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# Hydrothermal liquefaction of high protein microalgae *via* clay material catalysts†

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Hydrothermal liquefaction (HTL) is a promising method to convert wet microalgae into a petroleum-like biocrude. In this paper, we demonstrated the HTL of microalgae *Spirulina* over various clay material catalysts and open up interesting perspectives for efficiently liquefaction processes. The clay catalyst successfully improved the biocrude yield, the liquefaction conversion, and energy recovery. Investigation into optimizing operation conditions suggested that the reaction temperature and holding time significantly influenced the HTL process. The highest biocrude yields were obtained at a reaction temperature within the range of 220–250 °C and a holding time of 30 min with montmorillonite (46.56%), kaolinite (44.07%) and dolomitic limestone (46.35%). Adding clay catalysts changed the element content and produced more asphaltene content in the biocrude. The GC-MS analysis revealed that the catalyst greatly influenced the composition of the hydrocarbons, acids, esters, amides and heterocyclic compounds in the biocrude.

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## 1. Introduction

Environmental concerns about greenhouse gas emission and finite fossil fuel reserves persuaded humans to search for alternative renewable liquid fuels.<sup>1</sup> As the third generation of biomass, microalgae can efficiently convert sunlight into chemical energy and presents the potential to be the feedstock in further thermochemical or biological conversion processes.<sup>2–4</sup> Among various kinds of processes, hydrothermal liquefaction (HTL) could be a promising method to produce liquid biofuels from microalgae.<sup>5</sup>

The HTL of microalgae is the thermochemical process which turns microalgae biomass into liquid biocrude and other byproducts in an oxygen-isolated reactor with inert gases or reducing gases, at certain temperature (200–400 °C) and pressure (5–28 MPa).<sup>6–8</sup> The hot pressure water in the HTL is used for both reaction medium and solvent. The most important product is the high-energy-density black sticky liquid biocrude, which is mainly consisted by hydrocarbons and N/O-containing organic compounds.<sup>9,10</sup> Previous works have proven that the HTL process can convert up 20–70% of microalgae (dry weight) into biocrude with a heating value between 28 and 38 MJ kg<sup>-1</sup>, which is 66–90% of the heating value of petroleum. However, at present, the obtained biocrude unsatisfactorily contains a relatively high contents of oxygen (10–20%), nitrogen (3–10%) and

sulfur (1–2%), which were much higher than those in petroleum.<sup>11–13</sup> When directly applied to petroleum refining fuel or as the fuel, the high oxygen and nitrogen content would be damage to production facility or cause air pollution.<sup>14–16</sup> To improve the biocrude yield and reduce the heteroatom content, researchers started the catalyst screening work.

Many catalysts from petroleum industry such as Pd/C, Pt/C, Ru/C, Ni/SiO<sub>2</sub>-Al<sub>2</sub>O<sub>3</sub>, CoMo/γ-Al<sub>2</sub>O<sub>3</sub>, Raney-Ni, HZSM-5 and zeolite have been applied in the HTL of microalgae and the liquefaction process has been intensively investigated.<sup>17–22</sup> However, these synthetic industrial catalysts required extra cost and caused pollution during the catalyst production. In that case, we believed that a natural clay material catalyst could be greener. Besides, it is widely believed that the fossil fuel petroleum came from ancient fossilized organic materials like algae through the thermochemical process with intense heat and pressure in the lithosphere. During the process, rocks and clays might play a role as catalyst.<sup>23–27</sup> Since the HTL of microalgae to produce biocrude is much like the formation of petroleum, natural clay materials could be the catalyst to produce the petroleum-like liquid biocrude and provide a better way for catalyst screening.

In this paper, we reported a detailed study on hydrothermal liquefaction of high protein microalgae *Spirulina* over three kinds of conventional clay catalysts. Effects of catalyst types, reaction temperature, and holding time on the biocrude yield and liquefaction conversion were investigated. The characterization of target product biocrude was tested by element analysis, group component analysis, and GC-MS.

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## 2. Materials and methods

### 2.1. Materials

The HTL feedstock microalgae *Spirulina* were purchased from Shandong Binzhou Tianjian Biotechnology Co., Ltd. (Shandong, China). Table 1 showed the biochemical information of feedstock. The clay catalysts montmorillonite KSF (CAS 1318-93-0), kaolinite (CAS 1318-74-7), and dolomitic limestone (CAS 1317-65-3) were purchased from Sigma-Aldrich (Shanghai, China). Deionized water was prepared by a Millipore Milli Q UV Plus system. All other chemicals used were of analytic reagent grade and without any further purification.

### 2.2. HTL process and product separation

The HTL experiments were carried out in a 1.8 L high-pressure stainless reactor (Parr Instruments Co., Moline, PA). In a single round, 120 g of microalgae, 12 g of catalyst and 480 g of deionized water were loaded into the reactor, and the mixtures were well-stirred. After the loading, the reactor was sealed and pumped with N<sub>2</sub> three times to remove the air and provide an inert atmosphere. The reactor was heated to the set temperature and held for a certain time.

