



Cite this: *Chem. Commun.*, 2018, 54, 12840

Received 18th September 2018,  
Accepted 16th October 2018

DOI: 10.1039/c8cc07572g

rs.c.li/chemcomm

# Ruthenium-catalyzed C–H oxygenation of quinones by weak O-coordination for potent trypanocidal agents†

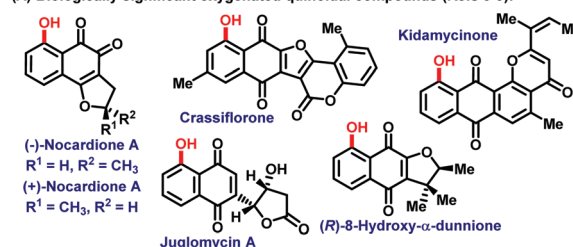
Gleiston G. Dias,<sup>ab</sup> Torben Rogge,<sup>b</sup> Rositha Kuniyil,<sup>b</sup> Claus Jacob,<sup>id c</sup>  
Rubem F. S. Menna-Barreto,<sup>d</sup> Eufânio N. da Silva Júnior<sup>id \*a</sup> and  
Lutz Ackermann<sup>id \*b</sup>

Ruthenium-catalysis enabled the C-5 selective C–H oxygenation of naphthoquinones, and also sets the stage for the site-selective introduction of a hydroxyl group into anthraquinones. A-ring modified naphthoquinoidal compounds represent an important class of bioactive quinones for which the present study encompasses the first C–H oxygenation strategy by weak O-coordination.

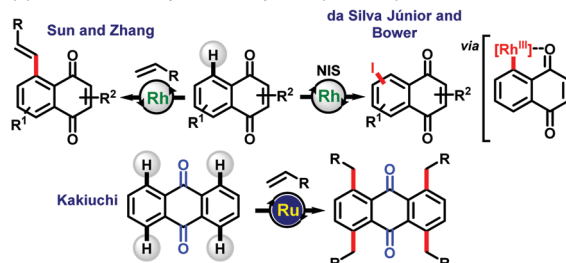
Among the versatile scaffolds studied in Medicinal Chemistry, quinones represent a privileged class of substance.<sup>1</sup> Thus, quinoidal compounds are involved in biologically relevant processes, such as in the electron transport chain, redox mediators, and as alkylating agents in a broad spectrum of biological processes.<sup>2</sup> For example, nocardiones A are notable bioactive naphthoquinoidal molecules. (–)-Nocardione A is a Cdc25B tyrosine phosphatase inhibitor, while (+)-nocardione A presents antifungal and cytotoxic activity with the ability to kill human myeloid leukemia cell lines *via* a mechanism of action associated with apoptosis.<sup>3</sup> Remarkable 1,4-naphthoquinones (1,4-NQs) include crassiflorone,<sup>4</sup> a natural product isolated from the stem bark of the African ebony (persimmon) tree *Diospyros crassiflora* with anti-mycobacterial and anti-gonorrhoeal properties,<sup>5</sup> among others. Furthermore, juglomycin A,<sup>6</sup> (R)-8-hydroxy-R-dunnione<sup>7</sup> and kidamycinone<sup>8</sup> are examples of important quinoidal compounds with diverse bioactivity<sup>9</sup> (Scheme 1A).

The quinones depicted in Scheme 1A bear a hydroxyl group as a recurring structural motif in the benzenoid A-ring, which is

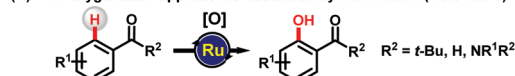
(A) Biologically significant oxygenated quinoidal compounds (Refs 3–9):



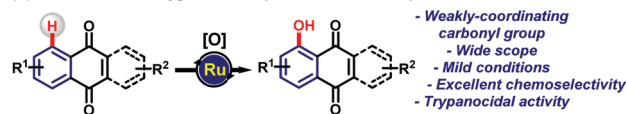
(B) C–H activation on quinoidal compounds (Refs 11a–f):



