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# Cochinchinenin C, a potential nonpeptide anti-diabetic drug, targets a glucagon-like peptide-1 receptor

Yijie Sha, Yunfeng Shi, Bing Niu\* and Qin Chen \*

The glucagon-like peptide-1 (GLP-1) receptor is currently being explored as a therapeutic target for anti-diabetic drugs. GLP-1 analogs possess therapeutic effects similar to those of other anti-diabetic drugs such as guanidine and sulfonylureas but do not cause hypoglycemia and gastrointestinal discomfort. GLP-1 has the ability to reduce blood glucose in a glucose-dependent manner. Several GLP-1 analog agonists have been developed. However, polypeptide drugs are easily degraded by DPP4 *in vivo*. Therefore, the focus is now on the development of nonpeptide anti-diabetic drugs targeting the GLP-1 receptor. In this study, computer-aided drug design was applied to search for potential molecules of this type. Cochinchinenin C, extracted from *sanguis draconis*, interacted well with GLP-1 receptor via hydrophobic interaction, which was confirmed by fluorescence spectroscopy and molecular simulation. In cell experiments, it was demonstrated that pancreatic beta cells promoted insulin secretion upon treatment with cochinchinenin C, and increases of intracellular cAMP and ATP levels also occurred, indicating GLP-1 receptor activation and glucose metabolism. These results showed that cochinchinenin C has potential for the development of drugs for treating diabetes.

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## 1 Introduction

Type 2 diabetes mellitus, the most common type of diabetes mellitus, is one of the most prevalent noncommunicable diseases globally. According to a report from the International Diabetes Federation, there are more than 415 million diabetes patients worldwide. It has also been estimated that there will be more than 642 million of such patients by 2040. Diabetes occurs in adults due to insulin resistance and inadequate insulin secretion, which result in elevated blood glucose levels.<sup>1,2</sup> Drugs for treating diabetes based on different targets have been developed for the treatment of type 2 diabetes mellitus. Currently, the most widely used therapeutic drugs include guanidine and sulfonylureas, but they also have disadvantages.<sup>3,4</sup> For example, guanidine causes the adverse effect of gastrointestinal discomfort, while sulfonylureas can cause hypoglycemia owing to a sustained effect of insulin secretion.<sup>5</sup> Hence, researchers are looking for safer, more curative diabetes drugs with fewer side effects. Glucagon-like peptide-1 receptor (GLP-1R) belongs to the glucagon receptor subfamily in the G-protein-coupled receptor B family, which is distributed in pancreatic cells, and the gastrointestinal, cardiovascular, pulmonary, and central nervous systems.<sup>6,7</sup> In early 1979, an incretin called GLP-1 was identified, which is a peptide secreted

by Langerhans cells derived from the end of the jejunum, ileum, and colon. GLP-1 do not promote formation of insulin, but promote excretion of the hormone.<sup>8</sup> One of the main disadvantages of GLP-1 analogs is that they do cause gastrointestinal discomfort like guanidine,<sup>9,10</sup> but GLP-1 analogs have the better therapeutic effect. As a glucose-dependent hormone, GLP-1 does not exhibit hypoglycemic effects when the body's blood sugar level is normal. A study found that GLP-1 activated intracellular cAMP in a glucose-dependent manner and then promoted the synthesis of PDX-1 transcription factors, finally promoting the secretion of insulin.<sup>11</sup> A series of GLP-1 analog agonists have been developed, such as exendin-4, eventide, and liraglutide.<sup>12,13</sup> However, these GLP-1 receptor agonists are easily degraded by DPP4 *in vivo* and lose their efficacy.<sup>14</sup> Therefore, the exploration of nonpeptide GLP-1 receptor agonists has gradually become a focus in the development of GLP-1 receptor agonists. Research on nonpeptide GLP-1 receptor agonists is currently underway. In support of this, Wang *et al.* established a high-throughput drug screening model for searching for potential nonpeptide GLP-1 receptor agonists.

*Sanguis draconis*, derived from *Daemonorops* spp., is mainly composed of flavonoids. Among them, more than 60% of active compounds belong to dihydrochalcone, including cochinchinenin C (Fig. 1)<sup>15</sup>. *Sanguis draconis* has numerous pharmacological functions, such as anti-blood-stasis, anticoagulation, and blood-sugar-lowering effects.<sup>16,17</sup> Increasing research is now focusing on how *sanguis draconis* can play a role in the

Shanghai Key Laboratory of Bio-Energy Crops, School of Life Sciences, Shanghai University, Shanghai, P. R. China. E-mail: chenqincc@staff.shu.edu.cn; Bingniu@shu.edu.cn



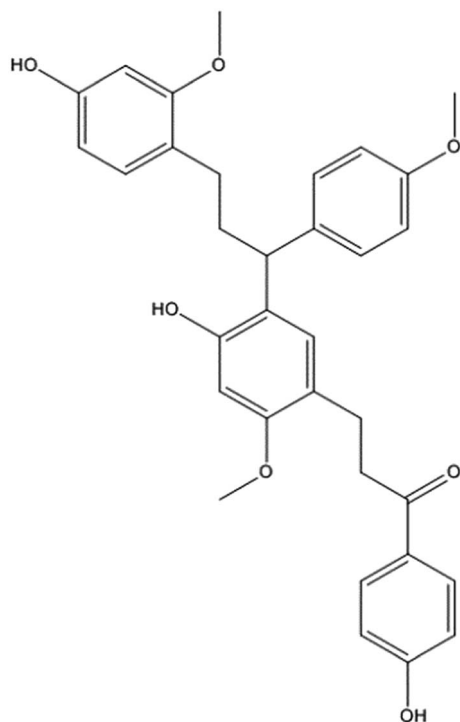


Fig. 1 The molecular structure of cochinchinenin C.

treatment of diabetes. Zhang *et al.* used a rat hyperglycemia model to reveal that *sanguis draconis* had a good hypoglycemic effect and could increase insulin secretion.<sup>18</sup> Research on the mechanism behind *sanguis draconis*'s hypoglycemic effect is currently ongoing.

As the structures of biological macromolecules are continually being determined and computer science continues to develop, computer-aided drug design (CADD) has become widely used in the research and development of leading compounds.<sup>19,20</sup> According to the structure of the drug target, the interaction between a small-molecule drug and the target protein can be studied by the molecular docking technique, and then the structure of the small molecule can be optimized. Hence, in this study, the interaction between cochinchinenin C and GLP-1 receptor was first determined through computer simulation. Then, this interaction was verified by fluorescence spectroscopy. The levels of insulin secretion, intracellular ATP, cAMP, and glucose in medium and mRNA gene insulin secretion were also determined after incubation of cochinchinenin C in a glucose-dependent manner. Finally, we proposed that cochinchinenin C is a potential nonpeptide GLP-1 receptor agonist.

