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Revival of the nearly extinct fluorescence of coumarin 6 in water and complete transfer of energy to rhodamine 123†

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The nearly extinct fluorescence of coumarin 6 in water due to microcrystal formation is revived by micelles. Practically complete transfer of energy from coumarin 6 to rhodamine 123 through resonance energy transfer could be achieved.

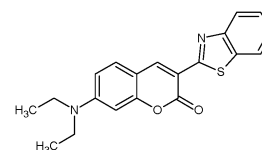


Fig. 1 Molecular structure of C6.

Introduction

An antenna system for light harvesting is considered to be efficient if it has a very high molar extinction coefficient, commendable photostability, and an ability to transfer its energy. Based on this concept many artificial light harvesting systems have been developed using dendrimers,^{1–3} biopolymers,^{4–6} porphyrin aggregates,^{7–9} nanocrystalline materials,^{10,11} etc. The antenna materials absorb photons and transfer the excited state energy to an acceptor through Förster resonance energy transfer (FRET) as one of the options. This non-radiative excited state energy transfer phenomenon depends on the distance between donor and acceptor molecules and the overlap between the emission spectrum of the donor and the absorbance spectrum of the acceptor.^{12,13}

On several occasions coumarins have been used in polymers to harvest and transfer solar radiation energy.^{14–16} Systems were developed using fluorescent coumarin dyes (donors) covalently linked through an amide spacer to inorganic Ru(II) complexes (acceptors) where the peripheral coumarin antenna units sensitize the production of the metal-to-ligand charge transfer (MLCT) states of the complex.¹⁷ New coumarin derivatives were designed to produce light harvesting devices for constructing stable dye-sensitized solar cells.¹⁸ Ismail *et al.* included coumarin 6 (C6) (Fig. 1) in an active layer of a poly(3-hexylthiophene)(P3HT):[6,6]-phenyl-C₆₁ butyric acid

methyl ester (PCBM) to provide bulk heterojunction solar cell.¹⁹ They varied the amount of C6 in the active layer blends and recorded the optical absorption and photocurrent of the device.

C6, however, has a tendency to form microcrystals through H-type aggregation in solution.^{20–22} Early reports suggested the generation of C6 microcrystals through the reprecipitation method using a concentrated solution of the dye in acetone at room temperature with a large volume of water.²⁰ Later, our group established that even at micromolar concentration and in an aqueous medium, C6 quickly forms microcrystals (Fig. 2).²¹ The C6 microcrystals formed in water lead to quick loss in C6 fluorescence and hence their applications become limited. In some of our previous studies, we have reported ways to retrieve C6 fluorescence through guest–host chemistry in an aqueous medium.^{21,23} Herein, we state that C6 fluorescence can be revived by using micelles in water followed by nearly complete transfer of the energy absorbed from light to an acceptor through Förster resonance energy transfer (FRET).

Results and discussion

The aggregated C6 molecules (microcrystals) were treated with sodium dodecyl sulphate (SDS) in an aqueous medium. The absorption spectrum of C6 shows bands at around 460 and 500 nm in water after the microcrystal formation (Fig. 3). Absorbance at 500 nm, due to the aggregates,^{20–22} remains structurally unchanged upon addition of SDS up to ~6 mM, indicating the existence of the aggregates under pre-micellar conditions. The spectrum changes abruptly at and above 8 mM SDS concentration where the absorption band due to the aggregates starts disappearing.

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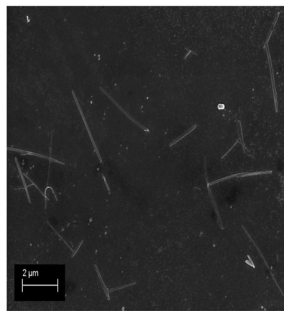


Fig. 2 Scanning electron microscopy image of C6 nanocrystals in water.

