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Protein-free ribosomal RNA folds to a near-native state in the presence of Mg^{2+} †

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The assembled bacterial ribosome contains around 50 proteins and many counterions. Here, focusing on rRNA from the large ribosomal subunit, we demonstrate that Mg^{2+} causes structural collapse in the absence of ribosomal proteins. The collapsed rRNA forms many native-like RNA–RNA interactions, similar to those observed in the assembled ribosome. We assayed rRNA structure by chemical footprinting in the presence and absence of Mg^{2+} . Our results indicate that Mg^{2+} -dependent conformational change is focused in non-helical regions, consistent with tertiary interactions. In the presence of Mg^{2+} , the large subunit rRNA adopts a state that includes the core inter-domain architecture of the assembled ribosome. We infer that the rRNA– Mg^{2+} state represents the core architecture of the LSU which, while not catalytically active, positions the residues of the LSU rRNA in such a way as to promote native interactions with rProteins to ultimately form a functional LSU.

Introduction

Cations accumulate around RNAs and facilitate folding.^{1,2} Monovalent cations promote formation of RNA secondary structure (2° structure). Mg^{2+} is often necessary for adoption of native states, characterized by long-range tertiary interactions. Mg^{2+} can increase local RNA density and rigidity by packing RNA functional groups in its first coordination shell.

Ribosomal structure is maintained by complex networks of rRNA–rRNA, rRNA–protein and rRNA–cation interactions.^{3,4} Mg^{2+} is required for peptidyl transferase activity of the ribosome.⁵ Over a hundred Mg^{2+} ions are observed in the X-ray structure of the large ribosomal subunit (LSU) of *H. marismortui*.^{3,6} The LSU contains four dinuclear Mg^{2+} complexes⁷ and over 20 bidentate Mg^{2+} clamps.⁸ Nucleotides that directly contact Mg^{2+} are dispersed throughout the 2° structure, in all seven domains of the LSU rRNA (Fig. S3†).

Focusing on the LSU, we infer roles of Mg^{2+} in the formation of native rRNA structure, and determine the degree to which rRNA sequence governs folding in the absence of ribosomal proteins (rProteins). We investigate collapse of rRNA from a 2° structural state in the presence of Na^+ alone into a compacted state upon addition of Mg^{2+} . We characterize rRNA folding at nucleotide resolution in the absence of rProteins, 5S rRNA, or base modifications.

Results and discussion

Chemical footprinting

We assayed rRNA folding by comparing footprinting data obtained in the presence and absence of Mg^{2+} . SHAPE (Selective 2'-Hydroxyl Acylation analyzed by Primer Extension) provides quantitative information about nucleotide flexibility and base pairing.⁹ SHAPE was used previously to characterize 2° and tertiary interactions in crystalline P4–P6 domain of the tetrahymena ribozyme.¹⁰ SHAPE has been performed previously on intact yeast ribosomes¹¹ and deproteinized *E. coli*, *C. difficile*, and *H. volcanii* rRNAs.¹²

Comparison of SHAPE data collected under various conditions can be used to characterize structural transitions. SHAPE has been used previously to monitor RNA structural transitions induced by cations, small molecules, or proteins.^{9,13} We have previously utilized this technique to monitor Mg^{2+} -induced transitions of domains and subdomains of LSU rRNA.^{14–16}

Here SHAPE was performed, in the absence and presence of Mg^{2+} , on the intact ~2900 nt *T. thermophilus* 23S rRNA (LSU rRNA). Experiments were performed in the presence of 250 mM Na^+ (abbreviated Na^+ conditions), which favors formation of 2° structure,^{1,2} and in 250 mM Na^+ plus 10 mM Mg^{2+} (Na^+/Mg^{2+} conditions), which favors formation of tertiary interactions. SHAPE data were collected for 2890 of the 2911 LSU rRNA nucleotides (>99%). Na^+ conditions yielded low SHAPE reactivities for helical nucleotides (Fig. S2, ESI†). Reactive sites are observed at non-helical regions of the canonical LSU rRNA 2° structure.^{17,18}

SHAPE data under different conditions were quantitatively compared showing broadly distributed changes in reactivity upon addition of Mg^{2+} to the LSU rRNA (Fig. 1). For this work we

