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Structural effects of ditopic azoprobe–cyclodextrin complexes on the selectivity of guest-induced supramolecular chirality

The selectivities of guest-induced supramolecular chirality for cations and anions were dramatically altered by a slight change in the spacer length of  $(15C5-Azo-n-dpa)_2-\gamma$ -cyclodextrin complexes in water.

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# Structural effects of ditopic azoprobe–cyclodextrin complexes on the selectivity of guest-induced supramolecular chirality†

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**Benzo-15-crown-5 and dipicolylamine are contained as the binding sites in a ditopic azoprobe (15C5-Azo-*n*-dpa). However, the selectivities of guest-induced supramolecular chirality for cations and anions were dramatically altered by a slight change in the spacer length of (15C5-Azo-*n*-dpa)<sub>2</sub>- $\gamma$ -CyD complexes in water.**

Chirality control by supramolecular assemblies and helical polymers based on chiral templates has attracted much attention in recent years.<sup>1</sup> Especially guest-induced chirality control is expected to apply for the development of versatile chiral switching and sensing systems.<sup>2</sup> To obtain the supramolecular chirality function, cyclodextrins (CyDs) are quite attractive host molecules.<sup>3</sup> Optically inert CyDs have a chiral nature in their cavities and can be efficiently combined with various achiral chromoionophores and fluoroionophores to induce chiral nature by forming an inclusion complex with CyDs.<sup>4</sup> In the previous study, we have reported a ditopic azoprobe (15C5-Azo-2-dpa) bearing benzo-15-crown-5 (B15C5) and dipicolylamine (dpa) as recognition sites. 15C5-Azo-2-dpa was found to form a 2:1 complex with  $\gamma$ -CyD and show a unique response function based on guest-induced supramolecular chirality in water.<sup>5</sup> By allowing ditopic azoprobes to be incorporated into  $\gamma$ -CyD, we revealed the response behavior of the supramolecular (15C5-Azo-2-dpa)<sub>2</sub>- $\gamma$ -CyD complex in the presence of each cationic and anionic species by measuring induced circular dichroism (ICD) spectra and UV-visible (Vis) absorption spectra. We confirmed that only when K<sup>+</sup>, Zn<sup>2+</sup>, and CO<sub>3</sub><sup>2-</sup> were all present, a large split-type Cotton effect appeared in the measured ICD spectra and a significant short-wavelength shift took place in the measured UV-Vis spectra. The result clearly demonstrates that the (15C5-Azo-2-dpa)<sub>2</sub>- $\gamma$ -CyD complex can exhibit supramolecular chirality due to the twisted

structure of the azoprobe dimer inside the  $\gamma$ -CyD cavity, only when it recognizes K<sup>+</sup> and Zn<sup>2+</sup> in the presence of CO<sub>3</sub><sup>2-</sup>.<sup>5</sup> From the ICD spectral change, we can estimate the spatial changes of the azoprobe dimer inside  $\gamma$ -CyD, which induce a change in the UV-Vis spectra.<sup>6</sup> Therefore each guest ion can be selectively detected in the presence of the other guest ions by measuring the spectral changes.

Herein we report how the 15C5-Azo-*n*-dpa structure affects the selectivity of guest-induced supramolecular chirality. The dramatic selectivity changes of supramolecular chirality were found to be noted for (15C5-Azo-*n*-dpa)<sub>2</sub>- $\gamma$ -CyD complexes by controlling the spacer length of 15C5-Azo-*n*-dpa from ethylene (*n* = 2) to propylene (*n* = 3) to butylene (*n* = 4) units (Fig. 1). Also, while 15C5-Azo-*n*-dpa has a dpa binding site for heavy metal ions, the selectivity of guest-induced supramolecular chirality changed from Zn<sup>2+</sup> for the (15C5-Azo-2-dpa)<sub>2</sub>- $\gamma$ -CyD complex to

