C. Zhao, Maltol, a food flavoring agent, attenuates acute alcohol-induced oxidative damage in mice, *Nutrients*, 2015, **7**, 682–696.
- 9 Y. Song, S. Hong, Y. Iizuka, C. Y. Kim and G. J. Seong, The neuroprotective effect of maltol against oxidative stress on rat retinal neuronal cells, *Korean J. Ophthalmol.*, 2015, **29**, 58–65.
- 10 Y. Yang, J. Wang, C. Xu, H. Pan and Z. Zhang, Maltol inhibits apoptosis of human neuroblastoma cells induced by hydrogen peroxide, *J. Biochem. Mol. Biol.*, 2006, **39**, 145–149.
- 11 X. Yu, L. Zhang, X. Yang, H. Huang, Z. Huang, L. Shi, H. Zhang and G. Du, Salvianolic acid A protects the peripheral nerve function in diabetic rats through regulation of the AMPK-PGC1 α -Sirt3 axis, *Molecules*, 2012, **17**, 11216–11228.
- 12 H. N. Kim, Y. R. Kim, J. Y. Jang, H. K. Shin and B. T. Choi, Electroacupuncture inhibits phosphorylation of spinal phosphatidylinositol 3-kinase/Akt in a carrageenan-induced inflammatory rat model, *Brain Res. Bull.*, 2012, **87**, 199–204.
- 13 S. Kumar, K. H. Arun, C. L. Kaul and S. S. Sharma, Effects of adenosine and adenosine A2A receptor agonist on motor nerve conduction velocity and nerve blood flow in experimental diabetic neuropathy, *Neurol. Res.*, 2005, **27**, 60–66.
- 14 R. Stavniichuk, V. R. Drel, H. Shevalye, Y. Maksimchyk, T. M. Kuchmerovska, J. L. Nadler and I. G. Obrosova, Baicalein alleviates diabetic peripheral neuropathy through inhibition of oxidative-nitrosative stress and p38 MAPK activation, *Exp. Neurol.*, 2011, **230**, 106–113.
- 15 T. Brismar, A. A. Sima and D. A. Greene, Reversible and irreversible nodal dysfunction in diabetic neuropathy, *Ann. Neurol.*, 1987, **21**, 504–507.
- 16 C. G. Jolival, K. E. Frizzi, L. Guernsey, A. Marquez, J. Ochoa, M. Rodriguez and N. A. Calcutt, Peripheral Neuropathy in Mouse Models of Diabetes, *Curr. Protoc. Mouse Biol.*, 2016, **6**, 223–255.
- 17 N. Papanas and D. Ziegler, Efficacy of alpha-lipoic acid in diabetic neuropathy, *Expert Opin. Pharmacother.*, 2014, **15**, 2721–2731.
- 18 I. G. Obrosova, Diabetic painful and insensate neuropathy: pathogenesis and potential treatments, *Neurotherapeutics*, 2009, **6**, 638–647.
- 19 S. O. Kim and H. J. Kim, Berberine ameliorates cold and mechanical allodynia in a rat model of diabetic neuropathy, *J. Med. Food*, 2013, **16**, 511–517.
- 20 J. Han, P. Tan, Z. Li, Y. Wu, C. Li, Y. Wang, B. Wang, S. Zhao and Y. Liu, Fuzi attenuates diabetic neuropathy in rats and protects Schwann cells from apoptosis induced by high glucose, *PLoS One*, 2014, **9**, e86539.
- 21 X. Yang, F. Liu, J. Guo, W. Yao, Q. Li, T. Liu and L. Xu, Antioxidation and anti-inflammatory activity of Tang Bi



- Kang in rats with diabetic peripheral neuropathy, *BMC Complementary Altern. Med.*, 2015, **15**, 66–75.
- 22 E. P. Davidson, L. J. Coppey, B. Dake and M. A. Yorek, Treatment of streptozotocin-induced diabetic rats with alogliptin: effect on vascular and neural complications, *Exp. Diabetes Res.*, 2011, **2011**, 810469.
 - 23 E. P. Davidson, L. J. Coppey, A. Holmes and M. A. Yorek, Effect of inhibition of angiotensin converting enzyme and/or neutral endopeptidase on vascular and neural complications in high fat fed/low dose streptozotocin-diabetic rats, *Eur. J. Pharmacol.*, 2012, **677**, 180–187.
 - 24 N. A. Calcutt, Modeling diabetic sensory neuropathy in rats, *Methods Mol. Med.*, 2004, **99**, 55–65.
 - 25 N. A. Calcutt, M. C. Jorge, T. L. Yaksh and S. R. Chaplan, Tactile allodynia and formalin hyperalgesia in streptozotocin-diabetic rats: effects of insulin, aldose reductase inhibition and lidocaine, *Pain*, 1996, **68**, 293–299.
 - 26 S. Kambiz, J. W. van Neck, S. G. Cosgun, M. H. van Velzen, J. A. Janssen, N. Avazverdi, S. E. Hovius and E. T. Walbeehm, An early diagnostic tool for diabetic peripheral neuropathy in rats, *PLoS One*, 2015, **10**, e0126892.
 - 27 D. Raccach, M. F. Lamotte-Jannot, T. Issautier and P. Vague, Effect of experimental diabetes on Na/K-ATPase activity in red blood cells, peripheral nerve and kidney, *Diabetes Metab.*, 1994, **20**, 271–274.
 - 28 P. Vague, T. C. Coste, M. F. Jannot, D. Raccach and M. Tsimaratos, C-peptide, Na⁺/K⁺-ATPase, and diabetes, *Exp. Diabetes Res.*, 2004, **5**, 37–50.