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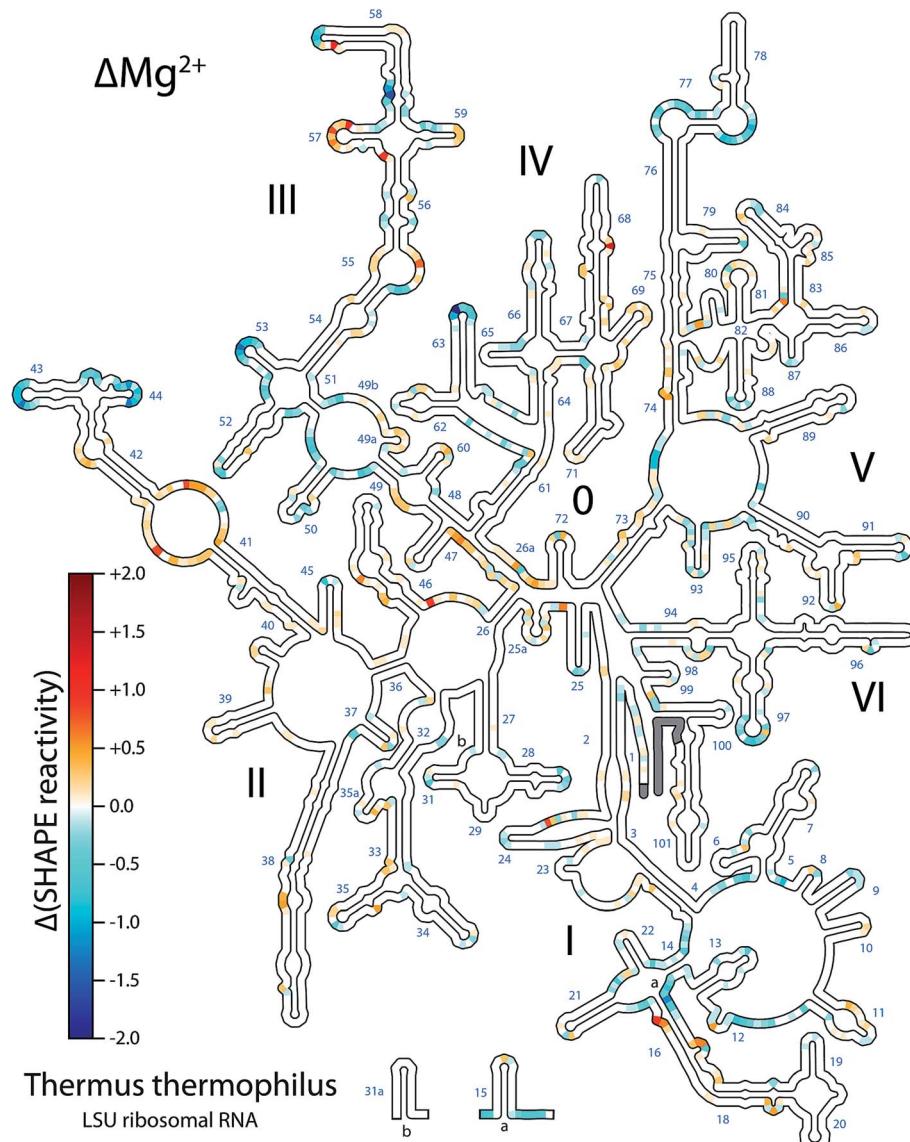


Fig. 1 *T. thermophilus* LSU rRNA exhibits Mg^{2+} -dependent changes in structure. Changes in SHAPE reactivity caused by the addition of Mg^{2+} (ΔMg^{2+} reactivity) are mapped onto LSU rRNA 2° structure. Positive values (red) indicate increased SHAPE reactivity upon addition of 10 mM Mg^{2+} . Negative values (blue) indicate decreased reactivity. Grey; data not available for those nucleotides. Domain numbers and helix numbers are indicated. Methods are described in the ESI† and full numeric SHAPE data are available in ESI† Dataset 1. This figure was generated using RiboVision; a full-page version with greater detail is provided in the ESI (Fig. S4†).

have collected, quantitatively analyzed and visualized an extensive amount of data. Na^+ and Na^+/Mg^{2+} data were normalized, and Na^+ SHAPE reactivities were subtracted from Na^+/Mg^{2+} reactivities at each nucleotide position to obtain ΔMg^{2+} data (Fig. 1 and ESI† Dataset 1). In analysis of Mg^{2+} -dependent SHAPE changes, values >0.3 SHAPE units (positive or negative) are termed ‘ ΔMg^{2+} sites’, *i.e.*, sites which experience a significant Mg^{2+} -dependent alteration in SHAPE reactivity; $\sim 7.5\%$ of LSU rRNA nucleotides exceed this threshold. Overall, the locations of ΔMg^{2+} sites are consistent with Mg^{2+} -induced formation of tertiary interactions observed in the three-dimensional structure of the ribosome. The extent and distribution of ΔMg^{2+} sites suggest a global transition from a state

comprising 2° structure with limited tertiary interactions (the rRNA- Na^+ state) to a state containing a significant extent of tertiary interactions, plus additional local interactions (the rRNA- Mg^{2+} state).

Addition of Mg^{2+} causes changes in SHAPE reactivity that are dispersed throughout the LSU rRNA. However, nucleotides within helical regions in the 2° structure¹⁹ are excluded from these changes (Fig. 1). The overall absence of ΔMg^{2+} sites from helices confirms that addition of Mg^{2+} does not change RNA 2° structure of the LSU rRNA.

