

Cite this: RSC Adv., 2013, 3, 23146

Received 4th July 2013

Accepted 24th September 2013

DOI: 10.1039/c3ra43417f

www.rsc.org/advances

Introduction

Cabergoline (**1**) is a natural product-based drug, representing a complex, branched amide of 6-*N*-allylated 9,10-dihydrolysergic

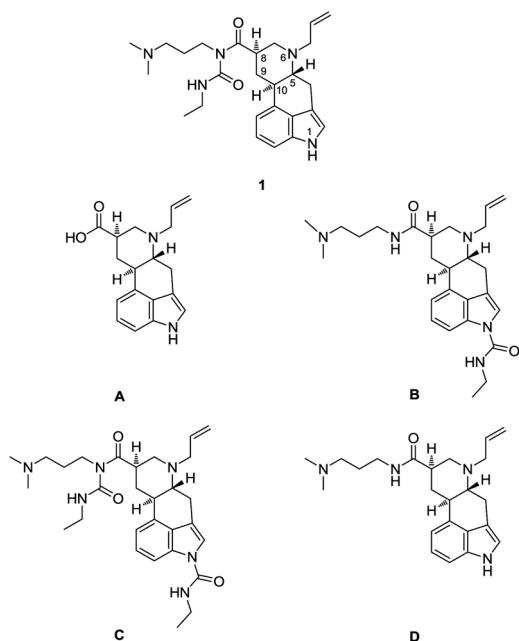


Fig. 1 Cabergoline (**1**) and *Ph. Eur.* impurities of cabergoline **A**, **B**, **C** and **D**.

^aLek d.d., a Sandoz Company, Kolodvorska cesta 27, 1234 Mengš, Slovenia. E-mail: jernej.wagger@sandoz.com; Tel: +386 1 721 7206

^bFaculty of Chemistry and Chemical Technology, University of Ljubljana, Aškerčeva 5, 1000 Ljubljana, Slovenia

^cEN→FIST Centre of Excellence, Dunajska c. 156, 1000 Ljubljana, Slovenia

† Electronic supplementary information (ESI) available: Copies of ¹H and ¹³C NMR spectra of all synthesised compounds. See DOI: 10.1039/c3ra43417f

Synthesis of European pharmacopoeial impurities **A**, **B**, **C**, and **D** of cabergoline†

Jernej Wagger,^{*a} Aljaž Požes^b and Franc Požgan^{bc}

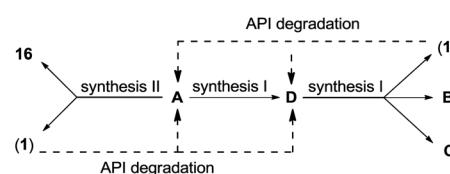
For the use of analytics, European pharmacopoeial impurities **A**, **B**, **C**, and **D** of cabergoline were synthesized. Ergocryptine was chosen as a starting material and synthesis was accomplished *via* two approaches, different in length and stereochemical outcome. A longer, indirect approach was realized through otherwise problematic oxidations of the 9,10-dihydrolysergol derivative, to the corresponding aldehyde and carboxylic acid. This was achieved by the use of activated DMSO and a Pinnick oxidation sequence. All four synthesized impurities are used as analytical standards in cabergoline manufacturing processes.

acid (cabergolinic acid) **A** (Fig. 1). Primarily acting as a dopaminergic D₂ receptor agonist, cabergoline is most widely used for treatment of hyperprolactinaemic disorders and both early and advanced Parkinson's disease.¹ The structure of cabergoline also encodes for different biogenic amines than dopamine, therefore displaying modest pharmacodynamic properties towards adrenergic and serotonergic receptors.¹

One of the fundamental roles of the pharmaceutical industry is to develop new drugs that are safe, effective and of high quality when they reach the patients. With respect to this, delivering an impurity profile of an active pharmaceutical ingredient (API) is a must for fulfilling the aforementioned criteria.² For the purpose of qualifying and/or quantifying the impurity profile of cabergoline (**1**) as the API, we have synthesized four known impurities, qualified by *European Pharmacopoeia (Ph. Eur.)*³ as impurities **A**, **B**, **C** and **D** (Fig. 1).

The genesis of these impurities can be found in the syntheses and other manufacturing process activities of cabergoline (Scheme 1).⁴

Since cabergoline (**1**), with the maximum weekly dosage of 2.0 mg, falls into low dosing drugs, the threshold for identification and qualification of impurities in API is usually 0.10% and 0.15% of mass, respectively.² These limits are low and efficient production processes should not exceed them. Therefore production syntheses are usually neither suitable nor cost



Scheme 1 Possible routes of cabergoline impurities formation. Some impurities (**A**, **D**) can represent syntheses intermediates or are formed *via* hydrolysis of amide bonds from cabergoline (**1**) or other impurities (**B**, **C**).



effective means to produce impurities for the use of analytics. Often, new synthesis routes have to be designed to obtain desired impurities and this was also our task. Synthesis routes for *Ph. Eur.* impurities **A**, **B**, **C** and **D** of cabergoline (**1**) were designed and results are presented in this paper.

Results and discussion

9,10-Dihydrolysergic acid (**2**) was selected as one of the key synthetic intermediates. Although there are several total syntheses of lysergic acid available, through which 9,10-dihydrolysergic acid (**2**) could be prepared, α -ergocryptine (**3**) was chosen as starting material. The reason is that α -ergocryptine (**3**) is biosynthetically produced at our production site and way of preparing 9,10-dihydrolysergic acid (**2**) from α -ergocryptine (**3**) is advantageous over total synthesis because it is shorter, easier and more economical. Also, there is a longstanding tradition of ergot alkaloids chemistry within Sandoz company, pioneered by Albert Hofmann.

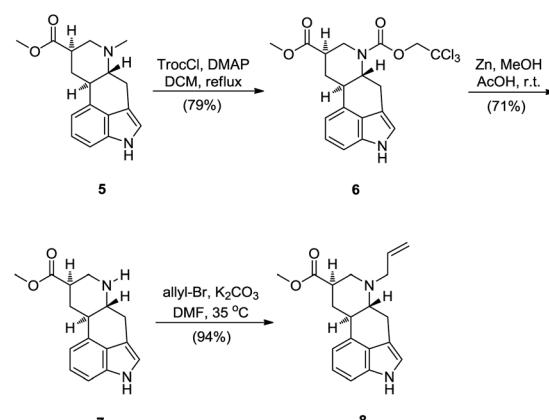
Starting with α -ergocryptine (**3**), the double bond between C9–C10 was diastereoselectively hydrogenated and the reduced ergopeptide **4** was cleaved by alkaline hydrolysis to give 9,10-dihydrolysergic acid (**2**).⁵ To increase solubility of 9,10-dihydrolysergic acid (**2**) in organic solvents, the corresponding methyl ester **5** was prepared (Scheme 2).⁶

In the next steps allylic substituent on 6-*N* of **5** had to be introduced. To introduce the allylic substituent on 6-*N* of **5**, 9,10-dihydrolysergic acid (**2**) had to be *N*-demethylated first. 2,2,2-Trichloroethyl chloroformate (TrocCl) was used, in a transformation mechanistically analogous to the von Braun reaction,⁷ to *N*-demethylate 9,10-dihydrolysergic acid methyl ester (**5**). Subsequent zinc mediated reductive cleavage of 6-*N*-Troc substituted compound **6** gave the desired 6-*N*-demethylated intermediate **7**. This *N*-demethylation sequence was already reported earlier and slight procedure modifications were used in our case.⁸ We have noticed that intermediate **7** decomposes slowly in acetic acid and is photolabile and therefore, the reaction

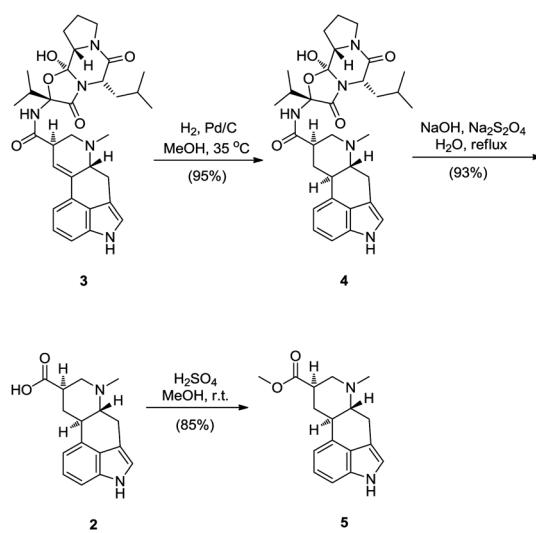
yield depends on the swiftness of work-up and isolation. 6-*N*-Allylation of **7** was then realized by reaction with allyl bromide and potassium carbonate in dimethylformamide, giving allylated intermediate **8** in a high yield (Scheme 3).⁹

At this point, intermediate **8** served us for the synthesis of impurities **A** and **D**. Alkaline hydrolysis of methyl ester **8** gave cabergolinic acid, termed as impurity **A** by *Ph. Eur.*, in 82% yield. The impurity **D** was prepared directly from methyl ester **8** as well. By reacting **8** with 3-(dimethylamino)-1-propylamine in acetic acid, direct amidation of ester gave impurity **D** in 74% yield (Scheme 4).⁴

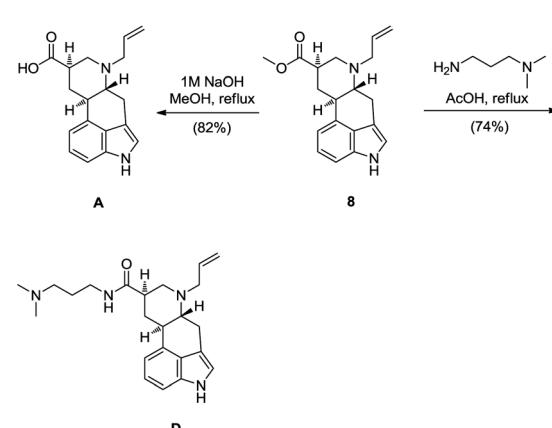
Compared to **A** and **D**, both impurities **B** and **C** have an ethylcarbamoyl functionality on the indole nitrogen. Since such functionalization of indole nitrogen requires the use of base, epimerization at the C8 position of ester **8** was possible. Although the stereocenter at C8 in 9,10-dihydrolysergic acid (**2**) is less prone to epimerization under basic conditions than in lysergic acid, due to the absence of C9–C10 double bond, epimerization does occur.^{10,11} For example, it is known from the literature, that exposing compound **5** to 4 equivalents of NaH in THF at 20 °C, app. 30% of the opposite, thermodynamically less favourable, axial epimer is observed, and by exposing compound **8** to 4 equivalents of LDA in THF at –20 °C, almost



Scheme 3 Synthesis of 6-*N*-allylated 9,10-dihydrolysergic acid methyl ester **8**.



Scheme 2 Synthesis of 9,10-dihydrolysergic acid methyl ester **5**.

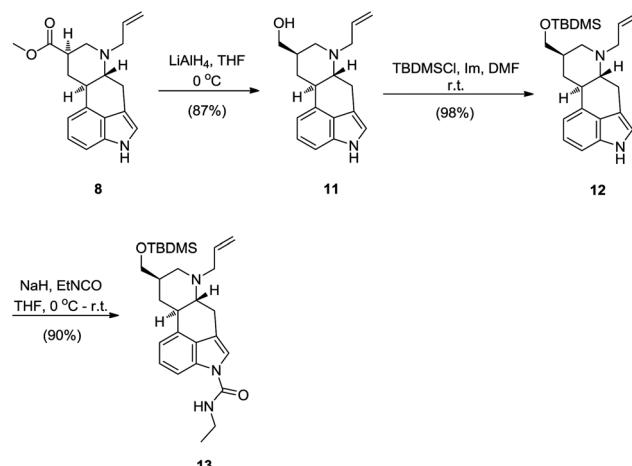


Scheme 4 Synthesis of *Ph. Eur.* impurities **A** and **D** from intermediate **8**.



