Drawing on biology to inspire molecular design: a redox-responsive MRI probe based on Gd(III)-nicotinamide†

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A novel, reversible redox-active MRI probe, GdNR1, has been developed for the study of redox changes associated with diseased states. This system exhibits switching in relaxivity upon reduction and oxidation of the appended nicotinimidium. Relaxivity studies and cyclic voltammetry confirmed the impressive reversibility of this system, at a biologically-relevant reduction potential. A 2.5-fold increase in relaxivity was observed upon reduction of the complex, which corresponds to a change in the number of inner-sphere water molecules, as confirmed by luminescence lifetimes of the Eu(III) analogue and NMRD studies. This is the first example of a redox-responsive MRI probe utilising the biologically-inspired nicotinimidium redox switch. In the future this strategy could enable the non-invasive identification of hypoxic tissue and related cardiovascular disease.

The non-ionizing, non-invasive and unlimited tissue penetration of magnetic resonance imaging (MRI) offers an exceptional opportunity to visualize biological events with probes that modulate the MRI signal upon a signaling event.1 Numerous complexes have been developed for chemical stimuli such as enzymatic activity,2,3 Ca2+, Cu2+, Zn2+,4,5 or neurotransmitters,6 or complexes have been developed for chemical stimuli such as quinols,15 thiols16 and merocyanine/spyrooxazine.17 Herein, we have used the nicotinimidium moiety to append a redox switch to a Gd-based MRI contrast agent. The appended nicotinimidium is a redox active group that operates at a physiologically-relevant reduction potential and is increasingly being understood to play a role in various pathologies, including Alzheimer’s disease, cardiovascular disease, obesity, diabetes and arthritis.20,21 On the other hand, hypoxia occurs when blood supply to tissues is restricted in a variety of pathologies including solid tumors and atherosclerosis. We are particularly interested in the preparation of reversible contrast agents that can sense dynamic changes in biological redox environments,10,24 and here sought to develop a Gd-based reversible redox sensor.

In this study we report Gd-nicotinamide redox sensor 1 (GdNR1), a Gd(III) contrast agent with a redox-sensitive switch that operates at a physiologically-relevant reduction potential (Scheme 1). We chose a nicotinimidium-based redox sensor as this moiety is used in the redox-active NAD/NADH cofactor in biological systems.

DO3A derivatives are commonly synthesized starting from DO3A-OBu, but due to challenges in purification of the ligand, a less-commonly utilized approach was employed to synthesize GdNR1, enabling a facile final purification step (Scheme S1, ESI†). After alkylating nicotinamide with 1,3-dibromopropane and tetra-substituted cyclen (DOTA-OBu), which could be separated using aqueous/organic liquid–liquid extraction. TFA deprotection of the tBu ester groups and a Cl⁻–exchange column gave the free ligand with a single counter-ion. Irrespective of whether lanthanide(III) complexes were then prepared by complexation with lanthanide(III) chlorides in water.

† Electronic supplementary information (ESI) available: Characterisation and further NMRD details. See DOI: 10.1039/c8cc07092j

Scheme 1 GdNR1 in oxidized (left) and reduced (right) forms.
Electrospray ionization mass spectrometry (ESI-MS) of the ligand was performed to confirm the nature of reduced and oxidized species, with studies carried out before deprotection of the tertiary butyl ester groups due to very poor ionization of the reduced-neutral free ligand or complex. The charged nicotinimidium appeared at 677.4 (+ve m/z) and the reduced ligand was found as a sodium adduct (+ve m/z 701.4) (Fig. S9 and S10, ESI†). This single proton difference confirms the nicotinimidium-to-dihydronicotinamide reduction. \(^1\)H and \(^13\)C NMR characterization of the analogous diamagnetic La(III) complex showed the loss of the aromatic peaks and appearance of the expected reduced nicotinamide peaks upon reduction of the system (Fig. S11–S14, ESI†).

Cyclic voltammetry (CV) of the complex revealed well-separated redox peaks consistent with the two electron reduction of the nicotinimidium ring to dihydronicotinamide (Fig. 1). The electrochemical reversibility of the system was confirmed over 12 cycles, highlighting the advantages of our system over previously-reported contrast agents that exhibit only quasi-reversibility.\(^{25}\)

To investigate redox-induced changes in the hydration state of the complex, a luminescent Eu(III) analogue was synthesized and the luminescence lifetimes measured in H\(_2\)O and D\(_2\)O with addition of reductant, oxidant and bicarbonate. The hydration number \((q)\) of Eu(III) complexes can be estimated (±0.1 to 0.3) using the empirical formula:\(^{26}\)

\[
q_{\text{Eu}(\text{H}_2\text{O})} = 1.2(\Delta k_{\text{obs}} - 0.25 - 1.2q^{\text{NH}} - 0.075q^{\text{CONH}}),
\]

where \(q^{\text{NH}}\) and \(q^{\text{CONH}}\), the number OH or CONH groups participating in lanthanide coordination are 0 for this complex, and \(\Delta k_{\text{obs}}\) is calculated from the decay rate constants \(1/\tau_{\text{H}_2\text{O}} - 1/\tau_{\text{D}_2\text{O}}\), expressed in ms\(^{-1}\).