## 2 Material and methods

### 2.1 Materials

The recombinant protein GLP1 receptor was purchased from Creative BioMart and used without further purification, configured as 25 mM. Cochinchinenin C was purchased from National Institutes for Food and Drug Control, and the stock

solution of Cochinchinenin C (CC) was prepared in DMSO at 0.01 mol L<sup>-1</sup>. Here, 50 mM Tris-HCl buffer was used to adjust the pH to 7.4. KRB buffer was composed of 129 mM NaCl, 4.8 mM KCl, 1.2 mM MgSO<sub>4</sub>, 1.2 mM KH<sub>2</sub>PO<sub>4</sub>, 2.5 mM CaCl<sub>2</sub>, 5 mM NaHCO<sub>3</sub>, 10 mM HEPES, and 0.1% BSA, pH 7.4.

### 2.2 Molecular docking

The structure of GLP1 receptor (4ZGM) was obtained from the PDB database (<http://www.rcsb.org/pdb>). The molecular structure of cochinchinenin C was obtained from the PubChem database (<https://pubchem.ncbi.nlm.nih.gov>). Discovery Studio 4.1 software was used to simulate the interaction between cochinchinenin C and GLP1 receptor.

### 2.3 Three-dimensional and two-dimensional fluorescence spectrometry

Three-dimensional fluorescence spectra were obtained at a constant GLP1 receptor concentration (0.025 μmol L<sup>-1</sup>) in the absence or presence of cochinchinenin C (0.05 μmol L<sup>-1</sup>). Two-dimensional fluorescence spectra were also obtained at a constant GLP-1 receptor concentration (0.025 μmol L<sup>-1</sup>) in the absence or presence of cochinchinenin C (0–0.045 μmol L<sup>-1</sup>). Emission spectra were recorded from 290 to 500 nm when excitation was applied at 280 nm.  $F_0$  and  $F$  represent the fluorescence intensity of GLP-1 receptor in the absence and presence of cochinchinenin C at an emission wavelength of 340 nm, respectively. Other experimental parameters were as follows: slit width, 5 nm; and voltage, 950 V.

### 2.4 The quenching mechanism

**Stern-Volmer quenching constant ( $K_{sv}$ ) and bimolecular quenching rate constant ( $K_q$ ).** Fluorescence spectrometry was performed at different temperatures (300, 305, and 310 K) to obtain the fluorescence spectra. Here, the Stern-Volmer equation (eqn (1)) was used,

$$F_0/F = 1 + K_{sv}[C] = 1 + K_q\tau_0[C], \quad (1)$$

where the slope of the equation represents the Stern-Volmer quenching constant ( $K_{sv}$ ). Eqn (2) was also used to calculate  $K_q$ :

$$K_{sv} = K_q\tau_0 \quad (2)$$

( $\tau_0$  is the average lifetime of the protein without quencher,  $\tau_0 = 10^{-8}$  s).

**Binding constant ( $K_a$ ) and the number of binding sites ( $n$ ).** Eqn (3) was used:

$$\lg[(F_0 - F)/F] = \lg K_a + n \lg[C], \quad (3)$$

plotted to  $\lg[C]$  with  $\lg[(F_0 - F)/F]$ , and a linear equation was obtained. The slope of the equation is the number of binding sites ( $n$ ) and the intercept of the equation is  $\lg K_a$ , from which  $K_a$  can be obtained.

**Thermodynamic parameters.** Eqn (4) was used:

$$\ln K_a = -\Delta H/RT + \Delta S/R, \quad (4)$$



( $T$ , absolute temperature;  $R$ , gas constant =  $8.3145 \text{ J mol}^{-1} \text{ K}^{-1}$ ). Plotted to  $1/T$  with  $\ln K_a$  and obtained the linear equation. The slope of the equation is  $-\Delta H/R$ , and the intercept of the equation is  $\Delta S/R$ . The enthalpy change ( $\Delta H$ ) and entropy change ( $\Delta S$ ) were calculated.  $\Delta G$  can be calculated from the following equation:  $\Delta G = \Delta H - T\Delta S$ .

## 2.5 Ins-1 cell culture

Ins-1 cells, a rat pancreatic  $\beta$ -cell line purchased from ATCC (US state), were cultured in DMEM high-glucose medium with 10% fetal bovine serum (FBS),  $100 \text{ U mL}^{-1}$  penicillin,  $100 \mu\text{g mL}^{-1}$  streptomycin,  $10 \text{ mmol L}^{-1}$  HEPES,  $2 \text{ mmol L}^{-1}$  L-glutamine,  $1 \text{ mmol L}^{-1}$  sodium pyruvate, and  $50 \mu\text{mol L}^{-1}$   $\beta$ -mercaptoethanol at  $37^\circ\text{C}$  in a humidified atmosphere and 5%  $\text{CO}_2$ . The culture medium was replaced every 2 days and passaged every four days following trypsinization. The subculture cells were seeded at a density of  $5.0 \times 10^5$  cells per mL.

## 2.6 Insulin secretion

To measure insulin secretion, Ins-1 cells were first seeded onto 24-well plates and cultured for 48 h to approximately 80–90% confluence. The cells were then washed twice with KRB buffer and starved for 2 h in KRB buffer. Next, cells were incubated in fresh KRB buffer containing  $10^{-8} \text{ mol L}^{-1}$  cochininenin C at different glucose concentrations (0, 5, 10, 20, and 30 mM). Cellular supernatant was sampled for insulin secretion. Insulin was measured using Iodine [ $^{125}\text{I}$ ]-Insulin Radioimmunoassay Kit (Shanghai Xinfan Biotechnology Co., Ltd.).

## 2.7 Cell viability

To determine cell viability, Ins-1 cells were seeded onto 96-well plates, where they were incubated in culture medium containing  $10^{-8} \text{ mol L}^{-1}$  cochininenin C for different incubation times (1, 2, 4, and 8 h). Cell viability was measured using Cell Counting Kit-8 (CCK-8) (Sangon Biotech (Shanghai) Co., Ltd.).

