



Cite this: *RSC Adv.*, 2018, 8, 18734

Fluorinated anionic fenugreek gum: their self-assembly behaviors and use as a novel thickening agent in fracturing gel

Chen Wang, * Xia Wang and Liewei Qiu

A novel fluorinated anionic fenugreek gum (FAFG) was obtained with excellent solution, gel, and broken-gel liquid properties in tertiary oil recovery, which were studied in detail and fenugreek gum (FG) and anionic fenugreek gum (AFG) were used as contrast samples. The results show that, compared to the FG and AFG solution properties, FAFG shows self-assembly properties, and inclusion of more fluorinated groups further reduced the critical associating concentration, yielding better associating properties. Compared to the FG and AFG gels, the FAFG gels exhibit good temperature resistance, shear resistance, rheological behaviors, and excellent salt tolerance. These improvements are attributed to the introduction of fluorinated groups and sulfonic groups. Compared to the FG and AFG broken-gel solutions, the FAFG broken-gel solutions presented slightly higher apparent viscosities. Moreover, the FAFG broken-gel solutions had profoundly lower surface tension and interfacial tension and extremely lower formation damage rates. They can hence serve as clean-up additives themselves after gel breaking. This automatically improves the flowback efficiency and reduces the formation damage. The residue rates of the FAFG broken-gel liquids were lower than those of the AFG and FG broken-gel liquids. Combined with its other excellent properties, FAFG would be an ideal component in fracturing gel.

Received 12th April 2018

Accepted 15th May 2018

DOI: 10.1039/c8ra03144d

rsc.li/rsc-advances

1. Introduction

Fenugreek gum (FG), which is derived from the endosperm of the seeds of fenugreek, is a highly branched polysaccharide. It is a galactomannan composed of an $\alpha(1/4)$ - β -D-mannan backbone attached to single α -D-galactopyranosyl groups at the O-6 positions of D-mannopyranosyl residues.^{1,2} FG has galactose and mannose residues in a 1 : 1.02 ratio; thus, it is a galactomannan with the highest amount of galactose. Thus, it has the highest water solubility and binding capacity among the galactomannans and yields high-viscosity aqueous solutions.³⁻⁵ Therefore, FG is used in myriad applications as binding, disintegrating, suspending, thickening, gelling, stabilizing, and protective agents.⁶⁻⁸

Anionic FG (AFG) is a kind of modified FG in which partial hydroxyl groups are replaced by carboxymethyl groups or sulfonic groups. While there are a few reports on AFG, Bassi & Kaur⁹ have obtained a carboxymethyl derivative of FG, which was found to possess bioadhesive properties and was therefore used for drug delivery. Unlike the carboxymethyl group, the sulfonic salt remains ionized over a wide range of pH values, does not form insoluble salts with bivalent metals such as Ca^{2+} or Mg^{2+} , and shows better tolerance to hard water. This makes

sulfonic acid derivatives of FG suitable for hydraulic fracturing.¹⁰

Hydraulic fracturing is widely used to stimulate oil and gas production from a reservoir. This technique improves well productivity by removing near wellbore damage and by increasing reservoir permeability in low as well as high permeability formations.¹¹ A hydraulic fracture is formed when a fluid is pumped down the well at high pressures for short periods of time. The high pressure fluid exceeds the rock strength and opens a fracture in the rock. A proppant is pumped into the fractures, and the mixture fills the fracture. The pressure is then released, allowing closure of the fracture onto the fluid/proppant mixture. After the treatment, the proppant remains in the created fracture in the form of permeable pack that serves to keep the fracture open. These proppant packs form conductive pathways for the hydrocarbons to flow into the wellbore, which will allow more extensive production at higher flow rates than otherwise possible. Leaving the gel in the fracture zone would cause formation damage by decreasing oil or gas production. Thus to complete the fracturing process, the fracturing fluid must be broken into broken-gel liquid. And the viscosity of broken-gel liquid should similar to that of water in order to recover from the formation.¹²

Nowadays, galactomannans and their derivatives are widely used as thickening agents for fracturing fluids to stimulate the oil and gas wells.¹³ However, fracturing fluids based on galactomannans and their derivatives, after a fracturing treatment,

Key Laboratory of Auxiliary Chemistry & Technology for Chemical Industry, Ministry of Education, Shaanxi University of Science & Technology, Xi'an 710021, P. R. China. E-mail: wangchenhg@sust.edu.cn



leave an insoluble polymer residue in the proppant pack inside the fracture and in the reservoir formation around the fracture.¹⁴ This residue reduces the permeability of the reservoir and the propped fracture. Hence, it damages the formation and limits the full potential that could be achieved with the stimulation operation.

Hydrophobically associating polysaccharides can aggregate together, assemble, and form a dynamic physical cross-linking network, which increases the hydrodynamic volume.^{15–17} Thus, it can substantially enhance the solution's viscosity and induce special rheological behavior in the aqueous solutions.¹⁸ Fluorocarbon chains possess a lower surface energy and a stronger associative effect than those of hydrocarbon chains.¹⁹ Therefore, their properties of thickening effects, salt resistance, and heat resistance are preferable when the fluorinated monomers are used as hydrophobic groups.^{20,21} Broken-gel liquids containing fluorinated groups possess a much lower surface tension and interfacial tension, which help produce more oil. In addition, when the fluorinated groups are incorporated into the molecules, they play the role of clean-up additives. This improves the efficiency of flowback and automatically reduces the formation damage. Therefore, fluorinated FG presents outstanding broken-gel performance.

This study presents a novel fluorinated AFG (FAFG) with excellent salt tolerance and outstanding broken-gel performance in hydraulic fracturing. The AFG is obtained by the modification of FG with 3-chloro-2-hydroxy propanesulfonate (CHP). Then, the FAFG was synthesized from a fluorinated reactive monomer (FAM) and AFG. The molecular structure of the product was characterized through Fourier-transform infrared (FT-IR) and fluorine-19 nuclear magnetic resonance (¹⁹F NMR) spectroscopy. The rheological and associating behaviors of FAFG gels and the properties of broken-gel liquids are reported in detail.

2. Materials and methods

2.1. Materials

FG—whose average molecular weight, M_w , was approximately 1.0×10^5 —was purchased from Beijing Jiade Biochemistry Co., Ltd. (Beijing, China). Hexamethylene diisocyanate (HDI) was purchased from Tianjin Kermel Chemical Reagent Co., Ltd. (Tianjin, China). Dibutyltin dilaurate (T-12) was purchased from Tianjing Hongyan Chemical Reagent Co., Ltd. (Tianjin, China). 1H,1H,2H,2H-perfluoro-1-decanol (PFD) was purchased from Harbin Xeogia Group Co., Ltd. (Harbin, China). Acetone, ethanol, tetrahydrofuran, petroleum ether, and sodium hydroxide were purchased from Zhonghai Chemical Industry (Shandong, China). CHP, sodium tetraborate, and ammonium persulfate were purchased from Shanghai Haoye Chemical Co., Ltd. (Shanghai, China). The FG was extracted using acetone for 24 h. Distilled water was used for all experiments.