(C) C–H oxygenation approaches described by Ackermann (Refs 12a–c):



(D) This work: C–H oxygenation of quinones and anthraquinones



Scheme 1 Overview.

usually installed in the first steps of their previously reported respective syntheses.<sup>3,4,8</sup> The preparation of such molecules *via* late-stage functionalization by C–H activation logic has thus far not been addressed.<sup>10</sup> This C–H activation strategy can be an instrument to maximize the synthesis process with the preparation of the target molecules in a reduced number of steps with overall significantly improved yields. Late-stage functionalization of quinones is not easily achieved due to the following aspects, including: (a) generally deactivated quinoidal systems present low nucleophilicity at the benzenoid A-ring; (b) the

<sup>a</sup> Institute of Exact Sciences, Department of Chemistry, Federal University of Minas Gerais, Belo Horizonte, MG, 31270-901, Brazil. E-mail: eufanio@ufmg.br

<sup>b</sup> Institut für Organische und Biomolekulare Chemie, Georg-August-Universität, Göttingen, Tammannstraße 2, 37077 Göttingen, Germany. E-mail: lutz.ackermann@chemie.uni-goettingen.de

<sup>c</sup> Division of Bioorganic Chemistry, School of Pharmacy, University of Saarland, D-66123 Saarbrücken, Germany

<sup>d</sup> Oswaldo Cruz Institute, FIOCRUZ, Rio de Janeiro, RJ, 21045-900, Brazil

† Electronic supplementary information (ESI) available: Experimental procedures and characterisation data for all compounds. CCDC 1859287 (2i), 1859288 (2f), 1859289 (2c), 1859290 (2d), 1859453 (2j), 1859454 (2e) and 1859455 (2h). For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c8cc07572g

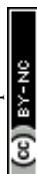
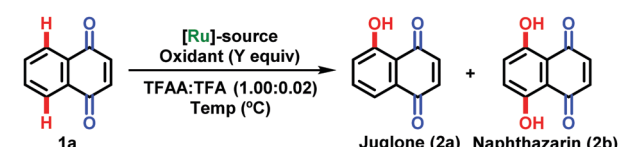


Table 1 Optimization of C–H oxygenation<sup>a</sup>


Reaction scheme showing the C–H oxygenation of 1a (1,4-naphthoquinone) to Juglone (2a) and Naphthazarin (2b) using a [Ru]-source, Oxidant (Y equiv), TFAA:TFA (1.00:0.02), and Temp (°C).

Entry	[Ru]-Source (mol%)	Oxidant	Y	Temp (°C)	2a	2b
1	[RuCl <sub>2</sub> ( <i>p</i> -cymene)] <sub>2</sub> (5)	PIFA	2.0	110	53	42
2	[RuCl <sub>2</sub> ( <i>p</i> -cymene)] <sub>2</sub> (2)	PIFA	1.2	80	87	5
3	[RuCl <sub>2</sub> ( <i>p</i> -cymene)] <sub>2</sub> (2)	PIDA	1.2	80	71	Traces
4	[RuCl <sub>2</sub> ( <i>p</i> -cymene)] <sub>2</sub> (2)	NH <sub>4</sub> S <sub>2</sub> O <sub>8</sub>	1.2	80	NR	NR
5	RuCl <sub>3</sub> ·H <sub>2</sub> O (5)	PIFA	1.2	80	NR	NR
6	[RhCp*Cl <sub>2</sub> ] <sub>2</sub> (5)	PIFA	1.2	80	NR	NR
7	CoCp*(CO)I <sub>2</sub> (5)	PIFA	1.2	80	NR	NR
8	Pd(OAc) <sub>2</sub> (5)	PIFA	1.2	80	NR	NR
9 <sup>b</sup>	—	PIFA	1.2	80	NR	NR
10 <sup>b</sup>	[RuCl <sub>2</sub> ( <i>p</i> -cymene)] <sub>2</sub> (2)	PIFA	1.2	80	92	3

<sup>a</sup> General reaction conditions: (1a) (0.4 mmol), PIFA (0.5 mmol), [RuCl<sub>2</sub>(*p*-cymene)]<sub>2</sub> (5.0 or 2.0 mol%); TFAA (1 mL) and TFA (0.02 mL), 18 h. <sup>b</sup> 12 h. PIFA = PhI(CF<sub>3</sub>CO<sub>2</sub>)<sub>2</sub>. PIDA = PhI(CH<sub>3</sub>CO<sub>2</sub>)<sub>2</sub>. NR = for all cases starting material 1a was recovered, with the exception of the reaction with Pd(OAc)<sub>2</sub>, here degradation of the products was observed. Yields of isolated products.