By contrast, nucleotides found in loops, bulges and otherwise unpaired regions in the 2° structure are responsive to Mg^{2+} (Fig. 1 and S2†). The observed changes in SHAPE reactivity are

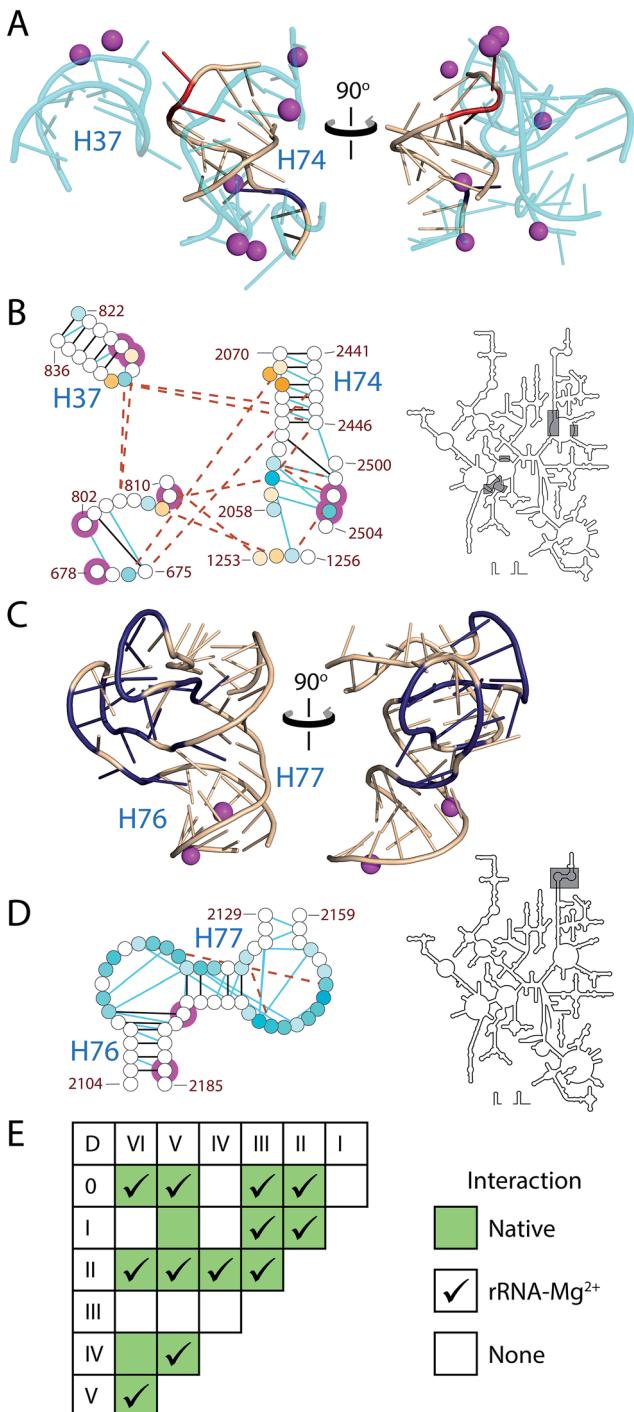


Fig. 2 Mg²⁺-induced changes in SHAPE reactivity in LSU rRNA are consistent with formation of tertiary interactions. (A) The 3D structure of H74 and the surrounding rRNA (cyan). H74 nucleotides are colored by ΔMg²⁺ values (red, increased reactivity; dark blue, decreased reactivity; beige, little/no change in reactivity). (B) 2° Structure, tertiary interactions and Mg²⁺ interactions of H74 in the assembled ribosome. (C) 3D structure of the uL1 protuberance. (D) 2° Structure, tertiary interactions and Mg²⁺ interactions of the uL1 protuberance. Nucleotides of the uL1 protuberance are colored by ΔMg²⁺ values as in Fig. 1. (E) Inter-domain interactions within the LSU rRNA. Green boxes indicate domain pairs that interact through long-range RNA–RNA contacts in the assembled LSU. Checkmarks indicate where data supports presence of inter-domain interactions in the rRNA–Mg²⁺ state. In (B and D), lines indicate RNA–RNA interactions in the 3D

consistent with a model in which Mg²⁺ induces native-like tertiary interactions. Six lines of evidence support this hypothesis: (i) ΔMg²⁺ sites are highly focused in rRNA loops, bulges, and other non-helical regions, which in general are expected to form tertiary interactions, (ii) ΔMg²⁺ sites cluster at or near nucleotides involved in tertiary interactions in the assembled LSU, (iii) ΔMg²⁺ sites cluster at or near rRNA regions involved in inter-domain interactions, (iv) ΔMg²⁺ sites are absent from regions that do not form tertiary interactions, (v) ΔMg²⁺ sites do not cluster exclusively at or near sites of first-shell RNA–Mg²⁺ interactions (ESI†), obviating the possibility that the signal arises predominantly from direct Mg²⁺ interactions, and (vi) it has been demonstrated previously that Mg²⁺ effects on SHAPE reactivity are related to formation of tertiary interactions.^{9,20}

Structural analysis

Using the RiboVision visualization suite,²¹ we compared RNA tertiary interactions²² in the assembled *T. thermophilus* ribosome⁴ to ΔMg²⁺ sites observed here by SHAPE. Regions with the greatest density of ΔMg²⁺ sites were manually inspected in both two and three dimensions. The ΔMg²⁺ sites correlate with locations of tertiary interactions in the assembled LSU (Fig. 2 and 3); nearly all ΔMg²⁺ sites occur at or directly adjacent to one or more nucleotides that participate in tertiary interactions in the LSU. Conversely, loops that do not participate in RNA–RNA tertiary interactions in the assembled LSU are not responsive to Mg²⁺ (ESI†).