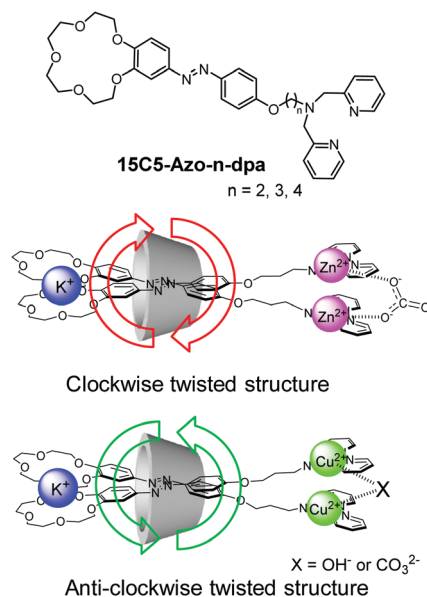


Fig. 1 Structure of 15C5-Azo-*n*-dpa and the twisted structures.

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$\text{Cu}^{2+}$  for the  $(15\text{C5-Azo-4-dpa})_2\text{-}\gamma\text{-CyD}$  complex. However, most dpa-based chemosensors display  $\text{Zn}^{2+}$  and/or  $\text{Cd}^{2+}$  selectivity in water.<sup>7</sup> Thus this is a unique example where the dpa-based supramolecular sensor exhibits a selectivity change from  $\text{Zn}^{2+}$  to  $\text{Cu}^{2+}$  caused by a change in the spacer length. We also report the specific selectivity changes of guest-induced supramolecular chirality for alkali-metal cations and anions based on the change in the spacer length of  $(15\text{C5-Azo-}n\text{-dpa})_2\text{-}\gamma\text{-CyD}$  complexes in water.

The synthesis of  $15\text{C5-Azo-}n\text{-dpa}$  was carried out by the azocoupling of 4'-aminobenzo-15-crown-5 with phenol, followed by the introduction of bromoethylene, bromopropylene, and bromobutylene spacers using the Williamson ether synthesis.<sup>8</sup> Then a dpa moiety was introduced under basic conditions with  $\text{K}_2\text{CO}_3$ , and the obtained products were purified using silica gel column chromatography. The structures of  $15\text{C5-Azo-}n\text{-dpa}$  were confirmed *via*  $^1\text{H}$  NMR and combustion analyses. Details of the synthesis are available in the ESI.<sup>†</sup>