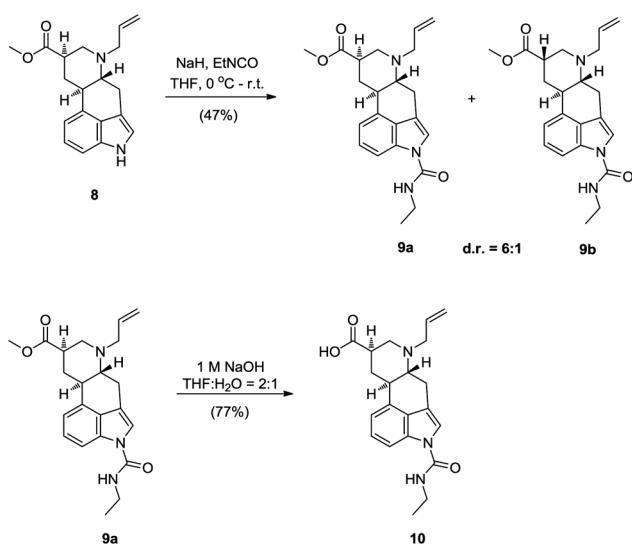
71% of axial epimer is generated.¹¹ Therefore, for installing the ethylcarbamoyl fragment on the indole nitrogen *N*-1 we have envisioned two different approaches. In the first one, a direct functionalization of *N*-1 of **8**, under basic conditions with EtNCO was planned. Being aware of the potential epimerisation problems under these conditions, a second indirect approach was envisioned. In this, indirect approach it was planned to reduce C8 carbonyl group of **8** to corresponding alcohol, do the functionalization of *N*-1 with EtNCO under basic conditions and then reoxidise the alcohol back to the corresponding carboxylic acid. Although longer, this reduction/oxidation approach, in comparison to the first direct one, would ensure us with the epimerisation free preparation of the desired carboxylic acid. Since both approaches for functionalization of *N*-1, direct and indirect one, have advantages and disadvantages, we have decided to test both of them. Subjecting methyl ester **8** to the 1.1 equivalents of NaH and EtNCO afforded the desired epimer **9a** in 6 : 1 diastereomeric ratio and with 47% isolated yield (Scheme 5).¹² With the methyl ester **9a** in hand we were only one step short of the key intermediate 9,10-dihydrolysergic acid **10**, which would enable us to prepare impurities **B** and **C**. Luckily saponification of the methyl ester **9a** proceeded selectively with respect to the *N*-1 ethylcarbamoyl fragment. This has provided us with the desired 9,10-dihydrolysergic acid derivative **10** in 77% yield. We found 9,10-dihydrolysergic acid **10** rather difficult to isolate, since it displays modest solubility in both organic solvents and water, but with acid **10** in hand, the door towards completion of the synthesis plan was open (Scheme 5).

For the indirect approach, methyl ester **8** had to be reduced first. Reacting ester **8** with the LiAlH₄ in THF proceeded smoothly and in high yield to give 9,10-dihydrolysergol derivative **11**.¹³ Hydroxy group in 9,10-dihydrolysergol derivative **11** was protected as TBDSM ether **12** and a carbamoyl moiety was introduced by reacting **12** with sodium hydride and ethyl-isocyanate to give TBDSM-protected 9,10-dihydrolysergol derivative **13** (Scheme 6).



Scheme 6 Synthesis of **13**—introduction of an ethylcarbamoyl fragment onto the indole nitrogen of lysergol derivative **12**.

For the next steps, TBDSM-protected alcohol group in **13** had to be removed and free alcohol reoxidized back to carboxylic acid. Deprotection of silyl ether in **13** with TBAF gave 9,10-dihydrolysergol derivative **14** in very good yield, although some temperature dependent chemoselectivity was observed. When silyl deprotection was performed at 0 °C, slow but clean conversion of **13** to **14** was achieved. On the other hand, when the reaction was done at room temperature, removal of the ethylcarbamoyl group was observed in addition to silyl deprotection. 9,10-Dihydrolysergol derivative **14** was then used for subsequent oxidation reaction. Ideally a one-step oxidation procedure from primary alcohol **14** to the corresponding carboxylic acid would be a method of choice. But surprisingly, while searching the literature, we learned that examples covering oxidations of 9,10-dihydrolysergols and their derivatives to the corresponding 9,10-dihydrolysergols and/or 9,10-dihydrolysergic acids are very scarce. In the most recent literature, Křen *et al.* reported on their efforts to oxidize *N*-protected lysergol derivatives to aldehydes and acids.¹⁴ Beside that, patent literature covers two examples of 9,10-dihydrolysergol oxidations. In the first one BAIB/TEMPO was used for direct oxidation of 9,10-dihydrolysergol derivative to the corresponding 9,10-dihydrolysergic acid.¹⁵ Unfortunately, BAIB/TEMPO mediated oxidation did not work in our case, giving only traces of the desired acid, and failure to oxidize the alcohol **14** is in agreement with the results reported in the literature.¹⁴ In the second literature example, slightly modified Parikh-Doering oxidation (SO₃·TEA, DMSO, TEA) was successfully used to oxidize 9,10-dihydrolysergol to the 9,10-dihydrolysergal,¹⁶ but as reported by Křen, they were unsuccessful in repeating this type of oxidation under standard Parikh-Doering conditions (SO₃·Py, DMSO, TEA).¹⁴ Since 9,10-dihydrolysergol derivatives are poorly soluble in various solvents, we anticipated that DMSO mediated oxidations could be a good choice amongst the plethora of alcohol oxidation methods. DMSO should address the solubility issue of 9,10-dihydrolysergol derivative and concurrently serve as an oxidant. Since contrary reports about effectiveness of the Parikh-Doering oxidation of 9,10-dihydrolysergols were



Scheme 5 Direct synthesis of the key 9,10-dihydrolysergic acid **10**—functionalization of the 9,10-dihydrolysergic acid methyl ester **8**.



available in the literature, we have decided to try other analogues reaction. Luckily one can chose among different DMSO mediated oxidations. Indeed, when alcohol **14** was subjected to Pfitzner–Moffatt oxidation conditions, a clean and high yielding conversion to the corresponding 9,10-dihydrolysergal **15** was observed.¹⁷ Here again temperature dependence on reaction selectivity was observed. Below 5 °C the reaction was clean, giving only aldehyde as a product, while at room temperature or higher several side products could be observed by TLC. Since we were successful with Pfitzner–Moffatt oxidation of **14**, the question was raised, whether the outcome of Parikh–Doering reaction, on the same alcohol substrate would be analogues? Oxidation of the alcohol **14**, under the Parikh–Doering conditions, proceeded smoothly, affording aldehyde **15** in slightly lower yield than *via* Pfitzner–Moffatt reaction (Scheme 7).¹⁸

An added value of this oxidation transformations also lies in the 9,10-dihydrolysergal itself. Knowing that aldehydes of this type are difficult to prepare and that pharmacological activity of synthetically modified ergot alkaloids varies mainly because of the nature and configuration of substituents on C8, such aldehydes also represent valuable medicinal chemistry intermediates.^{4a,16a,19}

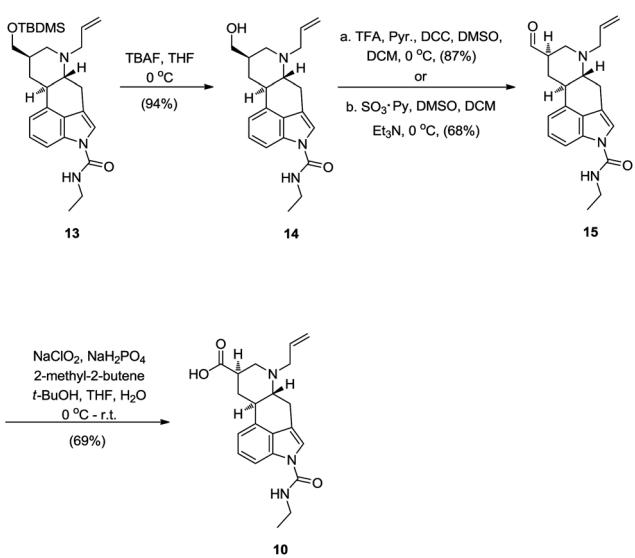
Oxidations of 9,10-dihydrolysergals to the corresponding 9,10-dihydrolysergic acids are hardly reported in the literature. This is mainly due to the fact that 9,10-dihydrolysergic acid derivatives are easily accessible from naturally occurring lysergic acid. The literature reports on oxidation of 9,10-dihydrolysergal with Tollens's reagent or MCPBA, but both methods have their limitations.¹⁴ The first one unfortunately gives poor yields while the second one suffers from chemoselectivity. Peroxy acid oxidizes not only the aldehyde, but also the nitrogen on position 6 to the corresponding *N*-oxide. Although reduction of *N*-oxide to the desired 9,10-dihydrolysergic acid was demonstrated,¹⁴ this method suffers from functional group compatibility, since allylated lysergal derivatives are not tolerated by peroxy acids nor

catalytic hydrogenation. On the other hand there are some examples of lysergal oxidations to lysergic acid derivatives. In the final steps of total synthesis of lysergic acid by Fujii and Ohno, Pinnick oxidation was used to oxidize 1-N-6-N-ditosylated iso-lysergal into the corresponding lysergic acid.²⁰ Indeed, upon subjecting aldehyde **15** to Pinnick oxidation, the desired 9,10-dihydrolysergic acid **10** was obtained, in 69% yield (Scheme 7).²¹

9,10-Dihydrolysergic acid **10** was then used to synthesize both impurities **B** and **C**. The coupling of **10** with 3-(dimethylamino)-1-propylamine by the use of CDI furnished the impurity **B** in 54% yield. For the synthesis of impurity **C** a more complex *N*-acylurea appendage on C8 had to be installed. In the original synthesis of cabergoline, this side chain was installed with the use of EDC·HCl, by exploiting the usually undesired ability of carbodiimide coupling reagents to form *N*-acylureas as intramolecular rearrangement side products.²² Reacting cabergolinic acid **A** with EDC·HCl, in the absence of an external nucleophile, Mantegani and co-workers synthesized cabergoline (**1**).⁴ Although the use of EDC·HCl gives the desired *N*-acylurea appendage in one step, a disadvantage of using this asymmetric carbodiimide is concomitant formation of regioisomeric *N*-acylurea and tedious separation of both isomers.