Relationships between ΔMg²⁺ sites and local and tertiary interactions are illustrated in Fig. 2 and 3. Helix 74 (H74), which lies at the functional and structural core of the LSU, contains ΔMg²⁺ sites consistent with formation of native tertiary interactions (Fig. 2A and B). In the assembled LSU, H74 interacts with RNA from Domain 0 (D0), DII and DV, in addition to Mg²⁺ ions to form a portion of the LSU functional core, the peptidyl transferase center. ΔMg²⁺ sites in H74 correlate with nucleotides involved in tertiary interactions that connect DV to distant nucleotides of D0 and DII (ESI†) that are remote from H74 in the primary and 2° structure. Formation of these tertiary interactions would necessarily cause collapse of the global RNA structure, decreasing the radius of gyration, as previously observed upon addition of Mg²⁺ to the LSU rRNA.²³

ΔMg²⁺ sites in the uL1 protuberance are consistent with Mg²⁺-induced formation of loop–loop interactions between nucleotides 2109–2119 and 2161–2173 (Fig. 2C and D). The uL1 protuberance extends from the exterior of the LSU and interacts with rProtein uL1.²⁴ The loop regions of the uL1 protuberance become unreactive to SHAPE upon introduction of Mg²⁺ (Fig. S2†). In the same way, virtually all ΔMg²⁺ sites in the LSU

structure. Solid black lines are 2° hydrogen bonding interactions. Dashed brown lines are tertiary hydrogen bonding interactions. Cyan lines are stacking interactions. Magenta circles are nucleotides in first-shell interactions with Mg²⁺ ions. Gray boxes on intact 2° structure (right) highlight the region displayed on the left. Three dimensional figures were generated with PyMol using the *T. thermophilus* ribosome (PDB entries 2J00 and 2J01).