2.2. Synthesis of AFG and FAFG

First, the FAM was synthesized from HDI and PFD through solution polymerization. A dry vessel fitted with a reflux

condenser, mechanical stirrer, and digital thermometer, and a nitrogen gas inlet was charged with 0.11 mol of HDI and 50 mL of acetone. A solution of 20 mL of acetone with 0.1 mol of PFD was added dropwise into the vessel for 1.0 h. Then, the T-12 (0.02 wt% based on the total reactant mass) was added, and the contents were stirred for 6.0 h under circulation reflux. After removal of the acetone by distillation, a viscous liquid was obtained, and FAM was finally obtained by washing the viscous liquid with petroleum ether three times (100 mL each time).

Second, AFG was synthesized from FG and CHP. Native FG powder (4.0 g) was suspended in a 200 mL mixture of solvents (the volume ratio of distilled water to ethanol was 1 : 2) and stirred for 30 min at room temperature. Next, 5 mL of NaOH solution (20%) was added in intervals of 1.0 mL every 10 min under continuous magnetic stirring at 60 °C. CHP of a specified weight was then added portion-wise to the reaction mixture over a period of 20 min. The reaction mixture was then heated to 70 °C for 4 h. The pH of the mixture was neutralized before the product was filtered *via* vacuum filtration; subsequently, it was washed three times with 50 mL of a hydro-alcoholic mixture (50% ethanol). The obtained product was then oven-dried at 105 °C for 24 h and powdered in a glass mortar.

Finally, a dry vessel fitted with the same equipment as described in the first step was charged with 5.0 g of the dried AFG and 200 mL of tetrahydrofuran. The solution of 20 mL of tetrahydrofuran with different amounts of FAM was added dropwise into the vessel for 1.0 h. Then, T-12 (0.02 wt% based on the total reactant mass) was added, and the contents were stirred for another 12.0 h under circulation reflux. The product was extracted from the acetone for approximately 24 h and dried in a vacuum. The degree of substitution (DS) was determined by nitrogen analysis. The synthetic processes are shown in Scheme 1.

2.3. Preparation of solutions and gels

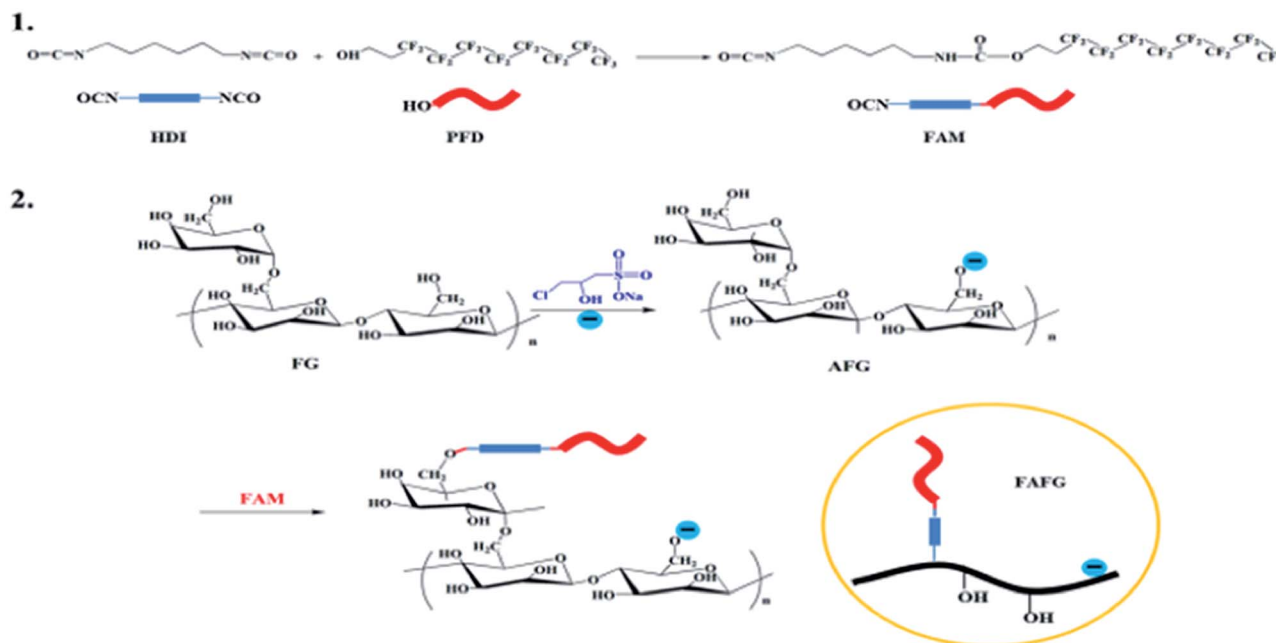
2.3.1. Preparation of solutions. Certain concentrations of FG, AFG, and FAFG solutions were prepared. FAFG powder was gradually dissolved in water by stirring at 300 rpm at 25 °C for 2.0 h to form a homogeneous mixture. FG and AFG solutions were prepared under the same conditions. The solutions were stored in containers under room temperature until use.

2.3.2. Preparation of gel. A cross-linking agent (sodium tetraborate) was added to the FAFG solutions at a volume ratio of active solution to cross-linking agent of 100 : 0.4. The mixtures were stirred with a glass rod until homogeneous gels were formed. FG and AFG gels were prepared under the same conditions. The gels were stored in containers under room temperature until use.

2.4. Preparation of broken-gel liquid

The gel breaker (ammonium persulfate) was added to the FAFG gels at a mass ratio of gel to gel breaker of 1000 : 0.5. The gel-breaking time and temperature were 2 h and 60 °C, respectively. The obtained solution was the broken-gel liquid. The broken-gel liquids of FG and AFG were prepared under the same





Scheme 1 Synthesis of FAM and FAFG.

conditions. The solutions were stored in containers under room temperature until use.

2.5. Preparation of standard mineralized water

The degree of mineralization, M , is defined as the total mass concentration of all salt in water (mg L^{-1}). The standard mineralized water was prepared as follows: 4902.68 g of distilled water was added to a 10 L beaker, which was continuously stirred by vigorous mechanical working to form a vortex. Then, 5.7165 g of anhydrous calcium chloride, 4.3150 g of magnesium chloride hexahydrate, and 87.2890 g of sodium chloride were added slowly in sequence—each must be completely dissolved before adding the next. The value of M was 19 334 mg L^{-1} . The total concentration of calcium and magnesium ions was 514 mg L^{-1} . Other mineralized waters were prepared by increasing or decreasing the various salt concentrations.

2.6. Test methods

2.6.1. Characterization. FT-IR spectroscopy of FG, AFG, and FAFG was performed using an FT-IR spectrometer (Model V70, Bruker) in dry air at room temperature. The gum samples were pressed directly on to attenuated reflectance KBr crystal as the sampling unit.

^{19}F -NMR spectroscopy of FAFG was performed using an NMR spectrometer (DPX-400 spectrometer, Bruker). The digital resolution was ± 0.01 ppm, with 0.1% D_2O as solvent.

