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Norborn-2-en-7-ones as physiologically-triggered carbon monoxide-releasing prodrugs†

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A prodrug strategy for the release of the gasotransmitter CO at physiological pH, based upon 3a-bromo-norborn-2-en-7-one Diels–Alder cycloadducts of 2-bromomaleimides and 2,5-dimethyl-3,4-diphenylcyclopentadienone has been developed. Examples possessing protonated amine and diamine groups showed good water solubility and thermal stability. Half-lives for CO-release in TRIS-sucrose buffer at pH 7.4 ranged from 19 to 75 min at 37 °C and 31 to 32 h at 4 °C. Bioavailability in rats was demonstrated by oral gavage and oCOM-21 showed a dose dependent vasorelaxant effect in pre-contracted rat aortic rings with an EC₅₀ of 1.6 ± 0.9 μM. Increased intracellular CO levels following oCOM-21 exposure were confirmed using a CO specific fluorescent probe.

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Introduction

Carbon monoxide (CO) is a key player in heme oxygenase mediated cellular regulation and protection in health and disease. Evidence, by our group and others,^{1,2} has established the endogenous existence of critical, CO-activated protective signaling pathways (vasodilatory, anti-apoptotic, anti-thrombotic and anti-inflammatory). For example, delivery of CO gas at low concentrations protects organs from ischaemic injury by decreasing cell death and most significantly inducing a preconditioning response.^{3,4} Carbon monoxide binds to hemeproteins, to modulate the activity or level of expression of key cellular targets such as soluble guanylate cyclase (sGC), and inducible nitric oxide synthase to produce a plethora of downstream signaling events.^{5,6} The anti-inflammatory effects of CO are mediated by p38 MAPK signaling and inhibition of the translocation of the potent damage-associated molecular pattern recognition molecule HMBG1. This results in a reduction of pro-inflammatory cytokines, such as TNF-α, and an increase in the expression of the anti-inflammatory IL-10.^{7,8}

The protective and physiological effects of low levels of CO have been studied in a number of both animal and clinical models with the literature extensively reviewed.^{5,8–12} The results of these studies have been so compelling that the FDA has granted

approval for the application of low dose CO gas in a range of clinical application trials such as CO gas delivery by inhalation in heart valve replacement surgery¹³ and in renal transplant procedure recipients.¹⁴ Inhalation of CO (100–125 ppm) in chronic obstructive pulmonary disease (COPD) patients, reduced sputum neutrophils and improved bronchial responsiveness.¹⁵ In renal transplant procedures low dose CO gas during surgery improves post-transplant kidney function.¹⁶ However, the difficulty of controlling gas delivery in a clinical setting, combined with the hazardous consequences of any gas leak, have been acknowledged as significant impediments in the use of CO in gaseous form.^{6,17} CO-releasing molecules (CORMs) have been developed as an alternative CO delivery system allowing a greater ease of administration and control, and potentially enabling a safer, tightly regulated method of low dose CO delivery. These molecules have been extensively studied and their beneficial effects¹⁸ have been demonstrated in transplant and disease models.^{19–22} A large proportion of existing CORMs are based on transition-metal carbonyl complexes [M_xLy(CO)_z, where M = Mn, Ru, Fe, W, Mo, Cr, Ir or Re].^{23–25} Early metal carbonyl complexes suffered from low water solubility and high toxicity. The attachment of organic ligands to the metal centre resulted in a newer generation of metal carbonyl complexes, such as CORM-3,²⁶ with increased water solubility. However, an issue with metal carbonyl complexes is that the residual by-products (iCORMs) obtained after CO release contain transition metals which is undesirable and could contribute to toxicity.^{27–29}

Several CORMs that do not contain transition metals have been reported.³⁰ One example is sodium boranocarbonate (CORM-A1),³¹ which is water soluble, capable of liberating CO at pH 7.4 and was found to improve the renal function of rabbit kidney transplants upon reperfusion.³² More recent organic

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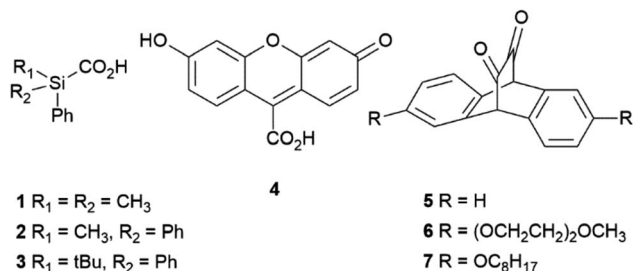


Fig. 1 Selected examples of organic CORMs.

examples (Fig. 1) include the silacarboxylic acids 1–3³³ the fluorescein analogue 4³⁴ and the cyclic α -diketones 5–7.³⁵

Silacarboxylic acids 1–3 have only been employed as a source of CO gas for carbonylation reactions and require the addition of fluoride-based desilylating reagents to induce CO release.^{33,36} The fluorescein analogue 4 requires exposure to visible light (500 nm) for CO release,³⁴ while the cyclic α -diketones 5–7 need to be encapsulated within pluronic micelles so that photoactivation (400–550 nm) can result in effective CO release. However, these examples have yet to show clinical relevance because of issues associated with the mechanism of CO release, solubility and cellular toxicity.