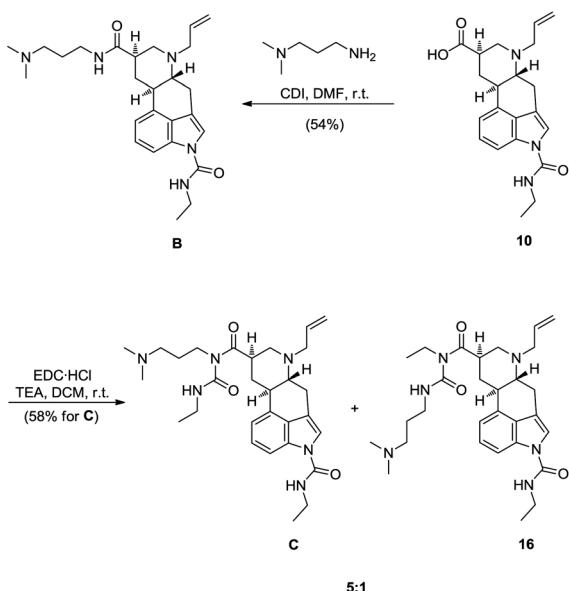
To compensate for disadvantage of applying EDC·HCl, different synthetic strategy was utilized in which excess ethylisocyanate was reacted with compound **D** to install *N*-acylurea appendage. Unfortunately the use of ethylisocyanate is also the root cause for formation of impurities **B** and **C** as side products in cabergoline (**1**) synthesis (Scheme 1).⁴ Other methods for *N*-acylurea installation in cabergoline (**1**) synthesis were also developed, although none of them introduces *N*-acylurea appendage in one step from the corresponding 9,10-dihydrolysergic acid.²³ Therefore, compromising lower yield, we have used EDC·HCl in DCM²⁴ for the synthesis of impurity **C**, thus preparing the desired impurity and its regioisomer **16** in ratio 5 : 1 respectively.²⁵ After separation of regioisomers, the impurity **C** was obtained in 58% yield (Scheme 8).

For the synthesis of **C**, another synthetic strategy would be viable, by following linear route **10** → **B** → **C**. By reacting **B** with an excess of EtNCO, impurity **C** could be prepared. In the original synthesis of cabergoline (**1**) up to 40 equivalents of EtNCO were reacted with compound **D** to get the desired API. The amount of EtNCO was later reduced in consequent synthesis optimizations, by the use of CuI and Ph₃P catalysis, to as low as 3 equivalents. Under these, optimised conditions cabergoline (**1**) was prepared along with the impurities **B** and **C** in the ratio of 82 : 6 : 12 respectively.²⁶ As mentioned earlier, other syntheses of cabergoline (**1**) were developed to avoid the use of toxic EtNCO and they are all based on the functionalization of the carboxamide side chain of **D**.²³ In our case the impurity **B** was prepared from precursor **10** in a modest 54% yield. Therefore, if the reaction of **B** with EtNCO, or any other suitable reagent, would work quantitatively, the impurity **C**, starting linearly from **10**, would be prepared in not greater than 54% yield. Following this, we have never tried to prepare impurity **C** from **10** *via* impurity **B**. Instead, the impurity **C** was prepared in one step, directly from carboxylic acid **10** with 58% yield (Scheme 8).



Scheme 7 Indirect, oxidative route to the 9,10-dihydrolysergic acid **10**.





Scheme 8 Synthesis of *Ph. Eur.* impurities **B** and **C**.

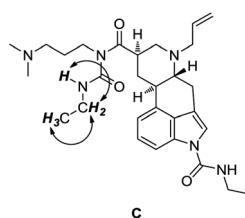


Fig. 2 Detected COSY interactions in ethylamido side chain (CH_3 , 1.18 ppm; CH_2 , 3.30 ppm; NH , 9.45 ppm) in impurity **C** are indicated by arrows.

The structure of proper *N*-acylurea appendage in the impurity **C** was determined using 2D COSY NMR technique, where expected interactions between ethylamido protons were observed (Fig. 2). The proton NMR of the impurity **C** is also in accordance with the previous report of Mantegani and co-workers.⁴

Conclusions

A synthesis of four European pharmacopoeial impurities **A**, **B**, **C** and **D** of cabergoline (**1**) was demonstrated and the synthesized impurities are used as HPLC analytical standards. To the best of our knowledge, this is the first report on selective synthesis of these impurities. From α -ergocryptine (**3**), 9,10-dihydrolysergic acid methyl ester (**5**) was prepared, which served as the key starting material. Using **5**, *N*-1 ethylcarbamoyl 9,10-dihydrolysergic acid **10** was prepared *via* two different routes. First route, a direct one, proceeds through direct *N*-1 ethylcarbamoylation of **10**, causing partial epimerisation at C8 position. In spite of epimerisation, this route is shorter and therefore advantageous over the indirect one. Second route, an indirect one, follows reduction-oxidation of 9,10-dihydrolysergic acid derivative **8**, for epimerisation free installation of *N*-1 ethylcarbamoyl moiety. Notwithstanding the fact that oxidations of 9,10-dihydrolysergol derivatives to the

corresponding 9,10-dihydrolysergals and further on to 9,10-dihydrolysergic acids are very rare, we have shown that by combination of activated DMSO and Pinnick reactions this oxidation sequence can be achieved successfully. Furthermore, such oxidation method of preparing otherwise scarcely presented 9,10-dihydrolysergals, can be useful in medicinal chemistry, for preparing novel pharmacologically active C8-substituted derivatives of ergoline.

Experimental

General experimental details

Where stated, anhydrous reactions were carried out in vacuum oven dried glassware, under stream of nitrogen, passed through Drierite™ drying column. Anhydrous solvents were purchased from Sigma-Aldrich and all reagents were commercial grade, used without any purification. Exception was α -ergocryptine which was acquired through in-house biofermentation process and was purified by crystallization from toluene. Reactions were monitored by thin layer chromatography (TLC) performed on pre-coated aluminum-baked silica gel (60 F₂₅₄, Merck) or aluminum oxide (60 F₂₅₄, Merck) plates. Developed TLC plates were visualized under UV light and/or by aqueous cerium ammonium molybdate and aqueous potassium permanganate. Flash chromatography was performed on ZEOPrep 60 Eco 40–63 μm silica gel and aluminum oxide activated, basic, with indicated mobile phase. Lower-temperature reactions were performed on ice/water bath or on Mettler-Toledo EasyMax™ synthesis workstation. Infrared spectra were recorded on a Bruker Alpha (Platinum ATR) FT-IR spectrometer and are reported in reciprocal centimeters (cm^{-1}). Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker Avance III 500 and Bruker Avance DPX 300 spectrometers. Chemical shifts for ¹H NMR spectra are reported in parts per million from tetramethylsilane, with the solvent resonance as the internal standard (CDCl_3 , δ 7.26 ppm, $\text{DMSO-}d_6$, δ 2.50 ppm). Data are reported as follows: chemical shift, multiplicity, (s = singlet, d = doublet, t = triplet, q = quartet, qn = quintet, m = multiplet, br = broad), coupling constant in Hz and integration. Chemical shifts for ¹³C NMR spectra are reported in parts per million from tetramethylsilane, using solvent central peak as the internal standard (CDCl_3 , δ 77.16 ppm, $\text{DMSO-}d_6$, δ 39.52 ppm). Where stated 2D NMR spectra were recorded. For pH measurements of water solutions, Seven Easy Mettler-Toledo pH meter was used. Optical rotations were determined at 589 nm at ambient temperature on Perkin-Elmer 2400 241MC polarimeter. Data reported as follows: $[\alpha]_D$, concentration (c in g per 100 mL) and solvent. Elemental microanalyses were performed on Perkin-Elmer Analyser 2400. Mass spectra were obtained using an AutoSpecQ, Q-TOF Premier and Agilent 6224 Accurate TOF LC/MS spectrometers. Melting points were determined on a Kofler micro hot stage and are uncorrected.

9,10-Dihydro- α -ergocryptine (4). In a 1000 mL round-bottom flask α -ergocryptine (**3**) was weighed (30 g, 52 mmol) and dissolved in MeOH (500 mL) at 35 °C. Then, while stirring, Pd (3.9 g, 10% on carbon) suspended in MeOH (143 mL) was added and reaction mixture was purged with N_2 . After that reaction



mixture was evacuated, purged with H_2 and put under H_2 atmosphere (balloon). Reaction mixture was stirred at 35 °C for 3 h and after that it was purged with N_2 and heated to 55 °C. At 55 °C the Pd catalyst was filtered off and filtrate was ran through aluminum oxide (30 g, basic) column and column was washed with MeOH (2 × 200 mL) heated to 55 °C. Combined MeOH fractions were evaporated at reduced pressure to dryness and resuspended in MeOH (450 mL). Suspension was heated to 55 °C and while stirring, H_2O (900 mL) was added. Stirred suspension was cooled to ambient temperature and further to 0 °C, at which it was stirred for 0.5 h. Precipitate, was filtered off and dried in vacuum to give the titled compound as an off white solid (28.5 g, 95%). Optionally, product could be further crystallized from MeOH. R_f : 0.33 (EtOAc–MeOH = 90% : 10%); mp = 224–226 °C, ref. 5a = 235 °C; $[\alpha]_D$ –30.1 (c 0.53; pyr.), ref. 5a $[\alpha]_D$ –41 (Pyr.); IR: ν_{max} 3627, 3369, 3354, 3198, 3064, 2955, 2847, 2792, 1720, 1657, 1626, 1546, 1432, 1218, 1191, 1003, 755 cm^{-1} ; 1H NMR (DMSO- d_6 , 500 MHz) δ 0.86 (d, J = 6.7 Hz, 3H); 0.90 (d, J = 6.8 Hz, 3H); 0.94 (d, J = 6.5 Hz, 3H); 1.05 (d, J = 6.7 Hz, 3H); 1.40 (q, J = 12.4 Hz, 1H); 1.64–1.82 (m, 3H); 1.85–2.05 (m, 5H); 2.12 (dt, J_1 = 13.5 Hz, J_2 = 6.7 Hz; 1H); 2.22 (t, J = 12.2 Hz, 1H); 2.36 (s, 3H); 2.48–2.54 (m, 1H); 2.63 (bd, J = 12.2 Hz, 1H); 2.78–2.84 (m, 1H); 2.94–3.02 (m, 2H), 3.30 (dd, J_1 = 14.7 Hz, J_2 = 4.1 Hz, 1H); 3.35–3.41 (m, 2H); 3.72 (dd, J_1 = 8.4 Hz, J_2 = 5.9 Hz, 1H); 4.32 (dd, J_1 = 7.6 Hz, J_2 = 5.8 Hz, 1H); 6.74 (d, J = 7.02 Hz, 1H); 6.98 (bs, 1H); 7.02 (t, J = 7.9 Hz, 1H); 7.13 (d, J = 8.1 Hz, 1H); 7.57 (d, J = 1.58 Hz, 1H); 9.05 (s, 1H); 10.66 (d, J = 1.55 Hz, 1H). ^{13}C NMR (DMSO- d_6 , 125 MHz) δ 15.7, 16.6, 21.5, 22.2, 22.7, 24.3, 26.0, 26.6, 30.5, 33.0, 39.4, 41.8, 42.6, 43.0, 45.7, 52.4, 58.8, 63.5, 66.5, 90.15, 103.3, 108.9, 110.0, 111.7, 118.6, 122.1, 125.9, 132.16, 133.20, 164.9, 165.2, 177.1; HRMS (ESI): m/z calcd for $C_{32}H_{44}N_5O_5$: [M + H]⁺, 578.3337; found: 578.3337.

