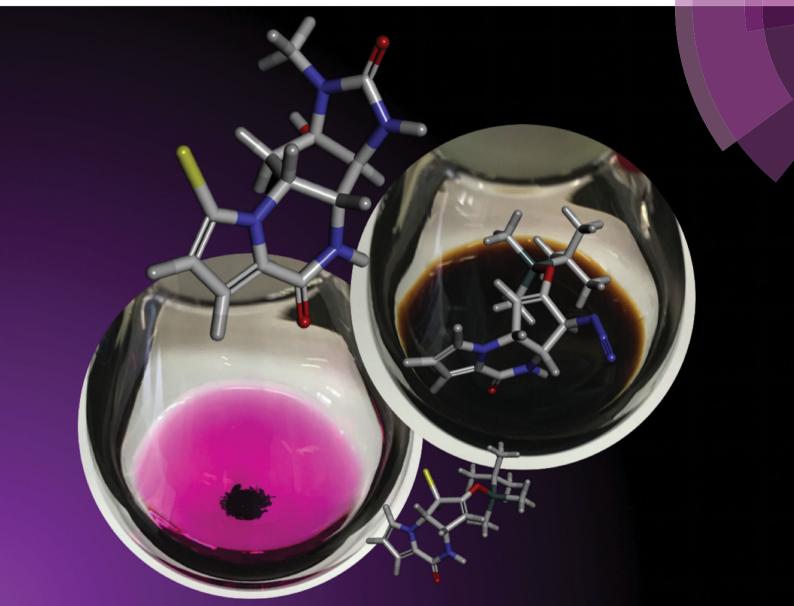
ChemComm

Chemical Communications

rsc.li/chemcomm



ISSN 1359-7345



ChemComm



COMMUNICATION

View Article Online



Cite this: Chem. Commun., 2018, 54 9893

Received 14th July 2018, Accepted 1st August 2018

DOI: 10.1039/c8cc05697h

rsc.li/chemcomm

Total synthesis of (–)-agelastatin A: an $S_H 2'$ radical azidation strategy†

Izuru Tsuchimochi, ab Yuta Kitamura, b Hiroshi Aoyama, b Shuji Akai, b Keiyo Nakai a and Takehiko Yoshimitsu*a

A reagent generated from TMSN₃/KMnO₄/BnEt₃NCl was found to promote an S_H2' radical azidation of a bromo silyl enol ether to furnish an azido silyl enol ether via olefin transposition. With the present azidation protocol, a new synthetic approach to agelastatin A, a potent antitumor marine alkaloid, has been established.

(-)-Agelastatin A (1), along with its congener agelastatin B (2), was first isolated as a cytotoxic constituent from the Coral Sea sponge Agelas dendromorpha by Pietra and co-workers in 1993 (Fig. 1). Thereafter, Molinski and co-workers identified the Indian Ocean sponge Cymbastela sp. as another source that produces 1 along with agelastatins C (3) and D (4), two additional agelastatin members.² In 2010, Al-Mourabit and co-workers reported the isolation of agelastatins E (5) and F (6) from the New Caledonian sponge A. dendromorpha.3 Early biological assessments of agelastatins conducted by the aforementioned laboratories have revealed that compound 1 exhibits remarkable properties, including antitumor activity, 1,3 brine shrimp toxicity, 2 and insecticidal activity.2 In addition, Meijer and Pettit have found that agelastatin A (1) is a potent inhibitor of GSK-3\beta, a pivotal serine/threonine kinase.4 Hale and El-Tanani have reported that agelastatin A (1) dramatically decreases β-catenin levels in cancer cells and inhibits cancer cell proliferation by arresting cell cycle at G2 phase.⁵

The biological significance of agelastatin A (1) has made it an attractive target for medicinal studies.^{6,7} For instance, Movassaghi's comparative cytotoxicity assay of all agelastatin members, i.e., A (1) to F (6), has successfully validated the relevance of agelastatin A (1)



Fig. 1 Agelastatin alkaloids.

as a promising anticancer agent.78 In addition, structure-activity relationship (SAR) studies on agelastatin analogues have recently been disclosed by the groups of Molinski,8 Romo/Liu,9 and Movassaghi, ¹⁰ boosting the applications of agelastatin particularly to blood cancer chemotherapy.