## 2.8 Intracellular cAMP and ATP levels

To measure the intracellular levels of cAMP and ATP, Ins-1 cells were seeded onto 96-well plates, where they were washed twice with KRB buffer and starved for 2 h in KRB buffer. Then, the cells were incubated in fresh KRB buffer containing  $10^{-8} \text{ mol L}^{-1}$  cochininenin C at different glucose concentrations (0, 5, and 20 mM) for 2 h. Intracellular cAMP was measured using the cAMP-Glo™ Assay Kit and the ATP level was determined using the CellTiter-Glo® Luminescent Kit (Promega Corporation).

## 2.9 Glucose absorption

To assay glucose absorption, Ins-1 cells were seeded onto 24-well plates, where they were incubated in fresh KRB buffer containing  $10^{-8} \text{ mol L}^{-1}$  cochininenin C at different glucose concentrations (0, 5, and 20 mM) for 4 and 8 h. Cellular supernatant was sampled to detect the change of glucose content in the medium. Cellular supernatant were measured using Glucose Assay Kit (Sigma-Aldrich).

## 2.10 Gene mRNA expression

The relative expression of genes associated with insulin secretion was determined by quantitative RT-PCR using the Bio-Rad CFX96 PCR System. All data are expressed as the ratio of the target gene to  $\beta$ -actin expression.

The primer sequences used for PCR were as follows: pdx-1, 5'-aaatccaccaaagctcagc-3' (forward) and 5'-aagttgagcatcactgcagc-3' (reverse); ins, 5'-gaccgcaagtgccaca-3' (forward) and 5'-tccacaagccagctcttg-3' (reverse); and  $\beta$ -actin, 5'-cccatctatgagggt-tacgc-3' (forward) and 5'-ttaaagtgcagcagatttc-3' (reverse).

## 2.11 Statistical analysis

All statistical analyses were conducted using the software IBM SPSS Statistical 20. Data were analyzed using one-way ANOVA. The threshold for statistical significance was set at  $P < 0.05$ . Results are presented as mean  $\pm$  SD.

# 3 Results and discussion

## 3.1 The interaction between cochininenin C and GLP1 receptor

In our study, the interaction between cochininenin C and GLP-1 receptor was first determined based on molecular docking. As seen in Fig. 2A, cochininenin C could bind to the extracellular domain of the GLP1 receptor. The site of action of cochininenin C–GLP-1 receptor was further analyzed. Cochininenin C mainly bound to amino acid residues Ser31, Leu32, Thr35, Val36, Tyr88 and Trp39 of the GLP-1 receptor through hydrophobic interactions (Fig. 2B and C). Hydrogen bonds also formed between cochininenin C and GLP-1 receptor (Table 1). As one of the small-molecule GLP-1 agonists, Boc5 was discovered. It was reported that Boc5 has the ability to increase the intracellular level of cAMP.<sup>21</sup> Hence, the interaction between Boc5 and GLP-1 receptor was measured to compare the result for cochininenin C–GLP-1 receptor (Fig. 2D–F). Surprisingly, it could be found that amino acid residues Trp39, Trp88 and Leu32 were also the key residues between Boc5 and GLP-1 receptor. What's more, hydrophobic interactions and hydrogen bonds were also formed between Boc5 and GLP-1 receptor (Table 2). Therefore, it could be seen that cochininenin C might be the potential agonist of GLP-1 receptor.

Based on the results of molecular docking analysis, further study of the interaction between cochininenin C and GLP-1 receptor was performed by three-dimensional fluorescence spectroscopy. As shown in Fig. 3A, there were two characteristic fluorescence peaks upon excitation at 230 nm and 280 nm. The luminescence at the excitation at 230 nm resulted from the structure of the protein polypeptide chain,<sup>22</sup> while that at the excitation at 280 nm resulted from spontaneous fluorescence amino acid residues, tyrosine, tryptophan, and phenylalanine.<sup>23</sup> However, cochininenin C has very weak fluorescence in solution. Hence, the fluorescence of GLP-1 receptor can be used as a probe to determine the interaction between cochininenin C and GLP-1 receptor based on the change of fluorescence intensity. The fluorescence peak at 347 nm upon excitation at