The focus of the current work is the development of organic CORMs that would release CO on physiological stimulus. Furthermore, the formation of an easily characterized organic by-product after CO release would allow determination of the CO-release kinetics whilst potentially reducing residual compound toxicity. Our approach is based upon the bridged carbonyl compound norbornadien-7-one (**8**)³⁷ which undergoes cheletropic loss of carbon monoxide (CO) giving benzene as a by-product (Fig. 2A).

It is well known that compound **8** is remarkably prone to fragmentation with loss of CO and several computation studies^{37,39} have been carried out on this reaction where the activation energy for this process was calculated to be between 11 to 12 kcal mol^{−1} with the corresponding rate constant at 300 K (27 °C) of $4.2 \times 10^5 \text{ s}^{-1}$ ($t_{1/2} = 1.6 \times 10^{-5} \text{ s}$).

With this in mind, Wang *et al.* have recently developed an elegant “click and release” CO prodrug utilising intermolecular Diels–Alder reactions of functionalised tetraphenylcyclopentadienones with strained alkynes to generate unstable norbornadienone intermediates with concomitant loss of CO.⁴⁰ An intramolecular variant, where the alkyne was tethered to the diene (Fig. 2B, **9**) provided a series of CORMs with CO release half-lives, controlled by the rate of the Diels–Alder reaction, ranging from 2 minutes to one week.³⁸ These compounds demonstrated possible therapeutic applications in an anti-inflammatory assay and a colitis animal model, however DMSO was required to solubilize them.

We sought an alternative approach where physiological pH controls the release of CO using bridged carbonyl compounds based upon a norbornenone substructure. These well-known compounds are generally prepared using Diels–Alder reactions of substituted cyclopentadienones and activated dienophiles, such as maleic anhydride, giving stable cycloadducts. Early work on these compounds carried out by Allen *et al.*⁴¹ and Fuch *et al.*^{42,43} showed that compounds such as **10** could decompose under thermal and/or photochemical conditions to release carbon monoxide (Scheme 1).

However, the conditions for CO release by heating (200 °C)⁴⁰ or photochemical activation^{42,43} as reported for **10** preclude the use of such compounds as CORMs in a biological setting. Given this we have developed a strategy that utilises the properties of both the norbornenone (stability) and norbornadienone (rapid decomposition releasing CO) class of compound.

This approach provides a prodrug for an organic CO donor molecule, the general strategy of which utilizes compounds with the functionalised norbornenone structure represented by **12** (Fig. 3). Elimination of HX would produce the transient norbornadienone which would undergo rapid/spontaneous cheletropic loss of CO and concomitant aromatisation to by-product **13**. X would be a halogen (Cl, Br or I) and R_4 an electron withdrawing group. The rate determining step, an E1cB process, would be affected by the substituents R_3 and R_4 and thus would control the rate of CO release. This could be achieved when R_3/R_4 was a cyclic imide, which would be less

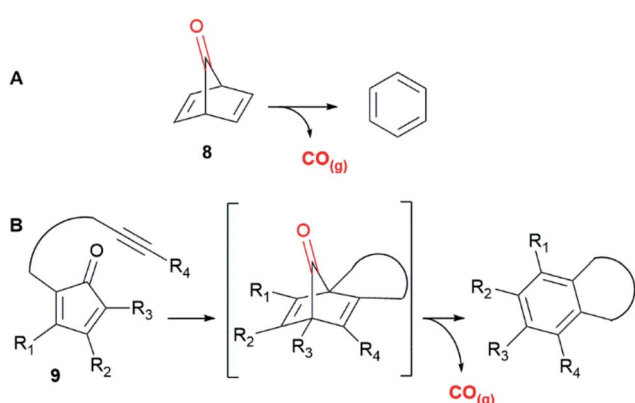
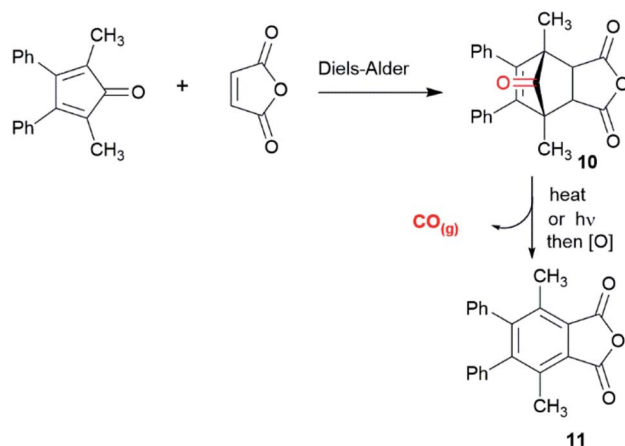