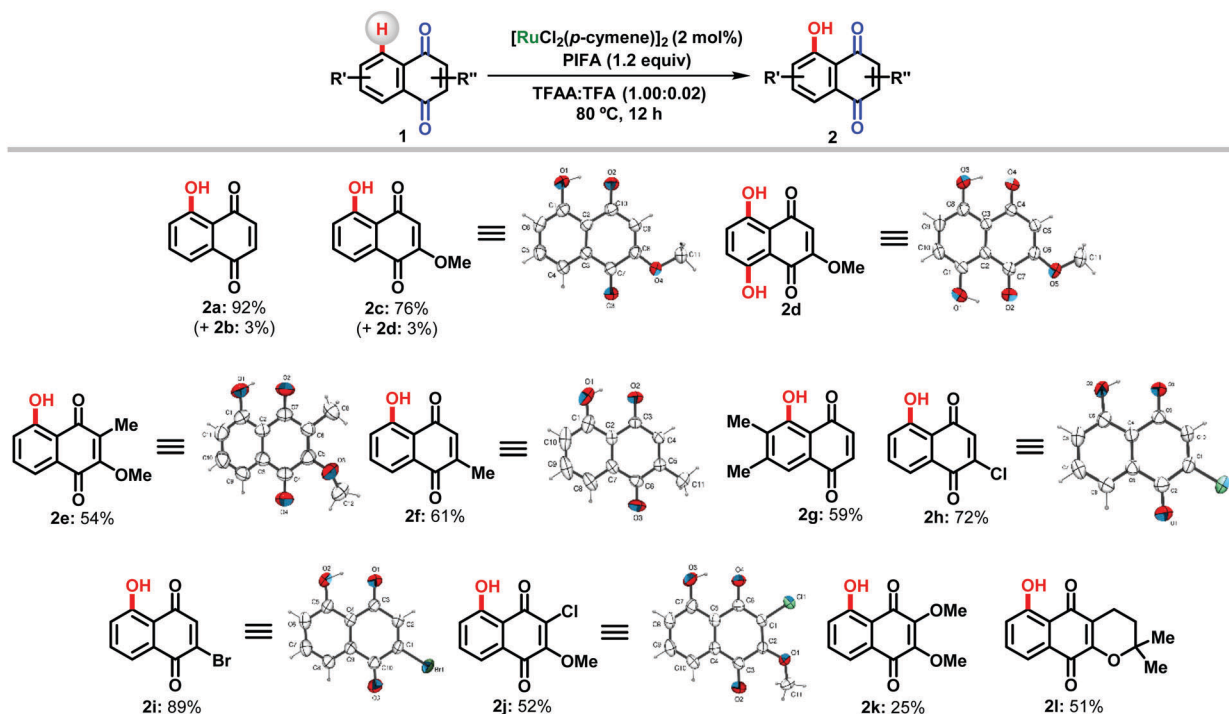
weakly coordinating ability of the B-ring carbonyls towards transition metal catalysts and (c) the difficulty in developing general reaction protocols capable of providing favorable results with the use of different classes of quinonoid systems.<sup>10</sup>

In this context, Sun and Zhang, Kakiuchi, Bower and da Silva Júnior groups<sup>11</sup> have recently developed useful methods for C–H functionalization of naphthoquinones and anthraquinones (Scheme 1B). Despite the recent development in the area, direct

C–H oxygenations of quinonoid compounds have been proven to be elusive. Given our recent success in terms of ruthenium-catalyzed C–H oxygenations with weakly coordinating ketones, amides and aldehydes<sup>12</sup> (Scheme 1C), herein, we describe an efficient process for the direct, site-selective installation of a hydroxyl group in a broad range of naphthoquinones and anthraquinones. For the first time, a C–H bond oxygenation strategy was thus established for the synthesis of quinonoid derivatives with unique bioactivity (Scheme 1D).