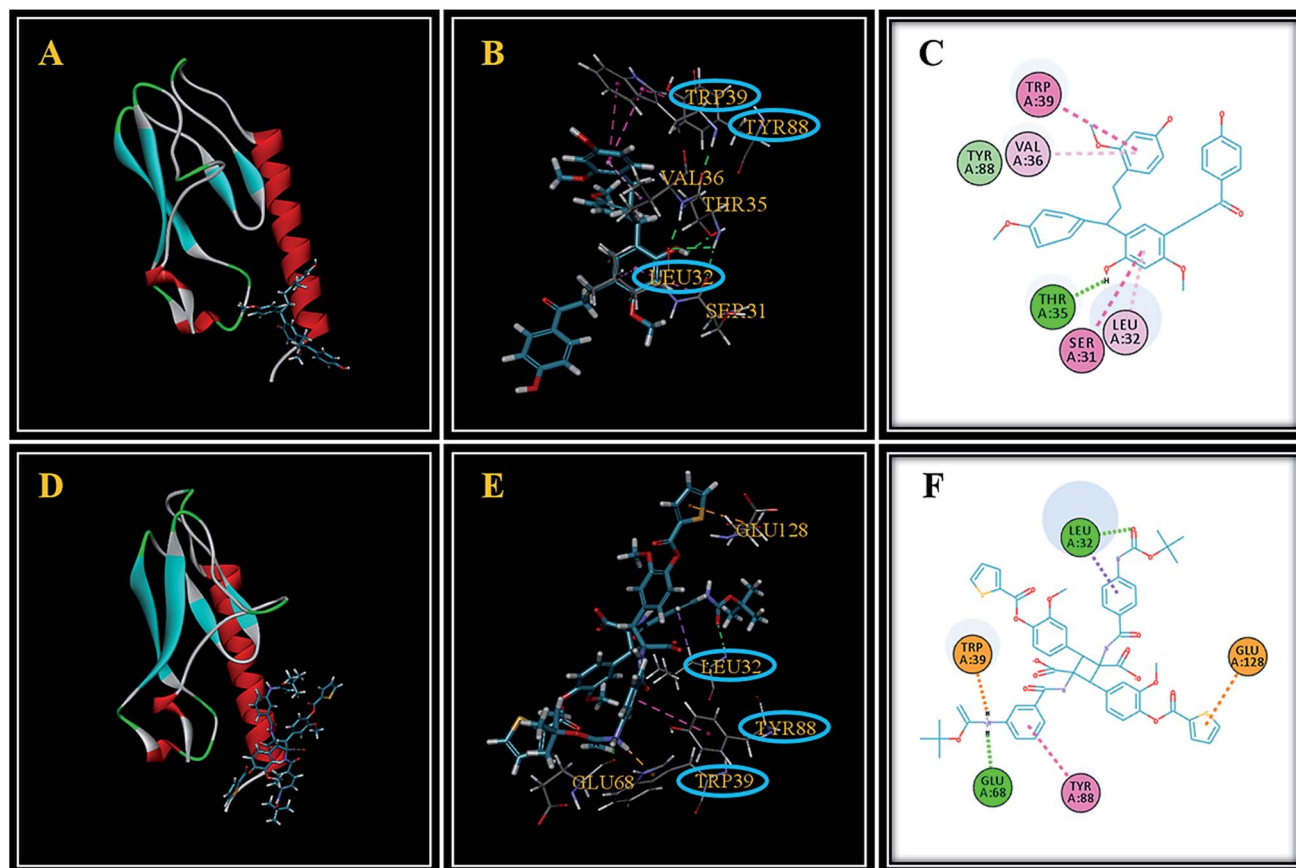


Fig. 2 Interaction between drugs and GLP-1 receptor through molecular docking. (A–C) Cochininenin C–GLP-1 receptor (D–F) Boc5–GLP-1 receptor.

Table 1 Parameters of the interaction between cochininenin C and GLP-1 receptor

Acting force	Bond length (Å)	From	To
4.74948	Hydrophobic	GLP1R:TRP39	CC
5.02218	Hydrophobic	GLP1R:TRP39	CC
4.94458	Hydrophobic	GLP1R:SER31:C,O; LEU32:N	CC
4.34227	Hydrophobic	CC	GLP1R:LEU32
4.84843	Hydrophobic	CC	GLP1R:VAL36
2.19292	Hydrogen bond	CC	GLP1R:THR35:OG1

Table 2 Parameters of the interaction between BOC5 and GLP-1 receptor

Acting force	Bond length (Å)	From	To
2.30382	Hydrogen bond	GLP1R:LEU32:HN	BOC5
1.96945	Hydrogen bond	BOC5	GLP1R:GLU68:O
2.38363	Hydrogen bond	BOC5	GLP1R:TRP39
4.80812	Electrostatic	GLP1R:GLU128:OE2	BOC5
2.85583	Hydrophobic	GLP1R:LEU32:HB2	BOC5
5.68481	Hydrophobic	GLP1R:TYR88	BOC5

280 nm was selected. The fluorescence peak position remained unchanged after adding cochininenin C, but the fluorescence intensity decreased by about 53.9% (Fig. 3B). This showed that

cochininenin C affected the fluorescence intensity of the GLP-1 receptor. Therefore, it was confirmed that cochininenin C did indeed interact with GLP-1 receptor. The interaction between cochininenin C and GLP-1 receptor was also studied by two-dimensional fluorescence spectroscopy. The fluorescence of GLP-1 receptor decreased upon the addition of cochininenin C, which showed that there was an interaction between cochininenin C and GLP-1 receptor (Fig. 3C).

The overlap integral of the fluorescence emission spectra of GLP-1 receptor and the UV-vis spectra of cochininenin C (Fig. 3D) was calculated to obtain the critical energy transfer distance  $R_0$ . The following formula was used to determine the efficiency of nonradiative energy transfer  $E$  between GLP-1 receptor and cochininenin C:  $E = 1 - F/F_0 = (R_0)^6 / ((R_0)^6 +$



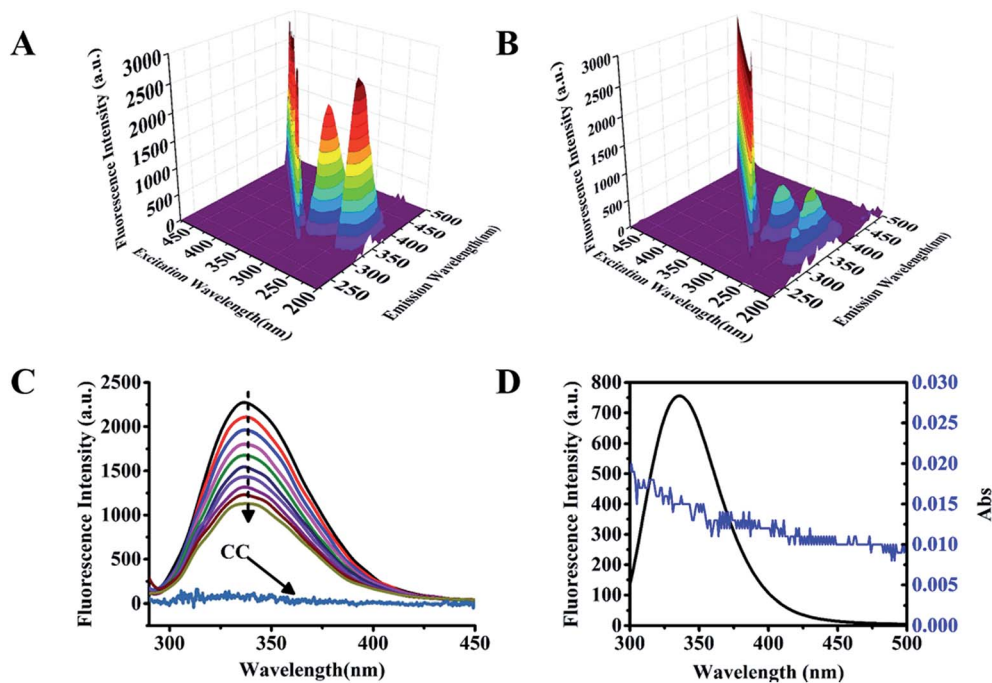


Fig. 3 Interaction between cochininenin C and GLP-1 receptor through spectral method. (A and B) GLP-1 receptor's three-dimensional fluorescence spectroscopy in the presence or absence of cochininenin C. (C) Two-dimensional fluorescence spectroscopy, keeping GLP-1 receptor' concentration constant ( $0.025 \text{ mol L}^{-1}$ ) while varying the cochininenin C concentration from  $0 \text{ mol L}^{-1}$  to  $0.045 \text{ mol L}^{-1}$ ; fluorescence intensity of cochininenin C ( $0.025 \text{ mol L}^{-1}$ ) alone, line CC. (D) Overlap of the fluorescence spectra of GLP-1 receptor ( $0.025 \text{ mol L}^{-1}$ ) and the absorption spectra of cochininenin C ( $0.05 \text{ mol L}^{-1}$ ).