Fig. 2 (A) Cheletropic loss of carbon monoxide from norbornadien-7-one (**8**). (B) Strategy reported by Wang *et al.*³⁸



Scheme 1 Formation and aromatisation of cycloadduct **10**.^{41,42}



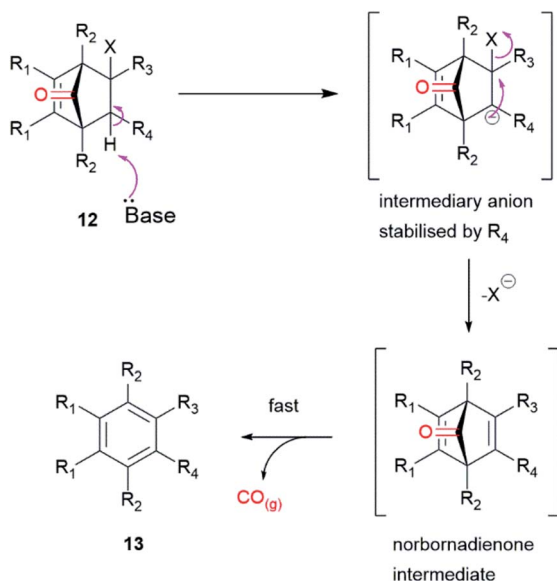
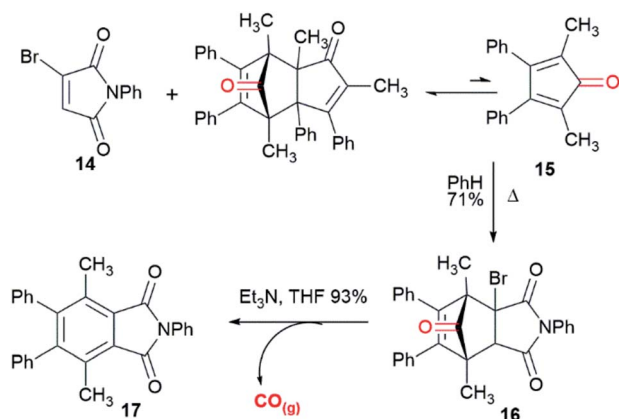


Fig. 3 Strategy for CO-release from a functionalized norbornen-7-one.

susceptible to hydrolysis than an anhydride and also provide a handle for further functionalization. This structural motif would allow the process to be triggered by raising the pH to give an intermediary anion, stabilized by the imide, followed by loss of halide to give a norbornadienone intermediate.

Discussion

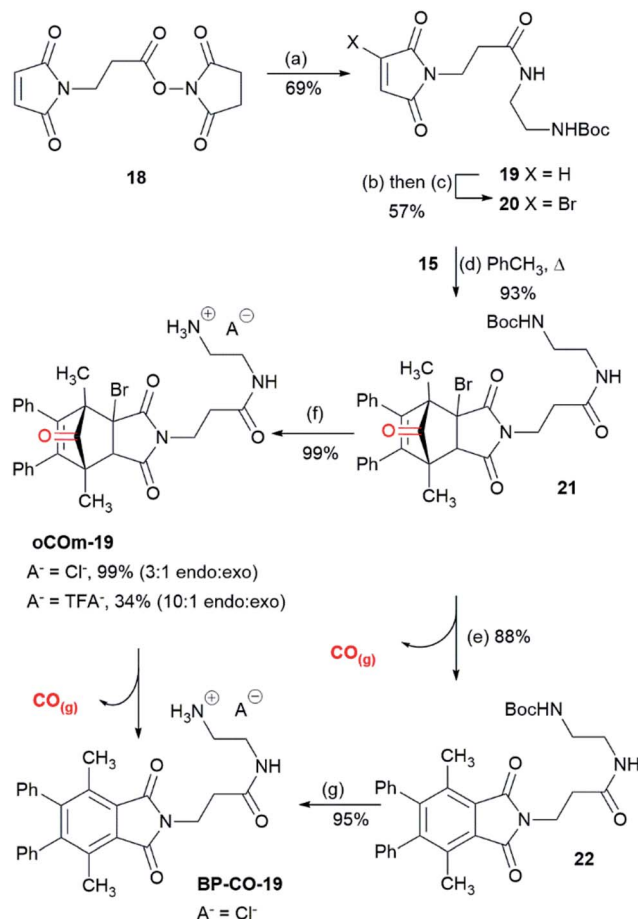
Einhorn *et al.*⁴⁴ reported the synthesis of polyarylphthalimides from the Diels–Alder reaction of tetraarylcyclopentadienones and 2-bromomaleimide in refluxing bromobenzene which occurred with concomitant decarbonylation. Our attempts to carry out the reaction of tetraphenylcyclopentadienone and 2-bromo-*N*-phenylmaleimide (**14**) at lower temperatures in order to isolate the Diels–Alder cycloadduct were unsuccessful due to the slow rate of reaction in refluxing toluene. We surmised that this was due to steric interactions between the 2- and 5- phenyl groups and the bromine substituent of the maleimide.