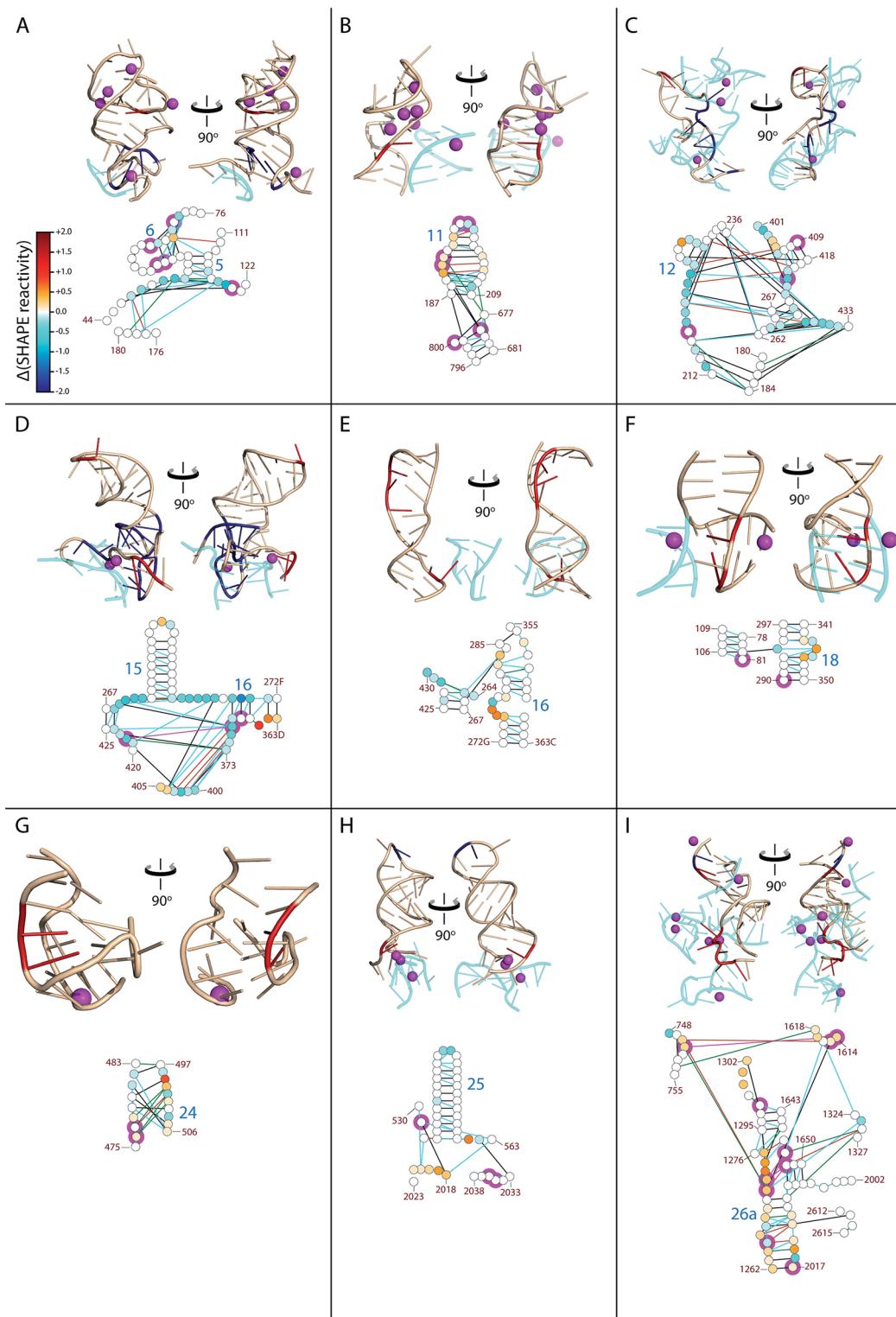


Fig. 3 ΔMg^{2+} regions of the LSU rRNA. LSU rRNA with high density of ΔMg^{2+} sites is shown with regions of rRNA that interact at or near the ΔMg^{2+} sites. A–Z Top panels: 3D structures of selected regions of LSU rRNA (adjacent ΔMg^{2+} sites) from the *T. thermophilus* ribosome crystal structure (PDB IDs: 2J00 and 2J01). Nucleotides are colored by ΔMg^{2+} values (red, increased reactivity; dark blue, decreased reactivity; wheat, little to no change in reactivity). LSU rRNA segments that form tertiary interactions at or near these ΔMg^{2+} sites are colored cyan. Mg^{2+} cations that form first shell interactions with RNA (2.4 Å cut-off) are represented by magenta spheres. RNA is in cartoon representation. A–Z Bottom panels: Mg^{2+} -induced changes in SHAPE reactivity mapped onto LSU rRNA 2° structure. Nucleotides are represented as circles, colored as in Fig. 1. Lines represent RNA–RNA interactions observed in the *T. thermophilus* ribosome structure (determined by FR3D; black, base–base; orange, base–phosphate; blue, base–stacking; and green, base–sugar). Magenta halos indicate nucleotides that interact directly with Mg^{2+} ions. The same nucleotides are displayed in corresponding 3D and 2D representations. Helix and nucleotide numbers are indicated.

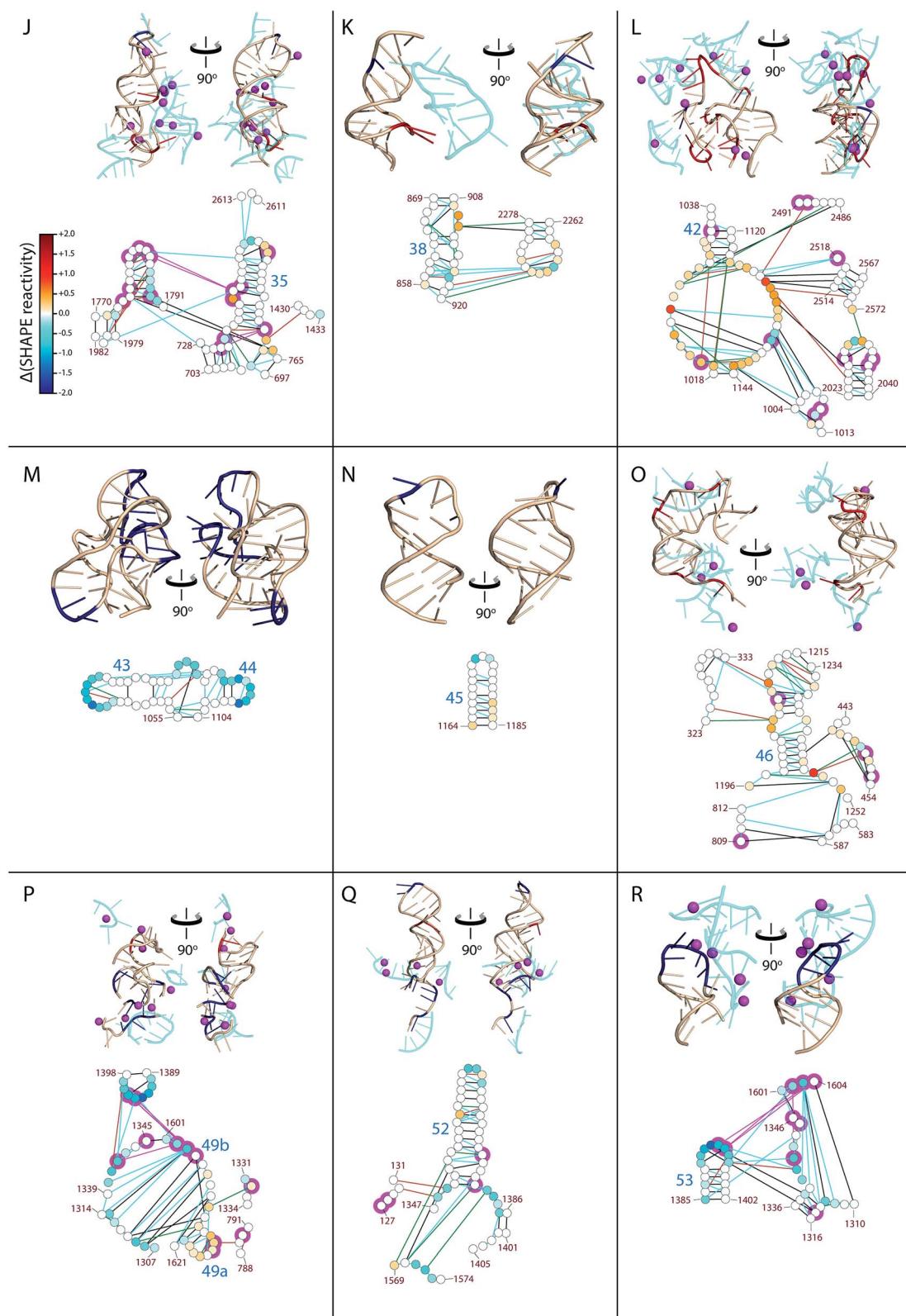


Fig. 3 (continued).