After completion of the holding time, the reactor was cooled to room temperature (25 °C), and then the exhaust valve was opened to release the pressure. The mixture of liquid and solid products were poured out from the reactor and collected. Dichloromethane (DCM) was used to wash the reactor, and the washing liquor was collected and mixed with products. The products and the washing DCM were filtered, and the solid residue was washed with DCM for three times to remove the DCM-soluble product. After filtering, the liquid mixture was separated into the water-soluble phase and DCM-soluble phase in a separating funnel. The DCM-soluble phase was treated by vacuum distillation at 60 °C, 0.05 MPa to remove the DCM. After

the evaporation, the obtained DCM-removed black viscous liquid product was defined as the aim product biocrude and then was weighted. The solid residue was moved into a drying oven at 105 °C for 24 h to remove the water and DCM and then weighted. The weight of organic residue was calculated by the mass difference between the solid residue and the sum of the weight of ash in dry microalgae and the weight of the clay catalyst. The analysis of the gaseous and water-soluble reaction products is beyond the scope of this paper. The experiment without catalyst was named as blank to be used for comparison in this paper. All the liquefaction experiments were repeated for at least three times, and only the average values were presented.

### 2.3. Analytic methods of product

The liquefaction conversion (%) and biocrude yield ( $Y_{\text{biocrude}}$ , %) were calculated by the following formulas (1) and (2), respectively.

$$\text{Liquefaction conversion(\%)} = \left(1 - \frac{\text{mass of organic solid residue}}{\text{mass of microalgae}}\right) \times 100\% \quad (1)$$

$$Y_{\text{biocrude}}(\%) = \frac{\text{mass of biocrude}}{\text{mass of microalgae}} \times 100\% \quad (2)$$

The content of asphaltene in biocrude sample was analyzed by ASTM D2007-03(2008) methods. Elements composition of feedstock and biocrude were analyzed by a VARIO EL III elemental analyzer to determine the content of C, H, N, S element. The O content was determined by difference. The higher heating value (HHV) of the sample was calculated as described literature,<sup>28</sup> as shown in formula (3):

$$\text{HHV (MJ kg}^{-1}\text{)} = (34C + 124.3H + 6.3N + 19.3S - 9.8O)/100 \quad (3)$$

where C, H, O, N, S are the element content of carbon, hydrogen, nitrogen, sulfur, and oxygen, respectively.

Energy recovery (ER) was calculated as the formula below to analyze the energy balance,<sup>29</sup> as shown in formula (4):

$$\text{ER(\%)} = \frac{(\text{HHV}_{\text{biocrude}} \times m_{\text{biocrude}})}{(\text{HHV}_{\text{algae}} \times m_{\text{algae}})} \times 100\% \quad (4)$$

The organic composition of biocrude was analyzed with a gas chromatography-mass spectrometry (GC-MS, QP2010, Shimadzu Co., Tokyo, Japan). GC-MS was equipped with a Varian DB-5 column (30 m × 0.25 mm × 0.25 μm). Helium was used as the carrier gas. The injection temperature and interface temperature were set at 250 °C and 320 °C. The ion source was adjusted to 200 °C. The mass spectrometer was operated in positive electron impact mode (EI) at 70 eV. Scan range of mass spectrum was in  $m/z$  of 20–650. All chromatogram peaks in spectra were compared with the electron impact mass spectrum from NIST Database (NIST11). Samples were directly diluted with acetone and filtered through a 0.45 μm filter. The column temperature was set at 50 °C for 2 min, then ramped up at a rate

Table 1 Analysis results of feedstock<sup>a</sup>

Analyses	<i>Spirulina</i>
<b>Proximate analysis (wt%)</b>	
Moisture W <sub>ar</sub>	6.5
Ash A <sub>ar</sub>	6.8
<b>Biochemical composition analysis (wt%, daf)<sub>ar</sub></b>	
Carbohydrates	23.7
Protein	66.0
Lipids	10.3
<b>Elemental analysis (wt% daf)<sub>ar</sub></b>	
C	47.08
H	8.77
O <sup>b</sup>	30.58
N	9.97
S	2.08
H/C (molar ratio)	2.23
O/C (molar ratio)	0.49
HHV (MJ kg <sup>-1</sup> )	24.94

<sup>a</sup> ar = as received. daf = dry ash free. <sup>b</sup> Determined by difference.



of 10 °C min<sup>-1</sup> to 120 °C and maintained for 1 min, afterward increased to 250 °C at the same heating rate and maintained for 20 min.

### 3. Results and discussions

#### 3.1. Effect of catalyst on hydrothermal liquefaction of *Spirulina*

After the HTL process, the microalgae was converted into various phase products like biocrude, solid residue, water-soluble aqueous phase product and gas phase product. However, the yields of biocrude were emphasized in recent studies.<sup>30</sup> This section provides the information about the effect of adding different clay material catalysts on in the hydrothermal liquefaction of *Spirulina*. The liquefaction processes were carried out at a reaction temperature of 270 °C and with a holding time of 30 min. Experiment data and further comparison were listed in Table 2.