Scheme 2 Synthesis and decarbonylation of CORM 16.

Pleasingly, the less sterically-encumbered 2,5-dimethyl-3,4-diphenylcyclopentadienone (**15**), which exists as a reversible dimer, reacted smoothly with **14** to give endo-**16** in 71% yield after crystallisation from diethyl ether (Scheme 2). To test whether this compound could release CO it was treated with triethylamine in THF. Effervescence occurred and phthalimide **17** was isolated in 93% yield. GC/MS headspace analysis of a solution of **16** in DMSO in a sealed tube after treatment with DBU confirmed that the gas released was CO. ¹H NMR analysis of the solution indicated the complete conversion of **16** into **17**.

With the proof of principle that cycloadducts like **16** can release CO on treatment with base, attention turned towards making water soluble analogues. To this end a series of CORMs functionalized with amine salts were prepared. Maleimide **18**,⁴⁵ with a pendant activated ester was reacted with mono-Boc protected ethylene diamine giving **19** which was subjected to a bromination–dehydrobromination sequence to give bromo-maleimide **20** in 57% yield over the two steps (Scheme 3). Cycloaddition of diene **15** and **20** gave adduct **21** as a 3 : 1 mixture of endo and exo-isomers in 93% yield. Adduct **21** was



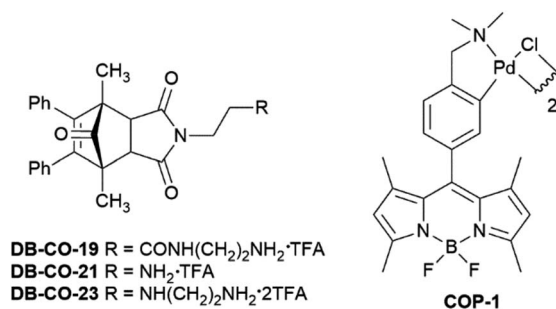
Scheme 3 Synthesis of oCOM-19 reagents and conditions: (a) BocNHCH₂CH₂NH₂, CH₂Cl₂, 0 °C to rt (69%). (b) Br₂, CCl₄, reflux. (c) NEt₃, THF, 0 °C (57% over two steps). (d) PhCH₃, **15**, reflux, (93%). (e) DBU, THF, rt, (88%). (f) 6 M HCl, 1,4-dioxane, 0 °C to rt, $A^- = Cl^-$ (99%), $A^- = TFA^-$, (34% + inseparable mixture of oCOM-19 and BP-CO-19). (g) 6 M HCl, 1,4-dioxane, 0 °C to rt, (95%).

sensitive to base and on treatment with DBU in THF gave phthalimide **22** (88%). Removal of the Boc-group of adduct **21** was achieved on reaction with 6 M HCl in dioxane to give **oCom-19** as its HCl salt (99%, 3 : 1 mixture of endo and exo isomers). In a separate reaction, the reaction product was subjected to reverse phase chromatography on a C-18 solid phase extraction cartridge using 1% aqueous TFA and acetonitrile which resulted in partial separation of the endo- and exo-isomers (10 : 1, 34%) of **oCom-19** as the TFA salt, with the remainder of the material as a mixture of **oCom-19** and aromatised product, iCORM **BP-CO-19**. The latter compound was also prepared in 95% yield by treatment of **22** with 6 M HCl in dioxane.

Using a similar process ethylenediamine and diethylenetriamine derived CORMs were prepared from carboethoxymaleimide **23** (Scheme 4).

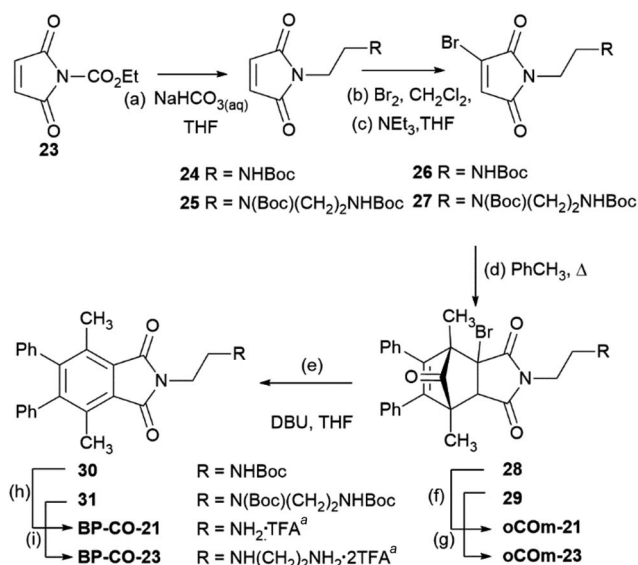
Reaction of bromomaleimide **26** with diene dimer **15** gave cycloadduct **28** (endo : exo = 2.5 : 1) in 84% yield after purification by column chromatography. Partial separation was achieved with a sample of endo-**28** isolated in 32% yield with the remaining 52% a 1 : 1 mixture of endo- and exo-isomers. Similarly, reaction of bromomaleimide **27** with **15** gave a 3 : 1 mixture of endo- and exo-adducts **29** which after column chromatography afforded the endo-isomer (60%) along with a mixture of the isomers in 27% yield. Removal of the Boc-protecting groups of endo-**28** and -**29** was achieved on treatment with 6 M HCl in dioxane and gave, after purification on reverse-phase solid-phase cartridges and lyophilization, **oCom-**