Job's plot analyses clearly revealed that  $15\text{C5-Azo-}n\text{-dpa}$  formed a 2 : 1 inclusion complex with  $\gamma\text{-CyD}$  in 4% DMSO–96% water (v/v) (Fig. S6, ESI<sup>†</sup>). The ICD spectra of  $(15\text{C5-Azo-}n\text{-dpa})_2\text{-}\gamma\text{-CyD}$  complexes are depicted in Fig. 2. For  $15\text{C5-Azo-2-dpa}$  (Fig. 2a), the selective ICD response was only noted for  $\text{Zn}^{2+}$  over other metal ions ( $\text{Mg}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Cd}^{2+}$ , and  $\text{Pb}^{2+}$ , as nitrate salts) in the presence of 50 mM  $\text{K}_2\text{CO}_3$ . The split ICD from the negative peak at 351 nm to the positive peak at 394 nm indicates the clockwise twisted structure of the two azoprobes inside the  $\gamma\text{-CyD}$  cavity.<sup>6</sup> For  $15\text{C5-Azo-3-dpa}$ , however, the ICD response was found to be observed not only for  $\text{Zn}^{2+}$  but also for  $\text{Cu}^{2+}$  in the presence of 50 mM  $\text{K}_2\text{CO}_3$ . Interestingly the ICD peak shape for  $\text{Cu}^{2+}$  was opposite compared with that for  $\text{Zn}^{2+}$ , indicating that the  $\text{Cu}^{2+}$  complex formed an anti-clockwise twisted structure inside the  $\gamma\text{-CyD}$  cavity. This change in the twisted structure may be due to the difference in the coordination configuration;  $\text{Cu}^{2+}$  was capable of forming a coordination bond with the phenoxy ether oxygen in the dpa complexes,<sup>9</sup> whereas only a few coordination bonds with the phenoxy ether oxygen were noted for the  $\text{Zn}^{2+}$ –dpa complexes.<sup>10</sup> For  $15\text{C5-Azo-4-dpa}$ , the selective ICD response was only noted for  $\text{Cu}^{2+}$  over other metal ions ( $\text{Mg}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ , and  $\text{Pb}^{2+}$ , as nitrate salts) in the presence of 50 mM  $\text{K}_2\text{CO}_3$ . Although  $15\text{C5-Azo-}n\text{-dpa}$  possesses the same dpa binding site for heavy metal ions, the selectivity was dramatically changed from  $\text{Zn}^{2+}$  for the  $(15\text{C5-Azo-2-dpa})_2\text{-}\gamma\text{-CyD}$  complex to  $\text{Cu}^{2+}$  for the  $(15\text{C5-Azo-4-dpa})_2\text{-}\gamma\text{-CyD}$  complex in the presence of  $\text{K}_2\text{CO}_3$ . The  $(15\text{C5-Azo-3-dpa})_2\text{-}\gamma\text{-CyD}$  complex exhibited selectivity for both  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$ , indicating an intermediate selectivity between  $15\text{C5-Azo-2-dpa}$  and  $15\text{C5-Azo-4-dpa}$ .<sup>11</sup> It is evident that the spacer length of  $15\text{C5-Azo-}n\text{-dpa}$  played an important role in controlling the selectivity of guest-induced supramolecular chirality. As shown in Fig. 2, it should be noted that the shapes of split Cotton effects based on  $\pi\text{-}\pi^*$  transition are not symmetric, indicating the overlap of the Cotton effect based on  $n\text{-}\pi^*$  transition at the longer wavelength. In addition, the location of azobenzenes along the z-axis of CyD is known to strongly affect the sign and intensity of the Cotton effect.<sup>6</sup> Although we consider that the bulky and hydrophobic B15C5 moieties restrict the movement of azobenzenes along the

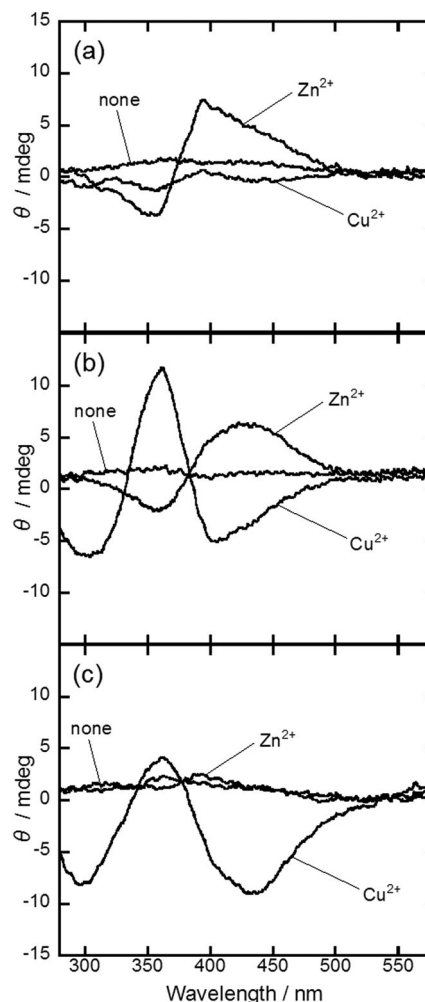


Fig. 2 ICD spectra of  $15\text{C5-Azo-}n\text{-dpa}/\gamma\text{-CyD}$  sensors in 4% DMSO aq.: (a)  $15\text{C5-Azo-2-dpa}$ ; (b)  $15\text{C5-Azo-3-dpa}$ ; (c)  $15\text{C5-Azo-4-dpa}$  = 0.04 mM,  $[\text{Zn}(\text{NO}_3)_2]$  = 0.04 mM,  $[\text{Cu}(\text{NO}_3)_2]$  = 0.04 mM,  $[\gamma\text{-CyD}]$  = 5 mM,  $[\text{K}_2\text{CO}_3]$  = 50 mM.

