Photolabile coumarins with improved efficiency through azetidinyl substitution†

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Azetidinyl substituents have been recently used to improve the fluorescence quantum yield of several classes of fluorophores. Herein, we demonstrate that other useful photochemical processes can be modulated using this strategy. In particular, we prepared and measured the quantum yield of photorelease of a series of 7-azetidinyl-4-methyl coumarin esters and compared it to their 7-diethylamino and julolidine-fused analogues. The efficiency of the photorelease reactions of the azetidinyl-substituted compounds was 2- to 5-fold higher than the corresponding diethylylamino coumarins. We investigated the origin of this effect in model fluorophores and in the photoactivatable esters, and found that H-bonding with the solvent seems to be the prominent deactivation channel inhibited upon substitution with an azetidinyl ring. We anticipate that this substitution strategy could be used to modulate other photochemical processes with applications in chemical biology, catalysis and materials science.

Introduction

Photochemical processes have been used extensively in a wide range of fields, including chemical biology,1 organic synthesis,2,3 and materials science,3 to gain selectivity and spatiotemporal control over the studied phenomena. The successful application of photochemical strategies relies on the ability of researchers to control the outcome of photoinduced reactions, by tuning chromophore parameters like absorption maxima, reaction selectivity or quantum efficiencies. A commonly adopted approach to modulate the photochemical evolutions of molecules consists in exploring structural variations of an already functioning scaffold4,5 to look for new motifs or substituents that enhance the desired properties.

Recently, Lavis and co-workers demonstrated that azetidinyl-substituted fluorophores such as 1 (λabs = 355 nm; λem = 471 nm; Chart 1) display larger quantum yields of fluorescence (Φf) compared to those substituted with open-chain analogues, such as dimethylmino or diethylamino (compound 2, λabs = 381 nm; λem = 468 nm), or larger cyclic amines, such as pyrrolidinyl or piperidiny6. Subsequently, Xu and co-workers found that aziridinyl-substituted fluorophores behave similarly.7 This improvement in Φf values has been attributed to a decrease in the rate of population of twisted intramolecular charge transfer (TICT) states9 upon excitation.7,9 We hypothesized that if these small heterocyclic rings suppress competitive decay channels that affect emission, the same effect could be exploited to improve the efficiency of other important, non-emissive photochemical processes.

4-Methylcoumarin derivatives are widely used as fluorophores and photoactivable (also known as “caging”) groups.10,11 Electron-rich coumarins may undergo photoinduced heterolytic cleavage when C4 is substituted with a methylene bearing a good leaving group (Scheme 1). The quantum yields of photoactivation (ΦPA) of such coumarins are usually moderate to low11-17 and depend on the nucleofugality of the leaving group.14 It would be advantageous to be able to improve the efficiency of these photoactivatable groups, in particular in a manner that is independent from the nucleofugality of the leaving group. Herein we report that azetidinyl substitution improves the efficiency of ΦPA of a series of photocleavable coumarins with leaving groups of diverse nucleofugality.

Chart 1 Structures of azetidinyl- and diethylaminocoumarins. The fluorescence quantum yields (Φf) shown were measured in H2O. In ref. 7 the reported Φf in aqueous HEPES buffer (pH 7.3) for 1 and for 7-diethylamino-4-methylcoumarin are 0.96 and 0.19, respectively. HEPES = 4-(2-hydroxyethyl)-1-piperazoneethanesulfonic acid.
Results and discussion

We synthesized 7-azetidinylcoumarin derivatives 3a–d, 7-diethylaminocoumarin derivatives 4a–d and the julolidine-fused derivatives 5a–d (Chart 2). We prepared the julolidine series because for these compounds the involvement of the TICT state is prevented by incorporation of the nitrogen into a system of fused rings. The \( \phi_{\text{PA}} \) values of compounds 3a–d, 4a–d and 5a–d were measured in mixtures of phosphate-buffered saline (PBS, pH 7.4) and MeCN (3/7, v/v). The rates of photolysis of 0.3 mM solutions were determined via HPLC by following the disappearance of the starting material upon irradiation at 405 nm (7.5 mW, see ESI†).

Fig. 1 depicts the results of photorelease experiments, including kinetic traces for representative runs of compounds 3b, 4b and 5b (Fig. 1a) and a comparison of the measured \( \phi_{\text{PA}} \) values of all the derivatives studied (Fig. 1b). In general, the \( \phi_{\text{PA}} \) values of derivatives 3a–d and 5a–d are comparable and significantly larger than the ones measured for derivatives 4a–d.