As shown in Table 2, three kinds of clay material catalysts all increased the biocrude yield, compared with the blank experiments (30.51%). The biocrude yields at 270 °C with different clay catalyst were very close (39.67%, 39.71% and 40.19% for montmorillonite, kaolin and dolomitic limestone, respectively). All the clay catalysts showed a favorable effect on the improvement of biocrude yield. The increased liquefaction conversion shown in Table 2 also indicated that adding clay catalysts not only catalyzed the decomposition of the solid organic matters in the microalgae but also enhanced the liquefaction process to produce more biocrude. According to previous studies, the primary compounds in the microalgae are usually liquefied and converted into biocrude in the following order: lipid, protein, and carbohydrate.<sup>31</sup> As shown in Fig. 1, the obtained biocrude yield with/without clay catalysts were all much higher than the lipid content in the biocrude. The improved biocrude yield should come from the conversion of protein or carbohydrate. Also, after the dosage of clay catalysts, the liquefaction conversion increased about 5% while the improvement of biocrude

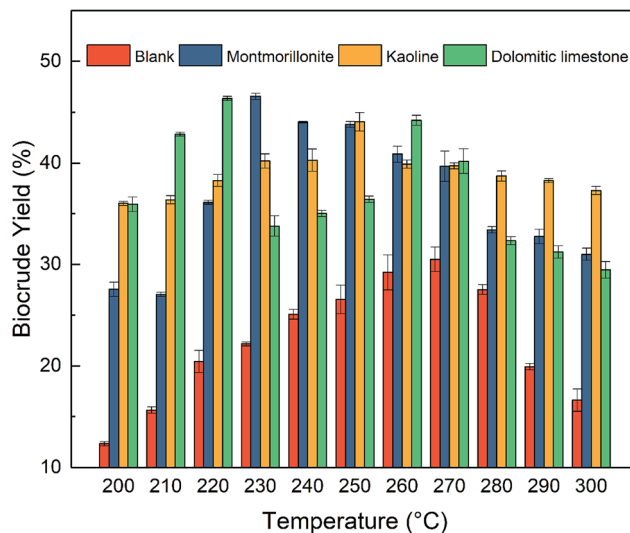


Fig. 1 Effect of reaction temperature on HTL of microalgae over different clay catalysts.

yield was around 10%. The difference suggested that the formation of more biocrude might not only come from the decomposition of microalgae but also be from the condensation reaction of small molecular water-soluble molecules.<sup>32</sup> As widely known, the asphaltene could be produced by the polymerization of small molecules. The growth of the asphaltene content in biocrude could prove that there must be more re-polymerization reactions after adding catalysts. Further element analysis suggested that adding clay catalysts resulted in a pleasing increase of the hydrogen content and slightly reduced the sulfur content in biocrude. The changes of N, S, and O heteroatom content could come from a series of reactions, like decarboxylation, deamination, desulfurization, esterification, dehydration condensation, and Maillard reactions, which were associated with the condensation reaction during the formation of asphaltene. Based on the element content in biocrude, the HHVs of biocrude samples were in the range of 34–36 MJ kg<sup>-1</sup>, which was about 85–90% of the HHV of petroleum. Adding clay catalysts slightly increased the HHV of the biocrude, as shown in Table 2. Moreover, the energy recoveries were all increased by more than 10% with the three kinds of catalysts. Therefore, clay catalyst did take a positive effect on HTL of microalgae *Spirulina*.

#### 3.2. Hydrothermal catalytic liquefaction results with different operating conditions

Many literature studies have indicated that HTL conditions, like reaction temperature and holding time, were the influential factors of the biocrude yield and the liquefaction conversion.<sup>33–35</sup> These condition parameters play a decisive role in the decomposing of microalgae and the formation of biocrude. Hence, the influence on the reaction temperature and holding time was studied.

**3.2.1. Effect of reaction temperature on HTL with different clay catalysts.** Fig. 1 presented the biocrude yield as the

Table 2 HTL results with different catalyst

	Blank	Montmorillonite	Kaoline	Dolomitic limestone
Biocrude yield (wt%)	30.51	39.67	39.71	40.19
Liquefaction conversion (%)	87.54	92.75	93.11	91.97
<b>Element analysis (wt%)</b>				
C	71.75	70.12	69.45	72.06
H	8.94	9.95	9.78	9.54
O <sup>a</sup>	11.95	12.15	13.36	10.91
N	6.83	7.42	7.16	6.97
S	0.53	0.36	0.25	0.52
Asphaltene content (%)	26.9	58.1	51.5	38.7
HHV (MJ kg <sup>-1</sup> )	34.87	35.55	34.96	35.83
Energy recovery (%)	42.66	58.28	55.66	57.74

<sup>a</sup> Determined by difference.



functions of temperature with different clay catalysts. The HTL experiments were performed with the individual catalyst at the temperature range of 200–280 °C, with an increment of 10 °C and a holding time of 30 min. The corresponding reaction temperature ranged from 1.98 to 9.85 MPa, depending on the final temperatures.

As shown in Fig. 1, with the reaction temperature increasing from 200 to 280 °C, the biocrude yield gradually increased and reached a maximum yield. After that, the biocrude yield gradually decreased with higher temperature. Consequently, from the perspective of biocrude yield, the optimum temperature for the highest biocrude yield with montmorillonite KSF (46.56%), kaolinite (44.07%) and dolomitic limestone (46.35%) were 230, 250 and 220 °C, respectively. The pre-set reaction temperature provided the necessary energy to convert the NAS into biocrude and other byproducts. Raising the reaction temperature in an appropriate range could enhance the conversion process to biocrude. However, when the higher temperature was supplied, the balance between the re-polymerization of small molecules and the cracking of big molecules could be reassigned. The decrease of biocrude yield suggested that the weight of biocrude production became lighter with higher temperature. However, the type of catalyst significantly affected the distribution of solid residue and products of other phases.