( $r$ )<sup>6</sup>, where  $F$  and  $F_0$  are the fluorescence GLP-1 receptor intensity in the presence and absence of cochininenin C. The distance  $r$  between GLP-1 receptor and cochininenin C could be calculated according to Forster's theory.<sup>24</sup> These parameters could be obtained:  $E = 0.39\%$ ,  $r = 2.88 \text{ nm}$ . The donor-acceptor distance between cochininenin C and the tryptophan residue distance  $R_0 = 2.79$ , which was less than  $7 \text{ nm}$ , and  $0.5R_0 < r < 1.5R_0$ , indicating that nonradiative energy transfer occurred between cochininenin C and GLP-1 receptor.

### 3.2 Quenching mechanism of the interaction between cochininenin C and GLP-1 receptor

Two-dimensional fluorescence spectroscopy was performed at different temperatures (300, 305, and 310 K) while varying the

concentration of cochininenin C from  $0$  to  $0.045 \mu\text{mol L}^{-1}$  (Fig. 4). As shown in Table 3, the  $K_{sv}$  value of cochininenin C-GLP-1 receptor gradually increased with increasing temperature. This suggested that the fluorescence quenching between cochininenin C and GLP-1 receptor might be dynamic because molecules moved faster in the solution with increasing temperature, which increased the probability of a collision between the excited-state fluorescence molecule and the quenching agent. In addition, the binding constant  $K_a$  and the number of binding sites  $n$  between cochininenin C and GLP-1 receptor were calculated. The results revealed that the binding constant between cochininenin C and GLP-1 receptor increased with increasing temperature and the binding constant is approximately 1.

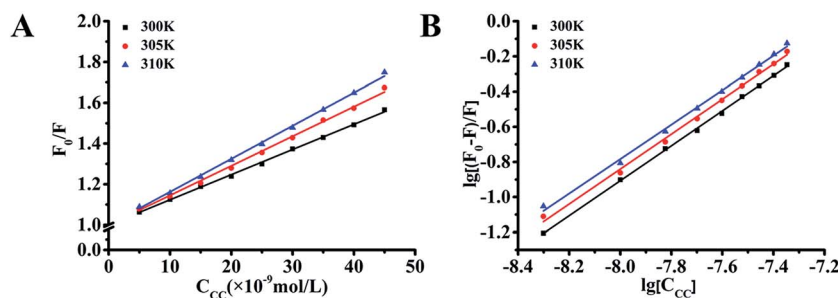


Fig. 4 Thermodynamic parameters of the interaction between cochininenin C and GLP-1 receptor. (A) Stern-Volmer curves of GLP-1 receptor fluorescence quenched cochininenin C. (B) Double-log plots of cochininenin C quenching effect on GLP-1 receptor's fluorescence.



**Table 3** Quenching constant  $K_{sv}$ , quenching rate constant  $K_q$ , binding constant  $K_a$ , and binding site  $n$  of cochininenin C with GLP1 receptor

Compound	$T$ (K)	$K_{sv}$ ( $\times 10^6$ L mol $^{-1}$ )	$K_q$ ( $\times 10^{14}$ L mol $^{-1}$ )	$K_a$ ( $\times 10^6$ L mol $^{-1}$ )	$n$
Cochininenin C	300	1.24	1.24	7.05	0.99
	305	1.45	1.45	7.12	0.99
	310	1.62	1.62	7.16	0.98

**Table 4** Thermodynamic parameters of cochininenin C–GLP1 receptor interaction

Compound	$T$ (K)	$\Delta H$ (K J $^{-1}$ mol)	$\Delta G$ (K J $^{-1}$ mol)	$\Delta S$ (J mol $^{-1}$ K $^{-1}$ )
Cochininenin C	300		−40.52	
	305	18.90	−41.51	198.05
	310		−42.50	

The thermodynamic parameters of the interaction between cochininenin C and GLP-1 receptor could be calculated using the following equation:  $\ln K_a = -\Delta H/RT + \Delta S/R$ .  $\Delta G$  could also be calculated using the following equation:  $\Delta G = \Delta H - T\Delta S$ . As shown in Table 4, enthalpy change ( $\Delta H$ ) and entropy change in the process of binding between cochininenin C and GLP-1 receptor were obtained, namely,  $\Delta H > 0$ ,  $\Delta S > 0$ . Based on the thermodynamics regular of the acting force between drug and biomacromolecule,<sup>25</sup> it was indicated that hydrophobic force was the main factor in the binding between cochininenin C and GLP-1 receptor, which is consistent with the results from molecular simulation. Then, the Gibbs free energy of cochininenin C–GLP-1 receptor was calculated to be less than 0, which proved that the interaction was spontaneous.

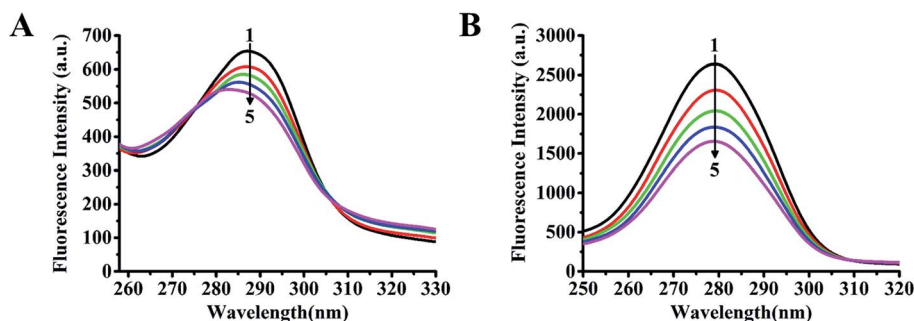
### 3.3 Effect of cochininenin C on the conformation of GLP-1 receptor

Synchronous fluorescence is usually used to determine the effects of drugs on the conformation of a target protein.<sup>26</sup> The synchronous fluorescence spectra of a protein upon addition of a compound at  $\Delta\lambda = 15$  and 60 nm show the spectral characteristics of tyrosine and tryptophan residues, respectively. In this experiment, the synchronous fluorescence spectra of GLP-1 receptor upon addition of cochininenin C at  $\Delta\lambda = 15$  and

