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Effect of borrelidin on hepatocellular carcinoma cells *in vitro* and *in vivo*†

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Hepatocellular carcinoma (HCC) is the most common type of liver cancer with high prevalence and mortality. Borrelidin, produced by several actinomycete bacteria of *Streptomyces* sp. exhibited diversiform activities including anti-bacterial, anti-viral, anti-angiogenic, and anti-tumor. However, the effect of borrelidin on HCC cells has not been characterized. The present study demonstrated borrelidin exhibited great potential to inhibit the growth of HCC cells, HepG2 and SMMC7721 *in vitro*. Western blot and real-time qPCR analysis revealed that borrelidin decreased the expressions of cyclin D1, cyclin D3, cyclin E1, CDK2, CDK4, and CDK6 and increased the expression of p21, thereby inducing G0/G1 cell cycle arrest. Moreover, borrelidin down-regulated expression of Bcl-2, up-regulated expression of Bax and increased cleavages of caspase-9 and caspase-3 to activate caspase-dependent apoptosis in HCC cells. Borrelidin inhibited migration and invasion through suppressing the expression of MMP-2 and MMP-9 in HCC cells. Further investigation indicated that the anti-tumor effect of borrelidin was mediated by MAPKs signaling pathway. In addition, an *in vivo* experiment revealed that borrelidin suppressed tumor growth in SMMC7721 xenograft model mice with few side effects. Cell cycle arrest and induced apoptosis were also observed in tumor tissues of model mice treated with borrelidin.

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1. Introduction

Hepatocellular carcinoma (HCC) is one type of liver cancer with a high incidence and mortality rate worldwide.¹ Despite tremendous progress in diagnosis and management, the present therapeutic strategies for treating HCC are still surgical resection or liver transplantation.² Moreover, HCC is highly resistant to conventional chemotherapeutic drugs such as anthracycline, fluoropyrimidine, camptothecin and platinum-based drugs.^{3,4} As a consequence, the development of new anti-tumor drugs is essential for improving the treatment of hepatic carcinoma.

In the process of our search for new bioactive natural products from actinomycetes associated with animal feces, the secondary metabolites of the fermentation broth of *Streptomyces vinaceusdrappus* (YIM 100880), which was isolated from *Hylobates hoolock* feces, were investigated. Borrelidin, a nitrile-

containing macrolide, was isolated from the fermentation broth of YIM 100880 by several chromatographic techniques and the structure was elucidated by ¹H-NMR, ¹³C-NMR, and MS spectroscopic data analysis. Borrelidin exhibited a wide spectrum of biological activities, such as anti-bacterial, anti-viral, and anti-tumor.⁵ It was proved that some of these activities were associated with the inhibition of threonyl-rRNA synthetase (ThrRS).⁶ Previous study had shown that borrelidin induced cell cycle arrest and apoptosis with activation of the GCN2 kinase pathway in acute lymphoblastic leukemia cells.⁷ Borrelidin also induced chop-dependent apoptosis through unfolded protein response (UPR) in oral cancer cell.⁸ In addition, borrelidin was an effective angiogenesis inhibitor through its inhibition of ThrRS and inducing caspases cascade in endothelial cells.⁹ The remarkable bioactivities have prompted the development of borrelidin as a potential anti-tumor candidate.

Currently, the effect of borrelidin on HCC remains unknown. In the present study, our investigation was designed to explore the effect of borrelidin on proliferation, cell cycle arrest, apoptosis, migration and invasion of two HCC cells, HepG2 and SMMC7721 *in vitro* and xenograft growth inhibition *in vivo*. Real-time qPCR and western blot analysis of molecular markers associated with cell cycle arrest and apoptosis as well as MAPKs cell signaling cascade were performed to identify the underlying mechanisms.

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† Electronic supplementary information (ESI) available: ¹H-NMR, ¹³C-NMR, ESI-MS spectra and HPLC chromatogram of borrelidin, and the relative quantitative analysis of related proteins assessed by western blot in Fig. 3(C and D), 4(E and F), 5(C), and 6. See DOI: 10.1039/c7ra08290h



2. Results and discussion

2.1. Borrelidin inhibited proliferation of HCC cells

Borrelidin was isolated by our co-author and provided with a purity of >98.0% determined by high-pressure liquid chromatography.