This analysis confirmed a decrease in the water coordination number from 1.9 to 0.3 upon oxidation (in non-degassed MES buffer, pH 7) (Table 1). This corresponds to a change from 2 bound water molecules in the reduced form to 0. We speculated that binding of a counteranion, promoted by the positively charged oxidized nicotinimidium, was responsible for occluding water binding. In non-degassed solutions and biological systems alike, bicarbonate is the major coordinating anion, with biological bicarbonate concentrations on the order of 20 mM, so we hypothesised that this ion binds to the oxidized form, as has previously been observed for responsive lanthanide complexes.\(^{27}\) Addition of HCO\(_3^−\) (10 eq.) to the buffered sample showed no change, and when this sample was oxidized, the \(q\) value decreased in the same manner. In comparison, in samples degassed by boiling and purging with argon to remove bicarbonate, the oxidized complex showed \(q = 2.1\), confirming our hypothesis that bicarbonate binding is responsible for this large change in hydration state. HEPES and MES buffers are reported to be mild reductants which can be used for synthesis of gold nanoparticles from AuCl\(_4^-\)\(^{28}\). This, together with a positive reduction peak observed in the CV (450 mV, Fig. 1) of GdNR1 could result in a partial reduction of the complex upon dissolution in the buffers, and subsequently higher \(r_1\) and \(q\) values observed by the luminescence lifetimes and NMRD measurements in buffered solution.

Nuclear magnetic resonance dispersion (NMRD) was used to study the relaxivity of the system in the oxidized and reduced forms in buffered non-degassed water. The \(r_1\) values of the native reduced complex in HEPES and MES buffers were 6.9 and 7.5 s\(^{-1}\) mM\(^{-1}\) respectively (Fig. 2 and Table S1, ESI†). Addition of the reductant sodium dithionite to GdNR1 in buffer showed no significant change to the NMRD profile (Fig. 2). H\(_2\)O\(_2\) was then used to oxidize the complex in buffered solution and the \(r_1\) was observed to decrease to 4.6 s\(^{-1}\) mM\(^{-1}\) at 60 MHz and 37 °C for both buffers.

The NMRD curve at low field confirmed the three-fold change in relaxivity from oxidized to reduced forms (Fig. 2). To further rule out any role for H\(_2\)O\(_2\) other than as an oxidant, we repeated the experiment with DOTAREM (Gd-DOTA) in clinical preparation, and observed no difference in the presence of excess H\(_2\)O\(_2\) (Fig. S16, ESI†).

The system could be reversibly switched between oxidized and reduced forms, although repeated cycling was limited by the buffering capacity due to the acidity of both H\(_2\)O\(_2\) and sodium dithionite (Fig. 3A). The synthesized version of the probe bearing reduced nicotinamidate showed a relaxation rate of 6.9 s\(^{-1}\) mM\(^{-1}\) at 60 MHz and 37 °C. Upon oxidation with H\(_2\)O\(_2\), the relaxation rate decreased to 3.7 s\(^{-1}\) mM\(^{-1}\). This clear change in relaxation rate with oxidation and reduction could also be observed in phantom images of tubes containing oxidized and reduced GdNR1 (Fig. 3B).

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**Table 1** Luminescence lifetimes and hydration numbers in water or MES buffer (20 mM, pH 7) at 298 K

<table>
<thead>
<tr>
<th></th>
<th>(\tau_{\text{H}_2\text{O}}) ms</th>
<th>(\tau_{\text{D}_2\text{O}}) ms</th>
<th>(q) H(_2)O</th>
</tr>
</thead>
<tbody>
<tr>
<td>EuNR1-reduced(^a) (MES, non-degassed)</td>
<td>0.32</td>
<td>0.80</td>
<td>1.9</td>
</tr>
<tr>
<td>EuNR1-reduced(^b) (MES, non-degassed)</td>
<td>0.44</td>
<td>0.55</td>
<td>0.3</td>
</tr>
<tr>
<td>EuNR1-oxidized(^a) + HCO(_3^−) (10 eq.)</td>
<td>0.34</td>
<td>0.95</td>
<td>1.9</td>
</tr>
<tr>
<td>EuNR1-oxidized(^a) (MES, non-degassed)</td>
<td>0.42</td>
<td>0.56</td>
<td>0.4</td>
</tr>
<tr>
<td>EuNR1-oxidized(^b) (H(_2)O &amp; D(_2)O degassed)</td>
<td>0.26</td>
<td>0.57</td>
<td>2.1</td>
</tr>
</tbody>
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\(^{a}\) Average of 3 measurements that vary by less 0.01 ms. \(^{b}\) Reduced by buffer.
Consistent with our studies of the Eu complex, the $r_1$ of the oxidized form was observed to be 7.6 s⁻¹M⁻¹ at 60 MHz and 37 °C when the water was degassed (boiling and argon purging) and the relaxivity did not change significantly upon addition of either oxidant ($H_2O_2$, 10 eq.) or reductant (sodium dithionite). In contrast, in the presence of bicarbonate, the $r_1$ decreased more than double upon oxidation (Fig. 4 inset). Upon reduction with sodium dithionite in the presence of bicarbonate, the $r_1$ value recovers to near that of the sample in high-purity degassed water of 7.1 s⁻¹M⁻¹ at 60 MHz and 37 °C. This is consistent with our hypothesis from the Eu lifetime studies that the changing charge of the complex from +1 to 0 modulates the binding and release of bicarbonate. This in turn affects water accessibility to the Gd(III) paramagnetic centre, one of the main parameters determining the relaxivity.

