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Orthogonal, dual protein labelling by tandem cycloaddition of strained alkenes and alkynes to *ortho*-quinones and azides†

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Reaction of cyclopropanated *trans*-cyclooctene (cpTCO) with *in situ* generated *ortho*-quinone is an efficient tool for bioorthogonal protein conjugation. The (4+2)-cycloaddition of cpTCO with *ortho*-quinone is significantly faster than its cyclooctyne counterpart (BCN). Orthogonal, tandem cpTCO–quinone and BCN–azide cycloadditions afforded a homogeneous, dual labelled antibody–drug conjugate.

Bioorthogonal chemistry can be defined as the combined arsenal of reactions between two (or more) unique molecular functionalities that together rapidly form a covalent bond, but are inert to the remaining repertoire of chemical groups present in the surroundings.¹ As a consequence of this high functional group compatibility, bioorthogonal reactions have *inter alia* been broadly applied for detection, immobilization and functionalization of biomolecules, such as carbohydrates, nucleic acid and proteins.² For example, the Staudinger ligation³ paved the way for the selective imaging of cell-surface azidosugars, azide–alkyne click chemistry is a well-established tool in the field of DNA/RNA origami,⁴ and oxime ligation⁵ has proven a powerful technology for protein modification.⁶ Arguably the most powerful bioorthogonal reactions are those defined by metal-free, strain-promoted cycloadditions of alkynes or alkenes,⁷ of which stand out the strain-promoted azide–alkyne cycloaddition (SPAAC)⁸ and the inverse electron-demand Diels–Alder cycloaddition (SPIEDAC)⁹ of strained alkenes and tetrazines. Conveniently, it has been established that with judicious choice of functionalities, a next level of orthogonality can be achieved, involving mutually exclusive, bioorthogonal reactions proceeding in tandem or even concurrently.¹⁰ Besides inherent orthogonality in reactivity, or the application of targeting agents,¹¹ the use of external stimuli to induce bioorthogonal reactivity with temporal

and spatial control has also been described, mostly based on photolysis of tetrazole^{10a} or cyclopropanone^{10b} to nitrile imides or cyclooctyne derivatives, respectively. We,¹² and others,¹³ recently described that enzymatic oxidation of aromatic rings can also be used for temporally controlled bioorthogonal chemistry. Specifically, we demonstrated that strain-promoted oxidation-controlled *ortho*-quinone cycloaddition (SPOCQ) with cyclooctyne,¹⁴ can be successfully applied for the site-specific conjugation of a cyclooctyne-functionalized fluorophore or a toxic payload to tyrosine-engineered proteins, including the monoclonal antibody trastuzumab,¹² as well as for highly efficient and complete surface modification.¹⁵ Interestingly, with these developments, tyrosine is emerging as a very versatile amino acid for various peptide¹⁶ and protein¹⁷ derivatization strategies. With the present study, we have expanded the arsenal of SPOCQ chemistry by demonstrating that besides cyclooctynes strained alkenes also display high reactivity for *ortho*-quinones. Specifically, we show that cyclopropanated *trans*-cyclooctene (cpTCO)¹⁸ rapidly reacts with *ortho*-quinones (a.k.a. 1,2-quinones),¹⁹ enabling clean and efficient tyrosinase-mediated protein conjugation. In addition, the inertness of cpTCO for azides provides a next level of dual, orthogonal bioconjugation, which may be executed without additional purification steps in between reactions.