z-axis of the  $\gamma\text{-CyD}$  cavity, the abovementioned factors make the detailed understanding of guest-induced ICD responses difficult. To obtain further evidence for the guest-induced ICD responses, additional analysis based on molecular mechanics and TD-DFT calculations as well as X-ray crystallography analysis are to be conducted.

In the presence of equivalent amounts of  $\text{Zn}^{2+}$  with  $15\text{C5-Azo-}n\text{-dpa}$  (20  $\mu\text{M}$ ), and 50 mM  $\text{CO}_3^{2-}$ , the ICD intensities at the maximum wavelength are plotted against the alkali metal ion diameter (Fig. 3a). As we reported previously, the  $(15\text{C5-Azo-2-dpa})_2\text{-}\gamma\text{-CyD}$  complex exhibited high  $\text{K}^+$  ion selectivity over other alkali metal ions in the presence of  $\text{Zn}^{2+}$  and  $\text{CO}_3^{2-}$ . This selectivity is consistent with the selectivity of sandwich complex formation of the two benzo-15-crown-5 derivatives with alkali metal ions.<sup>8,12</sup> However, for the  $(15\text{C5-Azo-3-dpa})_2\text{-}\gamma\text{-CyD}$  complex, the alkali metal ion selectivity was significantly reduced (Fig. 3a). This indicates that the formation of the clockwise twisted structure is dominated only by the bridge formation of  $\text{CO}_3^{2-}$  with the two dpa– $\text{Zn}^{2+}$  complexes in the



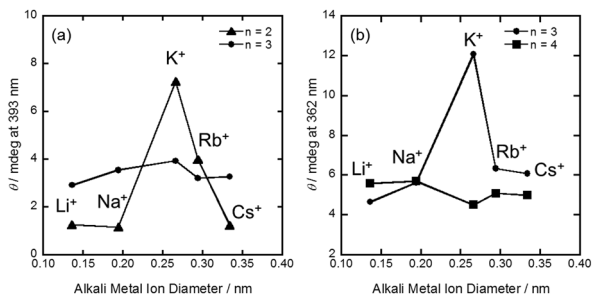


Fig. 3 Selectivity of **15C5-Azo-*n*-dpa**/γ-CyD sensors toward alkali metal ions in 4% DMSO aq., [γ-CyD] = 5 mM, [alkali metal ion] = 50 mM: (a) [**15C5-Azo-2-dpa**], [**15C5-Azo-3-dpa**] = 0.04 mM, [Zn(NO<sub>3</sub>)<sub>2</sub>] = 0.04 mM; (b) [**15C5-Azo-3-dpa**], [**15C5-Azo-4-dpa**] = 0.04 mM, [Cu(NO<sub>3</sub>)<sub>2</sub>] = 0.04 mM.