(6aR,10aR)-7-Methyl-4,6,6a,7,8,9,10,10a-octahydroindolo-[4,3-fg]quinoline-9-carboxylic acid (2). To NaOH (8.28 g, 207 mmol) in 250 mL flask, H_2O (96 mL) and $Na_2S_2O_4$ (1.2 g, 6.89 mmol) were added and solution was formed at room temperature. While stirring, 9,10-dihydro- α -ergocryptine (4) (20 g, 34.6 mmol) was suspended in the solution and reaction mixture was refluxed for 4 h. After that, dark brown solution was cooled to room temperature and acidified with AcOH to pH 6.2. Suspension formed was cooled bellow 15 °C and stirred for 2 h. Suspension was filtered off, thoroughly washed with H_2O and dried to give 9,10-dihydrolysergic acid (2) as a light grayish solid (8.7 g, 93%). R_f : 0.58 (THF–AcOH– H_2O = 75% : 13% : 12%); mp = decomposition at 250 °C, ref. 5a compound darkens at 250 °C and decomposes at 300 °C; $[\alpha]_D$ –120 (c 0.1; pyr.), ref. 5a $[\alpha]_D$ –122 (c 0.2; Pyr.); IR: ν_{max} 3370, 3240, 3194, 3117, 3048, 2948, 2852, 2348, 1577, 1445, 1390, 1345, 1090, 1029, 1007, 764 cm^{-1} ; 1H NMR (DMSO- d_6 , 500 MHz) δ 1.34 (q, J = 12.9 Hz, 1H); 2.00 (dt, J_1 = 10.9 Hz, J_2 = 4.1 Hz, 1H); 2.17 (t, J = 11.5 Hz, 1H); 2.37 (s, 3H); 2.48–2.55 (m, 1H); 2.70–2.85 (m, 3H); 3.12 (dd, J_1 = 11.0 Hz, J_2 = 2.1 Hz, 1H); 3.29 (dd, J_1 = 14.6 Hz, J_2 = 4.1 Hz, 1H); 6.79 (d, J = 7.1 Hz, 1H); 6.98 (bs, 1H); 7.01 (t, J = 7.6 Hz, 1H); 7.13 (d, J = 8.1 Hz, 1H); 10.65 (s, 1H). ^{13}C NMR (DMSO- d_6 , 125 MHz) δ 26.5, 30.5, 40.9, 42.6, 58.6, 66.6, 108.8, 110.0, 112.0, 118.6, 122.1, 125.9, 132.3, 133.2, 175.1; HRMS (ESI): m/z calcd for $C_{16}H_{19}N_2O_2$: [M + H]⁺, 271.1441; found: 271.1443.

Methyl (6aR,10aR)-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline-9-carboxylate (5). To 9,10-dihydrolysergic acid (2) (7.00 g, 25.9 mmol), MeOH (125 mL) was added and suspension was cooled to ~3 °C. Then slowly, concentrated H_2SO_4 (6.7 mL) was added to the suspension. After addition of H_2SO_4 , reaction mixture was allowed to warm up to room temperature and it was stirred for additional 15 h. After that MeOH (58 mL) was partly evaporated, H_2O (226 mL) was added and the solution was basified with 25% aqueous solution of NH_4OH to pH 8.80. Suspension formed, was stirred at room temperature for 2 h, after which it was filtered, washed with H_2O and dried to give titled compound as an off white solid (6.25 g, 85%). R_f : 0.45 (EtOAc–MeOH = 9 : 1); mp = 189–190 °C; $[\alpha]_D$ –66.1 (c 0.98; DCM); IR: ν_{max} 3142, 3094, 3002, 2947, 2864, 2834, 2804, 1729, 1608, 1443, 1238, 1096, 1025, 745 cm^{-1} ; 1H NMR (CDCl₃, 500 MHz) δ 1.56–1.66 (m, 1H); 2.21 (ddd, J_1 = 11.1 Hz, J_2 = 9.4 Hz, J_3 = 4.3 Hz, 1H); 2.34–2.42 (m, 1H); 2.52 (s, 3H); 2.66–2.75 (m, 1H); 2.94–3.04 (m, 3H); 3.26–3.32 (m, 1H); 3.42 (dd, J_1 = 14.7 Hz, J_2 = 4.3 Hz, 1H); 3.75 (s, 3H); 6.89 (t, J = 1.6 Hz, 1H); 6.95–6.98 (m, 1H); 7.16–7.21 (m, 2H); 8.07 (s, 1H). ^{13}C NMR (CDCl₃, 125 MHz) δ 27.0, 30.7, 40.3, 41.6, 43.2, 51.9, 58.8, 66.8, 108.8, 111.9, 113.4, 118.0, 123.3, 126.2, 132.8, 133.5, 174.5; HRMS (ESI): m/z calcd for $C_{17}H_{21}N_2O_2$: [M + H]⁺, 285.1598; found: 285.1595.

9-Methyl 7-(2,2,2-trichloroethyl) (6aR,10aR)-6,6a,8,-9,10,-10a-hexahydroindolo[4,3-fg]quinoline-7,9(4H)-dicarboxylate (6). To an oven dried glassware, dried compound 5 (8.51 g, 29.9 mmol) was weighed and anhydrous DCM (200 mL) was added under N_2 atmosphere. Mixture was stirred and TrocCl (18.7 mL, 136 mmol) and DMAP (0.162 g, 1.33 mmol) were added. Reaction mixture was heated to reflux and stirred for 16 h. After indicated time reaction mixture was cooled to room temperature, filtered and filtrate was concentrated under reduced pressure. Residue on evaporation was purified by column chromatography (EtOAc) to give titled compound as a white solid (10.54 g, 79%). R_f : 0.89 (EtOAc); mp = 211 °C, ref. 7 208–210 °C; $[\alpha]_D$ –60.8 (c 0.53; DCM); IR: ν_{max} 3382, 2955, 2847, 1717, 1598, 1436, 1374, 1266, 1187, 1146, 1030, 812, 756, 719 cm^{-1} ; 1H NMR (DMSO- d_6 , 500 MHz) δ 1.67 (m, 1H), 2.74–2.81 (m, 1H); 3.00–3.08 (m, 2H); 3.23–3.33 (m, 2H); 3.63–3.69 (m, 1H); 3.64 (s, 3H); 3.77 (dd, J_1 = 14.2, J_2 = 6.2 Hz, 1H); 4.05 (dd, J_1 = 14.2, J_2 = 2.6 Hz, 1H); 4.84 (d, J = 12.4 Hz, 1H); 4.87 (d, J = 12.4 Hz, 1H); 6.81 (d, J = 7.1 Hz, 1H); 7.00 (bs, 1H); 7.03 (m, 1H); 7.17 (d, J = 8.1 Hz, 1H); 10.73 (d, J = 1.5 Hz, 1H). ^{13}C NMR (DMSO- d_6 , 125 MHz) δ 25.2, 27.1, 36.2, 39.2, 41.8, 51.9, 60.1, 74.1, 96.0, 109.4, 110.2, 112.1, 118.9, 122.0, 125.9, 131.9, 133.4, 153.5, 174.2; HRMS (ESI): m/z calcd for $C_{19}H_{20}Cl_3N_2O_4$: [M + H]⁺, 445.0483; found: 445.0477.

Methyl (6aR,10aR)-4,6,6a,7,8,9,10,10a-octahydroindolo-[4,3-fg]quinoline-9-carboxylate (7). Compound 6 (20 g, 44.9 mmol) was suspended in AcOH (885 mL) and MeOH (140 mL) was added. Suspension was stirred vigorously at room temperature and Zn (20 g, 306 mmol) was added. After 65 min Zn was filtered off and filtrate was concentrated under reduced pressure. To the residue on concentration H_2O (500 mL) was added, suspension was stirred for 0.5 h and filtered. Filtrate was then basified with saturated aqueous NH_4OH to pH 8.1, DCM (400 mL) was added



and suspension was then acidified with glacial AcOH back to pH 6.6. Phases were separated and water phase was again basified with saturated aqueous NH_4OH to pH 8.1, DCM (200 mL) was added and pH was adjusted back to pH 6.6 with glacial AcOH. Phases were separated and combined organic phases were dried over MgSO_4 , concentrated under reduced pressure to give white solid that was suspended in heptane (225 mL), stirred for 1 h and filtered. White solid was washed with heptane and dried to give titled compound (8.6 g, 71%). R_f : 0.40 (AcOH-heptane = 1 : 2); mp = 173–175 °C, ref. 27 172 °C; $[\alpha]_D$ –24 (c 1.33; DCM); IR: ν_{max} 3402, 3261, 3144, 3093, 3003, 2946, 2861, 2801, 1728, 1601, 1548, 1517, 1435, 1248, 1234, 1199, 1109, 903, 732 cm^{-1} ; ^1H NMR (CDCl₃, 500 MHz) δ 1.63–1.72 (m, 1H); 2.74–2.92 (m, 6H); 2.96–3.02 (m, 1H); 3.05–3.12 (m, 1H); 3.46–3.52 (m, 1H); 3.74 (s, 3H); 6.88 (s, 1H); 6.94 (d, J = 6.7 Hz, 1H); 7.15–7.22 (m, 2H); 8.03 (bs, 1H). ^{13}C NMR (CDCl₃, 125 MHz) δ 29.6, 30.8, 41.3, 43.0, 48.4, 52.0, 59.6, 109.0, 111.8, 113.3, 117.8, 123.2, 126.6, 132.5, 133.6, 174.4; HRMS (ESI): m/z calcd for C₁₈H₂₁N₂O₂: [M + H]⁺, 297.1598; found: 297.1595.

Methyl (6aR,9R,10aR)-7-allyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline-9-carboxylate (8). Compound 7 (8.00 g, 29.6 mmol) was dissolved in DMF (58 mL), while stirring K_2CO_3 (8.17 g, 59.2 mmol) and allyl bromide (2.97 mL, 34.3 mmol) were added. The reaction mixture was stirred at 35 °C for 2.5 h. Then reaction mixture was filtered and EtOAc (350 mL) was added. Organic phase was washed with water (4 × 500 mL) and combined water phases were reextracted with EtOAc (150 mL). Combined organic phases were dried over Na_2SO_4 , concentrated under reduced pressure and dried to give titled compound as a white solid (8.68 g, 94%). R_f : 0.84 (EtOAc-EtOH = 88 : 12); mp = 146–149 °C, ref. 10 143–146 °C; $[\alpha]_D$ –117.1 (c 1.0; DCM), ref. 10 $[\alpha]_D$ –117.2; IR: ν_{max} 3402, 3147, 2948, 2838, 1729, 1641, 1610, 1547, 1443, 1365, 1201, 751 cm^{-1} ; ^1H NMR (CDCl₃, 500 MHz) δ 1.63–1.73 (m, 1H); 2.49 (t, J = 11.6 Hz, 1H); 2.54 (td, J_1 = 10.1 Hz, J_2 = 4.8 Hz, 1H); 2.75 (bt, J = 13.0 Hz, 1H); 2.90–3.05 (m, 3H); 3.34 (d, J = 11 Hz, 1H); 3.38–3.49 (m, 2H); 3.60 (dd, J_1 = 8.6 Hz, J_2 = 5.8 Hz, 1H); 3.75 (s, 3H); 5.23 (d, J = 10.2 Hz, 1H); 5.29 (d, J = 17.1 Hz, 1H); 5.99 (m, 1H); 6.89 (s, 1H); 6.95 (d, J = 6.3 Hz, 1H); 7.15–7.21 (m, 2H); 8.07 (s, 1H). ^{13}C NMR (CDCl₃, 125 MHz) δ 26.7, 30.8, 40.5, 41.7, 51.9, 54.8, 56.5, 63.7, 108.8, 111.9, 113.4, 118.0, 118.6, 123.3, 126.2, 132.9, 133.4, 133.8, 174.7; HRMS (ESI): m/z calcd for C₁₉H₂₃N₂O₂: [M + H]⁺, 311.1754; found: 311.1753.