Our group has also been engaged in synthetic and medicinal studies on 1 and has demonstrated that agelastatin analogues potentially attenuate brain cancer. 11 Furthermore, our SAR study has revealed that structural modifications of the N1-substituent of the D-ring of 1 could retain the in vitro and in vivo therapeutic efficacies of agelastatin analogues. 12,13 Movassaghi's group has further clarified that D-ring modifications expand the scope of derivatization of agelastatins to access potent analogues.¹⁰

In the present study, we have established a new route to agelastatin A (1) through an SH2' radical azidation protocol using TMSN₃/KMnO₄/BnEt₃NCl that enables the allylic transposition of a bromo silyl enol ether into an azido silyl enol ether, which serves as a useful D-ring precursor of the target natural product (Scheme 1).

The synthesis was commenced with Boc-protected aminoalcohol derivative 7 (>99% ee). 14 The Boc group of 7 was removed with hydrochloric acid (HCl) generated in situ from TMSCl in aq. MeCN to provide an ammonium salt (structure not shown). After evaporation of the solvents under reduced pressure, the resultant crude product was coupled with pyrrole-2-carboxylic acid using EDC, Et₃N, and DMAP in MeCN to furnish compound 8 in 84% yield. Then, compound 8 was hydrolyzed with LiOH in aq. EtOH to provide alcohol 9 in 88% yield. PDC oxidation of alcohol 9 in DMF delivered enone 10 in

^a Division of Pharmaceutical Sciences, Graduate School of Medicine, Dentistry, and Pharmaceutical Sciences, Okayama University, 1-1-1 Tsushima-naka, Kita-ku, Okayama 700-8530, Japan. E-mail: yoshimit@okayama-u.ac.jp; Tel: +81-86-251-7930

^b Graduate School of Pharmaceutical Sciences, Osaka University, 1-6 Yamadaoka, Suita, Osaka 565-0871, Japan

[†] Electronic supplementary information (ESI) available: Experimental procedures, characterization of new compounds including NMR spectra. CCDC 1852628. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c8cc05697h

Communication ChemComm

Scheme 1 Total synthesis of (-)-agelastatin A (1).

80% yield, which, upon treatment with Cs₂CO₃ in THF, gave tricyclic ketone 11 via a conjugate addition of the pyrrole nitrogen to the enone double bond. No racemization at C4 position took place in this transformation $(9 \rightarrow 10 \rightarrow 11)$, retaining the optical purity of 11 (>99% ee). Then, ketone 11 was subjected to enolization with NaHMDS followed by O-silylation with tert-butyldimethylsilyl chloride to produce silyl enol ether 12 along with its minor regioisomer 16 (12:16 = 8.3:1) (Fig. 2). Obviously, major product 12 was not ideal for further functionalization as it lacked a reactive alkene functionality at C5 position. However, we found that 12 and 16 underwent olefin isomerization with a trace acid probably due to their strained nature.16 Therefore, we envisioned that the brominative olefin transposition of 12 would take place via a bromonium formation followed by deprotonation to allow net olefin transposition that affords an enol ether suitable for C5 functionalization. To our delight, the treatment of silyl enol ether 12 with

Fig. 2 TBS enol ethers 12 and 16 generated from 11

N-bromophthalimide (NBP) was found to deliver allylic bromide 13 in stereoselective and regiospecific manners as we had expected.