DS, which is defined as the number of substituted hydroxyl groups per sugar unit of FG, was determined from the contents of nitrogen. The nitrogen content of the products (N wt%) was measured using an elemental analyzer (EA, Heraeus Co., Germany). The DS was calculated as described elsewhere.¹⁴ FAFG-1, FAFG-2, and FAFG-3 correspond to the cases in which the DS

values were 0.005, 0.01, and 0.02, respectively. The DS' (sulfonic acid groups) of AFG was 0.1.

2.6.2. Methods for testing solution and gel properties.

Multiangle dynamic light scattering measurements were carried out for the samples at five different scattering angles (50° , 70° , 90° , 110° , and 130°) using a Malvern 4800 photon correlation spectrometer.

Fluorescence spectroscopy (FS) measurements were performed using a Cary Eclipse Fluorophotometer at 25°C with excitation at 338 nm and a bandpass slit width of 3.0 nm in a scanning range of 350–550 nm. Pyrene was used as the fluorescent probe.

Scanning electron microscopy (SEM, VEGA3, TESCAN) was used to observe the sample microstructures. Prior to SEM imaging, the specimens were frozen in liquid nitrogen slush, and the ice sublimed away at low temperatures under a vacuum. Then, the freeze-dried samples were glued onto an aluminum stub, and their surfaces were coated with gold.

The rheological properties of the gels were determined using an RS150L Haake rheometer. The shear and temperature-resistant measurements were carried out with increasing temperature from 25°C to 70°C under a shear rate of 170 s^{-1} . The samples were then maintained at the test temperature (70°C) for an additional 3600 s. The dynamic measurement of FAFG gel ($\tan \delta$) was conducted in the temperature range of 25 – 70°C . These measurements were taken within the linear viscoelastic region. The apparent viscosity measurements of salt resistance were examined, while the test temperature and shear rate were set at 60°C and 7 s^{-1} , respectively.

2.6.3. Methods for testing broken-gel liquid properties.

Surface and interfacial tension measurements were carried out using a Theta Optical Tensiometer, which consists of a sample compartment, a light source, a lens, and an image capture camera. A pendant drop of a sample was formed within the



sample compartment by using a syringe, and the drop was recorded by the image capture camera. The surface tension was then obtained by analyzing the droplet with the help of the Young–Laplace equation. Measurements for each sample were repeated and then averaged. Measurements were conducted for the samples at 25 °C using the theta temperature control unit, which includes a heating plate in the sample compartment.

The residue content included the insoluble materials remaining in the broken-gel fluid after conventional gel breakage. A certain amount of broken-gel solution was poured into the centrifuge tube and centrifuged at 3000 rpm for 30 min. Then, the broken-gel residue was washed with 50 mL of distilled water and again centrifuged under the same condition after slowly pouring off the supernatant. This process was repeated three times. The obtained residue was dried to a constant weight and then weighed. The residue content was calculated as eqn (1):

$$R = m/V \quad (1)$$

where m is the mass of the residue (mg), V is the volume of the fracturing fluid (L), and R is the residue content of the fracturing fluid, mg L^{-1} .

The matrix permeability damage ratio was used to calculate the initial permeability ratio before damage. Then, the core was saturated again using the fracturing fluid filtrate, and the permeability ratio after damage was obtained. The matrix permeability damage ratio, D (%), was calculated as eqn (2):

$$D = (K_1 - K_2)/K_1 \times 100\% \quad (2)$$

where K_1 is the matrix permeability ratio before damage (μm^2), and K_2 is the matrix permeability ratio after damage (μm^2). The core was saturated again using salt water with the LDY32-300 core flow tester.

3. Results and discussion

3.1. Characterization of FAFG

FAM was successfully incorporated into FAFG, and this was verified by FT-IR and ^{19}F NMR (Fig. 1 and 2, respectively).

3.1.1. FT-IR. Fig. 1 presents the FT-IR spectra of FAM, FG, AFG, and FAFG. For FAM, there is a strong C–F absorption peak (1250 cm^{-1}) from PFD, and there are characteristic absorption bands of carbamate ($\nu_{\text{N-H}}$: 3340 cm^{-1} , 1558 cm^{-1} ; $\nu_{\text{C=O}}$: 1747 cm^{-1} ; $\nu_{\text{C-O}}$: 1145 cm^{-1}) in the spectrum of HDI.²² The carbimide group (2287 cm^{-1}) is still present, which indicates that FAM is synthesized successfully. Comparing the FT-IR spectra of FG and AFG, the characteristic absorption peaks of the sulfonated group (1051 cm^{-1} , 1217 cm^{-1}) appear in the spectrum of AFG.²³ These data indicate that AFG was prepared successfully.

Combined with the spectra of FAM, AFG, and FAFG, the characteristic absorption peaks of FAM appear in the FAFG spectrum. Simultaneously, the characteristic absorption of the aforementioned carbimide group vanishes. These data indicate that the FAM was successfully incorporated into the FAFG.

3.1.2. ^{19}F -NMR. As shown in the ^{19}F NMR spectrum (Fig. 2), the fluorine atoms are bonded in three different chemical environments in the FAFG molecules. Owing to the van der Waals force and coupling effect, the double peaks of the CF_3 groups appear in the low-field range (near -81.31 ppm) and the peaks of the CF_2 groups appear near -122.53 ppm . The peaks of CF_2 linked to CH_2 appear at -114.06 ppm .²⁴ The results of ^{19}F NMR spectroscopy confirm that the FAM was effectively introduced into FAFG.

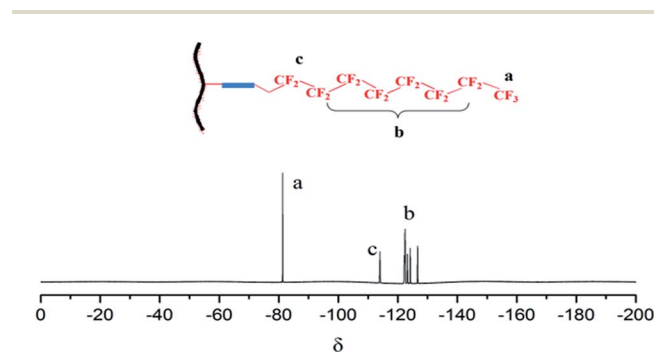


Fig. 2 ^{19}F -NMR spectrum of FAFG.