21 and -**23** both in 63% yield as their mono- and bis-trifluoroacetate salts as white powders. Aromatisation of **28** and **29** on treatment with DBU gave phthalimides **30** and **31** and subsequent removal of the Boc-group provided the respective iCORMs **BP-CO-21** and **BP-CO-23** in 48 and 62% yield respectively over the two steps. A series of desbromo-analogues of **oCom-19**, -**21** and -**23**, **DB-CO-19**, -**21** and -**23** respectively, which lack the ability to release CO under physiological conditions, were also synthesised by Diels-Alder reactions of diene **15** and maleimides **19**, **24**, and **25** and subsequent deprotection of the Boc groups for use as control compounds (Scheme S1, ESI†).



To assess whether CORMs **oCom-19**, -**21** and -**23** have therapeutic potential their water solubility and CO release profiles were measured at pH 7.4. Pleasingly all three were soluble in tissue culture water at 20 °C at 3.2, 10.8 and 18.2 mg mL⁻¹ respectively. Transplant organ preservation solutions are normally maintained at 8 °C. Unfortunately, the solubility of **oCom-19** at 8 °C was low and could not be determined, whereas those lacking the amide linking group, **oCom-21** and -**23**, were measured as 3.9 and 17.6 mg mL⁻¹ respectively. The CO release profiles for the more soluble CORMs **oCom-21** and -**23** were assessed by monitoring the formation of the corresponding iCORMs, **BP-CO-21** and **BP-CO-23** in TRIS-sucrose buffer at pH 7.4 using an HPLC method. **oCom-21** showed half-lives for CO release of 19 min at 37 °C (Fig. S1, ESI†) and 31 h at 4 to 5 °C. **oCom-23** half-lives at the same temperatures were 75 min and 32 h, respectively. The CO release was also supported by monitoring *via* a CO-selective electrode over a range of pH and myoglobin assays (Fig. S2a to c, ESI†). There was no sign of the formation of the corresponding iCORMs from solutions of **oCom-19**, -**21** and -**23** in aqueous media at pH < 7 after prolonged periods of time (>1 week). Samples showed no deterioration after being left at room temperature exposed to ambient light for over one month. The thermal stability of **oCom-21** was investigated by thermogravimetric analysis and was shown to undergo decomposition at 125 °C (Fig. S3a, ESI†). Further analysis showed that heating a sample of **oCom-21** to 90 °C and holding it at that temperature for 30 min resulted in a 4% loss of mass (Fig. S3b, ESI†). HPLC analysis of this sample showed that it was a 96 : 4 mixture of **oCom-21** and its corresponding iCORM (Fig. S3c, ESI†).

In context to applications such as organ transplantation, the toxicity of **oCom-19**, -**21** and -**23** was assessed using sensitive MDCK cells at 37 °C and EC₅₀ values were measured as 86, 11.2 and 10.1 μM, respectively (Fig. S4, ESI†). iCORMs **BP-CO-21** and



Scheme 4 Synthesis of **oCom-21** and -**23**. Reagents and conditions: (a) BocNH(CH₂)₂NH₂, THF, H₂O, 0 °C to rt, **24** (58%); BocNH(CH₂)₂-NBoc(CH₂)₂NH₂, THF, H₂O, 0 °C to rt, **25** (mixture with ethyl carbamate). (b) Br₂, CCl₄, reflux. (c) NEt₃, THF, 0 °C, **26** (73% over two steps), **27** (25% over two steps + 1 : 1 inseparable mixture of **27** and ethyl carbamate). (d) PhCH₃, **15**, reflux, **28** (84% 2.5 : 1 endo : exo), **29** (60% endo) (e) DBU, THF, rt, **30** (61%), **31** (76)%. (f) and (g) 6 M HCl, 1,4-dioxane, 0 °C to rt, **oCom-21** (63%), **oCom-23** (63%). (h) 6 M HCl, 1,4-dioxane, 0 °C to rt (78%). (i) 6 M HCl, 1,4-dioxane, rt to 50 °C (82%). ^aPurified by reverse phase C-18 solid phase extraction cartridge using 1% aqueous TFA and acetonitrile.