Interestingly, there was a second-highest peak of biocrude yield (44.21%) with dolomitic limestone as the catalyst at 260 °C. However, the biocrude obtained at a lower temperature with dolomitic limestone showed a better flowability than that obtained at a higher temperature. In the lower temperature

experiments, the obtained solid residue remained the form of microalgae while the dark green color of *Spirulina* changed into dark yellowish-brown. Obviously, the microalgae were not decomposed entirely at a lower temperature. The liquefaction conversions with dolomitic limestone at 220 °C was less than 80% also verified this phenomenon. The liquefaction conversion of 66.45% suggested that the unconverted organic matters in microalgae was more than the hard-to-be-converted carbohydrates. The liquefaction process did produce some DCM-soluble compounds, but the remained biochemical compounds like most carbohydrates or some proteins in the cell of microalgae were not entirely liquefied and converted into biocrude as the form obtained at high temperature. The higher active energy demand for liquefaction of carbohydrates or proteins might explain the results.<sup>32</sup>

The further comparison presented in Table 3 showed the influence of reaction temperature on element composition and asphaltene contents in biocrude. Asphaltene content data suggested that low reaction temperature led to a lower asphaltene content, and the increased temperature resulted in a higher asphaltene content. It seems that the higher temperature prompted the formation of macromolecule compounds like pitch, as described in literature.<sup>36</sup> Adding clay catalysts increased the asphaltene contents in the biocrude samples by more than 10% at 270 °C while the increase of reaction temperature from 220 to 270 °C led to an asphaltene content growth no more than 10%. The clay catalysts could take the positive catalytic role in the formation of asphaltene, and the influence was more important than the reaction temperature.

Table 3 HTL results under different reaction temperature

	Liquefaction conversion (%)	Element content (%)					ER (%)	Asphaltene content (%)
		C	H	O <sup>a</sup>	N	S		
<b>Blank</b>								
220 °C	55.82	62.65	8.82	21.28	5.98	1.27	25.3	25.4
230 °C	67.28	66.14	8.87	17.77	6.21	1.01	28.8	21.6
250 °C	82.55	71.22	8.95	12.19	6.78	0.86	37.0	25.7
270 °C	87.54	71.75	8.94	11.95	6.83	0.53	42.7	26.9
<b>Montmorillonite</b>								
220 °C	63.45	61.69	8.65	23.09	5.69	0.88	43.4	42.6
230 °C	71.23	67.84	9.35	16.67	5.62	0.52	62.6	53.1
250 °C	88.26	69.98	9.87	11.85	7.85	0.45	62.3	55.7
270 °C	92.75	70.12	9.95	12.15	7.42	0.36	58.3	58.1
<b>Kaoline</b>								
220 °C	68.14	59.25	8.66	25.91	5.43	0.75	44.3	41.8
230 °C	73.25	67.17	9.89	16.26	6.05	0.63	54.9	47.9
250 °C	94.84	68.84	9.92	14.1	6.46	0.68	61.7	48.4
270 °C	93.11	69.45	9.78	13.36	7.16	0.25	55.7	51.5
<b>Dolomitic limestone</b>								
220 °C	66.45	61.17	8.78	22.95	6.55	0.55	55.7	30.4
230 °C	78.25	68.24	8.86	15.82	6.63	0.45	44.9	34.2
250 °C	86.78	69.12	8.74	14.68	6.78	0.68	48.9	36.5
270 °C	91.97	72.06	9.54	10.91	6.97	0.52	57.7	38.7

<sup>a</sup> Determined by difference.



Element analysis presented an approximate increase of carbon and hydrogen content and a decrease of oxygen content, with the increase of the reaction temperature. The changing trend was pleasing for improving a higher heat value and stability of biocrude. Moreover, there was a trend of improvement of energy recovery with higher reaction temperature. Therefore, adding clay catalysts received a higher biocrude yield, a modified liquefaction conversion, and an improved biocrude quality. The

temperature significantly influenced the properties and characterization of biocrude.

**3.2.2. Effect of holding time on HTL with different clay catalysts.** Holding times are defined as a period of reacting at a maximum temperature, excluding heating and cooling times.<sup>36</sup> In our experiments, relations between holding time and biocrude yield or liquefaction conversion was investigated.

As shown in Fig. 2, the holding time experiments suggested that biocrude yields were closely linked to the length of holding time in an appropriate time range. When the holding time was shorter than 0.5 h, lengthening holding time significant improved the biocrude yield. Similar trends were discovered by Xu and Singh.<sup>37,38</sup> However, there was a certain threshold of holding time in the liquefaction process. As the biocrude yield peaked at 0.5 h, the further increase of holding time adverted results that the biocrude yield gradually decreased. A similar trend was found in the liquefaction conversion. A holding time expanding from 0 to 0.5 h increased the liquefaction significantly. However, the holding time between 0.5–8 h hardly affected the liquefaction conversion. Among various catalyst types, the longer holding time influenced the HTL under montmorillonite catalyst most.