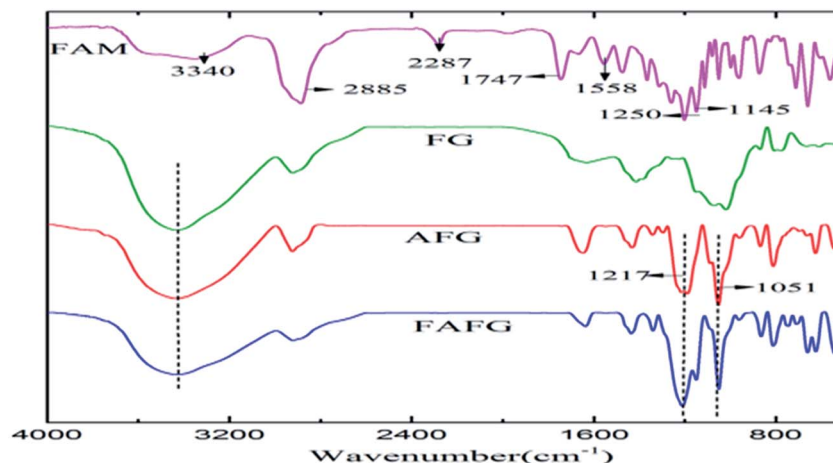


Fig. 1 FT-IR spectra of FAM, FG, AFG, and FAFG.



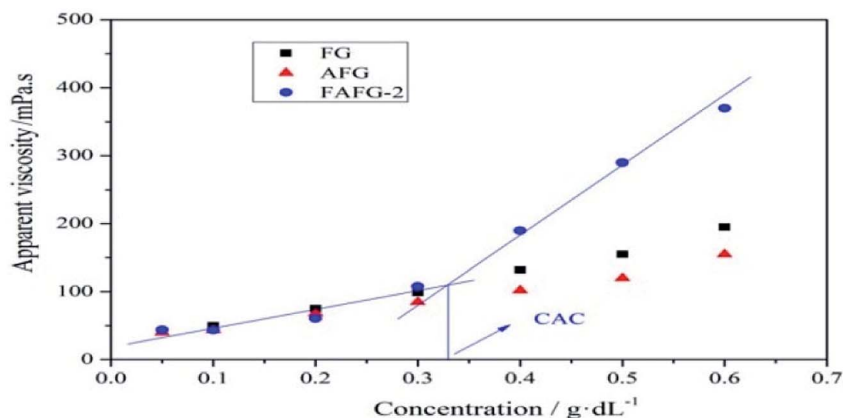


Fig. 3 Apparent viscosity of aqueous solutions (at room temperature).

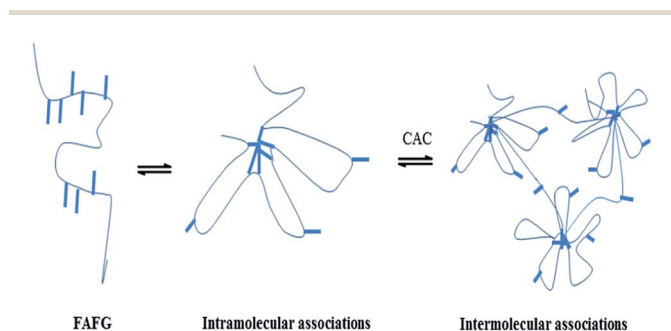
3.2. Properties of FAFG solutions

3.2.1. Critical associating concentration (CAC) of FAFG.

Fig. 3 definitively shows the special and unique associating behaviors of FAFG aqueous solutions from their macroscopic aspects. The data for FG and AFG are also shown for comparison. The change in the apparent viscosity of the sample solutions is the macroscopic behavior of their hydrophobic associative property, which presents the special thickening effect of FAFG.

The relationship between the concentration and the apparent viscosity is shown in Fig. 3. When the FAFG concentration increases, the apparent viscosities rise correspondingly, yet there is a sudden increase in the apparent viscosity. However, this abrupt change does not appear in the viscosity-concentration curves for FG and AFG. The resultant trend indicates that fluorinated groups are successfully introduced into AFG, which endow it with the special increasing pattern of apparent viscosity.

This nonlinear sudden increase of apparent viscosity of FAFG is the macroscopic expression of the transformation from intra-molecular aggregates to inter-molecular ones (Scheme 2), which results in the occurrence of the critical associating concentration (CAC). The CAC of FAFG-2 is 0.33 g dL⁻¹. The transformation causes the macromolecular chains to cross-link, increasing the hydrodynamic volume.



Scheme 2 Transformation from intramolecular associations to inter-molecular associations.

3.2.2. Fluorescence spectroscopy measurements.

In the fluorescence spectra of the pyrene probe, the first (I_1 : 371 nm) and the third (I_3 : 383 nm) emission peak intensities of pyrene were measured to calculate their ratio I_1/I_3 . This parameter strongly depends on the polarity around pyrene. Basically, a stronger polarity of the microenvironment around the pyrene probe leads to a higher value for I_1/I_3 . For example, the value of I_1/I_3 was 1.87 when the pyrene was in pure water and 0.58 in non-polar cyclohexane.

As seen in Fig. 4, the I_1/I_3 of the FAFG samples declines slightly from the beginning owing to the small amount of intramolecular hydrophobic micro-domains in their aqueous solutions. With increasing FAFG concentration, numerous hydrophobic micro-domains appear because of intermolecular associations, which drastically weakness the polarity of the microenvironment surrounding the pyrene, abruptly reducing the value of I_1/I_3 . Owing to the limited solubilization of pyrene in the solutions, the value of I_1/I_3 remains unchanged beyond certain concentrations. Additionally, the inclusion of more fluorinated groups further reduces the CAC values and yields better associating properties.

The value of I_1/I_3 in the FG and AFG aqueous solutions remains constant as their concentrations increase. It vividly displays that there are neither hydrophobic micro-domains nor associating effects in the FG and AFG solutions. However, the I_1/I_3 value of AFG is slightly higher than that of FG because the sulfonic acid groups in the AFGs result in a stronger polarity of the microenvironment surrounding pyrene. These conclusions confirm and supplement the results of the thickening effects by the measurement of apparent viscosity.

3.2.3. Apparent aggregate size.

Dynamic light scattering is an effective method for studying the solution properties of hydrophobic polymers, which can innately exhibit self-assembly behaviors in aqueous solutions. Fig. 5 shows the distributions of the apparent aggregate size of FG, AFG, and FAFG at concentrations of 0.1 g dL⁻¹ obtained *via* dynamic light scattering. FG and AFG show monodispersity, but FAFG shows polydispersity. The results demonstrate that associative behavior had already occurred in the FAFG aqueous solution. Numerous apparent aggregate sizes were detected because the



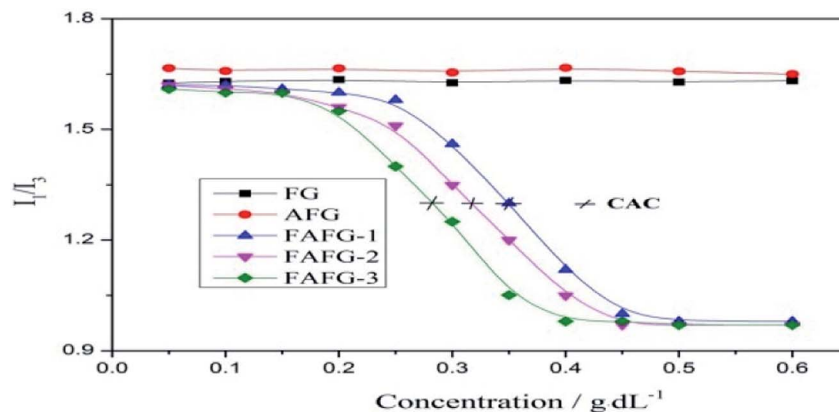


Fig. 4 I_1/I_3 values of aqueous solutions (at room temperature).