Preliminary studies involved the reaction of 1,4-naphthoquinone (1a) with PhI(CF<sub>3</sub>CO<sub>2</sub>)<sub>2</sub> (PIFA) as an oxidant and [RuCl<sub>2</sub>(*p*-cymene)]<sub>2</sub> as the catalyst (Table 1, entry 1). Here, juglone (2a) and naphthazarin (2b) were obtained in 53 and 42% yield, respectively. Further refinement aiming at minimizing the amount of the ruthenium-based catalyst afforded products 2a and 2b in 87 and 5% yield when 2 mol% of [RuCl<sub>2</sub>(*p*-cymene)]<sub>2</sub> and 1.2 equivalents of PIFA were used (entry 2). Despite the efficiency of the catalyst, we continued our efforts towards alternative oxidation sources. Hence, PIDA and NH<sub>4</sub>S<sub>2</sub>O<sub>8</sub> were also evaluated as oxygenation agents, albeit with limited success (entries 3 and 4). We also tested alternative sources of catalysts with the use of rhodium, cobalt and palladium complexes, but promising results were not observed (entries 5–8). A control experiment confirmed the essential role of the ruthenium catalyst (entry 9). Fine tuning of the reaction temperature enabled the formation of juglone (2a) in 92% yield (entry 10). Viable substrates for the new C–H oxygenation strategy are outlined in Scheme 2.

Structural assignments were based on detailed NMR analysis and X-ray diffraction analysis of products 2c–2f, 2h, 2i and 2j.



Scheme 2 Scope of the naphthoquinone C-5 oxygenation protocol.

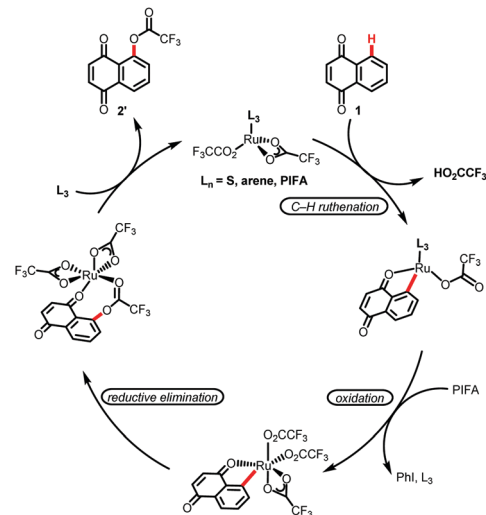


In general, quinones were prepared in moderate to high yields, tolerating the presence of electron withdrawing and donating groups. In most cases, the dihydroxylated product was not formed or was observed in only 3% yield (**2b** and **2d**). Our method proved effective even for the preparation of the hydroxylated derivative of  $\alpha$ -lapachone (**2l**), a natural product related to the lapachol family with remarkable antitumor activity.<sup>13</sup>

Subsequently, the synthetic utility of the ruthenium(II)-catalyzed C–H oxygenation was reflected by the chemo-selective diversification of anthraquinones **3**. Here, a slight modification of the reaction conditions featured the use of PIDA, allowing for the C–H oxygenations with excellent levels of positional and chemo-selectivities in terms of mono-functionalization, fully tolerating, *inter alia*, chloro and bromo functional groups (Scheme 3).