 - 29 D. Raccach, P. Gallice, J. Pouget and P. Vague, Hypothesis: low Na/K-ATPase activity in the red cell membrane, a potential marker of the predisposition to diabetic neuropathy, *Diabetes Metab.*, 1992, **18**, 236–241.
 - 30 A. M. Schmeichel, J. D. Schmelzer and P. A. Low, Oxidative injury and apoptosis of dorsal root ganglion neurons in chronic experimental diabetic neuropathy, *Diabetes*, 2003, **52**, 165–171.
 - 31 A. M. Vincent, J. W. Russell, P. Low and E. L. Feldman, Oxidative stress in the pathogenesis of diabetic neuropathy, *Endocr. Rev.*, 2004, **25**, 612–628.
 - 32 A. Hosseini and M. Abdollahi, Diabetic Neuropathy and Oxidative Stress: Therapeutic Perspectives, *Oxid. Med. Cell. Longevity*, 2013, **2013**, 168039.
 - 33 M. Rizzardini, M. Lupi, S. Bernasconi, A. Mangolini and L. Cantoni, Mitochondrial dysfunction and death in motor neurons exposed to the glutathione-depleting agent ethacrynic acid, *J. Neurol. Sci.*, 2003, **207**, 51–58.
 - 34 M. S. Kobayashi, D. Han and L. Packer, Antioxidants and herbal extracts protect HT-4 neuronal cells against glutamate-induced cytotoxicity, *Free Radical Res.*, 2000, **32**, 115–124.
 - 35 J. M. McHugh and W. B. McHugh, Diabetes and peripheral sensory neurons: what we don't know and how it can hurt us, *AACN Clin. Issues*, 2004, **15**, 136–149.
 - 36 T. Tosaki, H. Kamiya, Y. Yasuda, K. Naruse, K. Kato, M. Kozakae, N. Nakamura, T. Shibata, Y. Hamada, E. Nakashima, Y. Oiso and J. Nakamura, Reduced NGF secretion by Schwann cells under the high glucose condition decreases neurite outgrowth of DRG neurons, *Exp. Neurol.*, 2008, **213**, 381–387.
 - 37 A. M. Vincent, L. L. McLean, C. Backus and E. L. Feldman, Short-term hyperglycemia produces oxidative damage and apoptosis in neurons, *FASEB J.*, 2005, **19**, 638–640.
 - 38 A. M. Schmeichel, J. D. Schmelzer and P. A. Low, Oxidative injury and apoptosis of dorsal root ganglion neurons in chronic experimental diabetic neuropathy, *Diabetes*, 2003, **52**, 165–171.
 - 39 S. S. Kamboj, R. K. Vasishta and R. Sandhir, N-acetylcysteine inhibits hyperglycemia-induced oxidative stress and apoptosis markers in diabetic neuropathy, *J. Neurochem.*, 2010, **112**, 77–91.
 - 40 S. Murai, J. Matuszkiewicz, Y. Okuzono, H. Miya and R. De Jong, Aurora B Inhibitor TAK-901 Synergizes with BCL-xL Inhibition by Inducing Active BAX in Cancer Cells, *Anticancer Res.*, 2017, **37**, 437–444.
 - 41 K. R. Monk, M. L. Feltri and C. Taveggia, New insights on Schwann cell development, *Glia*, 2015, **63**, 1376–1393.
 - 42 N. P. Gonçalves, C. B. Vægter, H. Andersen, L. Østergaard, N. A. Calcutt and T. S. Jensen, Schwann cell interactions with axons and microvessels in diabetic neuropathy, *Nat. Rev. Neurol.*, 2017, **13**, 135–147.
 - 43 Z. L. Chen, W. M. Yu and S. Strickland, Peripheral regeneration, *Annu. Rev. Neurosci.*, 2007, **30**, 209–233.
 - 44 L. Eckersley, Role of the Schwann cell in diabetic neuropathy. International review of neurobiology, *Int. Rev. Neurobiol.*, 2002, **50**, 293–321.
 - 45 J. Ma, J. Liu, Q. Wang, H. Yu, Y. Chen and L. Xiang, The beneficial effect of ginsenoside Rg1 on Schwann cells subjected to hydrogen peroxide induced oxidative injury, *Int. J. Biol. Sci.*, 2013, **9**, 624–636.
 - 46 S. Hong, Y. Iizuka, T. Lee, C. Y. Kim and G. J. Seong, Neuroprotective and neurite outgrowth effects of maltol on retinal ganglion cells under oxidative stress, *Mol. Vision*, 2014, **20**, 1456–1462.
 - 47 K. Murakami, K. Ishida, K. Watakabe, R. Tsubouchi, M. Naruse and M. Yoshino, Maltol/iron-mediated apoptosis in HL60 cells: participation of reactive oxygen species, *Toxicol. Lett.*, 2006, **161**, 102–107.
 - 48 M. Hironishi, R. Kordek, R. Yanaqihara and R. M. Garruto, Maltol (3-hydroxy-2-methyl-4-pyrone) toxicity in neuroblastoma cell lines and primary murine fetal hippocampal neuronal cultures, *Neurodegeneration*, 1996, **5**, 325–329.
 - 49 E. Yasumoto, K. Nakano, T. Nakayachi, S. R. Morshed, K. Hashimoto, H. Kikuchi, H. Nishikawa, M. Kawase and H. Sakagami, Cytotoxic activity of deferiprone, maltol and related hydroxyketones against human tumor cell lines, *Anticancer Res.*, 2004, **24**, 755–762.
 - 50 Y. B. Kim, S. H. Oh, D. E. Sok and M. R. Kim, Neuroprotective effect of maltol against oxidative stress in brain of mice challenged with kainic acid, *Nutr. Neurosci.*, 2004, **7**, 33–39.