60 nm were obtained at a constant GLP-1 receptor concentration (0.025 mol L $^{-1}$ ), while varying the concentration of cochininenin C from 0 to 0.045  $\mu\text{mol L}^{-1}$ , as shown in Fig. 5A and B. In synchronous fluorescence spectra of GLP-1 receptor upon the addition of cochininenin C at  $\Delta\lambda = 15$ , the maximum excitation wavelength showed a blue shift accompanied by fluorescence quenching. When  $\Delta\lambda = 60$  nm, the maximum excitation wavelength hardly changed and the fluorescence intensity was still quenched. In summary, the degree of fluorescence quenching at  $\Delta\lambda = 60$  nm was higher than that at  $\Delta\lambda = 15$  nm, and it was also inferred that the fluorescence quenching of tryptophan residues was much greater than that of tyrosine residues. It was revealed that the fluorescence quenching between cochininenin C and GLP-1 receptor was mainly caused by the effect of cochininenin C on tryptophan residues. When  $\Delta\lambda = 15$  nm, the maximum excitation wavelength showed a red shift because the microenvironment of tyrosine residues in the GLP-1 receptor was affected by the addition of cochininenin C. Therefore, the results showed that cochininenin C affected the conformation of the GLP-1 receptor.

### 3.4 Effect of cochininenin C on insulin secretion and cell activity of Ins-1 cells

In this experiment, the effect of cochininenin C on insulin secretion was first measured at different glucose concentrations. Glucose-stimulated insulin secretion (GSIS) is an important element on the function of  $\beta$ -cell and for the cure of type 2 diabetes mellitus. As shown in Fig. 6A, cochininenin C significantly promoted insulin secretion in medium with 10 and 20 mM glucose. When the glucose concentration was 10 mM in medium, the promotion of insulin secretion in the cochininenin C-treatment group reached the highest value ( $3.79 \pm 0.18$  IU mL $^{-1}$ ), which was about 1.35-fold higher than that in the

**Fig. 5** Synchronous fluorescence of the interaction between cochininenin C and GLP-1 receptor. (A)  $\Delta\lambda = 15$  nm; (B)  $\Delta\lambda = 60$  nm.

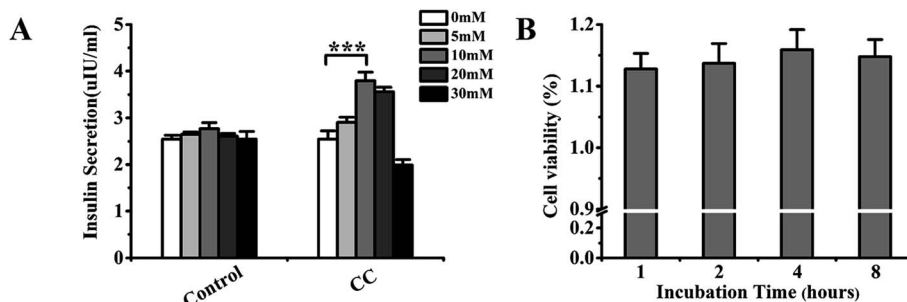


Fig. 6 Effect of cochinchinenin C on insulin secretion and cell viability of Ins-1 cells. (A) Effect of cochinchinenin C on insulin secretion at the condition of different glucose concentration. (B) The cell viability of Ins-1 cells with the incubation of cochinchinenin C compared with the control group. ( $n = 5$ ,  $***P < 0.005$ ).

control group. In contrast, when the glucose concentration was 30 mM in medium, insulin secretion was inhibited in the cochinchinenin C-treatment group. It could be speculated that the function of pancreatic beta cell was affected in such a high glucose concentration. Liu *et al.* found that geniposide, a potential anti-diabetic drug targeting for GLP-1 receptor, also inhibited insulin secretion at the condition of 33 mM glucose.<sup>27</sup> Therefore, it could be obtained that cochinchinenin C enhanced the promotion of insulin secretion in the presence of low or moderately high glucose concentration.

Cell activity tests are usually applied to estimate cytotoxicity upon treatment with drugs. Cell Counting Kit-8 (CCK-8) enables the sensitive determination of cell viability *via* cell proliferation and cytotoxicity assays based on the change of mitochondrial dehydrogenase to formazan. Specifically, cell proliferation and cytotoxicity could be determined through the detection of OD at 450 nm. In this experiment, cell viability was measured upon incubation for 1, 2, 4, and 8 h after adding  $10^{-8}$  M

cochinchinenin C to the culture medium (Fig. 6B). The results showed that the survival rate of Ins-1 cells increased with increasing incubation time. Therefore, it was revealed that cochinchinenin C exhibited almost no toxicity to cells and could be used in further experiments.

### 3.5 Mechanism of the effect of cochinchinenin C on insulin secretion in Ins-1 cells

Intracellular cAMP levels are commonly measured to determine whether extracellular signaling molecules are associated with cell surface receptors.<sup>28,29</sup> Patients are judged to be suffering from diabetes mellitus if their blood glucose concentration is greater than or equal to  $7 \text{ mmol L}^{-1}$ . Hence, 5 and 20 mM glucose levels were used in this experiment. The intracellular cAMP content was determined after incubation of cochinchinenin C for 2 h, so as to estimate whether cochinchinenin C activated the GLP-1 receptor on the cell surface. As shown in Fig. 7A, the content of cAMP increased with increasing glucose