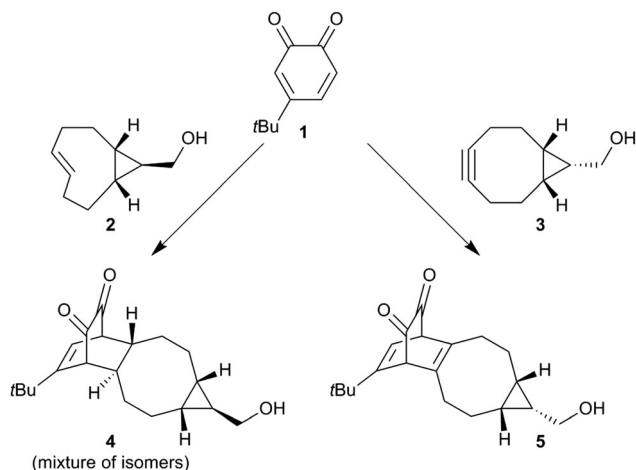
Before we embarked on the potential application of *trans*-cyclooctenes for SPOCQ modification of proteins, we tested if the strain-promoted click reaction between *trans*-cyclooctenes and *ortho*-quinone would take place. To this end, 4-*tert*-butyl-1,2-quinone **1** was mixed with cpTCO **2** (Scheme 1), and the disappearance of the absorbance that corresponds to the presence of *ortho*-quinone **1** at 395 nm was monitored.²⁰ Much to our delight, the reaction of quinone **1** with cpTCO **2** not only occurs (Fig. 1 and Fig. S3–S5, ESI†), but was actually found to take place faster than that with bicyclo[6.1.0]non-4-yne (BCN)²¹ **3**. Specifically, the SPOCQ reaction of quinone **1** with cpTCO **2** proceeded with a rate-constant of $2900 \pm 115 \text{ M}^{-1} \text{ s}^{-1}$, a near 3-fold increase *versus* the (corrected) rate constant of BCN **3** with quinone **1** ($1112 \pm 8 \text{ M}^{-1} \text{ s}^{-1}$, $N = 3$).¹² In the same setup we found a rate constant of $10.4 \pm 1.8 \text{ M}^{-1} \text{ s}^{-1}$ for the reaction of regular

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Scheme 1 Reaction between quinone **1** and either *exo*-cpTCO **2** or *endo*-BCN **3** to give alkene-SPOCQ product **4** (this work) or alkyne-SPOCQ product **5**.¹⁴

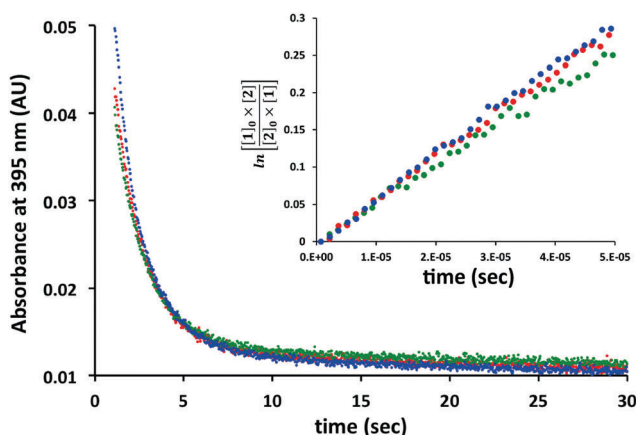


Fig. 1 Determination of the reaction rate constant of quinone **1** (100 μM) with cpTCO **2** (100 μM) in $\text{H}_2\text{O} : \text{MeOH}$ (1 : 1). Three separate experiments are depicted in the graph.

TCO with quinone **1**. Apparently, the fused cyclopropane ring increases the rate approximately 300-fold, in analogy with enhancement of reaction rate with tetrazine.¹⁸ For comparison, cpTCO-SPOCQ is more than 10^3 times faster than reaction of quinone **1** with a cyclopropene ($<1.7\text{--}1.9 \text{ M}^{-1} \text{ s}^{-1}$)²² and strain-promoted cycloadditions involving azides ($0.01\text{--}1 \text{ M}^{-1} \text{ s}^{-1}$),⁷ however slightly slower than SPIEDAC reaction of regular TCO with tetrazines (2000 to $>5000 \text{ M}^{-1} \text{ s}^{-1}$),¹³ and much slower than the fastest known cpTCO-tetrazine ligations (reaching above $10^6 \text{ M}^{-1} \text{ s}^{-1}$).²³ Nevertheless, in contrast to azide and tetrazine, an *ortho*-quinone is readily generated from a canonical amino acid (tyrosine), and therefore has the inherent advantage of (inducible) bioconjugation chemistry to native proteins.