Based on our previous reports, a plausible catalytic cycle is initiated by carboxylate-assisted C–H ruthenation,<sup>12,14</sup> along with subsequent oxidation-induced reductive elimination (Scheme 4).<sup>14,15</sup>

Neglected tropical diseases (NTDs) comprehend a group of seventeen illnesses that affect more than one billion people. Among NTDs, Chagas disease caused by the hemoflagellate protozoa *Trypanosoma cruzi* presents significant mortality and morbidity, especially in low-income populations in Latin America endemic countries.<sup>16,17</sup> Recently, the intense flux of immigration due to globalization has changed the distribution of Chagas disease patients, resulting in the presence of infected people in non-endemic countries such as European countries, the United States, Japan or Australia, where the transmission is sustained by blood transfusion or congenital routes.<sup>18,19</sup> Since the 1960s up to now, the clinical chemotherapy for Chagas disease has been based on two nitroderivatives, benznidazole and nifurtimox, that present important side effects and debatable anti-*T. cruzi* activity in chronic patients.<sup>20</sup> The absence of alternatives for the most severe phase of this illness leads to the continuous search for novel trypanocidal drugs.<sup>21</sup> In this context, in the last few years, we have evaluated diverse naphthoquinones against the parasite and proved the potential trypanocidal effect of

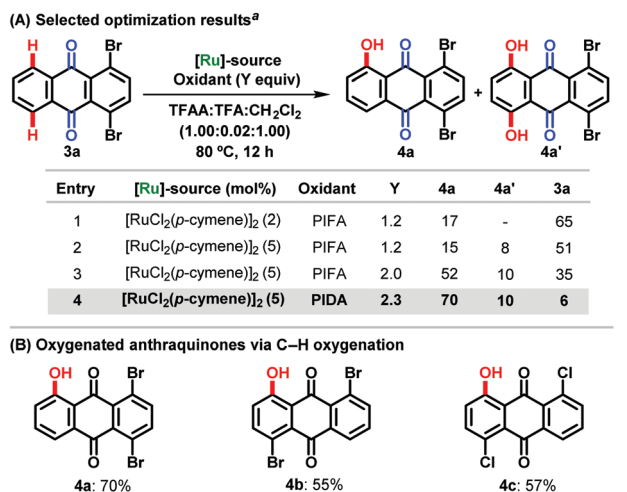


Scheme 4 Proposed mechanism.

these compounds.<sup>11f</sup> To understand the biological importance of A-ring modified quinoidal compounds and to continue our efforts towards the development of potent trypanocidal agents, we herein disclose the evaluation of 1,4-NQs against *T. cruzi*. Thus, compounds **2c**, **2e**, **2k** and **2l** were not active against the parasite with  $IC_{50}/24\text{ h} > 1000.0\text{ }\mu\text{M}$ . To our delight, six compounds showed activity against *T. cruzi* with  $IC_{50}$  in the range of 176.1 to 29.5  $\mu\text{M}$  (Table S1), highlighting **2h** ( $IC_{50} = 32.9\text{ }\mu\text{M}$ ) and **2i** ( $IC_{50} = 29.5\text{ }\mu\text{M}$ ). In comparison with the standard drug benznidazole ( $IC_{50} = 103.6\text{ }\mu\text{M}$ ), **2h** and **2i** are 3.1 and 3.5-fold more active, respectively (Fig. 1).

In conclusion, we have reported on an efficient C–H oxygenation strategy for naphthoquinones and anthraquinones. Thus, carboxylate-assisted<sup>22</sup> ruthenium(II) catalysis enabled the step-economical synthesis of potent bioactive compounds with ample scope. The obtained C–H hydroxylated naphthoquinones displayed unique trypanocidal activities.

E. N. S. J. acknowledges funding from CNPq 404466/2016-8 and PQ 305741/2017-9, CAPES and DAAD for exchange PROBRAL



<sup>a</sup> General reaction conditions: (**3a**) (0.3 mmol), PIDA (0.7 mmol), [RuCl<sub>2</sub>(*p*-cymene)]<sub>2</sub> (5.0 mol %); (TFAA) (1 mL), TFA (0.02 mL) and CH<sub>2</sub>Cl<sub>2</sub> (1 mL). Yields of isolated products.

Scheme 3 Synthesis of anthraquinone derivatives.

