

Cite this: *Chem. Commun.*, 2018, 54, 13355Received 19th July 2018,  
Accepted 3rd October 2018

DOI: 10.1039/c8cc05899g

rsc.li/chemcomm

## Size-controlled and water-soluble gold nanoparticles using UV-induced ligand exchange and phase transfer†

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**Oleylamine-capped gold nanoparticles (AuNPs) with sizes ranging from 5 to 13 nm and narrow size distributions (<10%) are synthesized by using a seeded growth approach. Water-solubility is achieved by using a UV-induced ligand exchange approach, resulting in transfer from the organic to an aqueous phase.**

Gold nanoparticles (AuNPs) are widely applied because of their characteristic size-dependent physical and chemical properties.<sup>1–5</sup> Some of their most important applications include staining in electron microscopy,<sup>6,7</sup> catalysis,<sup>8</sup> and biosensing.<sup>9–14</sup> Many of these applications – particularly the biological ones – need well-defined, water-dispersed AuNPs.<sup>15</sup> The most well-known method to synthesize water-dispersed AuNPs was developed by Turkevich in 1951.<sup>16</sup> Unfortunately, this method has inherent drawbacks such as the stability of AuNPs and the broad size distribution (>10%) of smaller (<20 nm) AuNPs.<sup>17</sup> Synthesizing AuNPs in an organic solvent, on the other hand, allows more control over the nanoparticle size and narrow size distributions (<10%) compared to similar-sized AuNPs synthesized in an aqueous phase.<sup>1,17–20</sup> The most widely used strategy to synthesize AuNPs in an organic solvent is the Schiffrin–B Brust method.<sup>21</sup> This method yields AuNPs with a narrow size distribution using a strong coordinating thiol ligand.<sup>18,22,23</sup> Peng *et al.* developed a method to synthesize 5 nm AuNPs with a narrow size distribution, by using a weakly coordinating oleylamine ligand.<sup>8</sup> However, as these AuNPs are exclusively soluble in organic solvents, their application in aqueous, *e.g.* biological, systems is restricted. Various methods have been developed to render hydrophobic nanoparticles water soluble, such as ligand exchange.<sup>24–28</sup> An interesting variation of this method is the UV-induced *in situ* ligand exchange and phase transfer approach as reported by the group of Mattoussi. Using this approach, the native, hydrophobic

and weakly coordinating ligand on semiconductor nanoparticles is replaced by a hydrophilic, multicoordinating lipoic acid derivative. These quantum dot nanoparticles then transferred from organic to aqueous solvents, and showed excellent colloidal stability.<sup>3,29,30</sup> We now show that oleylamine-capped AuNPs of well-defined and tunable size can be transformed into water-soluble gold nanoparticles of identical size by applying the UV-induced ligand exchange approach.

Here, we report a strategy to obtain water-soluble AuNPs with sizes ranging from 5 to 13 nm and narrow size distributions (<10%). Our strategy is as follows (Scheme 1): 4.5 nm oleylamine-capped AuNP seeds were synthesized using Peng *et al.*'s method.<sup>8</sup> Subsequently, these AuNPs were used in a seeded growth reaction to obtain sizes of 7–13 nm. The AuNP phase transferred quantitatively to the aqueous phase upon UV-induced photoligation of lipoic acid.<sup>29</sup> The process of phase transfer was indicated by the change of color of the bottom aqueous layer to deep-red and the completion by the loss of color of the top organic layer.

The AuNPs produced using the strategy presented in Scheme 1 are shown in Fig. 1. Starting from 4.5 nm oleylamine-capped AuNP seeds, we obtained AuNPs with sizes ranging from 7 to 13 nm (A–E) *via* a seeded growth approach. Seeded growth reactions rely on the slow reduction and deposition of a gold precursor on a seed, resulting in controlled growth of the seeds.<sup>31</sup> Transmission electron microscopy (TEM) images showed that the AuNPs form large, hexagonally ordered crystals when deposited on a TEM grid, indicating their narrow size distributions.<sup>31</sup> This infers that the narrow size distribution of the seeds is retained throughout the seeded growth approach (Fig. S1, ESI†). The UV-Vis absorbance spectra of these AuNPs showed sharp localized surface plasmon resonance (LSPR) peaks with peak maxima that characteristically redshift for bigger particles, from 513 nm (seeds) to 530 nm (E).<sup>8</sup> DLS characterization of the seeds and samples A–E showed the absence of aggregates in the correlation curves (Fig. S2, ESI†), and by fitting the data, we found for all samples that the sizes were in concordance with the TEM analysis and the UV-Vis measurements (Fig. S1, ESI†).