After this, we explored the suitability of cpTCO-quinone cycloaddition for protein conjugation (Fig. 2 and Fig. S6–S9, ESI[†]). To this end, a cpTCO-functionalized lissamine-PEG conjugate was synthesized and compared head-to-head to its BCN counterpart in labelling of a model antibody (trastuzumab), which contained an

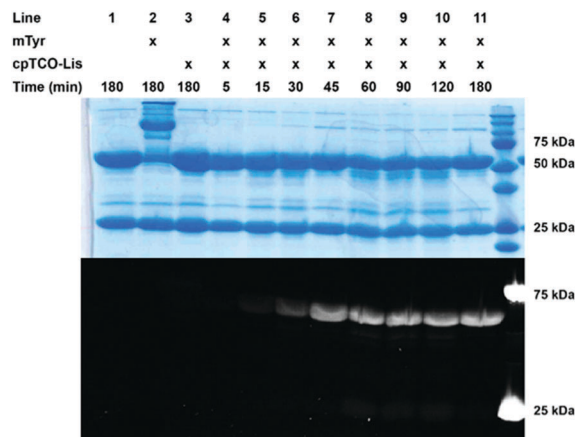


Fig. 2 Time-resolved SDS-PAGE analysis, based on heavy chain labelling, of cpTCO-PEG-lissamine SPOCQ with Tras[HC]G₄Y (lane 1) in the presence of mTyr (lanes 4–11). No reaction is apparent after 3 h in the absence of mTyr (lane 3), while the present of mTyr alone (absence of cpTCO reagent) leads to degradation/oligomer formation (lane 2).

exposed G₄Y-tag on each heavy chain (abbreviated as Tras[HC]G₄Y). Previously, we already showed that without this G₄Y-tag, labelling of a smaller protein or an antibody did not proceed.¹² In analogy to our previous work, we studied the alkene-SPOCQ reaction between cpTCO-PEG-lissamine and Tras[HC]G₄Y upon incubation with mushroom tyrosinase (mTyr) to oxidize the exposed tyrosine phenol ring of the G₄Y-tag to *ortho*-quinone.¹⁹ Indeed, when Tras[HC]G₄Y was mixed with mTyr in the presence of cpTCO-lissamine, rapid labelling of the antibody was observed that apparently levelled off after ~ 90 minutes (Fig. 2, lanes 4–11); no labelling was observed in the absence of mTyr (Fig. 2, lane 3). In the absence of a G₄Y-tag on the light chain, labelling of this part of the antibody was not observed (*i.e.* no fluorescent bands around 25 kDa were observed), thus demonstrating (1) the bioorthogonality of alkene-SPOCQ, and (2) the need for a G₄Y-tag on the protein. Based on HPLC analysis, a yield of $\sim 95\%$ may be calculated (Fig. S10–S12, ESI[†]), with reduced side-product formation in comparison to alkyne-SPOCQ (Fig. S9, ESI[†]), *i.e.* antibody oligomers formed by cross-linking of the *ortho*-quinone with endogenous protein nucleophiles (Fig. 2, lane 2). The reduced formation of protein oligomers for cpTCO *versus* BCN further corroborates the higher reaction rate with *ortho*-quinones. Since oxidation of tyrosine residues by mTyr is the rate-limiting step in either SPOCQ reaction,²⁰ the total reaction time does not decrease despite the higher reaction rate for cpTCO. However, the short lifetime of the generated quinone favours labelling with the fast reacting cpTCO by reducing side product formation.