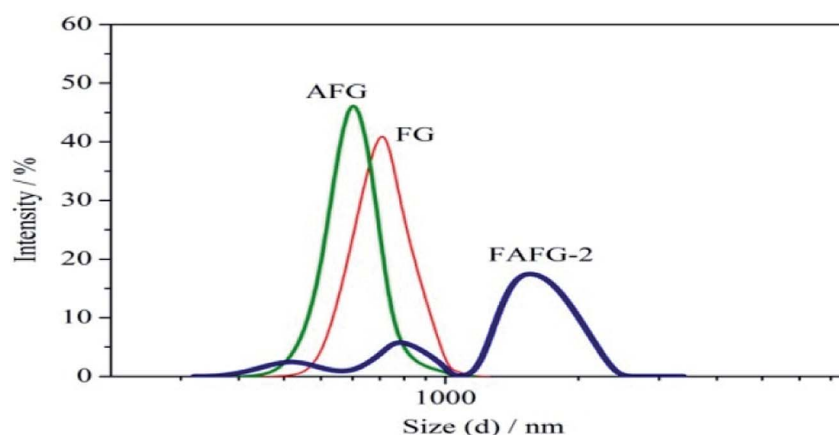


Fig. 5 Apparent aggregate size of different samples.

hydrophobic fluorinated groups can assemble together to form both intramolecular and intermolecular associations. Therefore, cross-linking structures formed, and each aggregate contains different numbers of FAFG molecules.

3.2.4. NaCl-resistant property. The concentration of NaCl had a noticeable effect on the apparent viscosity according to

the results in Fig. 6. Owing to the incorporation of sulfonic acid groups in FG, the AFG show improved salt resistant properties. Moreover, by incorporating fluorinated groups into the AFG, the viscosity reaches its maximum value with FAFG-2.

The non-polar hydrophobic groups in the FAFG tended to further increase the probability of association because of the

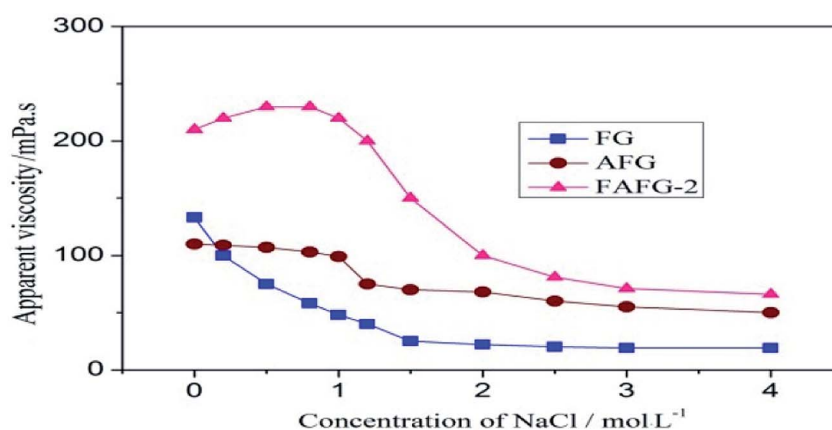


Fig. 6 Effect of NaCl concentration on apparent viscosity of various FG aqueous solutions.



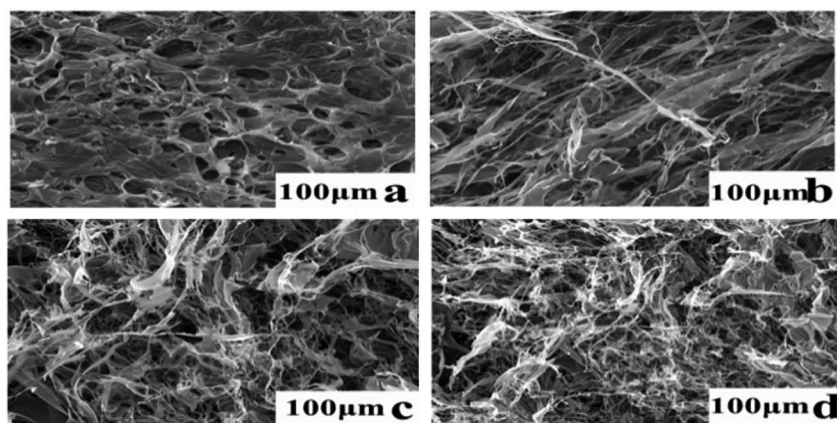


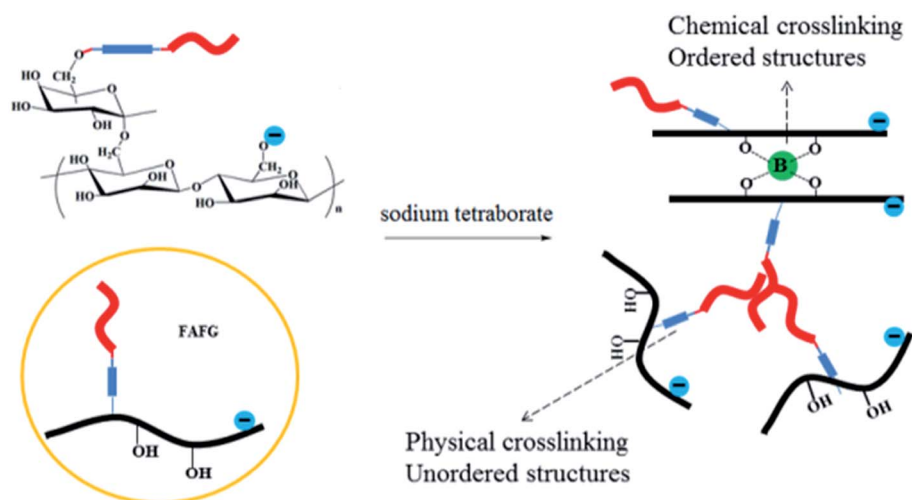
Fig. 7 Microstructures of the solutions: (a) AFG; (b) FAFG-1; (c) FAFG-2; and (d) FAFG-3.

enhancement of solution polarity with the relatively low concentration of NaCl solution; accordingly, a large increase in the number of the physical cross-linking points occurred and the apparent viscosity of the FAFG solution increased. However, when the concentration of NaCl was in the range of 0.8–2.0 mol L⁻¹, the apparent viscosity of the FAFG solution decreased rapidly. This result is attributed to the shielding effect of the negative charges in FAFG, which led in turn to a reduction in electro-static repulsion between polymer coils and between polymeric segments in the same coil. Consequently, there was a less remarkable expansion of the polymer coils in the solution. Additionally, the strong polarity brought about weak hydrogen bond actions. Both resulted in a relatively lower hydrodynamic volume, which was synonymous with a lower viscosity. In high concentrations of NaCl, the fluorinated hydrophobic groups in FAFG associated to maintain the apparent viscosity and did not decrease further. In addition, the branched structures were markedly improved in FAFG by the incorporation of sulfonic acid groups even if the strong polar ionic groups were shielded. These provided a larger steric

hindrance, enhancing the rigidity of the main chain and protecting FAFG from hydrolysis, which helped maintain the apparent viscosity of the solution. Thus, the self-assembly behavior of FAFG shows fine salt resistance.