(**15C5-Azo-3-dpa**)<sub>2</sub>-γ-CyD complex. The enhanced flexibility of **15C5-Azo-*n*-dpa** upon changing the spacer from ethylene (*n* = 2) to propylene (*n* = 3) should be the reason of this selectivity change. On the other hand, in the presence of equivalent amounts of Cu<sup>2+</sup> with **15C5-Azo-*n*-dpa** (20 μM), and 50 mM CO<sub>3</sub><sup>2-</sup>, the (**15C5-Azo-3-dpa**)<sub>2</sub>-γ-CyD complex exhibited high K<sup>+</sup> ion selectivity similar to the Zn<sup>2+</sup> system of the (**15C5-Azo-2-dpa**)<sub>2</sub>-γ-CyD complex (Fig. 3b). However, for the (**15C5-Azo-4-dpa**)<sub>2</sub>-γ-CyD complex, no alkali metal ion selectivity was observed in the anti-clockwise twisted structure of the two azoprobes inside the γ-CyD cavity (Fig. 3b). This is also ascribed to the enhanced flexibility of **15C5-Azo-*n*-dpa** upon changing the spacer from propylene (*n* = 3) to butylene (*n* = 4).

The effect of anion species on the ICD responses was also examined for (**15C5-Azo-*n*-dpa**)<sub>2</sub>-γ-CyD complexes. For the Zn<sup>2+</sup> complex system of (**15C5-Azo-*n*-dpa**)<sub>2</sub>-γ-CyD complexes (*n* = 2, 3), the effects of salt species on ICD responses were examined in the presence of 50 mM KX (X = NO<sub>3</sub><sup>-</sup>, CH<sub>3</sub>CO<sub>2</sub><sup>-</sup>, and OH<sup>-</sup>). The results are depicted in Fig. 4. Similar to the (**15C5-Azo-2-dpa**)<sub>2</sub>-γ-CyD complex, the (**15C5-Azo-3-dpa**)<sub>2</sub>-γ-CyD complex exhibited high CO<sub>3</sub><sup>2-</sup> selectivity, indicating that CO<sub>3</sub><sup>2-</sup> bridging with the two Zn<sup>2+</sup>-dpa binding sites induced the clockwise twisted structure of the azoprobe dimer inside the γ-CyD cavity. In addition, direct evidence of the relative orientation of the azoprobes and the macrocyclic ring was obtained *via* NOESY experiments. Cross-peaks between H3 protons inside the CyD cavity and protons of the azoprobes were clearly observed (Fig. S7, ESI†).

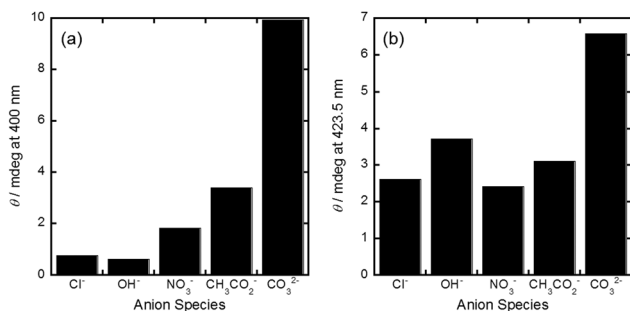


Fig. 4 Selectivity of (**15C5-Azo-*n*-dpa-Zn<sup>2+</sup>**)<sub>2</sub>-γ-CyD sensors toward anions in 4% DMSO aq.: (a) [**15C5-Azo-2-dpa**] = 0.04 mM; (b) [**15C5-Azo-3-dpa**] = 0.04 mM, [Zn(NO<sub>3</sub>)<sub>2</sub>] = 0.04 mM, [γ-CyD] = 5 mM, [Cl<sup>-</sup>], [OH<sup>-</sup>], [NO<sub>3</sub><sup>-</sup>], [CH<sub>3</sub>CO<sub>2</sub><sup>-</sup>], [CO<sub>3</sub><sup>2-</sup>] = 50 mM.