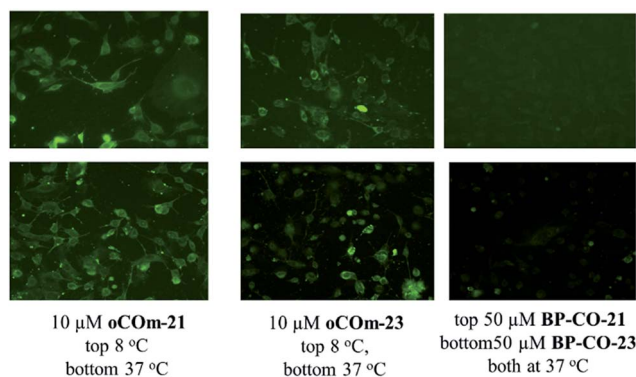


Fig. 4 CO specific fluorescent probe (COP-1) confirms increased intracellular CO levels following exposure to **oCOM-21** (10 μ M) or **oCOM-23** (10 μ M) against the respective **iCORM** (50 μ M) at both 37 $^{\circ}$ C and 8 $^{\circ}$ C.

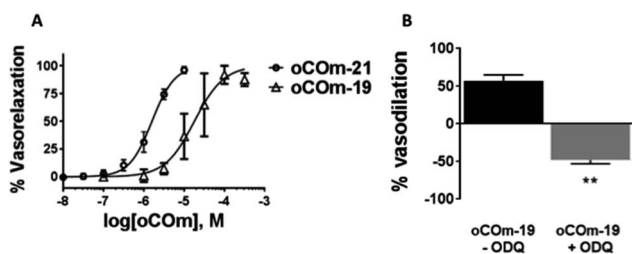


Fig. 5 Vasorelaxant effect of **oCOM-19** and **-21** in isolated rat aortas at 37 $^{\circ}$ C. (A) Relaxation responses in precontracted aortic rings expressed as a percent change of the contractile response to phenylephrine (100 nM). (B) Responses to EC_{50} concentrations of **oCOM-19** repeated in the presence of the guanylate cyclase inhibitor ODQ. Data was analysed using a One-way ANOVA with Bonferroni post-hoc test and expressed as mean \pm SEM. ** denotes $P < 0.01$ = vs. the non (–) ODQ inhibited group.

-23 gave EC_{50} values of 21 and 100 μ M, respectively. The toxicity of **oCOM-21** was also measured against this cell line at 8 $^{\circ}$ C and gave an $EC_{50} > 100 \mu$ M.

CO delivery to MDCK cells by **oCOM-21** and **-23** at 8 and 37 $^{\circ}$ C was confirmed using the CO-selective fluorescent probe **COP-1** developed by Chang *et al.*⁴⁶ (Fig. 4).

Oral bioavailability in rats was demonstrated by carboxyhaemoglobin analysis following administration of **oCOM-19** and **-21** by oral gavage. Moderate but significant increases in carboxyhaemoglobin levels in the plasma were obtained after 3 h of administration for the less soluble **oCOM-19** and within 1 h for **oCOM-21** (Table S1, ESI[†]). **BP-CO-19**, which cannot release CO, showed no increase in carboxyhaemoglobin at the same time points.

CO relaxes vascular tissues by acting on the cGMP pathway and high conductance calcium-activated K^+ (K_{Ca}) channels. To assess the vasorelaxant effect of our compounds, pre-contracted rat aortic rings were treated with **oCOM-19** and **-21**. A dose-dependent relationship was observed for both compounds with EC_{50} values of $18.0 \pm 0.8 \mu$ M and $1.6 \pm 0.9 \mu$ M respectively (Fig. 5). This vasodilatory effect was reversed by the addition of 1*H*-(1,2,4)oxadiazolo[4,3-*a*]quinoxalin-1-one (ODQ), an inhibitor of soluble guanylate cyclase (sGC).

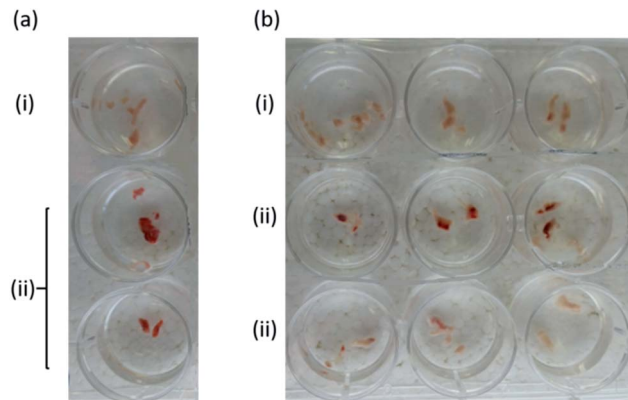


Fig. 6 Protective effect of **oCOM-21** on heart tissue (rat myocardium): (a) fresh heart tissue samples (i) untreated and not stained (ii) untreated and stained with TTC stain; (b) heart tissue samples after 48 hours incubation (i) control heart tissue samples stained with TTC stain (ii) heart tissue samples incubated with **oCOM-21** (3 μ M) after 48 hours and stained with TTC stain (iii) heart tissue samples incubated with **DB-CO-21** (3 μ M) after 48 hours and stained with TTC stain.

In a preliminary experiment, the preservation of tissue viability was demonstrated for rat myocardium. Tissue samples treated with **oCOM-21** stored in Dulbecco's modified Eagle medium appeared to be more viable after 48 h compared to an untreated control based upon dehydrogenase activity indicated by conversion of tetrazolium chloride (TTC, white) into the corresponding formazan (red) (Fig. 6).