For all liquefaction experiments with or without the clay catalyst, an appropriate holding time, like 0.5 h, could promote the biocrude yield and liquefaction conversion efficiently. It should be noted when the holding time was too short, the liquefaction conversion and biocrude yield were extremely low while the solid residue was hard for separation from the liquid

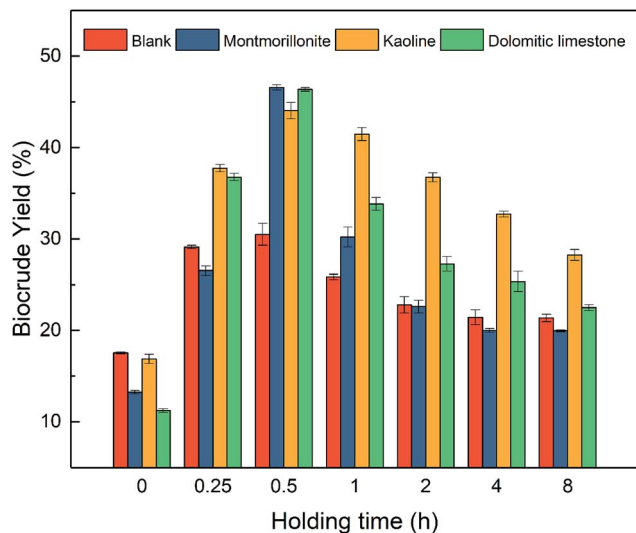


Fig. 2 Effect of holding time on HTL of microalgae over different clay catalysts.

Table 4 HTL result under different holding time

Holding Time	Liquefaction conversion (%)	Element content (%)					ER (%)	Asphaltene content (%)
		C	H	O <sup>a</sup>	N	S		
<b>Blank</b>								
0 h	79.64	68.99	9.05	14.31	6.98	0.67	23.84	21.3
0.5h	87.54	71.75	8.94	11.95	6.83	0.53	42.66	26.9
2 h	87.56	71.82	8.98	12.14	6.54	0.52	31.91	27.5
8h	88.01	72.68	9.01	11.21	6.58	0.52	30.27	26.8
<b>Montmorillonite</b>								
0 h	48.56	65.14	9.65	18.14	6.18	0.89	17.47	26.9
0.5h	71.23	67.84	9.35	16.67	5.62	0.52	62.56	53.1
2 h	71.22	68.93	8.94	15.21	6.27	0.65	30.45	53.4
8h	73.69	69.72	8.85	14.64	6.37	0.42	27.04	50.7
<b>Kaoline</b>								
0 h	75.65	64.68	9.97	18.61	5.98	0.76	22.39	32.1
0.5h	94.84	68.84	9.92	14.1	6.46	0.68	61.66	48.4
2 h	94.87	69.51	9.84	12.97	7.12	0.56	51.81	45.6
8h	93.78	69.67	9.76	12.28	7.68	0.61	39.89	47.8
<b>Dolomitic limestone</b>								
0 h	62.45	59.17	8.93	24.25	7.01	0.64	13.25	26.7
0.5h	66.45	61.17	8.78	22.95	6.55	0.55	55.72	30.4
2 h	66.68	62.88	8.77	20.93	6.86	0.56	33.64	31.2
8h	69.65	63.01	8.71	21.31	6.39	0.58	27.69	28.9

<sup>a</sup> Determined by difference.





products and kept in the form of microalgae, only with a changed color from dark green to brownish yellow. This results suggested that during the HTL process, most of the original biomolecules (lipids, proteins, and carbohydrates) in microalgae failed to convert into biocrude or any other product phases without enough holding time. On the contrary, if the holding time lasted too long, the decrease of biocrude yield could be observed. We believed this might be attributed to the cracking, steam reforming or decarboxylation reactions which turned the light oil or small molecule compounds in biocrude into the gaseous product, like methane, carbon oxide or carbon dioxide.<sup>39,40</sup> This explanation is corresponding to the growth of final pressure. At room temperature, the final pressure in the cooled reactor increased from 0.05 MPa (30 min) to 0.08 MPa (8 h). The increased final pressure suggested that more products of gaseous phase produced by expanding the holding time.<sup>41</sup> Meanwhile, the element analysis shown in Table 4 also

provided a signal about the decreasing trend of oxygen content in biocrude with a longer holding time. This signal could come from the synthesis of CO<sub>2</sub> by the decarboxylation of biocrude, as described in previous literature.<sup>32</sup> However, expanding the holding time requested more energy input for heat preservation, which made the process less economical. Therefore, an appropriate holding time, like 30 min, is favorite to the HTL of *Spirulina* over clay catalysts.

### 3.3. GC-MS characterization of biocrude and the possible reaction pathways

This section provides the information about molecules presented in the biocrude and how the clay catalysts changed the biocrude. The samples of biocrude obtained at the highest biocrude yield were analyzed by GC-MS to identify the chemical compounds. Obtained chemical compounds with a relative peak area more than 1% are listed in Table 5. It should be noted

Table 5 GC-MS results of the biocrude obtained from catalytic HTL over various catalysts