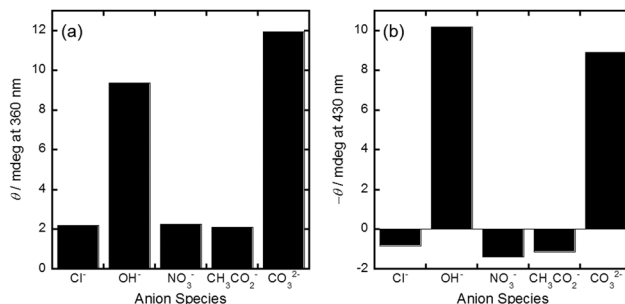


Fig. 5 Selectivity of (**15C5-Azo-*n*-dpa-Cu<sup>2+</sup>**)<sub>2</sub>-γ-CyD sensors toward anions in 4% DMSO aq.: (a) [**15C5-Azo-3-dpa**] = 0.04 mM; (b) [**15C5-Azo-4-dpa**] = 0.04 mM, [Cu(NO<sub>3</sub>)<sub>2</sub>] = 0.04 mM, [γ-CyD] = 5 mM, [Cl<sup>-</sup>], [OH<sup>-</sup>], [NO<sub>3</sub><sup>-</sup>], [CH<sub>3</sub>CO<sub>2</sub><sup>-</sup>], [CO<sub>3</sub><sup>2-</sup>] = 50 mM.

On the other hand, for the Cu<sup>2+</sup> complex system of (**15C5-Azo-*n*-dpa**)<sub>2</sub>-γ-CyD complexes (*n* = 3 and 4), the (**15C5-Azo-*n*-dpa**)<sub>2</sub>-γ-CyD complexes showed both CO<sub>3</sub><sup>2-</sup> and OH<sup>-</sup> selectivity (Fig. 5). This indicates that hydroxo-bridging between the two Cu<sup>2+</sup>-dpa binding sites induced the anti-clockwise twisted structure of the azoprobe dimer in the (**15C5-Azo-*n*-dpa**)<sub>2</sub>-γ-CyD complex. Thus, by changing the metal species of the dpa binding sites, the anion selectivity of the (**15C5-Azo-3-dpa**)<sub>2</sub>-γ-CyD complex can be easily controlled from the CO<sub>3</sub><sup>2-</sup> selectivity with the Zn<sup>2+</sup> system to both CO<sub>3</sub><sup>2-</sup> and OH<sup>-</sup> selectivity with the Cu<sup>2+</sup> system in water. These are apparently a unique function of guest-induced supramolecular chirality based on (**15C5-Azo-*n*-dpa**)<sub>2</sub>-γ-CyD complexes.

In conclusion, we have shown a novel guest-induced supramolecular chirality induced by twisted structural switching of the two **15C5-Azo-*n*-dpa** molecules inside the γ-CyD chiral cavity due to multi-point recognition of guest ions by the ditopic azoprobes in water. Although the two recognition sites are the same, a slight change in the spacer length of **15C5-Azo-*n*-dpa** was found to significantly affect the ICD response selectivity of (**15C5-Azo-*n*-dpa**)<sub>2</sub>-γ-CyD complexes. To the best of our knowledge, this is a novel selectivity control based on guest-induced supramolecular chirality which completely differs from the design strategy of conventional molecular recognition systems.

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## Conflicts of interest

There are no conflicts to declare.

## Notes and references

- Supramolecular Chirality, in *Top. Curr. Chem.*, ed. M. Crego-Calama and D. N. Reinhovdt, 2006, p. 26; Amplification of Chirality, in *Top. Curr. Chem.*, ed. K. Soai, 2008, p. 284; *Chirality at the Nanoscale*, ed. D. B. Amabilino, Wiley-VCH, Weinheim, 2009; R. Raval, *Chem. Soc. Rev.*, 2009, **38**, 707; J. Crassovs, *Chem. Soc. Rev.*, 2009, **38**, 830; M. Liu, L. Zhang and T. Wang, *Chem. Rev.*, 2015, **115**, 7304.