The most noticeable exceptions are compounds 3c, 4c and 5c, which have overall lower \( \phi_{\text{PA}} \), but even within this family, 3c and 5c seem to be photoreleased slightly more efficiently than 4c. For all other compounds, the 4- or 5-fold increase in \( \phi_{\text{PA}} \) observed is consistent with the increase observed between the \( \phi_{\text{f}} \) of compounds 1 and 2. These results confirm our hypothesis that azetidinyl substituents can be used to improve the efficiency of photochemical processes beyond fluorescence through inhibition of competitive deactivation pathways.

We then turned to investigate the nature of the decay channel inhibited by the azetidinyl substituent. Whereas it has been suggested that small-ring substituents inhibit the population of TICT states, it is not clear whether 7-substituted coumarins have never been firmly established. As a starting point, we performed femtosecond (fs) transient absorption (TA, 400 nm excitation and 100 fs time resolution, see ESI†) and fluorescence up-conversion spectroscopy measurements (FLUPS, 400 nm excitation, 1340 nm mixing pulse, 0.1 mm BBO, see ESI†) on the model fluorophores 1 and 2, to look for spectral signatures of a TICT state. We chose to work on the fluorophores because the measured \( \phi_{\text{f}} \) values (0.92 for 1 vs. 0.06 for 2) suggest that switching to the azetidinyl substituent renders radiative decay the predominant excited state process, whereas for the esters 3–5 heterolytic bond cleavage contributes to the kinetics of excited state decay, which may obscure the analysis.

The FLUPS and TA data were analyzed globally assuming a series of exponential steps to obtain evolution associated emission or differential absorption spectra (EAES or EADS), respectively. FLUPS data in H_{2}O and DMSO (Fig. 2) could be described for both dyes assuming three successive exponential steps. In H_{2}O (Fig. 2a–d), a two-step red shift was observed, with a magnitude twice as large for 2 than for 1 (~1200 and ~200 cm\(^{-1}\) vs. ~680 and ~100 cm\(^{-1}\), respectively). Furthermore, the longer time constant (reflecting the fluorescence lifetime, \( \tau_{\text{f}} \), also determined from time-correlated single photon counting experiments, see ESI†), is more than one order of

![Scheme 1](image-url)  
Scheme 1 Photoduced heterolytic cleavage of donor-substituted coumarins. R\(_{1}\), R\(_{2}\) = alkyl chains, cyclic or acyclic. LG = leaving group.

![Chart 2](image-url)  
Chart 2 Structures of compounds 3a–d, 4a–d and 5a–d. For details of the syntheses see the ESI†.
magnitude smaller for 2 than for 1 (0.40 ns vs. 5.0 ns). On the contrary, in DMSO (Fig. 2e-f) the red shifts (700 cm⁻¹ for both compounds) and the \( \tau_F \) (2.7 ns for 2 vs. 3.3 ns for 1) are comparable for the two dyes. The initial red shifts reflect the equilibration of the surrounding polar solvent molecules. The larger shifts observed with 2 in H₂O point to substantial solvent reorganization and stabilization of the excited state, such as those associated with hydrogen-bond (H-bond) interactions. The shorter \( \tau_F \) of 2 in H₂O and MeOH (see ESIT), but not in other highly polar solvents such as MeCN and DMSO (see ESIT), does not support involvement of a TICT state. The TA data did not reveal either any spectral signature attributable to any state other than the optically-populated excited state (see ESIT). Therefore, other deactivation pathways must be operative.

The shortening of \( \tau_F \) for 2 in polar protic solvents strongly suggests the possibility of H-bond induced non-radiative decay (HBIND) as deactivation mechanism of the excited state, a phenomenon already reported for other dyes. The efficiency of this process was shown to depend on both the H-bond donating strength of the solvent, described by the Kamlet–Taft parameter \( \alpha_s \), and the ability of the solvent to make an H-bond network, quantified by the density of OH groups \( \rho_{OH} \), itself approximated as \( 1/V_s \), where \( V_s \) is the volume of a solvent molecule. To test this hypothesis, we determined the non-radiative decay rates \( k_{nr} \) of 1 and 2 from the \( \tau_F \) and \( \phi_F \) values in a series of solvents of different H-bonding abilities (Fig. 3a, see ESIT). The correlation observed between \( k_{nr} \) and \( \alpha_{pOH} \) supports the involvement of HBIND as deactivation channel for 2 in protic solvents, but does not explain why the same pathway is not active for 1, which is structurally very similar.