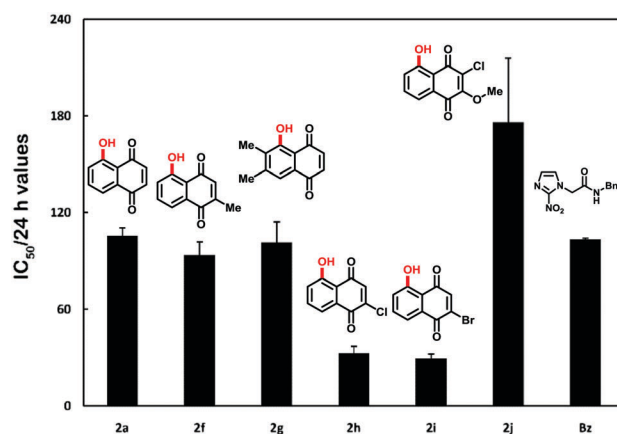


Fig. 1 Most active naphthoquinones on trypomastigote forms of *T. cruzi*. Benznidazole, a standard drug for the treatment of chagasic patients, was used as a control.  $IC_{50}/24\text{ h}$  values in  $\mu\text{M}$ .



program (99999.008126/2015-01) and Capes-Humboldt research fellowship programme for experienced researchers (Proc. No. 88881.145517/2017-01). L. A. acknowledges funding from the DFG (Gottfried-Wilhelm-Leibniz prize).

## Conflicts of interest

There are no conflicts to declare.