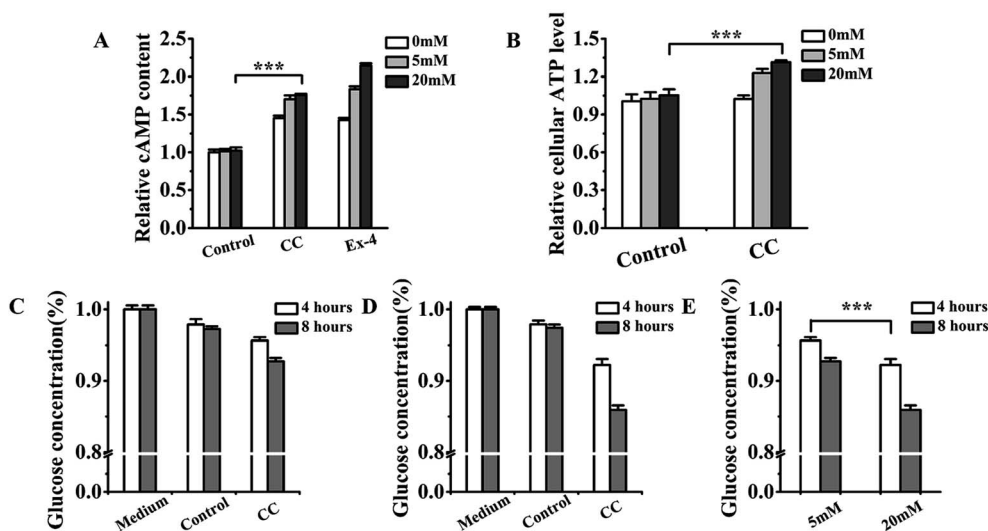


Fig. 7 Effect of cochinchinenin C on intracellular cAMP content, ATP level, and glucose absorption. (A) The change of intracellular cAMP content with the treatment of cochinchinenin C. (B) The change of intracellular ATP level with the treatment of cochinchinenin C. (C–E) The decrease of glucose concentration in medium was detected with the effect of cochinchinenin C. (C) 5 mM glucose-treated group; (D) 20 mM glucose-treated group; (E) comparison of capacity about glucose absorption in 5 mM glucose-treated group with 20 mM glucose-treated group. ( $n = 5$ ,  $***P < 0.005$ ).



concentration upon cochininenin C treatment, but lower than the Ex-4 group which is a GLP-1 receptor agonist. Therefore, it could be determined that the level of cAMP increased owing to the combination with GLP-1 receptor, then activated adenylate cyclase and finally caused the increase of intracellular cAMP.

In pancreatic beta cells, the level of intracellular ATP plays a critical role in insulin secretion.<sup>30</sup> The increase in ATP/ADP ratio leads to the closure of ATP-sensitive K channels ( $K_{ATP}$ ), causes the influx of calcium ions, and ultimately produces insulin secretion. Fig. 7B shows that the intracellular ATP level increased with increasing glucose concentration in the culture medium upon treatment with cochininenin C. Comparing the condition of 5 mM glucose with 20 mM glucose, the intracellular ATP level clearly increased with 20 mM glucose. The promotion of insulin secretion was related to the increase of intracellular ATP level. Furthermore, glucose content in cellular supernatant was measured before and after the treatment with cochininenin C.

As shown in Fig. 7C–E, upon treatment with cochininenin C for 4 and 8 h, glucose in cellular supernatant decreased at the conditions of 5 and 20 mM, respectively, which suggested that cochininenin C has the ability to promote glucose absorption. The glucose absorption by Ins-1 cells upon treatment with cochininenin C was also compared between the 5 and 20 mM conditions. Cochininenin C clearly promoted glucose absorption in the 20 mM condition upon incubation for 4 h. Hence, it could be concluded that elevated intracellular ATP levels were dependent on the constant metabolism of glucose in the medium upon treatment with cochininenin C. Because of the activation of GLP-1 receptor, some of the ATP generated by glucose metabolism changed into cAMP.

### 3.6 Effect of cochininenin C on the gene expression related to insulin secretion

In this experiment, the gene mRNA expression of Ins-1 cells was determined upon incubation with cochininenin C. Pdx-1, a transcription factor encoded by the *pdx-1* gene, plays an important role in the development and differentiation of pancreatic cells.<sup>31,32</sup> It can promote the proliferation of pancreatic beta cells. GLP-1 has been reported to promote Pdx-1 expression in a glucose-dependent manner. Upon treatment with cochininenin C, gene mRNA expression related to insulin secretion was measured. As shown in Fig. 8, the gene

mRNA expression of *pdx-1* and *ins* increased upon treatment with cochininenin C. In addition, the expression of *pdx-1* and *ins* was clearly enhanced by the treatment with cochininenin C compared with that in the control group under the condition of a glucose concentration of 20 mM. Therefore, it could be speculated that cochininenin C affected the expression of genes related to insulin secretion, thereby promoting such secretion.

As a class of B G protein-coupled receptor, GLP-1 receptor has an ability to mediate the action of GLP-1, which is useful in diabetes treatment. Cochininenin C, extracted from *sanguis draconis*, might be the potential GLP-1 receptor agonist through molecular docking, spectral experiment and cell experiment. Currently, the development of computer-aided drug design provides us with more convenience to estimate the interaction between drugs with target protein. Hence, the exploration of new drug discovery has become more rapidly than before. Fluorescence spectroscopy, a general method, was commonly applied to estimate the interaction between drug and biomolecule based on the changing of fluorescence. The results from spectrum could verify the interaction between cochininenin C and GLP-1 receptor obtained from computational simulation. Besides spectrum and molecular docking, the efficacy and mechanism of cochininenin C were also determined. Generally, the treatment of diabetes was dependent on how to control blood sugar. Glucose-stimulated insulin secretion is an important element on the function of  $\beta$ -cell. It is exciting to find that cochininenin C have the ability to promote insulin secretion with the increasing of blood sugar and also stop the efficacy at the normal blood sugar. What's more, the increasing of cAMP infers the activation of GLP-1 receptor with the incubation of cochininenin C. There were also obtained the results of the increase of ATP level, the absorption of glucose and the enhancement of gene about insulin secretion with the effect of cochininenin C. Since cochininenin C has the well therapeutic effect, new diabetes drug based on the structure of cochininenin C will be developed. The optimization of molecular structure, improvement in water solubility, enhancement of efficacy and reduction of side effects will be the focus for us to expand further researches in the future.

## 4 Conclusions

In this study, it could be obtained that cochininenin C interacted well with GLP-1 receptor agonist based on fluorescence spectrometry and molecular docking. In addition, cell experiments showed that cochininenin C activated the GLP-1 receptor and then enhanced intracellular cAMP level. Hence, we proposed that cochininenin C might be a potential nonpeptide GLP-1 receptor agonist.

Most of anti-diabetic drugs targeting for GLP-1 receptor were peptide agonists, such as exendin-4, eventide, and liraglutide. Although they have better therapeutic effect, the disadvantages of peptide agonists were that they could be easily degraded by DPP4 and needed to be injected intravenously or subcutaneously which might cause pain for patients. Therefore, the exploration of nonpeptide GLP-1 receptor agonist become

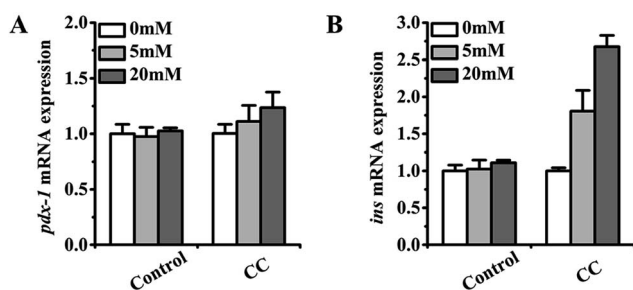


Fig. 8 Effect of cochininenin C on gene mRNA expression.



popular in research field for anti-diabetic drug. The aim of our study is to determine whether there is an interaction between cochininenin C and GLP-1 receptor *in vitro*. Actually, further experimental works including the interaction between cochininenin C and GLP-1 receptor *in vivo* should be designed. At the moment, the high-throughput screening model for GLP-1 receptor agonist is being established. With deep exploration, it is a hope that nonpeptide GLP-1 receptor agonist will be a more superior anti-diabetic drug choice and bring patients better therapeutic effect.