To shed light on the origin of these differences, we analyzed the solvatochromism of the absorption and emission bands of 1 and 2 in solvents of different polarities, both protic and aprotic (Fig. 3b and c, see ESIT). From these measurements it is possible to estimate the ground \( (\mu_g) \) and excited \( (\mu_e) \) state electric dipoles of the two dyes (see ESIT). We observed that the dipole moments of 1 \( (\mu_g = 3.9, \mu_e = 9.5 \text{ D}) \) are significantly smaller than for 2 \( (\mu_g = 7.5, \mu_e = 11.8 \text{ D}) \). Analysis of the solvatochromism of the absorption bands reveals further details. For 2 in protic solvents, a consistent bathochromic shift is observed (Fig. 3d), whereas for 1 both batho- and hypso-chromic shifts are found. These shifts can be rationalized considering the internal charge transfer (ICT) character of the first singlet excited state of coumarins, in which the substituent at C7 donates electron density to the carbonyl group at C2.

**Fig. 2** (a and b) Time-resolved fluorescence of 1 (a) and 2 (b) in H₂O. (c–f) EAES obtained from a global target analysis of the time-resolved fluorescence of 1 in H₂O (c) and DMSO (e) and of 2 in H₂O (d) and DMSO (f), assuming a series of three successive exponential steps with the time constants indicated in the legends.

**Fig. 3** (a) Non-radiative decay rate constant of 2 vs. solvent H-bonding ability. (b and c) Solvatochromism of 1 and 2. Absorption (b) and emission (c) maximum wavelengths vs. the solvent polarity function. (d) Solvatochromism of absorption of 1 and 2 in protic solvents. The points represent individual measurements, and the associated errors of the slopes represent the 95% confidence interval of the fits (see ESIT). TFE = trifluoroethanol; HFP = hexafluoroisopropanol.
of phenylazetidine (7.61 eV)\textsuperscript{28} compared to that of $N$,N-diethylthylaniline (6.95 eV).\textsuperscript{29}

To confirm that this mechanism also applies to the photolabile esters, we measured the $\phi_{PA}$ of compounds 3a–5a using n-pentanol (PeOH) and EtOH as co-solvents (see ESI\textsuperscript{†}) instead of PBS because both display lower H-bonding ability than H$_2$O. In EtOH mixture (for PeOH results, see ESI\textsuperscript{†}), the $\phi_{PA}$ of 3a, 4a and 5a became very similar ($\phi_{PA} = 0.007, 0.0037$, and 0.01, respectively). These efficiencies are in general lower than in aqueous mixture, which can be attributed to the reduced stabilization (and hence formation) of the ion pair in less polar solvents. These observations further support our working hypothesis that azetidinyl substituents shut down HBIND-associated decay of coumarin excited states, leading to increased fluorescence or photoactivation efficiency in H$_2$O.

Finally, the applicability of this improved photoremovable group was confirmed in live cells. We chose to prepare compound 6 (Fig. 4, for synthesis see ESI\textsuperscript{†}), which is a photoactivatable fluorescence probe. Photoactivatable dyes are useful for a number of applications including cell tracking,\textsuperscript{30} intracellular diffusion experiments\textsuperscript{31} and super-resolution microscopy.\textsuperscript{32} Live human cervical cancer (HeLa) cells were incubated with compound 6 in growth medium for 30 min. After this time, imaging in the green channel ($\lambda_{ex} = 488$ nm; $\lambda_{em} = 525 \pm 25$ nm) gave nearly no signal, confirming that the fluorescein fragment of compound 6 remains in the dark, spirolactone form (Fig. 4) and the coumarin is not excited at that wavelength. A region of interest comprising a single cell (Fig. 4) was irradiated with a 405 nm laser (120 mW, 50% power), which induced a 7-fold increase in fluorescence intensity. This experiment confirms that the azetidinylcoumarin protecting group can be removed with light in live cells.

Conclusions

To the best of our knowledge, azetidinyl substituents have been used, in the context of photochemistry, only to increase the $\phi_{PA}$ of fluorophores.\textsuperscript{28} We have demonstrated, however, that this simple substitution leads to the enhancement of other useful photoprocesses in the presence of H$_2$O. We also propose a mechanism of quenching that does not invoke TICT states, but rather HBIND as the unproductive decay channel that is shut down by azetidinyl donors in coumarins. Albeit beyond the scope of this work, we propose that azetidinyl substitution could enhance other important photochemical processes in H$_2$O; for instance, would an azetidinylated version of methylene blue be more efficient in photocatalysis\textsuperscript{33} or photodynamic therapy?\textsuperscript{24} Further studies are required to investigate the origin of the increase in $\phi_{PA}$ observed for other fluorophores and determine whether azetidinyl substitution affects the population of TICT states, as currently thought, or it deactivates other excited state decay pathways as it seems to be happening for the coumarins reported here. Testing these hypotheses might reserve interesting challenges and surprises.

Conflicts of interest

The authors declare no competing financial interests.

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References