(6aR,9R,10aR)-7-Allyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline-9-carboxylic acid A. Compound 8 (0.5 g, 1.6 mmol) was dissolved in MeOH (15 mL) and aqueous 1 M NaOH solution (4 mL) was added. Reaction mixture was refluxed for 45 min. The reaction mixture was concentrated under reduced pressure and residue was dissolved in H₂O (10 mL). Water solution was then acidified with AcOH to pH 5.2 and precipitate was stirred at ~3 °C (ice/water bath) for 3 h. Precipitate was then filtered off and washed with water to give impurity A as a white solid (0.388 g, 82%). R_f : 0.58 (THF-H₂O-AcOH = 98 : 1 : 1); mp = 246–247 °C; $[\alpha]_D$ –138.4 (c 0.72; DMSO); IR: ν_{max} 3208, 2942, 2322, 1650, 1589, 1448, 1365, 1337, 1283, 952, 757 cm^{-1} ; ^1H NMR (DMSO-*d*₆, 500 MHz) δ 1.31–1.41 (m, 1H); 2.30 (t, J = 11.5 Hz, 1H); 2.29–2.37 (m, 1H); 2.56 (bt, J = 12.8 Hz, 1H); 2.66–2.75 (m, 1H); 2.78–2.86 (m,

2H); 3.16–3.21 (m, 1H); 3.27 (dd, J_1 = 14.6 Hz, J_2 = 7.4 Hz, 1H); 3.36 (dd, J_1 = 14.6 Hz, J_2 = 3.9 Hz, 1H); 3.51 (dd, J_1 14.6 Hz, J_2 = 5.5 Hz, 1H); 5.17 (d, J = 10.2 Hz, 1H); 5.26 (d, J = 17.0 Hz, 1H); 5.90–6.00 (m, 1H); 6.79 (d, J = 7.1 Hz, 1H); 6.98 (s, 1H); 7.02 (t, J = 7.6 Hz, 1H); 7.14 (d, J = 8.1 Hz, 1H); 10.65 (s, 1H). ^{13}C NMR (DMSO-*d*₆, 125 MHz) δ 26.2, 30.6, 39.9, 40.9, 54.7, 55.6, 63.5, 108.8, 110.1, 112.1, 117.9, 118.6, 122.1, 125.9, 132.4, 133.2, 134.7, 175.2; HRMS (ESI): m/z calcd for C₁₈H₂₁N₂O₂: [M + H]⁺, 297.1598; found: 297.1595.

(6aR,9R,10aR)-7-Allyl-N-[3-(dimethylamino)propyl]-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline-9-carboxamide D. Compound 8 (0.500 g, 1.6 mmol) was dissolved in 3-dimethylamino-1-propylamine (3 mL) and glacial acetic acid was added (0.1 mL) and was stirred for 24 h at reflux temperature. Then the reaction mixture was cooled to room temperature and concentrated under reduced pressure to an oily brown residue. To the residue, EtOAc (100 mL) was added and organic phase was washed with 10% aqueous NaHCO₃ (30 mL) and phases were separated. Aqueous phase was reextracted with EtOAc (2 × 75 mL) and combined organic phases were washed with brine (2 × 50 mL). Organic phase was dried over anhydrous MgSO_4 , concentrated under reduced pressure and residue was crystallized from acetone to give titled compound as a light yellowish solid (450 mg, 74%). R_f : 0.18 (EtOAc-EtOH = 88 : 12); mp = 201–203 °C, ref. 4a 200–202 °C; $[\alpha]_D$ –63.0 (c 0.87; DCM); IR: ν_{max} 3365, 3296, 3067, 2941, 2795, 2719, 1623, 1545, 1443, 1417, 1292, 911, 739 cm^{-1} ; ^1H NMR (CDCl₃, 500 MHz) δ 1.60–1.71 (m, 3H); 2.26 (s, 6H), 2.41–2.45 (m, 2H); 2.48 (t, J = 11.5 Hz, 1H); 2.49–2.56 (m, 1H); 2.62–2.68 (m, 1H); 2.68–2.75 (m, 1H), 2.79–2.86 (m, 1H); 2.94–3.02 (m, 1H); 3.21–3.26 (m, 1H); 3.36–3.43 (m, 3H); 3.46 (dd, J_1 = 14.5 Hz, J_2 = 4.1 Hz, 1H); 3.58 (dd, J_1 = 14.4 Hz, J_2 = 5.9 Hz, 1H); 5.19 (d, J = 10.1, 1H); 5.25 (dd, J_1 = 17.1 Hz, J_2 = 0.8 Hz, 1H); 5.92–6.03 (m, 1H); 6.87–6.93 (m, 2H); 7.13–7.20 (m, 2H); 7.64 (bs, 1H); 8.19 (s, 1H). ^{13}C NMR (CDCl₃, 125 MHz) δ 25.5, 26.8, 31.1, 40.1, 40.7, 43.8, 45.6, 45.7, 55.8, 56.7, 59.4, 63.7, 108.8, 111.8, 113.2, 118.0, 118.7, 123.2, 126.2, 133.0, 133.4, 133.7, 173.7; HRMS (ESI): m/z calcd for C₂₃H₃₃N₄O: [M + H]⁺, 381.2649; found: 381.2647.

(6aR,9R,10aR)-Methyl 7-allyl-4-(ethylcarbamoyl)-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline-9-carboxylate (9a). Compound 8 (0.6 g, 1.90 mmol) in an oven dried round-bottom flask was dissolved in anhydrous THF (16 mL) under N₂ atmosphere and cooled to ~3 °C (ice/water bath). To the solution NaH (83.6 mg, 60% in mineral oil, 2.1 mmol) was added. Mixture was stirred for additional 10 min and EtNCO (0.225 mL, 2.8 mmol) was added. Reaction mixture was warmed to room temperature and stirred for 3 h. After 3 h reaction mixture was cooled to ~3 °C and quenched with brine (1 mL). Suspension was concentrated under reduced pressure and residue was suspended in DCM (100 mL). Water (75 mL) was added and phases were separated. Water phase was reextracted with ethyl acetate (40 mL) and combined organic phases were dried over MgSO_4 and concentrated in vacuum.¹² Residue on evaporation was purified using column chromatography to give title compound as a white solid (0.341 g, 47%). R_f : 0.18 (ethyl acetate-heptane = 1 : 1); mp = 176–178 °C; $[\alpha]_D$ –126.2 (c 0.16; DMF); IR: ν_{max} 3316, 3068, 3021, 2977, 2948, 2858, 2789, 1731, 1666, 1604, 1537, 1436, 1341, 1274, 1160,



1116, 1073, 929, 752, 635 cm^{-1} ; ^1H NMR (DMSO- d_6 , 500 MHz) δ 1.15 (t, J = 7.2 Hz, 1H); 1.32–1.42 (m, 1H); 2.28–2.36 (m, 2H); 2.48–2.56 (m, 1H); 2.77–2.86 (m, 3H); 3.15–3.21 (m, 1H); 3.24–3.30 (m, 3H); 3.31–3.36 (m, 1H); 3.48–3.55 (m, 1H); 3.65 (s, 3H); 5.18 (d, J = 10.3 Hz, 1H); 5.27 (d, J = 17.0 Hz, 1H); 5.87–6.00 (m, 1H); 7.00 (d, J = 7.4 Hz, 1H); 7.19 (t, J = 7.7 Hz, 1H); 7.52 (d, J = 1.3 Hz, 1H); 7.78 (d, J = 8.2 Hz, 1H); 7.99 (t, J = 5.4 Hz, 1H). ^{13}C NMR (DMSO- d_6 , 125 MHz) δ 15.0, 25.7, 30.3, 34.9, 40.6, 51.6, 54.3, 55.4, 62.8, 112.9, 115.1, 115.9, 117.9, 118.1, 124.7, 127.1, 132.8, 133.1, 134.6, 151.8, 173.7; HRMS (ESI): m/z calcd for $\text{C}_{22}\text{H}_{28}\text{N}_3\text{O}_3$: [M + H]⁺: 382.2125; 382.2118.

(6aR,9R,10aR)-7-Allyl-4-(ethylcarbamoyl)-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinolone-9-carboxylic acid (10)

(a) *Via saponification of 9a.* In a round-bottom flask compound 9a (100 mg, 0.272 mmol) was dissolved in THF–H₂O mixture (2 : 1, 6 mL). To the mixture aqueous solution of NaOH (1.0 M, 0.6 mL) was added and reaction mixture was stirred for 1 h. After that aqueous HCl (1.0 M, 0.9 mL) was added reaction mixture was concentrated under reduced pressure. The residue on evaporation was purified by column chromatography (THF–H₂O–AcOH = 98 : 1 : 1) to give titled compound as a yellowish solid (76.9 mg, 77%).

(b) *Via Pinnic oxidation of 15.* To aldehyde 15 (1.84 g, 5.20 mmol), *t*-BuOH (18 mL) and THF (18 mL) were added and solution was stirred and cooled to \sim 3 °C (ice/water bath). Then 2-methyl-2-butene (2.83 mL, 26.86 mmol) was added. To this a solution of NaH₂PO₄ (1.89 g, 15.7 mmol) and NaClO₂ (1.42 g, 12.6 mmol, 80%) in H₂O (18 mL) was added slowly. Reaction mixture was stirred for 1.5 h at \sim 3 °C (ice/water bath) and then it was stopped with addition of Na₂SO₃ (2.02 g, 16.0 mmol). Precipitate formed was filtered off and filtrate was concentrated under reduced pressure. Residue on concentration was purified by column chromatography with two different mobile phases used sequentially. First mobile phase (DCM–MeOH = 10 : 1) was used to separate yellow impurities and second mobile phase (THF–H₂O–AcOH = 98 : 1 : 1) was used to elute product which upon evaporation and drying gave light yellowish solid (1.33 g, 68%). R_f : 0.57 (THF–H₂O–AcOH = 98 : 1 : 1); mp = 155–158 °C (from EtOAc); $[\alpha]_D$ –73.3 (c 1.19; DMSO); calc. for: C₂₁H₂₅N₃O₃·H₂O: C, 65.44; H, 7.06; N, 10.90. Found: C, 65.03; H, 6.72; N, 10.58%; IR: ν_{max} 3308, 2962, 2930, 2872, 2854, 1677, 1586, 1532, 1439, 1390, 1274, 1035, 753 cm^{-1} . ^1H NMR (DMSO- d_6 , 500 MHz) δ 1.15 (t, J = 7.1 Hz, 3H); 1.27–1.38 (m, 1H); 2.25–2.37 (m, 2H); 2.68 (t, J = 10.8 Hz, 1H); 2.76–2.85 (m, 2H); 3.17 (d, J = 11.1 Hz, 1H); 3.20–3.48 (m, 5H); 3.52 (dd, J_1 = 14.6 Hz, J_2 = 3.3 Hz, 1H); 5.17 (d, J = 10.1 Hz, 1H); 5.27 (d, J = 16.9 Hz, 1H); 5.90–6.00 (m, 1H); 7.01 (d, J = 7.2 Hz, 1H); 7.19 (t, J = 7.7 Hz, 1H); 7.52 (s, 1H); 7.87 (d, J = 8.0 Hz, 1H); 7.99 (bs, 1H), 12.50 (bs, 1H). ^{13}C NMR (DMSO- d_6 , 125 MHz) δ 15.0, 25.7, 30.5, 34.9, 39.8, 40.8, 54.6, 55.5, 62.9, 112.9, 115.2, 115.9, 117.8, 118.1, 124.7, 127.2, 133.0, 133.1, 134.7, 151.9, 175.1; HRMS (ESI): m/z calcd for C₂₁H₂₆N₃O₃: [M + H]⁺, 368.1969; found: 368.1967.