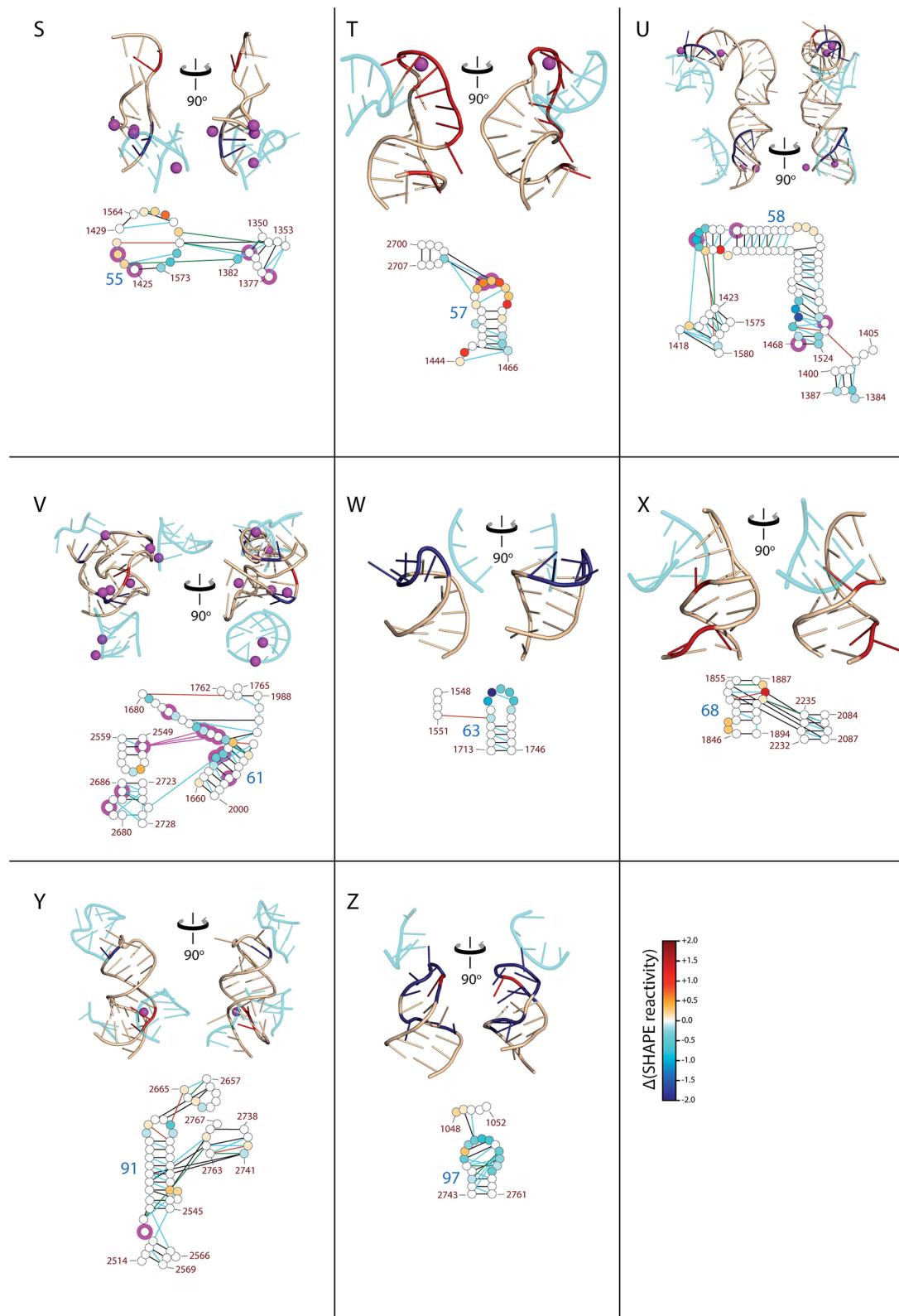


Fig. 3 (continued).

rRNA occur at or adjacent to nucleotides involved in tertiary interactions in the assembled LSU (Fig. 3). The locations of ΔMg^{2+} sites suggest formation of an intricate tertiary interaction network, corresponding with global collapse of the LSU rRNA into a native-like rRNA–Mg $^{2+}$ state. Further analysis of ΔMg^{2+} sites is provided in the ESI.[†]

In sum, the ΔMg^{2+} data support formation of native-like inter-domain architecture of protein-free LSU rRNA in the presence of Mg $^{2+}$. Most domain pairs observed in the assembled LSU are inferred to interact in the rRNA–Mg $^{2+}$ state (Fig. 2E). In the assembled ribosome, long-range inter-domain tertiary interactions connect disparate 2° regions of LSU rRNA.⁴ Mg $^{2+}$ has been implicated previously in facilitating these inter-domain contacts,⁶ whereas moderate monovalent cation concentrations induce 2° structure only.²⁵ SHAPE results here support formation of 12 out of 14 native domain pair interactions (Fig. 2E and Table S5, ESI[†]). The network of inter-domain interactions in the rRNA–Mg $^{2+}$ state supports formation of the core domain architecture of the LSU, in the absence of other interaction partners.