- 2 Z. Kokan, B. Peric, M. Vazdar, Z. Marinic, D. V. Topic, E. Mestrovic and S. I. Kirin, *Chem. Commun.*, 2017, **53**, 1945; *Comprehensive Chiroptical Spectroscopy*, ed. N. Berova, P. L. Polavarapu, K. Nakanishi and R. W. Woody, Wiley-VCH, New York, 2012; K. C.-F. Lrung, C.-P. Chak, C.-M. Lo, W.-Y. Wong, S. Xuan and C. H. K. Cheng, *Chem. – Asian J.*, 2009, **4**, 364; R. Klajn, J. F. Stoddart and B. A. Grzybowski, *Chem. Soc. Rev.*, 2010, **39**, 2203; J. W. Canary, S. Mortezeini and J. Liang, *Coord. Chem. Rev.*, 2010, **254**, 2249; C. Colvccini, A. Mazzrnti and D. Pasini, *Org. Biomol. Chem.*, 2010, **8**, 1807; M. Caricato, C. Colvccini, D. Dondi, D. A. Vander Griend and D. Pasini, *Org. Biomol. Chem.*, 2010, **8**, 3272; M. Caricato, A. Olmo, C. Gargivlli, G. Gattvso and D. Pasini, *Tetrahedron*, 2012, **68**, 7861; Y. Nakatani, Y. Furusho and E. Yashima, *Org. Biomol. Chem.*, 2013, **11**, 1614; M. Ziegler, A. V. Davis, D. W. Johnson and K. N. Raymond, *Angew. Chem., Int. Ed.*, 2003, **42**, 665; A. Mammanna, A. D'Vrso, R. Lauceri and R. Pvrrello, *J. Am. Chem. Soc.*, 2007, **129**, 8062; L. Rosaria, A. D'Vrso, A. Mammanna and R. Pvrrello, *Chirality*, 2008, **20**, 411; I. D. Cat, Z. Gvo, S. J. George, E. W. Meijer, A. P. H. J. Schenning and S. D. Feyter, *J. Am. Chem. Soc.*, 2012, **134**, 3171; P. A. Korevaar, S. J. George, A. J. Markvoort, M. M. J. Smulders, P. A. J. Hilbers, A. P. H. J. Schenning, T. F. A. de Greef and E. W. Meijer, *Nature*, 2012, **481**, 492; W. Zhang, W. Jin, T. Fukushima, N. Ishii and T. Aida, *J. Am. Chem. Soc.*, 2013, **135**(1), 114; M.-C. Li, N. Ousaka, H.-F. Wang, E. Yashima and R.-M. Ho, *Macro Lett.*, 2017, **6**, 980; M. Weis, C. Waloch, W. Seiche and B. Breif, *J. Am. Chem. Soc.*, 2006, **128**, 4188; J. Meevwissen and J. N. H. Reek, *Nat. Chem.*, 2010, **2**, 615; P. W. N. M. Van Leeuwen, D. Rivillo, M. Ranal and Z. Frexa, *J. Am. Chem. Soc.*, 2011, **133**, 18562; M. Hatano and K. Ishihara, *Chem. Commun.*, 2012, **48**, 4273; E. Yashima, K. Maeda and Y. Okamoto, *Nature*, 1999, **399**, 449; H. Ito, M. Ikeda, T. Hasegawa, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2011, **133**, 3419; H. Yamada, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2012, **134**, 7250; H. Yamada, Z. Q. Wu, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2012, **134**, 9506; M. Kumar, N. Jonnalagadda and S. J. George, *Chem. Commun.*, 2012, **48**, 10948; W. Makiguchi, S. Kobayashi, Y. Furusho and E. Yashima, *Angew. Chem., Int. Ed.*, 2013, **52**, 5275; S. Yamamoto, H. Iida and E. Yashima, *Angew. Chem., Int. Ed.*, 2013, **52**, 6849.