[(6aR,9R,10aR)-7-Allyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline-9-yl]methanol (11). In a dry round-bottom flask with compound 8 (0.200 g, 0.64 mmol), anhydrous THF (20 mL) was added under inert atmosphere. The solution was cooled to \sim 3 °C (ice/water bath) and with vigorous stirring

LiAlH₄ (0.97 mL, 0.97 mmol, 1 M/Et₂O) was added dropwise. Reaction was stopped after 45 min with addition of H₂O (0.1 mL) and was stirred for additional 20 min at indicated temperature. Suspension was then filtered through celite and washed with EtOAc (2 \times 5 mL). Filtrate was concentrated under reduced pressure and dried in vacuum to give titled compound as white solid (157.5 mg, 87%). R_f : 0.25 (EtOAc–EtOH = 88 : 12); mp = 206–209 °C, ref. 8 204–206 °C; $[\alpha]_D$ –74.4 (c 0.91, DMSO); IR: ν_{max} 3240, 3056, 2923, 2847, 1868, 1797, 1644, 1612, 1552, 1469, 1445, 1342, 1035, 733 cm^{-1} ; ^1H NMR (DMSO- d_6 , 500 MHz) δ 0.88–0.97 (m, 1H); 1.94 (t, J = 10.9 Hz, 1H); 2.27 (td, J_1 = 10.2 Hz, J_2 = 4.1 Hz, 1H); 2.52–2.59 (m, 2H); 2.73–2.80 (m, 1H); 3.07 (d, J = 9.9 Hz, 1H); 3.23 (dd, J_1 = 14.6 Hz, J_2 = 7.6 Hz, 1H); 3.26–3.31 (m, 1H); 3.36–3.43 (m, 3H); 3.49 (dd, J_1 = 14.6 Hz, J_2 = 5.6 Hz, 1H); 4.54 (t, J = 5.3 Hz, 1H); 5.15 (d, J = 10.4 Hz, 1H); 5.24 (d, J = 16.8 Hz, 1H); 5.95 (m, 1H); 6.77 (d, J = 7.1 Hz, 1H); 6.96 (bs, 1H); 7.00 (t, J = 7.6 Hz, 1H); 7.11 (d, J = 8.1 Hz, 1H); 10.60 (s, 1H). ^{13}C NMR (DMSO- d_6 , 125 MHz) δ 26.4, 30.9, 38.3, 40.5, 56.1, 56.7, 64.1, 64.6, 108.6, 110.3, 112.0, 117.4, 118.4, 122.0, 126.0, 133.1, 135.1; HRMS (ESI): m/z calcd for C₁₈H₂₃N₂O: [M + H]⁺, 283.1805; found: 283.1802.

(6aR,9R,10aR)-7-Allyl-9-[(*tert*-butyldimethylsilyl)oxy]-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline (12). To alcohol 11 (6.00 g, 21.2 mmol) in DMF (150 mL), imidazole (4.34 g, 63.8 mmol) and TBDMs-Cl (4.80 g, 31.9 mmol) were added. Reaction mixture was stirred for 2 h at room temperature after which it was quenched with saturated, aqueous solution of NaHCO₃ (150 mL). To a white suspension, H₂O (500 mL) and EtOAc (400 mL) were added and phases were separated after addition of brine (30 mL). Water phase was washed with EtOAc (2 \times 300 mL). Combined organic phases were reextracted with water (3 \times 400 mL), dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. Residue was dried at 40 °C for 19 h to give titled compound as white solid (8.39 g, 99%). R_f : 0.57 (EtOAc–EtOH = 88 : 12); mp = 225–227 °C (from MeOH); $[\alpha]_D$ –69.0 (c 1.19; DCM); calc. for: C₂₄H₃₆N₂OSi: C, 72.67; H, 9.15; N, 7.06. Found: C, 72.73; H, 9.39; N, 7.10%; IR: ν_{max} 3096, 3013, 2949, 2929, 2899, 2856, 2813, 2784, 1738, 1645, 1549, 1518, 1462, 1357, 1217, 1098, 834, 772, 742 cm^{-1} ; ^1H NMR (CDCl₃, 500 MHz) δ 0.08 (s, 3H); 0.09 (s, 3H); 0.93 (s, 9H); 1.09–1.18 (m, 1H); 2.02–2.16 (m, 2H); 2.49 (dt, J_1 = 3.8 Hz, J_2 = 10.9 Hz, 1H); 2.66 (d, J = 13.0 Hz, 1H); 2.75 (t, J = 12.7 Hz, 1H); 3.00 (bt, J = 9.1 Hz, 1H); 3.21 (d, J = 9.5 Hz, 1H); 3.38 (dd, J_1 = 14.2 Hz, J_2 = 7.5 Hz, 1H); 3.47 (dd, J_1 = 14.6 Hz, J_2 = 3.9 Hz, 1H); 3.51–3.56 (m, 1H); 3.58–3.63 (m, 2H); 5.20 (d, J = 10.1 Hz, 1H); 5.26 (d, J = 17.0 Hz, 1H); 5.95–6.06 (m, 1H); 6.89 (s, 1H); 6.92–6.95 (m, 1H); 7.15–7.20 (m, 2H), 8.01 (s, 1H). ^{13}C NMR (CDCl₃, 125 MHz) δ –5.2, –5.1, 1.2, 18.5, 26.1, 26.8, 29.8, 31.0, 38.7, 40.9, 56.8, 56.9, 64.3, 66.7, 108.58, 112.3, 113.3, 117.8, 118.2, 123.2, 126.4, 133.4, 134.0, 134.1; HRMS (ESI): m/z calcd for C₂₄H₃₇N₂OSi: [M + H]⁺, 397.2675; found: 397.2673.

(6aR,9R,10aR)-7-Allyl-9-[(*tert*-butyldimethylsilyl)oxy]-methyl-N-ethyl-6a,7,8,9,10,10a-hexahydroindolo[4,3-fg]quinoline-4(6*H*)-carboxamide (13). Compound 12 (16.00 g, 40.4 mmol) in an oven dried glassware was dissolved in anhydrous THF (400 mL) under N₂ atmosphere. Solution was cooled to \sim 3 °C (ice/water bath) and NaH (1.86 g, 60% in mineral oil, 44.4 mmol) was added.



Suspension was stirred for additional 10 min at indicated temperature and EtNCO (4.80 mL, 60.6 mmol) was added. Reaction mixture was then warmed to room temperature and stirred for 3 h. After 3 h reaction mixture was cooled to ~ 3 °C and quenched with brine (20.4 mL). Suspension was concentrated under reduced pressure, oily residue was dissolved in DCM (1000 mL) and washed with H₂O (400 mL). Water phase was then reextracted with DCM (2 \times 250 mL) and, combined organic phases were additionally washed with water (200 mL). Organic phase was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure to give brown solid, which was purified by column chromatography (EtOAc-heptane = 1 : 3) to give titled compound as pale yellow solid (16.94 g, 90%). *R*_f: 0.38 (EtOAc-heptane = 1 : 3); mp = 135–137 °C (from MeOH/H₂O); $[\alpha]$ _D –75.9 (*c* 1.40; DCM); calc. for: C₂₇H₄₁N₃O₂Si: C, 69.33; H, 8.84; N, 8.98. Found: C, 69.58; H, 9.11; N, 9.02%; IR: ν _{max} 3377, 2952, 2925, 2853, 2810, 1737, 1661, 1605, 1535, 1460, 1449, 1438, 1291, 1087, 837, 780, 754 cm^{–1}; ¹H NMR (CDCl₃, 500 MHz) δ 0.07 (s, 3H); 0.08 (s, 3H); 0.91 (s, 9H); 1.05–1.13 (m, 1H); 1.28 (t, *J* = 7.2 Hz, 3H); 2.00–2.13 (m, 2H); 2.40 (td, *J*₁ = 11.2 Hz, *J*₂ = 3.9 Hz, 1H); 2.58–2.63 (m, 2H); 2.91 (bt, *J* = 9.1 Hz, 1H); 3.18 (d, *J* = 8.6 Hz, 1H); 3.31 (dd, *J*₁ = 14.5 Hz, *J*₂ = 7.6 Hz, 1H); 3.36 (dd, *J*₁ = 15.0 Hz, *J*₂ = 3.9 Hz, 1H); 3.46–3.64 (m, 5H); 5.19 (d, *J* = 10.2 Hz, 1H); 5.24 (d, *J* = 17.1 Hz, 1H); 5.66 (bs, 1H); 5.90–6.01 (m, 1H); 7.04 (d, *J* = 7.3 Hz, 1H); 7.20 (s, 1H); 7.27 (t, *J* = 7.8 Hz, 1H); 7.72 (d, *J* = 8.2 Hz, 1H). ¹³C NMR (CDCl₃, 125 MHz) δ –5.23, –5.19, 15.1, 15.3, 18.4, 26.1, 26.5, 29.8, 30.9, 35.9, 38.5, 40.5, 56.6, 56.7, 63.8, 66.6, 111.7, 116.4, 116.5, 117.9, 118.4, 125.3, 128.4, 132.8, 133.8, 134.4, 152.5; HRMS (ESI): *m/z* calcd for C₂₇H₄₂N₃O₂Si: [M + H]⁺, 468.3041; found: 468.3040.

(6aR,9R,10aR)-7-Allyl-N-ethyl-9-(hydroxymethyl)-6a,7,8,9,10,10a-hexahydroindolo[4,3-fg]quinoline-4(6H)-carboxamide (14).

Compound 13 (4.5 g, 9.6 mmol) was dissolved in THF (50 mL). Solution was cooled to 0 °C, TBAF (19.2 mL, 19.2 mmol, 1 M in THF) was added and reaction mixture was stirred at indicated temperature for 45 h. After that H₂O (0.38 mL) was added and reaction mixture was concentrated under reduced pressure. Residue was purified by column chromatography (EtOH-EtOAc = 12 : 88), to give titled compound as a white solid (3.21 g, 94%). *R*_f: 0.38 (EtOH-EtOAc = 12 : 88); mp = 190–193 °C (from toluene); $[\alpha]$ _D –92.5 (*c* 1.26; DMSO); calc. for: C₂₁H₂₇N₃O₂: C, 71.36; H, 7.70; N, 11.89. Found: C, 71.65; H, 8.02; N, 11.89%; IR: ν _{max} 3424, 3303, 2932, 2913, 2856, 2812, 2784, 1664, 1603, 1533, 1439, 1243, 1142, 1039, 918, 812, 748 cm^{–1}; ¹H NMR (CDCl₃, 500 MHz) δ 1.08–1.18 (m, 1H); 1.30 (t, *J* = 7.2 Hz, 3H); 1.80 (bs, 1H); 2.08–2.17 (m, 2H); 2.44 (td, *J*₁ = 10.5 Hz, *J*₂ = 4.0 Hz, 1H); 2.63–2.71 (m, 2H); 2.92–2.93 (m, 1H); 3.16–3.24 (m, 1H); 3.34–3.43 (m, 2H); 3.49–3.55 (m, 2H); 3.55–3.62 (m, 2H); 3.66 (dd, *J*₁ = 10.8 Hz, *J*₂ = 4.7 Hz, 1H); 5.21 (d, *J* = 10.2 Hz, 1H); 5.26 (d, *J* = 16.8 Hz, 1H); 5.50 (t, *J* = 4.7 Hz, 1H); 5.92–6.02 (m, 1H); 7.06 (d, *J* = 7.4 Hz, 1H); 7.20 (d, *J* = 1.3 Hz, 1H); 7.28 (t, *J* = 8.0 Hz, 1H); 7.69 (d, *J* = 8.2 Hz, 1H); ¹H NMR (DMSO-*d*₆, 500 MHz) δ 0.85–0.95 (m, 1H); 1.15 (t, *J* = 7.2 Hz, 3H); 1.95 (dd, *J*₁ = 22.3 Hz, *J*₂ = 11.2 Hz, 1H); 2.25 (dt, *J*₁ = 10.3 Hz, *J*₂ = 3.8 Hz, 1H); 2.46–2.53 (m, 1H), 2.56 (bd, *J* = 12.7 Hz, 1H); 2.71–2.79 (m, 1H); 3.07 (d, *J* = 10.2 Hz, 1H); 3.22 (dd, *J*₁ = 14.6 Hz, *J*₂ = 7.7 Hz, 1H); 3.25–3.32 (m, 3H); 3.32–3.42 (m, 3H); 3.50 (dd, *J*₁ = 14.6 Hz, *J*₂ = 5.3 Hz, 1H); 4.57 (t, *J* = 5.1 Hz, 1H); 5.16 (d, *J* = 10.3 Hz, 1H); 5.24 (d, *J* = 17.1 Hz, 1H); 5.89–5.99 (m,