Conclusions

In conclusion, we have developed CO prodrugs based upon norbornenones which function *via* E1cB elimination of HBr under physiological conditions, producing a reactive norbornadienone intermediate that rapidly undergoes chelotropic loss of CO. The three prodrugs, **oCOM-19**, **-21**, and **-23** are thermally stable solids that show good water solubilities and CO release profiles. Under physiological conditions they have been shown to deliver CO to cells, have a vasorelaxant effect and have the potential to preserve tissue sample viability. We are developing new generations of these compounds and investigating their applications as additives for organ transplantation preservation solutions.

Acknowledgements

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Notes and references

- 1 I. A. Sammut, R. Foresti, J. E. Clark, D. J. Exon, M. J. J. Vesely, P. Sarathchandra, C. J. Green and R. Motterlini, *Br. J. Pharmacol.*, 1998, **125**, 1437–1444.



- 2 A. Ayer, A. Zarjou, A. Agarwal and R. Stocker, *Physiol. Rev.*, 2016, **96**, 1449–1508.
- 3 A. Bagul, S. A. Hosgood, M. Kaushik and M. L. Nicholson, *Transplantation*, 2008, **85**, 576–581.
- 4 I. C. Winburn, J. C. Harrison, R. J. MacGinley, R. J. Walker and I. A. Sammut, *Transplantation*, 2006, **82**, 1043.
- 5 S. Ghosh, J. Gal and N. Marczin, *Ann. Med.*, 2010, **42**, 1–12.
- 6 S. W. Ryter and L. E. Otterbein, *BioEssays*, 2004, **26**, 270–280.
- 7 L. E. Otterbein, F. H. Bach, J. Alam, M. Soares, L. H. Tao, M. Wysk, R. J. Davis, R. A. Flavell and A. M. Choi, *Nat. Med.*, 2000, **6**, 422–428.
- 8 Y. Ruan, L. Wang, Y. Zhao, Y. Yao, S. Chen, J. Li, H. Guo, C. Ming, S. Chen, F. Gong and G. Chen, *Kidney Int.*, 2014, **86**, 525–537.
- 9 M. Bilban, A. Haschemi, B. Wegiel, B. Y. Chin, O. Wagner and L. E. Otterbein, *J. Mol. Med.*, 2008, **86**, 267–279.
- 10 S. Brouard, L. E. Otterbein, J. Anrather, E. Tobiasch, F. H. Bach, A. M. Choi and M. P. Soares, *J. Exp. Med.*, 2000, **192**, 1015–1026.
- 11 N. Hill-Kapturczak and A. Agarwal, *Am. J. Physiol. Renal. Physiol.*, 2006, **290**, F787–F788.
- 12 S. W. Ryter, J. Alam and A. M. Choi, *Physiol. Rev.*, 2006, **86**, 583–650.
- 13 J. J. Freiburger, *CO as a Stimulant for Mitochondrial Biogenesis in Human Cardiac Muscle*, ClinicalTrials.gov Bethesda (MD): National Library of Medicine, US, 2012, <https://clinicaltrials.gov/ct2/show/NCT01727167>.
- 14 Mallinckrodt Pharmaceuticals, *Safety and Tolerability Study of Inhaled Carbon Monoxide in Kidney Transplant Patients*, ClinicalTrials.gov Bethesda (MD): National Library of Medicine, US, 2007, <https://clinicaltrials.gov/ct2/show/NCT00531856>.
- 15 E. Bathoorn, D. J. Slebos, D. S. Postma, G. H. Koeter, A. J. van Oosterhout, M. van der Toorn, H. M. Boezen and H. A. Kerstjens, *Eur. Respir. J.*, 2007, **30**, 1131–1137.
- 16 H. H. Usansky and K. Jamil, Dosing regimes and methods of treatment using carbon monoxide, US 2011/0280966, 5/12/2011, 1911.
- 17 S. R. Thom, L. K. Weaver and N. B. Hampson, *Am. J. Respir. Crit. Care Med.*, 2005, **171**, 1318.
- 18 C. Steiger, C. Hermann and L. Meinel, *Eur. J. Pharm. Biopharm.*, 2016, DOI: 10.1016/j.ejpb.2016.11.002, ahead of print.
- 19 A. Vannacci, C. Marzocca, L. Giannini, L. Mazzetti, S. Franchi-Micheli, P. Failli, E. Masini, R. Motterlini and P. F. Mannaioni, *Inflammation Res.*, 2006, **55**, S05–S06.