## Conflicts of interest

There are no conflicts to declare.

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## References

- 1 A. D. Assoc, *Diabetes care*, 2011, **34**, S62–S69.
- 2 Z. Fu, E. R. Gilbert and D. Liu, *Curr. Diabetes Rev.*, 2013, **9**, 25–53.
- 3 E. Renstrom, S. Barg, F. Thevenod and P. Rorsman, *Diabetes*, 2002, **51**, S33–S36.
- 4 M. B. Shaghafi, D. G. Barrett, F. S. Willard and L. E. Overman, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 1031–1036.
- 5 J. E. Schopman, A. C. R. Simon, S. J. M. Hoefnagel, J. B. L. Hoekstra, R. J. P. M. Scholten and F. Holleman, *Diabetes/Metab. Res. Rev.*, 2014, **30**, 11–22.
- 6 D. Tornehave, P. Kristensen, J. Romer, L. B. Knudsen and R. S. Heller, *J. Histochem. Cytochem.*, 2008, **56**, 841–851.
- 7 E. C. Tibaduiza, C. Chen and M. Beinborn, *J. Biol. Chem.*, 2001, **276**, 37787–37793.
- 8 J. M. Egan, C. Montrose-Rafizadeh, Y. Wang, M. Bernier and J. Roth, *Endocrinology*, 1994, **135**, 2070–2075.
- 9 A. A. Tahrani, A. H. Barnett and C. J. Bailey, *Nat. Rev. Endocrinol.*, 2016, **12**, 566–592.
- 10 A. Uccellatore, S. Genovese, I. Dicembrini, E. Mannucci and A. Ceriello, *Diabetes Ther.*, 2015, **6**, 239–256.
- 11 A. R. Meloni, M. B. DeYoung, C. Lowe and D. G. Parkes, *Diabetes, Obes. Metab.*, 2013, **15**, 15–27.
- 12 C. Alarcon, B. Wicksteed and C. J. Rhodes, *Diabetologia*, 2006, **49**, 2920–2929.
- 13 J. Nagakura, T. Yamakawa, M. Taguri, H. Tsuchiya, E. Shigematsu, J. Suzuki, S. Morita, K. Kadonosono and Y. Terauchi, *Endocr. J.*, 2016, **63**, 239–247.
- 14 C. F. Deacon, M. A. Nauck, M. Toft-Nielsen, L. Pridal, B. Willms and J. J. Holst, *Diabetes*, 1995, **44**, 1126–1131.
- 15 D. Gupta, B. Bleakley and R. K. Gupta, *J. Ethnopharmacol.*, 2008, **115**, 361–380.
- 16 C. M. Hu, J. S. Li, K. P. Cheah, C. W. Lin, W. Y. Yu, M. L. Chang, G. C. Yeh, S. H. Chen, H. W. Cheng and C. S. Choy, *Diabetes Res. Clin. Pract.*, 2011, **94**, 417–425.
- 17 J. Y. Fan, T. Yi, C. M. Sze-To, L. Zhu, W. L. Peng, Y. Z. Zhang, Z. Z. Zhao and H. B. Chen, *Molecules*, 2014, **19**, 10650–10669.
- 18 R. Zhang, *Tradit. Chin. Drug Res. Clin. Pharmacol.*, 2002, **13**, 3.
- 19 A. Tropsha and J. Bajorath, *J. Med. Chem.*, 2016, **59**, 1.
- 20 C. H. Reynolds, *Curr. Pharm. Des.*, 2014, **20**, 3380–3386.
- 21 F. Lin and R. X. Wang, *J. Mol. Model.*, 2009, **15**, 53–65.
- 22 B. Niu, M. Zhang, P. Du, L. Jiang, R. Qin, Q. Su, F. Chen, D. Du, Y. Shu and K. C. Chou, *Oncotargets Ther.*, 2017, **8**, 57149–57162.
- 23 X. Chen, K. Qian and Q. Chen, *Eur. J. Med. Chem.*, 2015, **93**, 492–500.
- 24 T. Forster and O. Sinanoglu, *Modern Quantum Chemistry*, 1996, pp. 93–138.
- 25 P. D. Ross and S. Subramanian, *Biochemistry*, 1981, **20**, 3096–3102.
- 26 L. Zhang, B. Liu, Z. Li and Y. Guo, *Luminescence*, 2015, **30**, 686–692.
- 27 J. H. Liu, L. X. Guo, F. Yin, Y. L. Zhang, Z. X. Liu and Y. W. Wang, *PLoS One*, 2013, **8**(10), e78315.
- 28 F. Schwede, O. G. Chepurny, M. Kaufholz, D. Bertinetti, C. A. Leech, O. Cabrera, Y. M. Zhu, F. Mei, X. D. Cheng, J. E. M. Fox, P. E. MacDonald, H. G. Genieser, F. W. Herberg and G. G. Holz, *Mol. Endocrinol.*, 2015, **29**, 988–1005.
- 29 E. P. Smith, Z. B. An, C. Wagner, A. G. Lewis, E. B. Cohen, B. L. Li, P. Mahbod, D. Sandoval, D. Perez-Tilve, N. Tamarina, L. H. Philipson, D. A. Stoffers, R. J. Seeley and D. A. D'Alessio, *Cell Metab.*, 2014, **19**, 1050–1057.
- 30 G. W. Cline, R. L. Pongratz, X. J. Zhao and K. K. Pappas, *Biochem. Biophys. Res. Commun.*, 2011, **415**, 30–35.
- 31 C. M. McKinnon and K. Docherty, *Diabetologia*, 2001, **44**, 1203–1214.
- 32 X. L. Wang, C. M. Cahill, M. A. Pineyro, J. Zhou, M. E. Doyle and J. M. Egan, *Endocrinology*, 1999, **140**, 4904–4907.