## References

- (a) R. W. DeSimone, K. S. Currie, S. A. Mitchell, J. W. Darrow and D. A. Pippin, *Comb. Chem. High Throughput Screening*, 2004, **7**, 473; (b) L. Costantino and D. Barlocco, *Curr. Med. Chem.*, 2010, **5**, 381; (c) C. D. Duarte, E. J. Barreiro and C. A. M. Fraga, *Mini-Rev. Med. Chem.*, 2007, **7**, 1108; (d) K. Bondensgaard, M. Ankersen, H. Thøgersen, B. S. Hansen, B. S. Wulff and R. P. Bywater, *J. Med. Chem.*, 2004, **47**, 888.
- (a) R. H. Thomson, *Naturally Occurring Quinones*, Academic Press, London, 2nd edn, 1971; (b) R. H. Thomson, *Naturally Occurring Quinones III Recent Advances*, Chapman and Hall, London, 3rd edn, 1987; (c) G. Powis, *Pharmacol. Ther.*, 1987, **35**, 57; (d) P. J. O'Brien, *Chem.-Biol. Interact.*, 1991, **80**, 1; (e) E. A. Hillard, F. C. Abreu, D. C. Ferreira, G. Jaouen, M. O. F. Goulart and C. Amatore, *Chem. Commun.*, 2008, 2612.
- (a) T. Otani, Y. Sugimoto, Y. Aoyagi, Y. Igarashi, T. Furumai, N. Saito, Y. Yamada, T. Asao and T. Oki, *J. Antibiot.*, 2000, **53**, 337; (b) D. L. J. Clive and S. P. Fletcher, *Chem. Commun.*, 2003, 2464.
- J. Padwal, W. Lewis and C. J. Moody, *Org. Biomol. Chem.*, 2011, **9**, 3484.
- E. Uliassi, G. Fiorani, R. L. Krauth-Siegel, C. Bergamini, R. Fato, G. Bianchini, J. C. Menéndez, M. T. Molina, E. L. Montero, F. Falchi, A. Cavalli, S. Gul, M. Kuzikov, B. Ellinger, G. Witt, C. B. Moraes, L. H. Freitas-Júnior, C. Borsari, M. P. Costi and M. L. Bolognesi, *Eur. J. Med. Chem.*, 2017, **141**, 138.
- H. Maeda and G. A. Kraus, *J. Org. Chem.*, 1996, **61**, 2986.
- X.-H. Cai, X.-D. Luo, J. Zhou and X.-J. Hao, *J. Nat. Prod.*, 2005, **68**, 797.
- T. Mabit, A. Siard, M. Pantin, D. Zon, L. Foulgoc, D. Sissouma, A. Guingant, M. Mathé-Allainmat, J. Lebreton, F. Carreaux, G. Dujardin and S. Collet, *J. Org. Chem.*, 2017, **82**, 5710.
- (a) K. Ushiyama, N. Tanaka, H. Ono and H. Ogata, *Jpn. J. Antibiot.*, 1971, **24**, 197; (b) A. S. Soares, F. L. Barbosa, A. L. Rüdiger, D. L. Hughes, M. J. Salvador, A. R. Zampronio and M. E. A. Stefanello, *J. Nat. Prod.*, 2017, **80**, 1837.
- (a) J. C. K. Chu and T. Rovis, *Angew. Chem., Int. Ed.*, 2018, **57**, 62–101; (b) E. N. da Silva Júnior, G. A. M. Jardim, R. S. Gomes, Y.-F. Liang and L. Ackermann, *Chem. Commun.*, 2018, **54**, 7398; (c) P. Gandeepan and L. Ackermann, *Chem.*, 2018, **4**, 199–222; (d) Z. Dong, Z. Ren, S. J. Thompson, Y. Xu and G. Dong, *Chem. Rev.*, 2017, **117**, 9333–9403; (e) D. L. Davies, S. A. Macgregor and C. L. McMullin, *Chem. Rev.*, 2017, **117**, 8649–8709; (f) C. Borie, L. Ackermann and M. Nechab, *Chem. Soc. Rev.*, 2016, **45**, 1368–1386; (g) M. H. Shaw and J. F. Bower, *Chem. Commun.*, 2016, **52**, 10817–10829; (h) J. Li and L. Ackermann, *Nat. Chem.*, 2015, **7**, 686–687; (i) K. Shin, H. Kim and S. Chang, *Acc. Chem. Res.*, 2015, **48**, 1040–1052; (j) S. I. Kozhushkov and L. Ackermann, *Chem. Sci.*, 2013, **4**, 886–896; (k) B. Li and P. H. Dixneuf, *Chem. Soc. Rev.*, 2013, **42**, 5744–5767; (l) P. B. Arockiam, C. Bruneau and P. H. Dixneuf, *Chem. Rev.*, 2012, **112**, 5879–5918; (m) O. Daugulis, H.-Q. Do and D. Shabashov, *Acc. Chem. Res.*, 2009, **42**, 1074–1086; (n) L. Ackermann, R. Vicente and A. R. Kapdi, *Angew. Chem., Int. Ed.*, 2009, **48**, 9792–9826; (o) H. M. Davies and J. R. Manning, *Nature*, 2008, **451**, 417–424, and references cited therein.