With compound 13 in possession, the nitrogen functionalization at C5 position was examined to access key intermediate 14 (Table 1). An attempted ionic S_N2' azidation of 13 with NaN₃ in DMF was unsuccessful (entry 5), giving rise to a desilylated product. To this end, we expected that the electrophilic nitrogen radical species would preferentially undergo an addition reaction with the electron-rich enol double bond to facilitate S_H2' radical azidation to deliver compound 14.

An azide radical is known to be generated from an anionic azide by oxidation processes. The Magnus protocol represents such an example, which utilizes trimethylsilylazide (TMSN₃) in combination with iodosylbenzene (PhIO) in CH2Cl2 at low temperature (-78 °C). The Magnus method was proved to afford desired product 14 albeit in moderate yield (entry 8).¹⁷ Therefore, we sought a new reagent system to deliver an azido radical and found that the treatment of 13 with TMSN₃ (10 equiv.)/ KMnO₄ (0.3 equiv.)/BnEt₃NCl (0.3 equiv.) successfully produced azide 14 in 50% yield along with regioisomeric azide 17 (21%) and bromide 18 (3%)¹⁸ (entry 1). In the absence of KMnO₄, no reaction took place and unreacted 13 was recovered (entries 6 and 7). When catalytic KMnO₄ (0.1 equiv.) was used in combination with TMSN₃ (10 equiv.) and BnEt₃NCl (0.1 equiv.) in either the presence or absence of molecular oxygen (O2), the chemical yield was low, suggesting that catalysis by O₂ in the present radical azidation was not operative (entries 2 and 3). Increasing the amount of Mn(vII) reagent was found to have no impact on the improvement of the chemical yields (entry 4).

It should be mentioned that the addition of TMSN3 to the mixture of KMnO₄ and BnEt₃NCl at 0 °C caused the evolution of molecular nitrogen (N2) accompanied by a color change of the solution from purple to dark brown, suggesting the production of low-valent manganese species from the Mn(vII) reagent. Although the reactive species responsible for the present radical azidation remains unclear, we assume that permanganate(vII) (MnO₄⁻) reacts with TMSN₃ to generate a low-valent mangan azide complex that serves as a metastable azide radical source. To clarify this hypothesis, we measured the amount of nitrogen gas (N2) that was generated from the reagent system. When KMnO₄ (0.33 mmol) was treated with BnEt₃NCl (0.33 mmol) and a large excess of TMSN₃ (11.1 mmol), 20-24 mL (ca. 0.9-1.1 mmol) of molecular nitrogen, which corresponds to ca. 3.0 equiv. relative to 1.0 equiv. of permanganate ion (MnO₄⁻), was generated. Assuming that 1.0 equiv. of permanganate reacts with 5.0 equiv. of TMSN₃ to produce 3.0 equiv. of molecular nitrogen, we propose that a pentavalent Mn(v) species is produced from the Mn(vII) species (Scheme 2).

Jiao and co-workers have reported that Mn(III) generated from MnBr₂ in the presence of molecular oxygen serves as an effective catalyst to generate an azide radical from TMSN₃. 19a We have examined Mn(OAc)₃·2H₂O in combination with TMSN₃ (6 equiv.) as a possible source of azido radical and found that desired material 14 could be similarly produced in 42% yield along with 17 (8%) (entry 9). 19b This result suggests that Mn(III) azide complex is likely responsible for the present radical azidation. Based on these observations, we currently assume that metastable Mn(v)