No.	Compounds	Retention time (min)	Relative content (%)			
			Blank	Montmorillonite	Kaoline	Dolomitic limestone
1	1,4-Dioxane	2.408	15.33	—	—	—
2	2-Methylpyrazine	3.691	3.67	—	—	—
3	4-Hydroxy-4-methylpentan-2-one	3.886	7.83	4.18	2.02	2.97
4	<i>o</i> -xylene	4.218	—	—	—	—
5	(2 <i>R</i> ,3 <i>R</i> )-1- <i>tert</i> -Butyl-2,3-dimethylaziridine	4.702	—	3.63	2.34	—
6	2,5-Dimethylpyrazine	4.746	3.17	—	—	—
7	3,3,4-Trimethyldecane	4.802	0	2.49	—	—
8	2-Methylpentane-2,4-diamine	5.734	2.42	—	—	—
9	( <i>E</i> )- <i>N</i> -Butylidenebutan-1-amine	6.475	1.94	—	—	—
10	2-Phenyl- <i>N</i> -(propan-2-ylidene)ethanamine	8.38	—	—	1.2	—
11	Dibenzofuran	10.124	—	—	—	4.06
12	9 <i>H</i> -Fluorene	10.595	—	—	—	2.75
13	Octadecane	11.05	—	7.29	—	10.16
14	Pentadecane	11.052	—	—	15.6	—
15	Hexadecane	11.057	16.82	—	—	—
16	5-Methyl-dihydropyrimidine-2,4(1 <i>H</i> ,3 <i>H</i> )-dione	11.389	4	—	—	—
17	(7 <i>R</i> ,11 <i>R</i> , <i>E</i> )-3,7,11,15-Tetramethylhexadec-2-ene	11.918	—	2.39	8.75	—
18	9-Methylundec-1-ene	11.924	2.12	—	—	—
19	Dodecyl pentan-2-yl sulfite	12.527	—	6.49	—	—
20	Hexadecanoic acid	12.604	—	—	—	15.78
21	Tetradecanoic acid	12.605	7.38	—	10.19	—
22	(9 <i>Z</i> ,12 <i>Z</i> )-Octadeca-9,12-dienoic acid	13.769	—	8.64	—	7.87
23	( <i>E</i> )-Octadec-9-enoic acid	13.777	10.1	—	—	—
24	Hexadecanamide	14.143	—	9.54	5.3	12.85
25	Tetradecanamide	14.151	12.69	—	—	—
26	<i>N</i> -Methyldodecanamide	14.414	—	8.8	7.16	7.13
27	Tridecanamide	14.424	2.75	—	—	—
28	<i>N</i> -Methyltridecanamide	14.767	—	5.7	5.12	4.82
29	Methyl stearate	15.871	—	—	—	3.16
30	(12 <i>E</i> ,15 <i>E</i> )-Methyl octadeca-12,15-dienoate	15.876	—	2.88	—	—
31	2-(Hex-5-enyl)oxirane	15.892	3.89	—	—	—
32	<i>N</i> -Butylstearamide	16.233	—	—	—	5.72
33	<i>N,N</i> -Diethyldodecanamide	16.236	—	8.6	19.57	—
34	1-Morpholinodecan-1-one	17.711	—	17.25	—	12.08
35	1-Isothiocyano-3-methylhexane	17.732	5.89	—	—	—
36	1-(Pyrrolidin-1-yl)dodecan-1-one	19.158	—	5.69	4.77	3.41
37	Undecanal	21.125	—	—	—	4.68
38	<i>N,N</i> -Bis(2-hydroxyethyl)stearamide	21.126	—	6.43	8.06	—



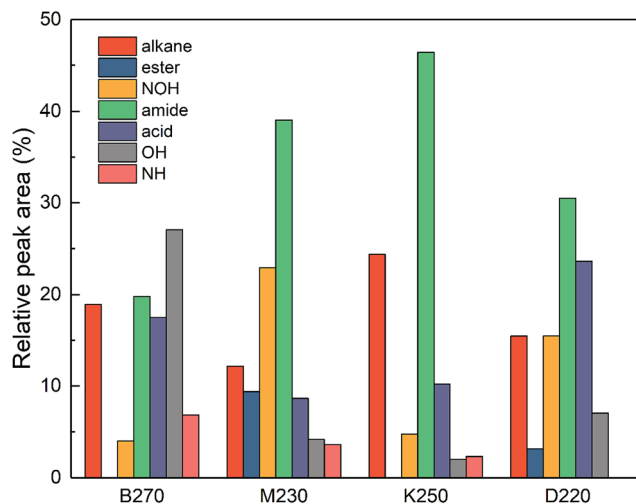


Fig. 3 Group fraction distribution of biocrude with the highest biocrude yield under different clay catalysts. B: blank experiment; M: montmorillonite catalyst; K: kaolin catalyst; D: dolomitic limestone catalyst. OH: oxygen-containing-only heterocyclic compound; NH: nitrogen-containing-only heterocyclic compound; NOH: oxygen-and-nitrogen-containing heterocyclic compound.

that the GC-MS analysis could only identify components which can vapor and pass through the GC column. Those high-boiling-point compounds like asphaltene or thermolabile components could not identify by GC-MS. To simplify the discussion, these identified compounds were classified into seven group: hydrocarbons, esters, fatty acids, fatty acid amides, oxygen-containing-only heterocyclic compounds (OH), nitrogen-containing-only heterocyclic compounds (NH) and oxygen-and-nitrogen-containing heterocyclic compounds (NOH). The group distribution of GC-MS identified compounds was presented in Fig. 3.