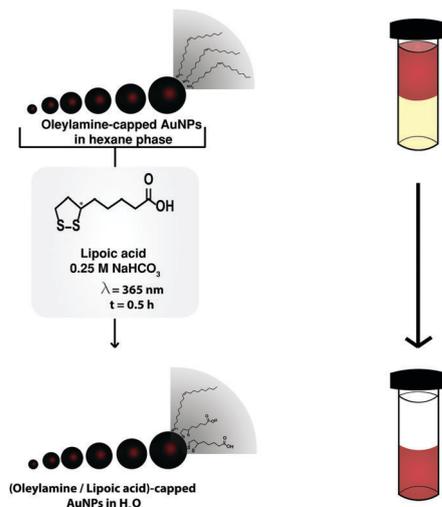
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† Electronic supplementary information (ESI) available. See DOI: 10.1039/c8cc05899g

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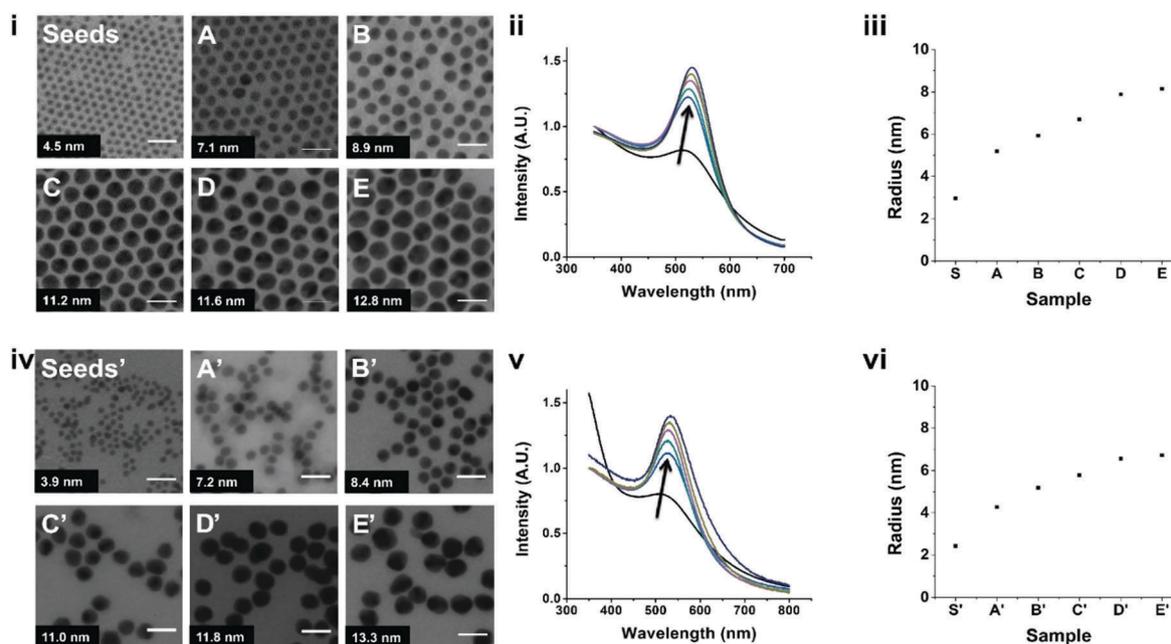


**Scheme 1** Schematic representation of UV-induced phase transfer of hydrophobic AuNPs. AuNPs, suspended in an organic phase, are mixed with an aqueous solution of lipoic acid in 0.25 M NaHCO<sub>3</sub>. Upon UV-irradiation for 30 minutes, the AuNPs quantitatively phase-transfer from the organic to the aqueous layer.