ChemComm Communication

Table 1 Azidation of bromide 13 with various reagents

			Yield ^a (%)			
Entry	Reagents (equiv.)	Time	14	17	18	13^{b}
1	KMnO ₄ (0.3), BnEt ₃ NCl (0.3), TMSN ₃ (10), MeCN	40 min	50	21	3	Trace
2	$KMnO_4$ (0.1), $BnEt_3NCl$ (0.1), $TMSN_3$ (10), $MeCN$	40 min	30	4	12	30
3	KMnO ₄ (0.1), BnEt ₃ NCl (0.1), TMSN ₃ (10), MeCN, O ₂	40 min	31	5	11	21
4	KMnO ₄ (0.6), BnEt ₃ NCl (0.6), TMSN ₃ (10), MeCN	40 min	43	11	4	9
5	NaN_3 (1.1), DMF^c	15 min	_	_	_	<u></u> d
6	$BnEt_3NCl$ (0.3), $TMSN_3$ (10), $MeCN$	75 min	_	_	_	90
7	TMSN ₃ (10), MeCN	70 min	_	_	_	89
8	PhIO (1.2), TMSN ₃ (2.4), $CH_2Cl_2^e$	40 min	24	17	14	6
9	$Mn(OAc)_3 \cdot 2H_2O(3)$, $TMSN_3(6)$, $MeCN^c$	11 h	42	8	_	_

^a Isolated yields after purification by column chromatography. ^b Recovered unreacted starting material. ^c The reaction was conducted at r.t. ^d Bromoketone (60%) was produced. ^e The reaction was conducted at -78 °C.

Scheme 2 Plausible generation of Mn(v) azide species

The formation of compounds 17 and 18, which provides an insight into the mechanism of the present azidation, also requires elaboration (Scheme 3). When azide 14 and isomeric azide 17 were separately subjected to the same reaction conditions for 1 h, only a trace amount of corresponding azide 17 and 14 was produced along with the unreacted starting azides, respectively. This indicates that both azides 14 and 17, once produced, were hardly susceptible to the $S_{\rm H}2'$ azidation. In contrast, when isomeric bromide 18 was treated with the reagent, compounds 14 (34%), 17 (27%), and 18 (12%) were obtained similar to the case of 13. Based on these results, we propose that the addition of an azide radical to bromide 13 generates a Br radical that undergoes rapid addition to substrate 13 to generate regioisomeric bromide 18. Then, 18 is further converted into compound 17 via a radical azidation.

With azide 14 in possession, we further endeavored to accomplish the total synthesis. Thus, azide 14 was subjected to catalytic hydrogenation followed by one-pot urea formation with Batey's reagent²⁰

Scheme 3 Plausible mechanisms of the production of regioisomeric byproducts **17** and **18**.

and subsequent desilylative cyclization with CsF to afford tetracyclic compound **15** in 51% yield over three steps. It should be mentioned that no purification was required in the three-step sequence, allowing ease of experimental operations. Finally, the known bromination protocol was applied to compound **15** to furnish (—)-agelastatin A (1).

In conclusion, we have established a new approach to (-)-agelastatin A (1) by the strategic implementation of brominative olefin transposition and subsequent S_H2' radical azidation. The present approach features a late-stage construction of D-ring that would allow facile production of D-ring analogues. We believe that the present synthesis would facilitate further development of new agelastatin analogues.

This work was supported by a grant [KAKENHI #15K14977] generously provided by the Ministry of Education, Culture, Sports, Science and Technology of Japan (MEXT), The NOVARTIS Foundation (Japan) for the Promotion of Science, and the Hoansha Foundation.

Conflicts of interest

The authors declare no conflicts of interest.

Notes and references

(a) M. D'Ambrosio, A. Guerriero, C. Debitus, O. Ribes, J. Pusset,
 S. Leroy and F. J. Pietra, J. Chem. Soc., Chem. Commun., 1993, 1305;
 (b) M. D'Ambrosio, A. Guerriero, G. Chiasera and F. Pietra,