In the rRNA–Mg $^{2+}$ state, tertiary interactions not supported by correlation of ΔMg^{2+} sites to assembled LSU-interacting regions must fall into one of three classes; (i) already present in Na $^+$ alone, (ii) present only in Na $^+/\text{Mg}^{2+}$, but undetectable by comparative SHAPE, or (iii) absent or unstable in Na $^+/\text{Mg}^{2+}$, in which case their formation is dependent on agents other than or in addition to Mg $^{2+}$. Inter-domain interactions that are not indicated by ΔMg^{2+} sites may be pre-organized for protein-mediated association. The extent of supported Mg $^{2+}$ -induced tertiary interactions (Fig. 2 and 3) suggests that most nucleotides involved in the absent interactions are pre-organized for association, contingent upon addition of a mediating rProtein (see ESI, Table S6[†]). In this model, formation of certain native tertiary interactions is more favorable in the rRNA–Mg $^{2+}$ state than in presence of Na $^+$ alone.

A role for rProteins is suggested by RNA–RNA interactions that appear to be absent from the rRNA–Mg $^{2+}$ state. These absent interactions occur at or adjacent to regions of native rRNA interaction with rProteins uL2, uL3, uL4, uL15, uL16 and bL20 (Table S6[†]). Based on this, we suggest that these rProteins foster formation of certain inter-domain RNA–RNA interactions in the assembled ribosome. Five of the six genes coding for these rProteins (uL2, uL3, uL4, uL16, and bL20) are essential for survival,²⁶ and all six rProteins exhibit strong interconnectivity in LSU assembly maps. The correlation between functional and structural importance of this set of rProteins implies that the interactions they mediate foster formation of the active, native LSU. It appears that the rRNA–Mg $^{2+}$ state represents the core architecture of the LSU which, while not catalytically active,²⁷ positions the residues of the LSU rRNA in such a way as to promote native interactions with rProteins to ultimately form a functional LSU.

Conclusions

The LSU rRNA interacts *in vivo* with the 5S rRNA and over 30 LSU proteins.⁴ Our experiments involve *in vitro*-transcribed LSU

rRNA in solutions containing only buffer and ions, within reasonable physiological ranges, with no additional components. We have used comparative SHAPE to provide single-nucleotide information regarding Mg $^{2+}$ -induced structural changes. We propose a model of the near-native LSU state in the presence of Mg $^{2+}$ based on correlations between our data and three-dimensional ribosomal structures. Our model, termed the rRNA–Mg $^{2+}$ state, is collapsed with a considerably reduced radius, and includes core inter-domain architecture similar to that in the native LSU, but is not expected to be catalytically competent. Protein-free rRNA is not capable of peptidyl transferase, even in presence of Mg $^{2+}$.²⁷ Mg $^{2+}$ interactions may be cooperative with rProtein interactions in formation of the native LSU. Certain rProtein–rRNA interactions are cation-mediated,⁶ underscoring the interdependence of rProteins and cations in the structure of the assembled LSU.

The results presented here are consistent with previous results, in which removal of Mg $^{2+}$ results in global unfolding of the LSU rRNA, as detected by several biophysical techniques.²³ Virtually all aspects of ribosome structure and function involve Mg $^{2+}$ to some degree.²⁸ Ribosome function can survive significant deproteinization, but is completely abolished upon exposure to divalent cation-chelating compounds.⁵ We suggest that Mg $^{2+}$ is crucial to the formation of local interactions, inter-domain contacts and core LSU architecture. When the assembled LSU is depleted of Mg $^{2+}$, this core architecture is lost, resulting in dissociation of domains and their unfolding, and loss of ribosomal function. Our results support a model in which the LSU rRNA requires only Mg $^{2+}$ to fold to a well-defined, collapsed near-native state, and elements such as rProteins induce local structural changes that foster formation of the remaining interactions to form a fully catalytic LSU. This model is consistent with the findings of Woodson and coworkers, who used chemical footprinting and molecular dynamics simulations of the small ribosomal subunit to show that ribosomal proteins dampen rRNA fluctuations at subdomain interfaces and cause switching of conformation of select regions of the rRNA.²⁹

Conflicts of interest

The authors of this paper declare that they have no conflict of interest.

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References

- 1 P. Brion and E. Westhof, *Annu. Rev. Biophys. Biomol. Struct.*, 1997, **26**, 113–137.
- 2 J. C. Bowman, T. K. Lenz, N. V. Hud and L. D. Williams, *Curr. Opin. Struct. Biol.*, 2012, **22**, 262–272.