- 20 Y. Caumartin, J. Stephen, J. P. Deng, D. Lian, Z. Lan, W. Liu, B. Garcia, A. M. Jevnikar, H. Wang, G. Cepinskas and P. P. Luke, *Kidney Int.*, 2011, **79**, 1080–1089.
- 21 A. Sener, K. C. Tran, J. P. Deng, B. Garcia, Z. Lan, W. Liu, T. Sun, J. Arp, M. Salna, P. Acott, G. Cepinskas, A. M. Jevnikar and P. P. Luke, *J. Urol.*, 2013, **190**, 772–778.
- 22 M. D. Pizarro, J. V. Rodriguez, M. E. Mamprin, B. J. Fuller, B. E. Mann, R. Motterlini and E. E. Guibert, *Cryobiology*, 2009, **58**, 248–255.
- 23 S. H. Heinemann, T. Hoshi, M. Westerhausen and A. Schiller, *Chem. Commun.*, 2014, **50**, 3644–3660.
- 24 R. Motterlini and L. E. Otterbein, *Nat. Rev. Drug Discovery*, 2010, **9**, 728–743.
- 25 A. C. Kautz, P. C. Kunz and C. Janiak, *Dalton Trans.*, 2016, **45**, 18045–18063.
- 26 J. E. Clark, P. Naughton, S. Shurey, C. J. Green, T. R. Johnson, B. E. Mann, R. Foresti and R. Motterlini, *Circ. Res.*, 2003, **93**, e2–e8.
- 27 M. Vadori, M. Seveso, F. Besenzone, E. Bosio, E. Tognato, F. Fante, M. Boldrin, S. Gavasso, L. Ravarotto, B. E. Mann, P. Simioni, E. Ancona, R. Motterlini and E. Cozzi, *Xenotransplantation*, 2009, **16**, 99–114.
- 28 I. C. Winburn, K. Gunatunga, R. D. McKernan, R. J. Walker, I. A. Sammut and J. C. Harrison, *Basic Clin. Pharmacol. Toxicol.*, 2012, **111**, 31–41.
- 29 S. Romanski, E. Stamellou, J. T. Jaraba, D. Storz, B. K. Kramer, M. Hafner, S. Amslinger, H. G. Schmalz and B. A. Yard, *Free Radical Biol. Med.*, 2013, **65**, 78–88.
- 30 S. H. Heinemann, T. Hoshi, M. Westerhausen and A. Schiller, *Chem. Commun.*, 2014, **50**, 3644–3660.
- 31 R. Motterlini, P. Sawle, S. Bains, J. Hammad, R. Alberto, R. Foresti and C. J. Green, *FASEB J.*, 2005, **19**, 284–286.
- 32 A. Sandouka, B. J. Fuller, B. E. Mann, C. J. Green, R. Foresti and R. Motterlini, *Kidney Int.*, 2006, **69**, 239–247.
- 33 S. D. Friis, R. H. Taaning, A. T. Lindhardt and T. Skrydstrup, *J. Am. Chem. Soc.*, 2011, **133**, 18114–18117.
- 34 L. A. P. Antony, T. Slanina, P. Sebej, T. Solomek and P. Klan, *Org. Lett.*, 2013, **15**, 4552–4555.
- 35 P. Peng, C. Wang, Z. Shi, V. K. Johns, L. Ma, J. Oyer, A. Copik, R. Igarashi and Y. Liao, *Org. Biomol. Chem.*, 2013, **11**, 6671–6674.
- 36 S. D. Friis, T. Skrydstrup and S. L. Buchwald, *Org. Lett.*, 2014, **16**, 4296–4299.
- 37 J. Zhang, D. M. Ho and R. A. Pascal Jr, *J. Am. Chem. Soc.*, 2001, **123**, 10919–10926.
- 38 X. Ji, C. Zhou, K. Ji, R. E. Aghoghovbia, Z. Pan, V. Chittavong, B. Ke and B. Wang, *Angew. Chem., Int. Ed.*, 2016, **55**, 15846–15851.
- 39 C.-H. Lai, E. Y. Li, K.-Y. Chen, T. J. Chow and P.-T. Chou, *J. Chem. Theory Comput.*, 2006, **2**, 1078–1084.
- 40 D. Wang, E. Viennois, K. Ji, K. Damera, A. Draganov, Y. Zheng, C. Dai, D. Merlin and B. Wang, *Chem. Commun.*, 2014, **50**, 15890–15893.
- 41 C. F. H. Allen and J. Van Allan, *J. Am. Chem. Soc.*, 1942, **64**, 1260–1267.
- 42 B. Fuchs, *J. Chem. Soc. C*, 1968, 68–71, DOI: 10.1039/j39680000068.
- 43 B. Fuchs and G. Scharf, *J. Org. Chem.*, 1981, **46**, 5395–5398.
- 44 R. Vanel, F. Berthiol, B. Bessieres, C. Einhorn and J. Einhorn, *Synlett*, 2011, 1293–1295.
- 45 Y.-C. Su, Y.-L. Lo, C.-C. Hwang, L.-F. Wang, M. H. Wu, E.-C. Wang, Y.-M. Wang and T.-P. Wang, *Org. Biomol. Chem.*, 2014, **12**, 6624–6633.
- 46 B. W. Michel, A. R. Lippert and C. J. Chang, *J. Am. Chem. Soc.*, 2012, **134**, 15668–15671.