Communication ChemComm

- Helv. Chim. Acta, 1994, 77, 1895; (c) M. D'Ambrosio, A. Guerriero, M. Ripamonti, C. Debitus, J. Waikedre and F. Pietra, Helv. Chim. Acta, 1996, 79, 727.
- 2 T. W. Hong, D. R. Jímenez and T. F. Molinski, J. Nat. Prod., 1998, 61, 158.
- 3 S. Tilvi, C. Moriou, M. Martin, J. Gallard, J. Sorres, K. Patel, S. Petek, C. Debitus, L. Ermolenko and A. Al-Mourabit, *J. Nat. Prod.*, 2010, 73, 720.
- 4 (a) L. Meijer, A. M. Thunnissen, A. W. White, M. Garnier, M. Nikolic, L. H. Tsai, J. Walter, K. E. Cleverley, P. C. Salinas, Y. Z. Wu, J. Biernat, E. M. Mandelkov, S. H. Kim and G. R. Pettit, *Chem. Biol.*, 2000, 7, 51; (b) G. R. Pettit, S. Ducki, D. L. Herald, D. L. Doubek, J. M. Schmidt and J.-C. Chapuis, *J. Oncol. Res.*, 2005, **15**, 11.
- 5 (a) C. K. Mason, S. McFarlane, P. G. Johnston, P. Crowe, P. J. Erwin, M. M. Domostoj, F. C. Campbell, S. Manaviazar, K. J. Hale and M. El-Tanani, Mol. Cancer Ther., 2008, 7, 548; (b) M. Harmata, Strategies and Tactics in Organic Synthesis, Elsevier Academic Press, London, 2005, ch. 11, vol. 6, pp. 352–394.
- 6 For selected reviews on total synthesis of agelastatins, see: (a) G. Dong, Pure Appl. Chem., 2010, 82, 2231; (b) T. Yamaoka, Y. Ichikawa and H. Kotsuki, J. Synth. Org. Chem., Jpn., 2012, 70, 615.
- 7 (a) D. Stien, G. T. Anderson, C. E. Chase, Y. Koh and S. M. Weinreb, J. Am. Chem. Soc., 1999, 121, 9574; (b) K. S. Feldman and J. C. Saunders, J. Am. Chem. Soc., 2002, 124, 9060; (c) K. S. Feldman, J. C. Saunders and M. L. Wrobleski, J. Org. Chem., 2002, 67, 7096; (d) K. J. Hale, M. M. Domostoj, D. A. Tocher, E. Irving and F. Scheinmann, Org. Lett., 2003, 5, 2927; (e) M. M. Domostoj, E. Irving, F. Scheinmann and K. J. Hale, Org. Lett., 2004, 6, 2615; (f) F. A. Davis and J. Deng, Org. Lett., 2005, 7, 621; (g) B. M. Trost and G. Dong, J. Am. Chem. Soc., 2006, 128, 6054; (h) Y. Ichikawa, T. Yamaoka, K. Nakano and H. Kotsuki, Org. Lett., 2007, 9, 2989; (i) D. P. Dickson and D. J. Wardrop, Org. Lett., 2009, 11, 13414; (j) N. Hama, T. Matsuda, T. Sato and N. Chida, Org. Lett., 2009, 11, 2687; (k) P. M. Wehn and J. Du Bois, Angew. Chem., Int. Ed., 2009, 48, 3802; (l) F. A. Davis, J. Zhang, Y. Zhang and H. Qiu, Synth. Commun., 2009, 39, 1914; (m) B. M. Trost and G. Dong, Chem. - Eur. J., 2009, **15**, 6910; (n) M. Movassaghi, D. S. Siegel and S. Han, Chem. Sci., 2010, 1, 561; (o) Y. Menjo, A. Hamajima, N. Sasaki and Y. Hamada, Org. Lett., 2011, 13, 5744; (p) T. Kano, R. Sakamoto, M. Akakura and K. Maruoka, J. Am. Chem. Soc., 2012, 134, 7516; (q) J. C. P. Reyes and D. Romo, Angew. Chem., Int. Ed., 2012, 51, 6870; (r) P. A. Duspara and R. A. Batey, Angew. Chem., Int. Ed., 2013, 52, 10862; (s) A. H. Antropow,