3 N. Ban, P. Nissen, J. Hansen, P. B. Moore and T. A. Steitz, *Science*, 2000, **289**, 905–920.

4 M. Selmer, C. M. Dunham, F. V. Murphy, A. Weixlbaumer, S. Petry, A. C. Kelley, J. R. Weir and V. Ramakrishnan, *Science*, 2006, **313**, 1935–1942.

5 H. F. Noller, V. Hoffarth and L. Zimniak, *Science*, 1992, **256**, 1416–1419.

6 D. J. Klein, P. B. Moore and T. A. Steitz, *RNA*, 2004, **10**, 1366–1379.

7 C. Hsiao and L. D. Williams, *Nucleic Acids Res.*, 2009, **37**, 3134–3142.

8 A. S. Petrov, J. C. Bowman, S. C. Harvey and L. D. Williams, *RNA*, 2011, **17**, 291–297.

9 S. A. Mortimer and K. M. Weeks, *J. Am. Chem. Soc.*, 2007, **129**, 4144–4145.

10 Q. Vicens, A. R. Gooding, A. Laederach and T. R. Cech, *RNA*, 2007, **13**, 536–548.

11 J. A. Leshin, R. Heselpoth, A. T. Belew and J. D. Dinman, *RNA Biol.*, 2011, **8**, 478–487.

12 C. A. Lavender, R. Lorenz, G. Zhang, R. Tamayo, I. L. Hofacker and K. M. Weeks, *PLoS Comput. Biol.*, 2015, **11**, e1004126.

13 C. E. Dann III, C. A. Wakeman, C. L. Sieling, S. C. Baker, I. Irnov and W. C. Winkler, *Cell*, 2007, **130**, 878–892.

14 S. S. Athavale, J. J. Gossett, C. Hsiao, J. C. Bowman, E. O'Neill, E. Hershkovitz, T. Preeprem, N. V. Hud, R. M. Wartell, S. C. Harvey and L. D. Williams, *RNA*, 2012, **18**, 752–758.

15 C. Hsiao, T. K. Lenz, J. K. Peters, P. Y. Fang, D. M. Schneider, E. J. Anderson, T. Preeprem, J. C. Bowman, E. B. O'Neill, L. Lie, S. S. Athavale, J. J. Gossett, C. Trippe, J. Murray, A. S. Petrov, R. M. Wartell, S. C. Harvey, N. V. Hud and L. D. Williams, *Nucleic Acids Res.*, 2013, **41**, 3373–3385.

16 K. A. Lanier, S. S. Athavale, A. S. Petrov, R. M. Wartell and L. D. Williams, *Biochemistry*, 2016, **55**, 4603–4613.

17 A. S. Petrov, C. R. Bernier, B. Gulen, C. C. Waterbury, E. Hershkovitz, C. Hsiao, S. C. Harvey, N. V. Hud,

18 G. E. Fox, R. M. Wartell and L. D. Williams, *PLoS One*, 2014, **9**, e88222.

19 H. F. Noller, J. Kop, V. Wheaton, J. Brosius, R. R. Gutell, A. M. Kopylov, F. Dohme, W. Herr, D. A. Stahl, R. Gupta and C. R. Woese, *Nucleic Acids Res.*, 1981, **9**, 6167–6189.

20 S. Blouin, R. Chinnappan and D. A. Lafontaine, *Nucleic Acids Res.*, 2011, **39**, 3373–3387.

21 C. Bernier, A. S. Petrov, C. Waterbury, J. Jett, F. Li, L. E. Freil, B. Xiong, L. Wang, A. Le, B. L. Milhouse, E. Hershkovitz, M. Grover, Y. Xue, C. Hsiao, J. C. Bowman, S. C. Harvey, J. Z. Wartel and L. D. Williams, *Faraday Discuss.*, 2014, **169**, 195–207.

22 M. Sarver, C. L. Zirbel, J. Stombaugh, A. Mokdad and N. B. Leontis, *J. Math. Biol.*, 2008, **56**, 215–252.

23 Q. M. Yi and K. P. Wong, *Biochem. Biophys. Res. Commun.*, 1982, **104**, 733–739.

24 A. Nikulin, I. Eliseikina, S. Tishchenko, N. Nevskaia, N. Davydova, O. Platonova, W. Piendl, M. Selmer, A. Liljas, D. Drygin, R. Zimmermann, M. Garber and S. Nikonorov, *Nat. Struct. Mol. Biol.*, 2003, **10**, 104–108.

25 J. K. Frederiksen, N. S. Li, R. Das, D. Herschlag and J. A. Piccirilli, *RNA*, 2012, **18**, 1123–1141.

26 S. Shoji, C. M. Dambacher, Z. Shahjani, J. R. Williamson and P. G. Schultz, *J. Mol. Biol.*, 2011, **413**, 751–761.

27 P. Khaitovich, T. Tenson, A. S. Mankin and R. Green, *RNA*, 1999, **5**, 605–608.

28 K. Trappl and N. Polacek, *Met. Ions Life Sci.*, 2011, **9**, 253–275.

29 S. C. Abeysirigunawardena, H. Kim, J. Lai, K. Ragunathan, M. C. Rappé, Z. Luthey-Schulten, T. Ha and S. A. Woodson, *Nat. Commun.*, 2017, **8**, 492.