The effects of borrelidin on the growth inhibition of two HCC cells, HepG2 and SMMC7721 were assessed by MTT assay. The results showed that borrelidin time- and dose-dependently inhibited the growth and proliferation of the two HCC cells (Fig. 1). Half inhibitory concentrations (IC_{50}) of borrelidin in HepG2 and SMMC7721 cells were 6.7 ± 0.3 and $1.5 \pm 0.2 \mu\text{M}$, respectively. According to the results, the concentrations below IC_{50} were used to conduct following-up studies.

2.2. Borrelidin caused HCC cells G0/G1 phase arrest

Oncogenic transformation or loss of tumor suppressor function drives abnormal growth and proliferation in cancer cells, which often results from uncontrolled cell cycle.¹⁰ In order to explore the effect of borrelidin on the cell cycle progression of HepG2 and SMMC7721 cells, the cells were incubated with borrelidin and then followed by flow cytometric analysis to examine the DNA distribution. The results suggested that borrelidin caused a G0/G1 phase cell cycle arrest in HCC cells (Fig. 2). Cell cycle progression was known to be tightly regulated by different cyclins, cyclin dependent kinases (CDKs) and cyclin dependent kinase inhibitors (CKIs) in different phase of cell cycle.¹¹ Among of cyclins, cyclin D and cyclin E, played important roles in the G1/S phase progression of cells. Cyclin D interacted with CDK4 and CDK6 to allow expression of cyclin E which bound and activated CDK2 to form CDK2/cyclin E complex, thus promoting cell cycle progression from G1 to S phase.

On the other hand, the activities of cyclins/CDKs complex were negatively regulated by CKIs, especially p21.^{11,12} The disorders of these biomarkers would result in the G0/G1 phase cell cycle arrest in cells.¹³ To disclose the underlying mechanism, we focused on the genes related to cell cycle progression by using real-time qPCR. As shown, borrelidin decreased the

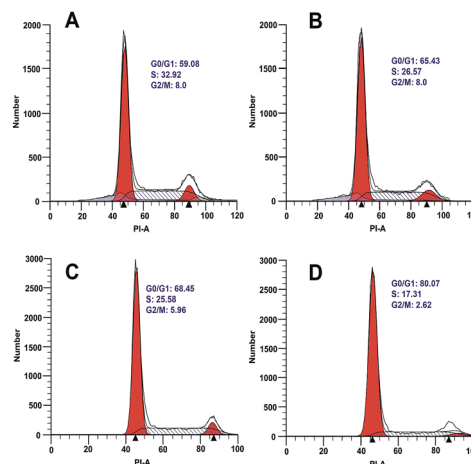


Fig. 2 Measurement of the cell cycle in HCC cells treated with borrelidin. Flow cytometric analysis on cell cycle arrest of HepG2 cells treated with (A) vehicle, (B) 5.0 μM borrelidin for 24 h, respectively. SMMC7721 cells treated with (C) vehicle, (D) 1.0 μM borrelidin for 24 h, respectively. The vehicle-treated group was the control (Con).

mRNA expressions of *cyclin D1*, *cyclin D3*, *cyclin E1*, *CDK2*, *CDK4* and *CDK6* and increased p21 (Fig. 3A and B). We further analyzed the protein expressions of these molecules by western