- (a) K. Kitazawa, T. Kochi, M. Sato and F. Kakiuchi, *Org. Lett.*, 2009, **11**, 195; (b) D. Matsumura, K. Kitazawa, S. Terai, T. Kochi, Y. Le, M. Nitani, Y. Aso and F. Kakiuchi, *Org. Lett.*, 2012, **14**, 3882; (c) C. Zhang, M. Wang, Z. Fan, L.-P. Sun and A. Zhang, *J. Org. Chem.*, 2014, **79**, 7626; (d) G. A. M. Jardim, E. N. da Silva Júnior and J. F. Bower, *Chem. Sci.*, 2016, **7**, 3780; (e) G. A. M. Jardim, T. L. Silva, M. O. F. Goulart, C. A. de Simone, J. M. C. Barbosa, K. Salomão, S. L. de Castro, J. F. Bower and E. N. da Silva, Júnior, *Eur. J. Med. Chem.*, 2017, **136**, 406; (f) G. A. M. Jardim, W. X. C. Oliveira, R. P. de Freitas, R. F. S. Menna-Barreto, T. L. Silva, M. O. F. Goulart and E. N. da Silva Júnior, *Org. Biomol. Chem.*, 2018, **16**, 1686.
- (a) V. S. Thirunavukkarasu and L. Ackermann, *Org. Lett.*, 2012, **14**, 6206; (b) F. Yang and L. Ackermann, *Org. Lett.*, 2013, **15**, 718; (c) F. Yang, K. Rauch, K. Kettelhoit and L. Ackermann, *Angew. Chem., Int. Ed.*, 2014, **53**, 11285–11288; (d) Y.-H. Sun, T.-Y. Sun, Y.-D. Wu, X. Zhang and Y. Rao, *Chem. Sci.*, 2016, **7**, 2229–2238; (e) X. Yang, G. Shan and Y. Rao, *Org. Lett.*, 2013, **15**, 2334–2337; (f) V. S. Thirunavukkarasu, S. I. Kozhushkova and L. Ackermann, *Chem. Commun.*, 2014, **50**, 29–39.
- D. C. M. Ferreira, I. Tapsoba, S. Arbault, Y. Bouret, M. S. A. Moreira, A. V. Pinto, M. O. F. Goulart and C. Amatore, *ChemBioChem*, 2009, **10**, 528.
- For examples of ruthenium-catalyzed C–H oxygenations under arene ligand-free conditions, see: V. S. Thirunavukkarasu, J. Hubrich and L. Ackermann, *Org. Lett.*, 2012, **14**, 4210–4213.
- For arene ligand-free ruthenium catalysts for C–H activations, see: (a) L. Ackermann, A. Althammer and R. Born, *Synlett*, 2007, 2833–2836; (b) L. Ackermann, R. Born and P. Álvarez-Bercedo, *Angew. Chem., Int. Ed.*, 2007, **46**, 6364–6367; (c) L. Ackermann, A. Althammer and R. Born, *Tetrahedron*, 2008, **64**, 6115–6124; (d) L. Ackermann and R. Vicente, *Org. Lett.*, 2009, **11**, 4922–4925; (e) P. Marce, A. J. Paterson, M. F. Mahon and C. G. Frost, *Catal. Sci. Technol.*, 2016, **6**, 7068–7076; (f) J. McIntyre, I. Mayoral-Soler, P. Salvador, A. Poater and D. J. Nelson, *Catal. Sci. Technol.*, 2018, **8**, 3174–3182; (g) M. Simonetti, D. M. Cannas, X. Just-Baringo, I. J. Vitorica-Yrezabal and I. Larrosa, *Nat. Chem.*, 2018, **10**, 724–731; (h) K. Korvorapun, N. Kaplaneris, T. Rogge, S. Warratz, A. C. Stückl and L. Ackermann, *ACS Catal.*, 2018, **8**, 886–892; (i) F. Fumagalli, S. Warratz, S.-K. Zhang, T. Rogge, C. Zhu, A. C. Stückl and L. Ackermann, *Chem. – Eur. J.*, 2018, **24**, 3984–3988.
- (a) G. A. Schmunis and Z. E. Yadon, *Acta Trop.*, 2010, **115**, 14; (b) Y. Jackson, V. M. Herrera and J. Gascon, *Bull. W. H. O.*, 2014, **92**, 771.
- (a) L. Conteh, T. Engels and D. H. Molyneux, *Lancet*, 2010, **375**, 239; (b) A. Rassi and J. A. Marin-Neto, *Lancet*, 2010, **375**, 1388.
- (a) C. Bern and S. P. Montgomery, *Clin. Infect. Dis.*, 2009, **49**, e52; (b) C. Bern, S. Kjos, M. J. Yabsley and S. P. Montgomery, *Clin. Microbiol. Rev.*, 2011, **24**, 655.
- WHO. *Expert Committee. Control of Chagas Disease*, World Health Organization, Brasilia, Brazil, 2002. WHO technical report series 905.
- (a) M. N. Soeiro and S. L. de Castro, *Expert Opin. Ther. Targets*, 2009, **13**, 105; (b) J. A. Urbina, *Acta Trop.*, 2010, **115**, 55.
- J. Rodrigues Coura and S. L. de Castro, *Mem. Inst. Oswaldo Cruz*, 2002, **97**, 3.
- L. Ackermann, *Chem. Rev.*, 2011, **111**, 1315–1345.