Next, these samples were subjected to the UV-induced ligand exchange and phase transfer. The AuNPs quantitatively phase transferred from the organic phase to water, which generally proceeded in less than 30 minutes. We do note here that with increasing AuNP size, the time to achieve complete phase transfer also increased from minutes to about an hour. TEM on these AuNPs after phase transfer (seeds' and A'-E') showed

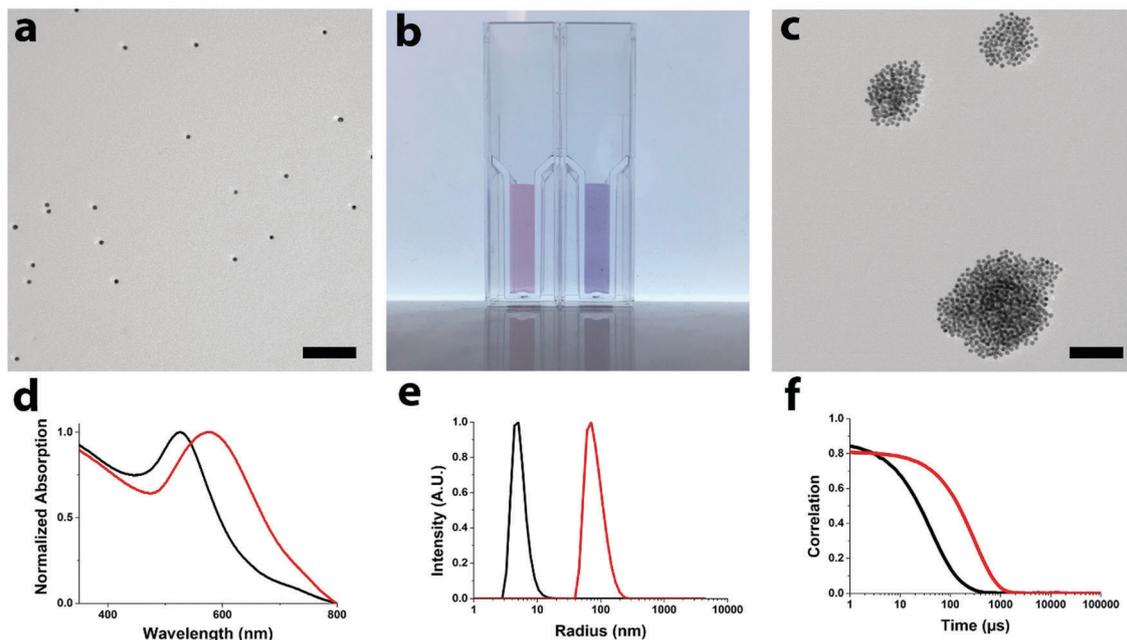
that the sizes and size distributions of the samples were comparable to those before ligand exchange (Fig. S3 and Table S1, ESI<sup>†</sup>). Characterization *via* UV-Vis spectroscopy showed the same trend as before phase transfer, with a slight redshift of the LSPR peak ( $\sim 3$  nm, Table S1, ESI<sup>†</sup>), which is attributed to the differences in solvent polarity and capping ligand.<sup>32,33</sup> Finally, DLS characterization (Fig. S4, ESI<sup>†</sup>) showed the absence of aggregates. The measured diameters were  $\pm 1$  nm smaller after phase transfer and we attribute this to the differences in size of the ligands. Finally, <sup>1</sup>H nuclear magnetic resonance (NMR) was used to analyze the surface composition of the AuNPs, after seeded growth and ligand exchange (A'-E'). The AuNPs were digested using KCN to release surface-bound ligands into solution, which eliminates effects of severe line broadening in the <sup>1</sup>H NMR spectra.<sup>34-37</sup> For relative ligand quantification, the integrated peak intensity associated with lipoic acid ( $\delta = 3.5-3.2$  ppm) was compared to the one of oleylamine ( $\delta = 1.15-0.85$  ppm).<sup>37</sup> These spectra showed that, after ligand exchange, the AuNPs are capped with lipoic acid with some (9-14%) residual oleylamine still present (Fig. S5 and Table S2, ESI<sup>†</sup>). We have also applied this method to sample A' with longer reaction times for the exchange and phase transfer. The spectra showed that increasing the exchange reaction time does not affect the amount of residual oleylamine on the AuNP surfaces (Table S3, ESI<sup>†</sup>).