1H); 6.98 (d, *J* = 7.3 Hz, 1H); 7.18 (t, *J* = 7.8 Hz, 1H); 7.50 (s, 1H); 7.85 (d, *J* = 8.2 Hz, 1H); 7.97 (t, *J* = 5.4 Hz, 1H). ¹³C NMR (CDCl₃, 125 MHz) δ 15.3, 26.5, 30.9, 36.0, 38.6, 40.5, 56.4, 56.8, 63.7, 66.5, 111.7, 116.3, 116.6, 118.4, 118.8, 125.4, 128.4, 132.7, 133.5, 134.2, 152.4; ¹³C NMR (DMSO-*d*₆, 125 MHz) δ 15.0, 25.9, 30.8, 34.9, 38.2, 40.1, 56.1, 56.5, 63.6, 64.6, 112.7, 115.4, 115.9, 117.6, 118.0, 124.7, 127.3, 133.1, 133.6, 134.9, 151.9; HRMS (ESI): *m/z* calcd for C₂₁H₂₈N₃O₂: [M + H]⁺, 354.2176; found: 354.2177.

(6aR,9R,10aR)-7-Allyl-N-ethyl-9-formyl-6a,7,8,9,10,10a-hexahydroindolo[4,3-fg]quinoline-4(6H)-carboxamide (15).^{17,18}

(a) *Via Pfitzner–Moffatt oxidation of 14.* Alcohol 14 (3.00 g, 8.49 mmol) was dissolved in anhydrous DMSO (20 mL) and anhydrous DCM (20 mL) under N₂ atmosphere. Solution was cooled to ~ 3 °C (ice/water bath) and with stirring DCC (5.26 g, 25.5 mmol), anhydrous pyridine (1.38 mL, 17 mmol) and TFA (0.99 mL, 12.7 mmol) were added sequentially. Reaction mixture was stirred for 1.5 h at ~ 3 °C (ice/water bath). Suspension formed was filtered and to the filtrate DCM (250 mL) was added and organic phase was washed with water (3 \times 250 mL). Combined water phases were reextracted with DCM (250 mL). Then combined organic phases were dried over anhydrous Na₂SO₄, concentrated under reduced pressure and residue was purified by column chromatography (MeOH–EtOAc = 10 : 90 and DCM–MeOH = 90 : 10) to give titled aldehyde as a white solid (2.60 g, 87%).

(b) *Via Parikh–Doering oxidation of 14.* In an oven dried, round-bottom flask, alcohol 14 (50 mg, 0.141 mmol) under N₂ atmosphere, was dissolved in anhydrous DMSO (2 mL) and to the solution anhydrous DCM (2 mL) was added. Et₃N (0.197 mL, 1.41 mmol) was added and reaction mixture was cooled to ~ 3 °C (ice/water bath). To this solution SO₃·Py (67.4 mg, 0.423 mmol) dissolved in anhydrous DMSO (0.25 mL) was added dropwise and reaction mixture was stirred for 80 min. After that time reaction mixture was poured in to H₂O (20 mL) and water phase was extracted with DCM (3 \times 15 mL). Combined organic phases were dried over MgSO₄ and concentrated under reduced pressure. Residue on concentration was purified by column chromatography (EtOH–EtOAc = 1 : 9) to give aldehyde 15 as a white solid (33.7 mg, 68%). *R*_f: 0.56 (MeOH–EtOAc = 10 : 90); mp = 165–167 °C (from DCM/heptane); $[\alpha]$ _D –70.0 (*c* 0.45; DCM); calc. for: C₂₁H₂₅N₃O₂: C, 71.77; H, 7.17; N, 11.96. Found: C, 71.61; H, 7.31; N, 11.84%; IR: ν _{max} 3320, 2927, 2849, 1726, 1668, 1623, 1566, 1535, 1437, 1242, 1085, 892, 752, 640 cm^{–1}; ¹H NMR (CDCl₃, 300 MHz) δ 1.30 (t, *J* = 7.2 Hz, 3H); 1.35–1.45 (m, 1H); 2.31 (t, *J* = 11.6 Hz, 1H); 2.42–2.50 (m, 1H); 2.65–2.75 (m, 1H); 2.85–3.05 (m, 3H); 3.30–3.44 (m, 3H); 3.45–3.63 (m, 3H); 5.19–5.32 (m, 2H); 5.59 (bt, *J* = 5.0 Hz, 1H); 5.90–6.00 (m, 1H); 7.07 (d, *J* = 7.4 Hz, 1H); 7.21 (d, *J* = 1.6 Hz, 1H); 7.30 (t, *J* = 7.6 Hz, 1H); 7.72 (d, *J* = 8.3 Hz, 1H); 9.76 (s, 1H). ¹³C NMR (CDCl₃, 75.5 MHz) δ 15.3, 26.4, 27.9, 36.0, 40.2, 48.6, 52.5, 56.5, 63.5, 112.0, 115.9, 116.5, 118.2, 118.9, 125.4, 128.3, 132.8, 133.3, 133.4, 152.4, 202.6; HRMS (ESI): *m/z* calcd for C₂₁H₂₆N₃O₂: [M + H]⁺, 352.2020; found: 352.2019.

(6aR,9R,10aR)-7-Allyl-N⁹-(3-(dimethylamino)propyl)-N⁴-ethyl-6a,7,8,9,10,10a-hexahydroindolo[4,3-fg]quinoline-4,9(6H)-dicarboxamide B. Carboxylic acid 10 (0.580 g, 1.58 mmol) in oven dried glassware was dissolved in anhydrous DMF (10 mL) under



N_2 atmosphere and CDI (0.267 g, 1.74 mmol) was added. Mixture was stirred at room temperature for 3.5 h and then 3-dimethylamino-1-propylamine (0.221 mL, 1.74 mmol) was added. Reaction mixture was stirred at room temperature for 20 h. After indicated time, H_2O (100 mL) was added and reaction mixture was extracted with DCM (4×75 mL). Combined organic fractions were washed with H_2O (3×100 mL) and concentrated under reduced pressure. Residue on evaporation was suspended in H_2O (75 mL). Suspension was cooled to ~ 3 °C (ice/water bath) and acidified with 1 M aqueous HCl to pH 3.75 to obtain solution which was filtered and extracted with DCM (2×75 mL). Organic phases were discarded and water phase was re-cooled to ~ 3 °C (ice/water bath), basified with 1 M aqueous NaOH to pH 9.60 and precipitate formed was filtered off and washed with H_2O (20 mL). Solid residue on filtration was dried to give titled compound as a pale yellowish solid (0.385 g, 54%). R_f : 0.25 (EtOAc–BuOH–pyr.–DMF = 40 : 25 : 10 : 25); mp = 205–207 °C; $[\alpha]_D$ –75.4 (*c* 0.83; DCM); IR: ν_{max} 3294, 3066, 2934, 2861, 2765, 2725, 1663, 1639, 1535, 1439, 1346, 1212, 919, 751 cm^{–1}; ¹H NMR (CDCl₃, 500 MHz) δ 1.28 (t, *J* = 7.2 Hz, 3H); 1.57 (m, 1H); 1.64–1.71 (m, 2H); 2.26 (s, 6H); 2.35–2.46 (m, 4H); 2.51–2.58 (m, 1H); 2.58–2.64 (m, 1H); 2.72–2.78 (m, 1H); 2.79–2.86 (m, 1H); 3.15–3.20 (m, 1H); 3.27 (dd, *J*₁ = 14.4 Hz, *J*₂ = 7.7 Hz, 1H); 3.33 (dd, *J*₁ = 14.9 Hz, *J*₂ = 4.0 Hz, 1H); 3.36–3.40 (m, 2H); 3.46–3.54 (m, 3H); 5.17 (d, *J* = 10.2 Hz, 1H); 5.22 (d, *J* = 17.1 Hz, 1H); 5.79 (t, *J* = 5.4 Hz, 1H); 5.86–5.96 (m, 1H); 7.00 (d, *J* = 7.3 Hz, 1H); 7.22 (bs, 1H); 7.23–7.27 (m, 1H); 7.69 (bt, *J* = 4.6 Hz, 1H); 7.71 (d, *J* = 8.2 Hz, 1H). ¹³C NMR (CDCl₃, 125 MHz) δ 15.3, 25.5, 26.4, 31.0, 35.9, 40.0, 40.3, 43.7, 45.5, 55.6, 56.5, 59.3, 63.1, 111.9, 116.0, 116.4, 118.1, 118.7, 125.3, 128.2, 132.8, 133.6, 152.5, 173.6; HRMS (ESI): *m/z* calcd for C₂₆H₃₈N₅O₂: [M + H]⁺, 452.3020; found: 452.3019.

(6aR,9R,10aR)-7-Allyl-*N*⁹-[3-(dimethylamino)propyl]-*N*⁴-ethyl-*N*⁹-(ethylcarbamoyl)-6a,7,8,9,10,10a[hexahydroindolo-[4,3-*fg*]-quinoline-4,9(6*H*)-dicarboxamide (C) and regioisomer (6aR,9R,10aR)-7-allyl-*N*⁹-[(3-(dimethylamino)propyl)carbamoyl]-*N*⁴,*N*⁹-diethyl-6a,7,8,9,10,10a-hexahydroindolo[4,3-*fg*]quinoline-4,9(6*H*)-dicarboxamide (16). Carboxylic acid **10** (0.490 g, 1.33 mmol) in an oven dried glassware, under N_2 atmosphere was suspended in anhydrous DCM (8 mL). TEA (0.37 mL, 2.66 mmol) and EDC·HCl were added and reaction mixture was stirred for 23 h at room temperature. After that reaction was stopped with addition of H_2O (30 mL) and solution was extracted with DCM (2×30 mL). Combined organic phases were concentrated under reduced pressure and residue was purified by column chromatography (pyr.–Me₂CO = 10 : 90). First the desired product, impurity C, was eluted and evaporation of mobile phase gave titled compound as light yellowish foam (0.405 g, 58%). Part of the foam (55 mg) was further crystallized from toluene (0.2 mL) and heptane (3 mL) at –40 °C to give light yellowish solid (45 mg, 82%). R_f : 0.46 (Pyr.–Me₂CO = 10% : 90%); mp = 109–110 °C, ref. 4a 125–127 °C (from diisopropyl ether); $[\alpha]_D$ –71.8 (*c* 0.19; DCM); IR: ν_{max} 3334, 3274, 2971, 2936, 2867, 2828, 2781, 2245, 1668, 1525, 1438, 1340, 1525, 1438, 1340, 1271, 1138, 913, 729 cm^{–1}; ¹H NMR (CDCl₃, 500 MHz) δ 1.18 (t, *J* = 7.2 Hz, 3H); 1.29 (t, *J* = 7.2 Hz, 3H); 1.73 (m, 1H); 1.83–1.91 (m, 2H); 2.28 (s, 6H); 2.34–2.44 (m, 2H); 2.49–2.58 (m, 2H); 2.63 (t, *J* = 12.9 Hz, 1H); 2.79 (d, *J* = 11.8 Hz, 1H); 2.95 (bt, *J* = 8.9 Hz, 1H); 3.16