According to Fig. 3, adding clay catalysts increased the amide and NOH content while reduced the OH and NH content in biocrude. montmorillonite reduced the hydrocarbons and fatty acids content by 35.7% and 50.6%, respectively. Kaoline catalyst increased the hydrocarbon yield by 28.6% but decreased the acid yield by 41.7%. The dolomitic limestone catalytic liquefied more fatty acids into biocrude while avoiding the synthesis of alkanes and esters. The possible reaction pathways would contain the next few types:<sup>12,42–44</sup> (i) hydrolysis of lipid, protein, and carbohydrates, which explained the formation of fatty acid and other reaction intermediates such as amino acids and alcohols;<sup>6,45</sup> (ii) esterification, ammonolysis and decarboxylation of fatty acid, which could be the cause of esters, amides and alkanes;<sup>22,46,47</sup> (iii) annulation reaction from the hydrolysis product of carbohydrates and proteins by themselves or between them, that is why there were plenty of heterocyclic compounds in biocrude;<sup>48–50</sup> (iv) polymerization to product asphaltene or other macromolecule compounds.<sup>51–53</sup> It should be noted that there was a obvious competitive relation between the formation of esters, fatty acid amides, fatty acids, and some alkanes. These compounds all come from the fatty acids from the hydrolysis of lipid and the three types of clay catalysts

(montmorillonite, kaoline, and dolomitic limestone) played a different role in the formation of these four kinds of organic compounds. However, it is a pity that only part of the components in biocrude could be identified by GC-MS and current analysis could only provide an approximate composition of the biocrude. Further analysis based on high resolution mass spectrum should be taken in the further.

## 4. Conclusion

In this paper, three kinds of clay materials were introduced as the catalyst into the HTL of high protein microalgae *Spirulina*. Adding clay catalyst improved the liquefaction conversion and biocrude yield. Research suggested that operating parameters like reaction temperature and holding time were significant for HTL products of *Spirulina* with clay catalysts. The optimized operating conditions were a holding time of 0.5 h and an appropriate reaction temperature for the different catalyst (230, 250 and 220 °C for montmorillonite, kaoline and dolomitic limestone, respectively). The catalyst screening studies would help researchers to understand and improve the catalytic liquefaction of microalgae better.

## Conflicts of interest

There are no conflicts to declare.

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## References

- 1 C. Y. Tian, B. M. Li, Z. D. Liu, Y. H. Zhang and H. F. Lu, *Renewable Sustainable Energy Rev.*, 2014, **38**, 933–950.
- 2 A. Demirbas, *Energy Convers. Manage.*, 2010, **51**, 2738–2749.
- 3 L. Gouveia and A. C. Oliveira, *J. Ind. Microbiol. Biotechnol.*, 2009, **36**, 269–274.
- 4 L. Brennan and P. Owende, *Renewable Sustainable Energy Rev.*, 2010, **14**, 557–577.
- 5 A. Marcilla, L. Catala, J. C. Garcia-Quesada, F. J. Valdes and M. R. Hernandez, *Renewable Sustainable Energy Rev.*, 2013, **27**, 11–19.
- 6 A. A. Peterson, F. Vogel, R. P. Lachance, M. Froling, M. J. Antal and J. W. Tester, *Energy Environ. Sci.*, 2008, **1**, 32–65.
- 7 D. López Barreiro, S. Riede, U. Hornung, A. Kruse and W. Prins, *Algal Res.*, 2015, **12**, 206–212.
- 8 S. S. Toor, L. Rosendahl and A. Rudolf, *Energy*, 2011, **36**, 2328–2342.
- 9 Y. H. Zhu, K. O. Albrecht, D. C. Elliott, R. T. Hallen and S. B. Jones, *Algal Res.*, 2013, **2**, 455–464.
- 10 C. Torri, D. Fabbri, L. Garcia-Alba and D. W. F. Brilman, *J. Anal. Appl. Pyrolysis*, 2013, **101**, 28–34.