3.2.5. Solution microstructures. The SEM images about freeze-dried AFG and FAFG solutions in Fig. 7 provide a direct visualization of these materials. To accomplish this, the species were frozen in liquid nitrogen slush, and the ice was sublimed away at a low temperature under a vacuum. Provided that the final structure, when coated with gold, is robust enough to sustain itself against gravity and weak electrostatic forces, it should be viewable in the SEM.

Fig. 7 shows four micrographs of AFG and FAFG solution samples with same sizes (100 μm). Fig. 7(a) shows that the AFG fibers appear uniformly distributed throughout the image, and void spaces exist among the fibers. However, as seen in Fig. 7(b), (c) and (d), the FAFG molecules were much more highly entangled and strongly interpenetrated than those of AFG. In addition, the higher DS yielded much tighter interpenetrations, indicating that the more fluorinated groups resulted in more



Scheme 3 Principle of gel formation.



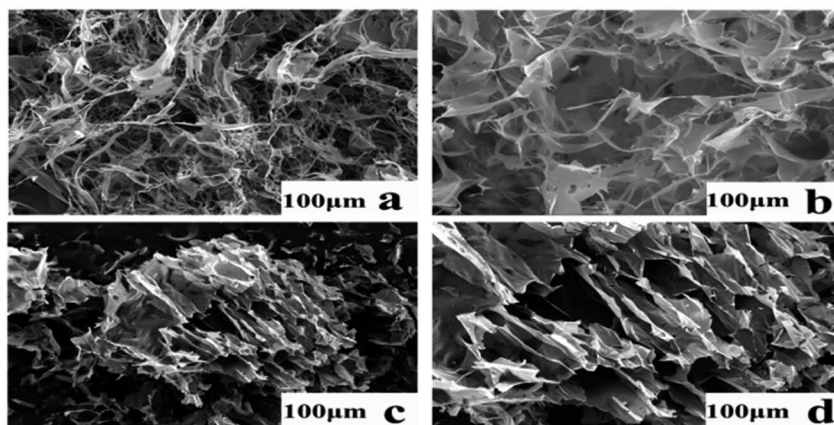


Fig. 8 Microstructures of the gels: (a) and (b) FAFG-2 solutions (counterpart); (c) and (d) FAFG-2 gels.

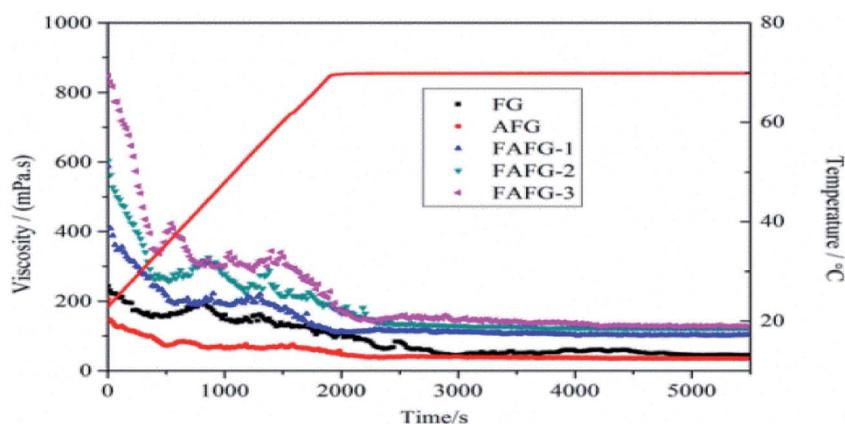


Fig. 9 Shear and temperature-resistant properties of gels.

physical networks in the solutions. The micrographs show that the FAFG solutions would be beneficial to fracturing operations.

3.3. Properties of FAFG gels

The cross-linking principles of FAFG and AFG are not the same. As shown in Scheme 3, the sodium tetraborate links FG macromolecules together through coordination bonds. Moreover, hydrophobic association occurs between FAFG macromolecules because they contain strong hydrophobic fluorinated groups. Thus, except for coordination cross-linking, physical cross-linking structures will also form in the gel system. The dynamic, three-dimensional networks endow the gel with a viscoelastic state.

3.3.1. Gel microstructures. Fig. 8 shows micrographs of FAFG-2 gel and its counterpart of FAFG-2 solutions with sizes of 100 and 50 μm . As seen from Fig. 8(a) and (b), the FAFG macromolecule in solutions were entangled uniformly and distributed throughout the image. However, for the micrographs of FAFG-2 gel in Fig. 8(c) and (d), well-ordered multi-layered structures appeared. It is well established that the requirements for coordination cross-linking is adjacent OH groups. Importantly, when the sodium tetraborate is used as

a crosslinker in the FAFG solution system, linear molecules (FAFG) are arranged and layered in order except for the networks formed by fluorinated association. The formation process is vividly interpreted in Scheme 3.

3.3.2. Shear resistance and heat resistance of FAFG gels.

The variations of viscosity with temperature and time are shown in Fig. 9. With the increase of shear time and temperature, the apparent viscosities of the FG, AFG, and FAFG gels decline, but the viscosities of the FAFG gels are greater than those of FG and AFG. Unlike the FG and AFG gels, hydrophobic associating physical cross-links formed by the fluorocarbon hydrophobic chains exist in the FAFG gels in addition to the chemical cross-linking formed by the cross-linking agent. As a result, the FAFG molecules are bonded by both chemical and physical cross-linking, resulting in more cross-links. Thus, the viscosity and shear resistance both increase accordingly.

As the temperature increases, the viscosities of the FAFG gels suddenly rise because the moderate heat makes the intramolecular associations transform into intermolecular associations. However, extending the shear time and further increasing temperature damage the association between the molecules, reduce the viscosity. Finally, a dynamic equilibrium between the entanglement and disentanglement of molecular chains is



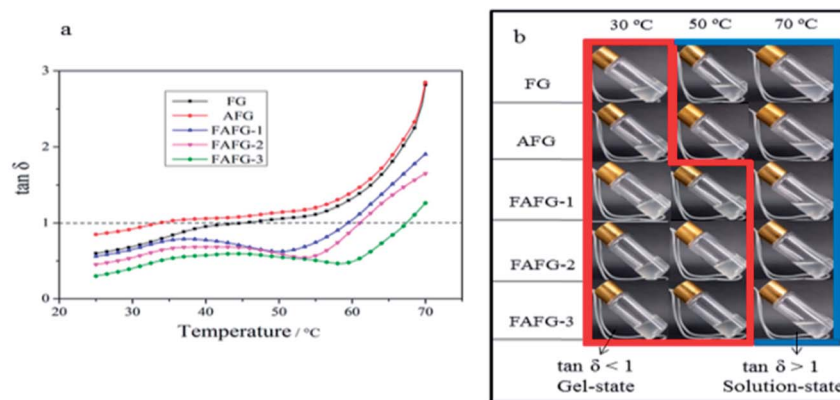


Fig. 10 Loss factors and digital photographs of FAFG gel at different temperatures: (a) loss factors; (b) digital photographs.