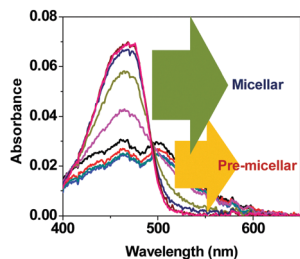


Fig. 3 Absorption spectra of C6 with added SDS (0–16 mM) in water.

Thus, it is evident that the C6 microcrystals begin to disaggregate in the presence of SDS micelles. This is understandable as C6 is a hydrophobic dye; hence the hydrophobic core of the micelle will attract the dye molecules. The predominating hydrophobic effect over the π - π stacking interaction leads to complete disintegration of the microcrystals.

Although the absorption spectrum of C6 did not show much change in structure at the pre-micellar concentration of SDS, the fluorescence spectrum shows noticeable sharpening with a hypsochromic shift (Fig. 4a). This is probably due to the SDS molecules surrounding the C6 microcrystals providing a relatively less polar environment in the vicinity. The fluorescence intensity of C6 increases substantially upon formation of the SDS micelles due to revival of the molecular characteristics of C6 (Fig. 4b and c). Time-resolved emission decay data of the excited state of C6 in water take a biexponential fit. The faster and slower components decay in 526 ps and 2.53 ns, respectively. The slower one is reported to be for the C6 molecule itself.²⁴ The faster component is for the intramolecular charge transfer (ICT) in C6.²¹ Table 1 provides the excited state lifetimes of C6 under different conditions in an aqueous environment. In water, the C6 microcrystals have π - π stacking interactions and hence a fast decay component is observed. However, upon encapsulation in SDS micelles, a growth component suggests the occurrence of ICT in the C6 molecules in the excited state. This is possible as the C6 molecules regain their molecular identity upon being disaggregated by the micelles.

Now as the nearly extinct fluorescence from the C6 microcrystals could be revived by applying micelles from SDS in an aqueous environment, we further intended to investigate execution of energy transfer by C6 in an aqueous medium, which is otherwise impossible due to microcrystal formation and loss in

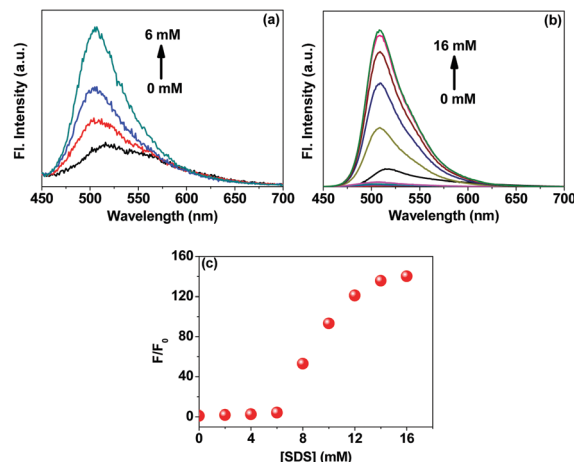


Fig. 4 Change in the fluorescence spectra of C6 in (a) pre-micellar and (b) micellar phases of SDS in water; and (c) relative change in fluorescence intensity with increase in SDS concentration. The excitation wavelength is 430 nm.

Table 1 Time-resolved emission decay data for C6 under different conditions at room temperature with and without added Rh123 in the presence of SDS micelles in water. χ^2 represents the goodness of the fits and the values in parentheses are the contributions of each of the components. The excitation wavelength is 405 nm