(d, *J* = 10.8 Hz, 1H); 3.26–3.40 (m, 5H); 3.47–3.58 (m, 3H); 3.72–3.90 (m, 2H); 5.19 (d, *J* = 10.0 Hz, 1H); 5.25 (d, *J* = 17.1 Hz, 1H); 5.62 (bt, *J* = 5.3 Hz, 1H); 5.88–5.99 (m, 1H), 7.02 (d, *J* = 7.2 Hz, 1H); 7.26 (t, *J* = 7.7 Hz, 1H), 7.70 (d, *J* = 8.2 Hz, 1H); 9.45 (bs, 1H). ¹³C NMR (CDCl₃, 125 MHz) δ 14.9, 15.3, 26.5, 29.8, 30.4, 31.3, 35.6, 36.0, 39.9, 42.3, 43.3, 44.9, 55.7, 56.2, 63.4, 111.8, 116.1, 116.6, 118.1, 118.7, 125.4, 128.3, 132.7, 133.5, 133.7, 152.4, 177.6; HRMS (ESI): *m/z* calcd for C₂₉H₄₃N₆O₃: [M + H]⁺, 523.3391; found: 523.3390.

Continuing elution, regioisomer **16** was isolated and evaporation of mobile phase gave (67 mg, 10%) yellow oil. R_f : 0.29 (pyr.–Me₂CO = 10 : 90); $[\alpha]_D$ –40.2 (*c* 0.10; DCM); IR: ν_{max} 3282, 2971, 2935, 2868, 2817, 2785, 2246, 1688, 1524, 1438, 1340, 1271, 1139, 917, 729 cm^{–1}; ¹H NMR (CDCl₃, 500 MHz) δ 1.28 (t, *J* = 7.2 Hz, 3H); 1.30 (t, *J* = 6.7 Hz, 3H); 1.72–1.83 (m, 3H); 2.27 (s, 6H); 2.40 (m, 2H); 2.50–2.66 (m, 3H); 2.74 (d, *J* = 13.0 Hz, 1H); 2.92–2.99 (m, 1H); 3.15 (d, *J* = 13.4 Hz, 1H); 3.21 (d, *J* = 11.4 Hz, 1H); 3.29–3.36 (m, 3H); 3.39 (dd, *J*₁ = 14.7 Hz, *J*₂ = 3.7 Hz, 1H); 3.47–3.53 (m, 2H); 3.56 (dd, *J*₁ = 15.2 Hz, *J*₂ = 6.4 Hz, 1H); 3.92 (q, *J* = 7.0 Hz, 2H); 5.21 (d, *J* = 10.1 Hz, 1H); 5.26 (d, *J* = 17.0 Hz, 1H); 5.64 (bt, *J* = 5.4 Hz, 1H); 5.88–5.98 (m, 1H); 7.02 (d, *J* = 7.3 Hz, 1H); 7.23 (bs, 1H); 7.27 (t, *J* = 7.8 Hz, 1H); 7.72 (d, *J* = 8.3 Hz, 1H); 9.33 (bs, 1H). ¹³C NMR (CDCl₃, 125 MHz) δ 15.3, 15.9, 26.5, 27.2, 29.4, 31.3, 35.9, 39.6, 40.1, 42.3, 45.3, 55.6, 56.4, 57.3, 63.4, 112.0, 115.9, 116.5, 118.2, 119.1 125.4, 128.2, 132.8, 133.1, 133.4, 152.4, 155.0, 177.9; HRMS (ESI): *m/z* calcd for C₂₉H₄₃N₆O₃: [M + H]⁺, 523.3391; found: 523.3385.

Acknowledgements

Financial support for this research was provided by Lek d.d., a Sandoz Company. We would like to acknowledge Roman Burja TLCM head for valuable help, Dr. Martin Črnugelj for NMR assistance and University of Ljubljana, Faculty of Chemistry and Chemical Technology for high resolution mass analyses and elemental microanalyses. Dr. Franc Požgan gratefully acknowledges The Ministry of Higher Education, Science and Technology of the Republic of Slovenia, the Slovenian Research Agency (P1-0230-0103) and EN→FIST Centre of Excellence for their partial financial support.

Notes and references

- 1 P. Del Dotto and U. Bonuccelli, *Clin. Pharmacokinet.*, 2003, **42**, 633.
- 2 ICH Q3A Guideline, Revision 2, 2008.
- 3 *European Pharmacopoeia*, Monograph C, 7th edn (7.5), 2012, p. 1535.
- 4 (a) E. Brambilla, E. di Salle, G. Briatico, S. Mantegani and A. Temperilli, *Eur. J. Med. Chem.*, 1989, **24**, 421; (b) P. Salvati, A. M. Caravaggi, A. Temperilli, G. Bosisio, O. Sapini and E. di Salle, *US Pat.* 4526892, 1985, *Chem. Abstr.*, 1985, **103**, 160754.
- 5 (a) A. Stoll and A. Hofmann, *Helv. Chim. Acta*, 1943, **26**, 2070; (b) A. Stoll, A. Hofmann and Th. Petrzilka, *Helv. Chim. Acta*, 1946, **29**, 635; (c) R. Ručman and N. Djordjević, *DE Pat.* 2708042, 1976, *Chem. Abstr.*, 1977, **88**, 158466.



6 G. Marzoni, W. L. Garbrecht, P. Fludzinski and M. L. Cohen, *J. Med. Chem.*, 1987, **30**, 1823.

7 J. V. Braun, K. Heider and E. Müller, *Chem. Ber.*, 1918, **51**, 273.

8 (a) A. M. Crider, R. Grubb, K. A. Bachmann and A. K. Rawat, *J. Pharm. Sci.*, 1981, **70**, 1319; (b) T. A. Montzka, J. D. Matiskella and R. A. Partyka, *Tetrahedron Lett.*, 1974, **15**, 1325; (c) Z. Časar and T. Mesar, *WO Pat.* 2009133097, 2009, *Chem. Abstr.*, 2009, **151**, 528967.

9 E. C. Kornfeld and N. J. Bach, *EP Pat.* 3667, 1979, *Chem. Abstr.*, 1979, **92**, 181450.

10 M. Hafner, M. Sulyok, R. Schuhmacher, C. Crews and R. Krska, *World Mycotoxin J.*, 2008, **1**, 67.

11 J. Beneš, A. Černý, V. Miller and S. Kudrnáč, *Collect. Czech. Chem. Commun.*, 1983, **48**, 1333.

12 Amount of epimerisation in transformation of **8** to **9** was determined on the worked up, crude reaction mixture. Recording ^1H NMR of the reaction mixture showed on 6 : 1 epimer ratio. Triplets of $\text{CH}_3\text{CH}_2\text{NCO}$ at δ [ppm] 1.15 and 1.12 of **9a** and **9b** were used. See ESI.†

13 S. Ohno, Y. Adachi, M. Koumori, K. Mizukoshi, M. Nagasaka and K. Ichihara, *Chem. Pharm. Bull.*, 1994, **42**, 1463.

14 R. Gažák, V. Křen, P. Sedmra, D. Passarella, M. Novotna and B. Danieli, *Tetrahedron*, 2007, **63**, 10466.

15 Z.-X. Wang, Y. Q. Li, M. Kondamreddy and X. Cai, *US Pat.* 20080275240, 2008, *Chem. Abstr.*, 2008, **149**, 534450.

16 (a) S. Mantegani, E. Brambilla, C. Caccia, E. di Salle, M. A. Cervini, R. A. McArthur, G. Traquandi and M. Varasi, *Il Farmaco*, 1998, **53**, 65; (b) L. Bernardi, S. Mantegani, A. Temperili and G. Traquandi, *GB Pat.* 2162173, 1986, *Chem. Abstr.*, 1986, **105**, 79216.

17 K. E. Pfitzner and J. G. Moffatt, *J. Am. Chem. Soc.*, 1963, **85**, 3027. In general DCC was used for DMSO activation, albeit more expensive EDC·HCl was also employed. There were no differences in reaction yield between the two, although aldehyde prepared using EDC·HCl did not require so thorough purification with respect to the residual urea by-product.

18 J. R. Parikh and W. v. E. Doering, *J. Am. Chem. Soc.*, 1967, **89**, 5505.

19 (a) P. A. Stadler and R. K. A. Giger, in *Natural Products and Drug Development*, ed. P. L. Krogsgaard, S. C. Brøgger and H. Kofod, Munksgaard, Copenhagen, 1984, pp. 463–484; (b) P. M. Dewick, in *Medicinal Natural Products*, John Wiley and Sons, Ltd, 2nd edn, 2002, pp. 368–376.

20 S. Inuki, A. Iwata, S. Oishi, N. Fujii and H. Ohno, *J. Org. Chem.*, 2011, **76**, 2072.

21 (a) B. O. Lindgren and T. Nilsson, *Acta Chem. Scand.*, 1973, **27**, 888; (b) B. S. Bal, W. E. Childers, Jr and H. W. Pinnick, *Tetrahedron*, 1981, **37**, 2091.

22 (a) B. F. DeTar and R. Silverstein, *J. Am. Chem. Soc.*, 1966, **88**, 1013; (b) J. Podlech, Synthesis of Peptides and Peptidomimetics, in *Houben-Weyl, Methods of Organic Chemistry, Additional and Supplementary Volume E22a to the 4th Edition*, ed. M. Goodman, A. Felix, L. Moroder and C. Toniolo, Georg Thieme Verlag, Stuttgart, New York, 2001, pp. 517–533.

23 (a) S. W. Ashford, K. E. Henegar, A. M. Anderson and P. G. M. Wuts, *J. Org. Chem.*, 2002, **67**, 7147; (b) A. L. Gutman, G. Nisnevich, I. Rukhman, B. Tishin, A. Vilensky and B. Pertsikov, *US Pat.* 20020177709, 2002, *Chem. Abstr.*, 2002, **138**, 4728; (c) J. Galambos, L. Czibula, F. Sebők, S. Bálint and F. Kassai, *WO Pat.* 2005085243, 2005, *Chem. Abstr.*, 2005, **143**, 286587.

24 A solvent screening was made by others (see ref. 15) with the aim to improve the ratio of cabergoline (**1**) vs. its regioisomer in the reaction of cabergolinic acid **A** with EDC·HCl. The best results were achieved with halogenated solvents like DCM, DCE and chlorobenzene with the ratios of cabergoline (**1**) vs. regioisomer of 4.1, 4.1 and 4.7 respectively.

25 Ratio of **C** vs. **15** was determined from ^1H NMR spectrum of evaporated crude reaction mixture. Singlets of NMe_2 at δ [ppm] 2.30 and 2.28 for **C** and **15** were used.

26 I. Candiani, W. Cabri, F. Zarini, A. Bedeschi and S. Penco, *Synlett*, 1995, 605.

27 T. Fehr, P. A. Stadler and A. Hofmann, *Helv. Chim. Acta*, 1970, **53**, 2197.