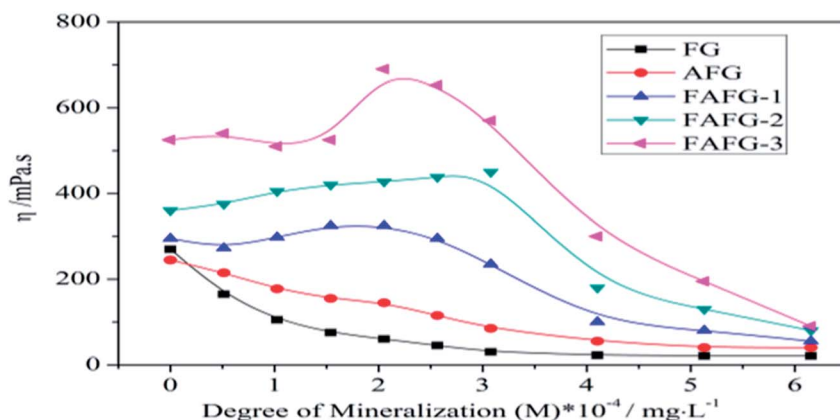


Fig. 11 Effects of M on the FG gels' viscosities.

reached, and the viscosity becomes constant. From the different DSs of the FAFG gels, it can be concluded that a higher DS improves the shear resistance and heat resistance.

3.3.3. Loss factor and digital photographs of FAFG gel. The loss factor ($\tan \delta$) is an important parameter for representing the viscoelasticity of a solution or a gel system. It exhibits viscosity when $\tan \delta > 1$ and elasticity when $\tan \delta < 1$. The loss factor ($\tan \delta$) and their digital photographs of FG, AFG and FAFGs at different temperatures are shown in Fig. 10.

The $\tan \delta$ of FG and AFG increases as the temperature increases, but $\tan \delta$ of FAFG shows different trends of decreasing at relative higher temperatures (40–60 °C). Correspondingly, the digital photographs in Fig. 10(b) presents that the FAFGs are gel-state and FG/AFG are solution-state at 50 °C. On the whole, $\tan \delta$ increases, showing the elasticity of the system decreases because the molecular thermal motions are increased. However, the $\tan \delta$ of FAFG shows decreasing in the temperature range of 40–50 °C (FAFG-1), 45–55 °C (FAFG-2), 50–60 °C (FAFG-3), suggesting that the FAFG can enhance the elasticity performances to some extent. These trends may be attributed to the temperature prompting “physical cross-linking” of hydrophobic groups in FAFG molecular fragments. They are thus associated together to improve the elasticity of the

system. Their digital photographs in Fig. 10(b) vividly complement the transforming process from gel to solution with the increase of temperature. In the red square box, the $\tan \delta < 1$ and digital photographs present gel-state. Otherwise, $\tan \delta > 1$ and digital photographs present solution-state in the blue square box. All the results show that the FAFG gels have better ability of temperature tolerance.

3.3.4. Salt resistance FAFG gel. Fig. 11 shows that the apparent viscosities of the FG and AFG gels decrease with the addition of minerals. The viscosity of the AFG gel is higher than that of the FG gel because AFG is an anionic polyelectrolyte. The molecules contain sulfonic groups ($-\text{SO}_3\text{Na}$), which are easily ionized to $-\text{SO}_3^-$ and $-\text{Na}^+$ in aqueous solutions. Thus, the macromolecules stretch owing to the strong electrostatic repulsions, so they can partially resist the salt.

However, the viscosity of the FAFG gel reaches a maximum value when M is near 2×10^4 mg L⁻¹. When the M value exceeds 3×10^4 mg L⁻¹, the FAFG gel's viscosity falls sharply. Additionally, a higher DS increases the FAFG gel's viscosity because it is influenced by the association between fluorinated molecular chains in the system. The gentle increase in M (gently increased polarity) is good for association. This leads to an increase in the number of physical cross-linking points,



Table 1 Properties of broken-gel liquids

Samples	Viscosities (mPa s)	Surface tensions (mN m ⁻¹)	Interfacial tensions (mN m ⁻¹)	Residue rates (%)
FG	5.4	40.99	4.88	245
AFG	2.9	47.12	6.77	175
FAFG-1	3.3	25.05	1.99	203
FAFG-2	3.9	22.23	1.37	218
FAFG-3	4.2	20.30	0.77	236

Table 2 Results of formation damage

Samples	Permeability of formation matrix (10 ⁻³ μm ²)		Formation damage rates (%)
	Before damage	After damage	
FG	3.28	2.88	12.19
AFG	3.28	2.97	9.45
FAFG-1	3.28	3.00	8.54
FAFG-2	3.28	3.06	6.71
FAFG-3	3.28	3.10	5.49

increasing the viscosity. When M continues to increase in the FAFG gel systems, the curl of the molecules is aggravated, and even macromolecules are degraded by high M .

3.4. Properties of FAFG broken-gel liquids

The performance of the broken-gel liquids directly affects the flowback of fracturing fluid, which is a principal factor for formation damage. The rational use of a gel breaker is helpful for accelerating gel breaking and expediting the flowback. Ammonium persulfate was used as the gel breaker, and the properties of the broken-gel liquid were studied.

3.4.1. Viscosity. The viscosities of the broken-gel liquids are shown in Table 1. The viscosity of FG was slightly lower than that of AFG because the introduction of $-\text{SO}_3^-$ into FG will enhance the repulsion between molecules, reducing the degree of entanglement between the molecules of the broken-gel liquid.

However, the viscosity of FAFG is higher than that of FG and AFG. Furthermore, a higher DS leads to higher viscosity. Because there are fluorinated groups in FAFG, these can be associated together to entangle the FAFG broken-gel pieces. When DS increases, the viscosities of the sample broken-gel liquids increase accordingly owing to the presence of more associating points.

3.4.2. Interfacial and surface tensions. The surface tension and interfacial tension of the broken-gel liquids were determined at room temperature. The results are shown in Table 1. It can be concluded that the FAFG broken-gel liquids possess much lower surface tension and interfacial tension. Moreover, the FAFG broken-gel solutions with a DS of 0.02 had profoundly lower surface tension (20.30 mN m⁻¹) and interfacial tension

(0.77 mN m⁻¹). The FAFG macromolecule breaks into smaller molecular fragments in this procedure, reducing its molecular mass. These molecular fragments contain sulfonic acid groups and fluorinated groups, which have good surface activity. In the process of flowback, they have a tendency for “active” flowback. This avoids the phenomenon of “chromatographic separation” of broken-gel liquids. Specifically, the clean-up additives used in oil fields are always fluorocarbon surfactants whose function is to hasten flowback of the broken-gel fluid. However, by directly incorporating the fluorinated groups in the molecules, they serve as clean-up additives after gel breaking. This automatically improves flowback efficiency and reduces formation damage.