In order to exploit the lack of reactivity of cpTCO for azides, and high reaction rate with *ortho*-quinones, we envisioned that tandem, dual labelling of an antibody should be accessible by orthogonal reactions of BCN and cpTCO with azide and (exposed) tyrosine, respectively. To test this, we explore the possibility of tandem installation of two different functional moieties onto a glycan-remodelled monoclonal antibody²⁴ by cpTCO-SPOCQ and BCN-SPAAC, executed in either order. We started by the cpTCO-SPOCQ reaction on the Tras[HC]G₄Y C-terminus followed by SPAAC



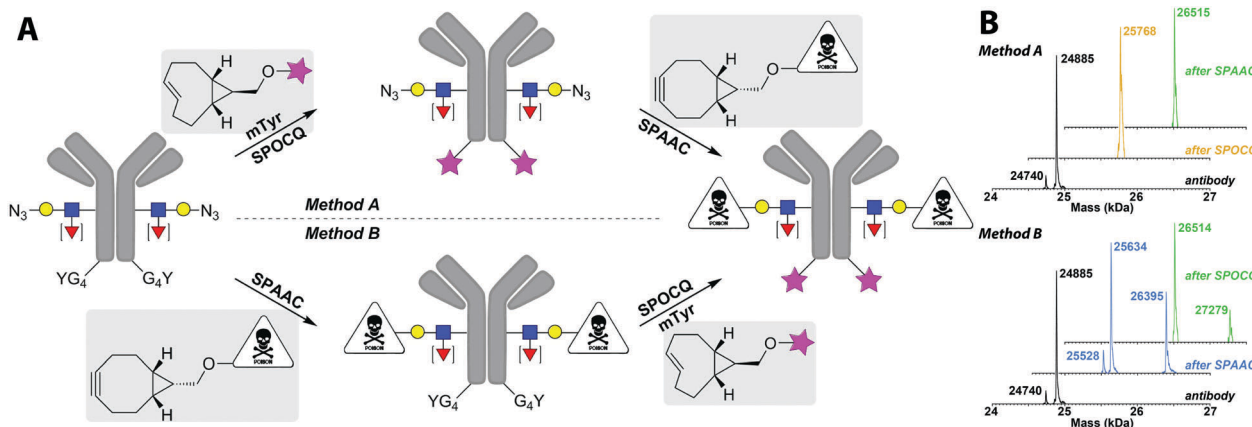


Fig. 3 (A) Schematic depiction of the two routes for dual labelling of antibodies with lissamine and MMAE. One route employs the modification of the G₄Y-tag using the cpTCO–SPOCQ reaction with cpTCO–lissamine, followed by SPAAC reaction with BCN–MMAE. The other route employs first a SPAAC reaction on the azido-glycan followed by SPOCQ on the G₄Y-tag of the heavy chain. (B) The deconvoluted mass spectra (crude) of the different steps of the tandem bioconjugation process using the two methods are shown. The main peak in the black trace identifies with GlcNAc-(1,6-Fuc)-6-N₃-GalNAc, the minor peak at Δ – 145: GlcNAc-6-N₃-GalNAc; in-line fragmentation of the MMAF toxin is also visible.

modification of azido-glycan with BCN–MMAE (method A), but also checked method B (Fig. 3A).

In order to obtain the same conjugate *via* both routes, we used cpTCO–PEG–lissamine and BCN–sulfamide–MMAE (see ESI† for structures) with a cathepsin-cleavable linker (MMAE = monomethyl auristatin E, a potent antimitotic agent that inhibits cell division). In addition, the sulfamide spacer in the BCN unit enhanced solubility and conjugation efficiency.²⁵