- K. Xu, R. J. Buchsbaum and M. Movassaghi, *J. Org. Chem.*, 2017, **82**, 7720; (t) Y. Yao, X. Wang and G. Liang, *Tetrahedron*, 2017, **73**, 4538.
- 8 E. P. Stout, M. Y. Choi, J. E. Castro and T. F. J. Molinski, J. Med. Chem., 2014, 57, 5085.
- 9 (a) M. Jouanneau, B. McClary, J. C. P. Reyes, R. Chen, Y. Chen, W. Plunkett, X. Cheng, A. Z. Milinichik, E. F. Albone, J. O. Liu and D. Romo, *Bioorg. Med. Chem. Lett.*, 2016, 26, 20927; (b) B. McClary, B. Zinshteyn, M. Meyer, M. Jouanneau, S. Pellegrino, G. Yusupova, A. Schuller, J. C. P. Reyes, J. Lu, Z. Gou, S. Ayinde, C. Luo, Y. Dang, D. Romo, M. Yusupov, R. Green and J. O. Liu, *Cell Chem. Biol.*, 2017, 24, 605.
- 10 A. H. Antropow, K. Xu, R. J. Buchsbaum and M. Movassaghi, J. Org. Chem., 2017, 82, 7720.
- (a) T. Yoshimitsu, T. Ino and T. Tanaka, Org. Lett., 2008, 10, 5457;
 (b) T. Yoshimitsu, T. Ino, N. Futamura, T. Kamon and T. Tanaka, Org. Lett., 2009, 11, 34025;
 (c) D. Shigeoka, T. Kamon and T. Yoshimitsu, Beilstein J. Org. Chem., 2013, 9, 860.
- (a) Z. Li, T. Kamon, D. A. Personett, T. Caulfield, J. A. Copland, T. Yoshimitsu and H. W. Tun, Med. Chem. Commun., 2012, 3, 233;
 (b) Z. Li, D. Shigeoka, T. R. Caulfield, T. Kawachi, Y. Qiu, T. Kamon, M. Arai, H. W. Tun and T. Yoshimitsu, Med. Chem. Commun., 2013, 4, 1093.
- 13 H. W. Tun, T. Yoshimitsu, D. Shigeoka, T. Kamon, Z. Li, Y. Qiu and T. R. Caulfield, US Pat., US9464093B2, 2016.
- 14 (a) M. J. Mulvihill, J. I. Gage and M. J. J. Miller, J. Org. Chem., 1998, 63, 3357; (b) C. Cesario, L. P. Tardibono and M. J. J. Miller, J. Org. Chem., 2009, 74, 448.
- 15 The optical purity was unambiguously confirmed by ¹H NMR analysis of Mosher esters derived from an alcohol that was prepared by reduction of ketone 11 with NaBH₄ (for the details, see the ESI†).
- 16 (a) A. Deyine, G. Dujardin, M. Mammeri and J.-M. Poirier, Synth. Commun., 1998, 28, 1817; (b) K. Inanaga, Y. Ogawa, Y. Nagamoto, A. Daigaku, H. Tokuyama, Y. Takemoto and K. Takasu, Beilstein J. Org. Chem., 2012, 8, 658.
- 17 P. Magnus, M. B. Roe and C. J. Hulme, J. Chem. Soc., Chem. Commun., 1995, 263.
- 18 The stucture of regioisomeric bromide 18 was unambiguously confirmed by X-ray crystallographic analysis (CCDC 1852628)†.
- 19 (a) X. Sun, X. Li, S. Song, Y. Zhu, Y.-F. Liang and N. Jiao, J. Am. Chem. Soc., 2015, 137, 6059; (b) Y. Zhao, Y. Hu, H. Wang, X. Li and B. Wan, J. Org. Chem., 2016, 81, 4412.
- 20 P. A. Duspara, Md. S. Islam, A. J. Lough and R. A. Batey, J. Org. Chem., 2012, 77, 10362.