Investigating the role of competing species in the relaxation process, we measured the changes in relaxivity upon the addition of citrate, lactate, HSA and in F12 (+10% FBS) cell culture medium (GdNR1 1 mM). Citrate (0.13 mM), lactate (2.3 mM) and cell culture medium did not disturb the relaxivity switching (Fig. S17, ESI†). In 4% HSA solution, relaxivity increased indicating non-covalent binding of GdNR1 to the protein associated with an increased rotational correlation time; however, the switching of the system was not impeded (Fig. S17, ESI†). Lowe et al. previously observed this phenomenon in DO3A derived complexes, where water coordination decreased up to five-fold upon binding of endogenous anions such as bicarbonate and relaxivity dramatically increased with HSA.²⁹

In HEPES and MES the oxidized complex showed an increase in relaxivity in the NMRD curve from 20 to 60 MHz, characteristic of aggregation.³⁰–³⁵ Dynamic light scattering confirmed the presence of ~100 nm aggregates in the MES buffered solution with $H_2O_2$ added (Fig. S19, ESI†). However, this aggregation was not visibly observed in the presence of HSA or in cell culture media, so it is likely that under these conditions the complex has sufficient solubility to overcome this problem.

Fitting of proton NMRD profiles to Solomon-Bloembergen-Morgan theory (ESI† text, Fig. 2 and 4) was performed to evaluate the effect of hydration number and other parameters (especially $\tau_R$) on relaxivity, using parameters previously described for Gd(III) complexes (Table S2, ESI†).²²,²³ As expected for a small DO3A derived complex the rotational correlation time ($\tau_R$) is short. Fitting the NMRD data from Fig. 4 before oxidation shows a very similar result to that observed for GdNR1 in high-purity water. Upon oxidation of GdNR1 with $H_2O_2$, the shape of the NMRD profile changed dramatically with a peak in relaxivity occurring at higher Larmor frequency (100 MHz). Such a profile is typical of a...
slow tumbling system, and the slow tumbling implies that aggregation has occurred. However, the relaxivity is much lower than expected for a slow tumbling Gd-complex with inner-sphere water ligand(s), and this low relaxivity implies that the hydration number is decreased and likely q is close to 0. We then investigated the effects of pH on relaxivity. \( r_1 \) is constant over \( pH 3-6 \), decreases by 25% at \( pH 7 \), and further at \( pH 9 \) (Fig. S18, ESI†). This relaxivity decrease between \( pH 6 \) and 7 correlates with the first pK\(_a\) of carboxic acid, at \(~6.4\), suggesting that it may be result from formation of bicarbonate from water-dissolved CO\(_2\) from air and its subsequent coordination to the Gd centre. At \( pH 9 \), the relaxivity drops to \(~4\ s^{-1} \cdot M^{-1} \), higher than the relaxivity of \(~2.5\ s^{-1} \cdot M^{-1} \) (Fig. 4 inset) measured for GdNR1 in pure water spiked with bicarbonate (pH 8–9), indicating that the effects of the environment on relaxivity of the complex are a combination of bicarbonate-related as well as other pH-related mechanisms, as described previously for Gd-based complexes.\(^{29,40}\)

In summary, we report the first example of a reversible redox-responsive MRI probe based on a Gd(m)-DO3A nicotinamide chelate, and provide a detailed insight into its mechanism and performance. Using a novel synthetic approach, it was possible to produce the final chelate in a few facile steps. The system shows response to sodium dithionite and H\(_2\)O\(_2\). There is a large signal enhancement upon reduction of the nitricimidium in the presence of bicarbonate. Luminescence lifetimes of the Eu(ni) analogue revealed a change in binding of inner-sphere water molecules upon oxidation and reduction events. Bicarbonate is endogenous to blood serum and would be available to function with this MRI probe for signal suppression of the oxidised form. GdNR1 represents a useful biological application of this probe, we have confirmed its non-suppression of the oxidised form. GdNR1 represents a useful responsive MRI probe based on a Gd(III)-DO3A nicotinamide chelate, and provide a detailed insight into its mechanism and performance.

Notes and references


Conflicts of interest
There are no conflicts to declare.