AuNPs find applications in a plethora of areas, including detection of analytes *via* aggregation of multivalent networks.<sup>10-13</sup> Here we present the sensing properties of the water-soluble AuNPs showing a color change from red to purple, corresponding to a red shift in optical absorption, upon aggregation



**Fig. 1** UV-induced ligand exchange and phase transfer of AuNPs from (i-iii) organic to (iv-vi) aqueous phase. (i and iv) TEM images of AuNP seeds and AuNPs obtained after the seeded growth (A-E). (iv) The respective seeds (seeds') and samples (A'-E') after phase transfer. Scale bars are 20 nm. (ii and v) Normalized UV-Visible absorption spectra before (ii) and (v) after phase transfer, showing a clear red shift of the LSPR peaks. The spectra are normalized at  $\lambda = 350$  nm and are shifted vertically to facilitate comparison between the samples. (iii and vi) Hydrodynamic radii of the AuNPs, as determined from number-averaged DLS fits of the AuNPs (iii) before and (vi) after ligand exchange.





**Fig. 2** Formation of PAMAM–AuNP composite upon the addition of  $\sim 3$  nmoles of PAMAM-G5-NH<sub>2</sub> (+) to a solution containing  $\sim 3$  nmoles of AuNPs (–). (a) TEM micrograph of suspended sample B'. (b) Photograph showing B' without (left) and with (right) PAMAM-G5-NH<sub>2</sub>. After the addition of PAMAM-G5-NH<sub>2</sub> the color of the solution changes from red to purple. (c) TEM micrograph of the formed PAMAM–AuNP composite, showing that extended networks formed. (d) UV-Visible absorption spectra of AuNPs with (red) and without (black) PAMAM-G5-NH<sub>2</sub>. These spectra show that the addition of PAMAM-G5-NH<sub>2</sub> results in a redshift of the LSPR peak. (e and f) DLS characterization of AuNPs with (red) and without (black) added PAMAM-G5-NH<sub>2</sub>. The number-weighted size fit (e) indicates that the AuNP hydrodynamic radius is  $\sim 5$  nm, whereas the hydrodynamic radius of the PAMAM–AuNP composite is  $\sim 80$  nm. The DLS correlation curve (f) shows that the addition of PAMAM-G5-NH<sub>2</sub> results in an increase of the correlation time, indicative of the formation of larger particles. Scale bars in (a) and (c) represent 100 nm.

with amine-terminated fifth generation poly(amidoamine), PAMAM-G5, dendrimers (see Fig. 2).<sup>38</sup> The formation of multivalent networks is further corroborated by TEM and DLS measurements. Control experiments with the addition of water, salt, and negatively and positively charged PAMAM, and in buffer (see the ESI,† S3) prove the selectivity of these AuNPs for positively charged PAMAM.

Here we showed a method to prepare size-controlled and water-soluble AuNPs. 4.5 nm oleylamine-capped AuNP seeds were synthesized in an organic solvent. The AuNP sizes were then tuned from 4.5 to 7–13 nm *via* a seeded growth reaction approach, while retaining the narrow size distributions. By combining these oleylamine-capped AuNPs with UV-induced photoligation of lipolic acid, we achieved quantitative phase transfer of the AuNPs to the aqueous phase, proceeding due to ligand exchange of oleylamine for lipolic acid. We believe that the here reported strategy is a straightforward and robust approach to transfer AuNPs from organic to aqueous solvents. This approach can be applied to obtain well-defined AuNPs in high concentrations, for use in studies where control over the nanoparticle size, size distribution and surface composition is of importance.

JBTH and AHV thank NWO for financial support of the MONALISA project 717.013.006. We thank the Wageningen Electron Microscopy Centre for their support with the TEM measurements.

## Conflicts of interest

There are no conflicts to declare.

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