Conditions	τ_1 (ps)	τ_2 (ns)	χ^2
Monitored at 506 nm			
C6 in H ₂ O	526 (18.68%)	2.53 (81.32%)	1.13
C6 in SDS micelles	680 (−5.49%)	2.62 (105.49%)	1.02
[Rh123] (μ M)			
1	416 (−4.97%)	2.69 (104.97%)	1.03
2.4	405 (−4.35%)	2.75 (104.35%)	1.04
3.4	367 (−3.8%)	2.79 (103.8%)	1.08
5.4	358 (−3.05%)	2.88 (103.05%)	1.12
6.6	352 (−1.73%)	2.93 (101.73%)	1.17
Monitored at 540 nm			
C6 in H ₂ O	648 (38%)	2.22 (62%)	1.11
C6 in SDS micelles	790 (−4.75%)	2.62 (104.75%)	0.98
[Rh123] (μ M)			
1	—	3.21	1.13
2.4	—	3.63	1.17
3.4	—	3.85	1.13
5.4	—	4.14	1.20
6.6	—	4.29	1.19

fluorescence yield. Upon revival of fluorescence in water, C6 retains its molecular identity and is fit to act as an antenna material for energy harvesting. The spectroscopic characteristics of C6 and rhodamine 123 (Rh123) make them an appropriate FRET pair as in SDS micelles. The emission spectrum of C6 has an appreciable overlap with the absorption spectrum of Rh123 (Fig. S1, ESI[†]). We excited C6 in SDS micelles at 430 nm and gradually added Rh123 that accepts the emitted energy from C6 and emits at 540 nm (Fig. 5a). The calculated FRET efficiency shows nearly 100% transfer of energy from C6 in micelles to Rh123 (Fig. 5b). Hence, in totality, the concept fulfils the aspects for an efficient energy harvesting system in an aqueous environment.



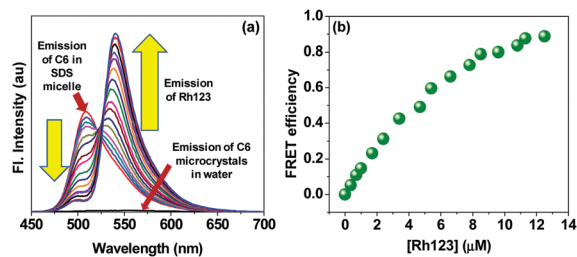


Fig. 5 (a) Förster resonance energy transfer (FRET) between C6 and Rh123 in SDS micelles in an aqueous environment, and (b) calculated FRET efficiency. The concentration of Rh123 was varied from 0 to 12.5 μM . The excitation wavelength is 430 nm.

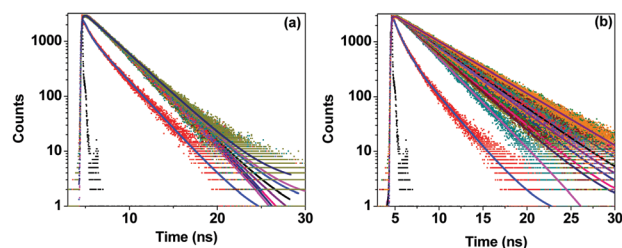


Fig. 6 Time resolved decay traces for C6 in water, SDS micelles and added Rh123 monitored at (a) 506 nm and (b) 540 nm. The solid lines are fits to the raw data. The sample was excited at 405 nm.

The emission from C6 is due to excited state intramolecular charge transfer (ICT) that is confirmed by the time resolved emission study. After exciting the donor (C6) using a suitable excitation source (405 nm), emissions at 506 nm (emission of C6) and 540 nm (emission of Rh123) were monitored. The decay profiles are shown in Fig. 6 and the lifetimes calculated from the fits are tabulated in Table 1. With gradual addition of the acceptor (Rh123), excited state ICT in C6 (growth component) becomes faster as the energy is donated by C6 to the acceptor. Monitoring the emission of Rh123 shows the stability of the acceptor as its excited state lifetime gradually increases.

Conclusion

In summary, we could devise a simple way to revive the fluorescence of C6 that disappears due to microcrystal formation in water through self-aggregation. Application of SDS micelles leads to disintegration of the aggregation by hydrophobic interaction and hence revives the lost fluorescence of C6. The energy from C6 could be transferred to Rh123 that acted as an acceptor in the FRET process. This work provides an efficient energy harvesting device in an aqueous environment under normal temperature and pressure.

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