3.4.3. Residue rates. The residue rates of the broken-gel liquids are shown in Table 1. The residue rate of AFG is substantially smaller than that of FG because AFG is more hydrophilic than FG, which leads to fewer residues. In contrast, the residue rate of FAFG is higher than that of AFG because of the strong hydrophobic groups introduced into FAFG. Furthermore, a higher DS leads to poorer hydrophilic behavior. Therefore, the residue rates of FAFG broken-gel liquids largely meet the standards.

3.4.4. Formation damages. Table 2 shows the damage rate of the broken-gel liquid on the matrix permeability at the tested temperature of 60 °C. The formation damage rate of the AFG broken-gel liquid is 22.48% smaller than that of the FG broken-gel liquid. This likely occurs because the sulfonic acid groups in the AFG yield repulsive force against the formation.

Likewise, the formation damage rates of the FAFG broken-gel liquids are much smaller than that of the FG broken-gel liquid (the formation damage rate of the FAFG-3 broken-gel liquid is 54.96% smaller than that of the FG broken-gel liquid). Moreover, the formation damage rate of the FAFG-1 broken-gel liquid is 35.71% smaller than that of the FAFG-3 broken-gel liquid, indicating that more fluorinated groups result in less formation damage. Because the crude oil in formation will be perfectly emulsified by the high surface activity of the broken-gel liquids, the formation damage is greatly reduced.

4. Conclusions

This study presented a novel FAFG with excellent salt tolerance and outstanding broken-gel performance in tertiary oil recovery. Compared to FG and AFG aqueous solutions, the FAFG aqueous solution showed self-assembly properties, and the CAC value of FAFG-2 was 0.33 g dL⁻¹. The inclusion of more fluorinated groups reduces the CAC, yielding better associating properties. Additionally, multiple apparent aggregate sizes of FAFG appeared owing to both intramolecular and intermolecular associations. The incorporation of sulfonic acid groups and fluorinated groups enhanced the salt resistance of the aqueous solution.

Compared to the FG and AFG gels, the FAFG gels exhibited good temperature resistance, shear resistance, rheological behavior, and excellent salt tolerance. These improvements are attributed to the introduction of fluorinated groups and



sulfonic groups. In addition, increasing the DS of the FAFG gel yielded better rheological behavior.

Compared to the FG and AFG broken-gel liquids, the FAFG broken-gel solutions presented slightly higher apparent viscosities, profoundly lower interfacial and surface tensions, and extremely lower formation damage rates. The residue rates of the FAFG broken-gel liquids were lower than those of the AFG and FG broken-gel liquids. Additionally, increasing the DS of the FAFG broken-gel liquid leads to better broken-gel performance. These results evince the potential applications of FAFG in fracturing fluids with its other excellent properties.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We would like to express gratitude to the Natural Science Foundation of China [grant number 5160030644]; and the Scientific Research Fund of Shaanxi Provincial Education Department [grant number 17JK0101]; and the Natural Science Foundation of Shaanxi Province (2017JM2017).

References

- 1 M. Kasran, S. W. Cui and H. D. Goff, *Food Hydrocolloids*, 2013, **30**, 552–558.
- 2 H. L. Jian, X. Lin, W. A. Zhang, W. M. Zhang, D. F. Sun and J. X. Jiang, *Food Hydrocolloids*, 2014, **40**, 115–121.
- 3 G. Ravindran and L. Matia-Merino, *Food Hydrocolloids*, 2009, **23**, 1047–1053.
- 4 Y. Wei, Y. Lin, R. Xie, Y. Xu, J. Yao and J. Zhang, *J. Food Eng.*, 2015, **166**, 21–28.
- 5 M. K. Youssef, Q. Wang, S. W. Cui and S. Barbut, *Food Hydrocolloids*, 2010, **23**, 2049–2053.
- 6 O. Kaltsa, N. Spiliopoulou, S. Yanniotis and I. Mandala, *Food Hydrocolloids*, 2016, **61**, 625–632.
- 7 M. Meghwal and T. K. Goswami, *J. Food Process. Technol.*, 2012, **3**, 1–10.
- 8 Y. Brummer, W. Cui and Q. Wang, *Food Hydrocolloids*, 2003, **17**, 229–236.
- 9 P. Bassi and G. Kaur, *Eur. J. Pharm. Biopharm.*, 2015, **96**, 173–184.
- 10 V. Mathur and N. K. Mathur, *J. Sci. Ind. Res.*, 2005, **64**, 475–481.
- 11 R. Barati and J. T. Liang, *J. Appl. Polym. Sci.*, 2014, **131**, 318–323.
- 12 M. Mage, *Chem. Eng. News*, 2013, **91**, 4.
- 13 S. Wang, Y. Zhang, J. Guo, J. Lai, D. Wang, L. He and Y. Y. Qin, *J. Pet. Sci. Eng.*, 2014, **124**, 432–435.
- 14 S. Yan, Y. Wang, J. He and H. Zhang, *Russ. J. Appl. Chem.*, 2015, **88**, 1884–1891.
- 15 D. Charpentiervalenza, L. Merle, G. Mocanu, L. Picton and G. Muller, *Carbohydr. Polym.*, 2005, **60**, 87–94.
- 16 Y. Feng, L. Billon, B. Grassl, G. Bastiat, O. Borisov and J. François, *Polymer*, 2005, **46**, 9283–9295.
- 17 X. Z. Yu, P. W. Fei, Z. L. Miao and E. J. Wang, *J. Phys. Chem. B*, 2005, **109**, 22250–22255.
- 18 C. Wang, X. Li, B. Du, P. Li and H. Li, *Carbohydr. Polym.*, 2013, **95**, 637–643.
- 19 Y. X. Zhang, A. H. Da, G. B. Butler and T. E. Hogen-Esch, *J. Fluorine Chem.*, 1991, **54**, 91–93.
- 20 H. Li, Z. Zhang, C. Hu, S. Wu and S. Ying, *Eur. Polym. J.*, 2004, **40**, 2195–2201.
- 21 J. Song, W. Fan, X. Long, L. Zhou, C. Wang and G. Li, *J. Pet. Sci. Eng.*, 2016, **146**, 999–1005.
- 22 E. Pretsch, P. Bühlmann and M. Badertscher, *Structure determination of organic compounds: tables of spectral data*, Springer, 4th edn, 2000, vol. 22, pp. 355–360.
- 23 L. Zhang, L. R. Kucera, S. Ummadisetty, J. R. Nykaza, Y. A. Elabd, R. F. Storey, K. A. Cavicchi and R. A. Weiss, *Macromolecules*, 2014, **47**, 4387–4396.
- 24 C. Schorn, D. Naumann, H. Scherer and J. Hahn, *J. Fluorine Chem.*, 2001, **107**, 159–169.