- 11 Y. Guo, T. Yeh, W. H. Song, D. H. Xu and S. Z. Wang, *Renewable Sustainable Energy Rev.*, 2015, **48**, 776–790.
- 12 Y. Chen, Y. L. Wu, D. R. Hua, C. Li, M. P. Harold, J. L. Wang and M. D. Yang, *RSC Adv.*, 2015, **5**, 18673–18701.
- 13 K. Kumar, S. Ghosh, I. Angelidaki, S. L. Holdt, D. B. Karakashev, M. A. Morales and D. Das, *Renewable Sustainable Energy Rev.*, 2016, **65**, 235–249.
- 14 S. Arvindnarayan, K. K. S. Prabhu, S. Shobana, G. Kumar and J. Dharmaraja, *Int. Biodeterior. Biodegrad.*, 2017, **119**, 260–272.
- 15 C. Y. Yang, R. Li, C. Cui, S. P. Liu, Q. Qiu, Y. G. Ding, Y. X. Wu and B. Zhang, *Green Chem.*, 2016, **18**, 3684–3699.
- 16 Z. Chang, P. Duan and Y. Xu, *Bioresour. Technol.*, 2015, **184**, 349–354.
- 17 L. Gao, J. Sun, W. Xu and G. Xiao, *Bioresour. Technol.*, 2017, **225**, 293–298.
- 18 D. L. Barreiro, B. R. Gomez, F. Ronsse, U. Hornung, A. Kruse and W. Prins, *Fuel Process. Technol.*, 2016, **148**, 117–127.
- 19 P. Biller, R. Riley and A. B. Ross, *Bioresour. Technol.*, 2011, **102**, 4841–4848.
- 20 P. G. Duan and P. E. Savage, *Ind. Eng. Chem. Res.*, 2011, **50**, 52–61.
- 21 G. W. Roberts, M. O. P. Fortier, B. S. M. Sturm and S. M. Stagg-Williams, *Energy Fuels*, 2013, **27**, 857–867.
- 22 J. Zhang, W. T. Chen, P. Zhang, Z. Luo and Y. Zhang, *Bioresour. Technol.*, 2013, **133**, 389–397.
- 23 K. A. Kvenvolden, *Org. Geochem.*, 2006, **37**, 1–11.
- 24 G. P. Glasby, *Resour. Geol.*, 2006, **56**, 83–96.
- 25 V. Simanzhenkov and R. Idem, *Crude oil chemistry*, CRC Press, 2003.
- 26 T. Gold, *Annu. Rev. Energy*, 1985, **10**, 53–77.
- 27 S. R. Hart, *Nature*, 1984, **309**, 753–757.
- 28 H. Zeb, J. Choi, Y. Kim and J. Kim, *Energy*, 2017, **118**, 116–126.
- 29 S. Yokoyama, A. Suzuki, M. Murakami, T. Ogi, K. Koguchi and E. Nakamura, *Fuel*, 1987, **66**, 1150–1155.
- 30 K. P. R. Dandamudi, T. Muppaneni, N. Sudasinghe, T. Schaub, F. O. Holguin, P. J. Lammers and S. Deng, *Bioresour. Technol.*, 2017, **236**, 129–137.
- 31 P. Biller and A. B. Ross, *Bioresour. Technol.*, 2011, **102**, 215–225.
- 32 P. J. Valdez, V. J. Tocco and P. E. Savage, *Bioresour. Technol.*, 2014, **163**, 123–127.
- 33 S. S. Toor, H. Reddy, S. Deng, J. Hoffmann, D. Spangsmark, L. B. Madsen, J. B. Holm-Nielsen and L. A. Rosendahl, *Bioresour. Technol.*, 2013, **131**, 413–419.
- 34 K. Anastasakis and A. B. Ross, *Bioresour. Technol.*, 2011, **102**, 4876–4883.
- 35 P. Fongarland, N. Essayem and F. Rataboul, *Ind. Eng. Chem. Res.*, 2017, **56**, 126–134.
- 36 Y. Xue, H. Y. Chen, W. N. Zhao, C. Yang, P. Ma and S. Han, *Int. J. Energy Res.*, 2016, **40**, 865–877.
- 37 C. Xu and J. Lancaster, *Water Res.*, 2008, **42**, 1571–1582.
- 38 R. Singh, K. Chaudhary, B. Biswas, B. Balagurumurthy and T. Bhaskar, *J. Supercrit. Fluids*, 2015, **104**, 70–75.
- 39 T. M. Brown, P. G. Duan and P. E. Savage, *Energy Fuels*, 2010, **24**, 3639–3646.
- 40 Y. F. Yang, C. P. Feng, Y. Inamori and T. Maekawa, *Resour., Conserv. Recycl.*, 2004, **43**, 21–33.
- 41 F. Wang, Z. Chang, P. Duan, W. Yan, Y. Xu, L. Zhang, J. Miao and Y. Fan, *Bioresour. Technol.*, 2013, **149**, 509–515.
- 42 C. Torri, L. G. Alba, C. Samori, D. Fabbri and D. W. F. Brilman, *Energy Fuels*, 2012, **26**, 658–671.
- 43 A. Demirbas, *Energy Convers. Manage.*, 2000, **41**, 633–646.
- 44 M. Balat, *Energy Sources, Part A*, 2008, **30**, 649–659.
- 45 S. M. Changi, J. L. Faeth, N. Mo and P. E. Savage, *Ind. Eng. Chem. Res.*, 2015, **54**, 11733–11758.
- 46 Y. Chen, Y. Wu, P. Zhang, D. Hua, M. Yang, C. Li, Z. Chen and J. Liu, *Bioresour. Technol.*, 2012, **124**, 190–198.
- 47 D. Zhou, L. A. Zhang, S. C. Zhang, H. B. Fu and J. M. Chen, *Energy Fuels*, 2010, **24**, 4054–4061.
- 48 A. Croce, E. Battistel, S. Chiaberge, S. Spera, F. De Angelis and S. Reale, *Chemsuschem*, 2017, **10**, 171–181.
- 49 C. Zhang, X. H. Tang, L. L. Sheng and X. Y. Yang, *Green Chem.*, 2016, **18**, 2542–2553.
- 50 A. A. Peterson, R. P. Lachance and J. W. Tester, *Ind. Eng. Chem. Res.*, 2010, **49**, 2107–2117.
- 51 J. Y. Chu, W. K. Jiang, S. B. Wu, L. A. Lucia and M. Lei, *Energy Fuels*, 2017, **31**, 1638–1643.
- 52 F. A. Atiku, K. D. Bartle, J. M. Jones, A. R. Lea-Langton and A. Williams, *Fuel*, 2016, **182**, 517–524.
- 53 L. Li, Q. You, S. Y. Wu, S. Huang, Y. Q. Wu and J. S. Gao, *J. Energy Inst.*, 2016, **89**, 335–345.