- 3 G. Wenz, *Angew. Chem., Int. Ed. Engl.*, 1994, **33**, 803.
- 4 U. Klein, G. Gimpl and F. Fahrenholz, *Biochemistry*, 1995, **34**, 13784; L. Szente and J. Szeman, *Anal. Chem.*, 2013, **85**, 8024; T. Kida, T. Iwamoto, H. Asahara, T. Hinoue and M. Akashi, *J. Am. Chem. Soc.*, 2013, **135**, 3371; C. Shimpuku, R. Ozawa, A. Sasaki, F. Sato, T. Hashimoto, A. Yamauchi, I. Suzuki and T. Hayashita, *Chem. Commun.*, 2009, 1709.
- 5 K. Nonaka, M. Yamaguchi, M. Yasui, S. Fujiwara, T. Hashimoto and T. Hayashita, *Chem. Commun.*, 2014, **50**, 10059.
- 6 B. Mayer, X. Zhang, W. M. Nau and G. Marconi, *J. Am. Chem. Soc.*, 2001, **123**, 5240; M. Kodaka and T. Furuya, *Bull. Chem. Soc. Jpn.*, 1989, **62**, 1154; M. Kodaka, *J. Am. Chem. Soc.*, 1993, **115**, 3702; M. Kodaka, *J. Phys. Chem.*, 1991, **95**, 2110; K. Harata and H. Uedaira, *Bull. Chem. Soc. Jpn.*, 1975, **48**, 375; I. Tinoco, Jr., *Adv. Chem. Phys.*, 1962, **4**, 113.
- 7 K. Ghosh, D. Tarafdar, A. Majumdar, C. G. Daniliuc, A. Samadder and A. R. Khuda-Bukhsh, *RSC Adv.*, 2016, **6**, 47802; S. Sumiya, Y. Shiraishi and T. Hirai, *J. Phys. Chem. A*, 2013, **117**, 1474; P. Das, S. Bhattacharya, S. Mishra and A. Das, *Chem. Commun.*, 2011, **47**, 8118; K. M. K. Swamy, M.-J. Kim, H.-R. Jeon, J.-Y. Jung and J. Yoon, *Bull. Korean Chem. Soc.*, 2010, **31**, 3611.
- 8 F. Sato, M. Tsukano, K. Sakamoto, W. Umamoto, T. Hashimoto and T. Hayashita, *Bull. Chem. Soc. Jpn.*, 2008, **81**, 1589; F. Sato, K. Sakamoto, W. Umamoto, T. Hashimoto and T. Hayashita, *Chem. Lett.*, 2007, **36**(7), 880.
- 9 Y. Mikata, T. Fujimoto, T. Fujiwara and S. Kondo, *Inorg. Chim. Acta*, 2011, **370**, 420; Đ. Škalamera, E. Sanders, R. Vianello, A. Maršavelski, A. Pevec, I. Turel and S. I. Kirin, *Dalton Trans.*, 2016, **45**, 2845; E. Y. Tirel, Z. Bellamy, H. Adams, V. Lebrun, F. Duarte and N. H. Williams, *Angew. Chem., Int. Ed.*, 2014, **53**, 8246.
- 10 B. Mayer, X. Zhang, W. M. Nau and G. Marconi, *J. Am. Chem. Soc.*, 2001, **123**, 5240; M. Kodaka, *J. Am. Chem. Soc.*, 1993, **115**, 3702; M. Kodaka, *J. Phys. Chem.*, 1991, **95**, 2110; K. Harata and H. Uedaira, *Bull. Chem. Soc. Jpn.*, 1975, **48**, 375; I. Tinoco, Jr., *Adv. Chem. Phys.*, 1962, **4**, 113.
- 11 To elucidate the metal ion selectivity, the effects of metal ion concentration on ICD responses are depicted in Fig. S4, ESI†.
- 12 A. Yamauchi, T. Hayashita, A. Kato, S. Nishizawa, M. Watanabe and N. Teramae, *Anal. Chem.*, 2000, **72**, 5841; A. Yamauchi, T. Hayashita, S. Nishizawa, M. Watanabe and N. Teramae, *J. Am. Chem. Soc.*, 1999, **121**, 2319.