Thus, clean and efficient enzymatic glycan remodelling was achieved by endoglycosidase and glycosyltransferase, to install 6-azido-GalNAc onto the core-GlcNAc of transiently expressed Tras[HC]G₄Y (Fig. 3A and Fig. S13, ESI†).¹⁴ Next, a SPOCQ reaction between cpTCO–lissamine and the G₄Y-tag on the antibody heavy chain was performed in the presence of mTyr (Fig. S14, ESI†). The observed mass of 25 768 Da (Fig. 3B, yellow peak) corresponds to the SPOCQ product of the azido-glycan antibody and cpTCO–lissamine (expected: 25 766 Da), and corroborates the complete lack of reactivity of the cpTCO moiety with the azido-group on the glycan chain. Subsequently, SPAAC modification of the azido-functionalized glycan chain with BCN–MMAE resulted in the clean formation of the dual labelled fluorescent antibody–drug conjugate. HPLC analysis showed almost complete conversion (Fig. S15, ESI†), and MS analysis identified the presence of the target compound with a FW of 26 516 Da (expected: 27 275 Da) (Fig. 3B, green peak in top spectrum). The difference between the observed and expected mass corresponds to loss of the C-terminal part of MMAE due to fragmentation in the mass chamber (Δ 762 Da).

Lastly, the tandem labelling steps were performed in reverse order, *i.e.* first SPAAC reaction on the remodelled azido-glycan chain using BCN–MMAE SPAAC (Fig. S16, ESI†), then cpTCO–lissamine SPOCQ reaction on the G₄Y-tag upon tyrosinase oxidation (Fig. S17, ESI†). As expected, the same final product is obtained, albeit *via* a different intermediate product. We were happy to find that in this case, besides the mass of the linker-fragmented final conjugate (at 26 514 Da), the mass of

the desired dual labelled ADC is also observed at 27 279 Da (expected: 27 275 Da) (Fig. 3B, lower set of MS spectra). This difference is presumably due to some experimental variability during the MS analysis.

In conclusion, we have uncovered a useful addition to the toolbox of copper-free click chemistry, *i.e.* the rapid reaction of strained alkenes and *ortho*-quinones, and used this reaction to establish a convenient and high-yielding new strategy for the labelling of antibodies. The rate of the SPOCQ reaction between cpTCO and *ortho*-quinone is ~ 2.5 times higher than that for BCN, ~ 300 times higher than that for TCO (Fig. S30 and S31, ESI†), and $>10^3$ times higher than that for cyclopropene; the last two strained alkenes would be too slow for efficient SPOCQ labelling of proteins, due to the high reactivity of the quinone, *e.g.* with competing nucleophiles. This high reactivity of cpTCO enables faster and cleaner conjugation, as superior labelling of a G₄Y-derivatized antibody with a fluorescent moiety was demonstrated when compared to its BCN counterpart. Ultimately, we developed a convenient protocol for the dual labelling of an antibody with a toxic payload and a dye using the orthogonal character of cpTCO and BCN, which may find application in the assessment of internalization potential of glycan-conjugated antibody–drug conjugates for targeted therapy.²⁵ Our setup allows a convenient preparation of a glycan-remodelled azido-containing antibody that also contains a peptidic G₄Y tag, and to use this in a two-step dual-labelling strategy to obtain homogeneous ADCs. We note that other methods have been reported for metal-free click conjugation to unnatural amino acids genetically encoded into monoclonal antibodies, for example *p*-acetylphenylalanine²⁶ or *p*-azidomethylphenylalanine,²⁷ followed by oxime ligation or SPAAC, respectively. However, low antibody titers (1 g L^{-1} maximum), which are significantly lower than expression titers on monoclonal antibodies based on canonical amino acids alone ($3\text{--}7 \text{ g L}^{-1}$), limit such applications. Similar arguments apply to genetic encoding of L-DOPA into proteins, which is also suitable for SPOCQ modification.²⁸ We expect that our current method will



find widespread application, not only in the preparation of ADCs but extending to other applications in the field of bioconjugation, such as PEGylation, immobilization, protein–protein dimer formation, and beyond. Efforts along those lines are currently ongoing in our laboratory.

Conflicts of interest

F. L. V. D. declares that he is also an employee and share-holder of Synaffix BV.

Notes and references

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