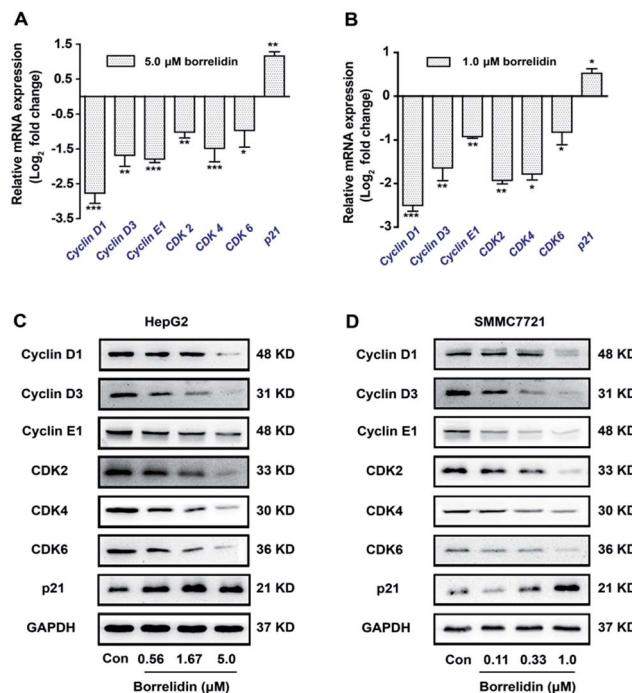


Fig. 3 Regulation of borrelidin on G0/G1 phase related molecules. The mRNA levels of *cyclin D1*, *cyclin D3*, *cyclin E1*, *CDK2*, *CDK4*, *CDK6* and *p21* of HepG2 cells (A) and SMMC7721 cells (B) treated with borrelidin were quantified by real-time qPCR. The protein levels of cyclin D1, cyclin D3, cyclin E1, CDK2, CDK4, CDK6 and p21 of HepG2 cells (C) and SMMC7721 cells (D) treated with borrelidin were assessed by western blot. The control (Con) group was treated with vehicle. Data were represented as means \pm SD ($n = 3$). * $p < 0.05$ vs. control; ** $p < 0.01$ vs. control; *** $p < 0.001$ vs. control.

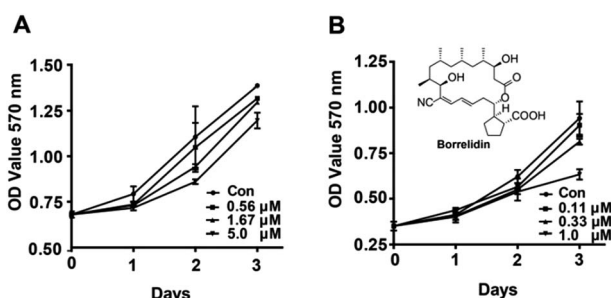


Fig. 1 Effect of borrelidin on growth and proliferation of HCC cells. The growth curve of HCC cells was determined by MTT assay (A) HepG2 cells were treated with borrelidin (0, 0.56, 1.67 and 5.0 μM) for 0 h, 24 h, 48 h and 72 h. (B) SMMC7721 cells were treated with borrelidin (0, 0.11, 0.33 and 1.0 μM) for 0 h, 24 h, 48 h and 72 h. The control (Con) group was treated with vehicle. Data were represented as means \pm SD ($n = 3$).



blot assay and the up-regulations and down-regulations of them in a dose-dependent manner were consistent with their transcriptional target genes (Fig. 3C and D). In summary, borrelidin caused G0/G1 phase cell cycle arrest in HCC cells through down-regulating cyclins and CDKs and enhancing p21 expression in cell cycle progression.

2.3. Borrelidin induced HCC cells apoptosis

Precious studies had shown that borrelidin could induce apoptosis in various cancer cells, including breast cancer, oral cancer and acute lymphoblastic leukemia cells.^{7,8,14} To observe

whether borrelidin could induce apoptosis of HCC cells, Hoechst 33258 staining was conducted in HepG2 and SMMC7721 cells with borrelidin treatment. As shown in Fig. 4C, borrelidin-treated group exhibited typical morphological features of apoptosis including shrinking nuclei and dense fluorescence emission in contrast with control group. Besides, flow cytometric analysis of annexin V/PI showed that significant increase in population of apoptotic cells were detected in borrelidin-treated HepG2 and SMMC7721 cells (Fig. 4A and B). The anti-apoptotic protein Bcl-2 and the pro-apoptotic protein Bax, were the well-characterized members of the Bcl-2 family that regulated cell apoptosis.¹⁵ Moreover, the activation of

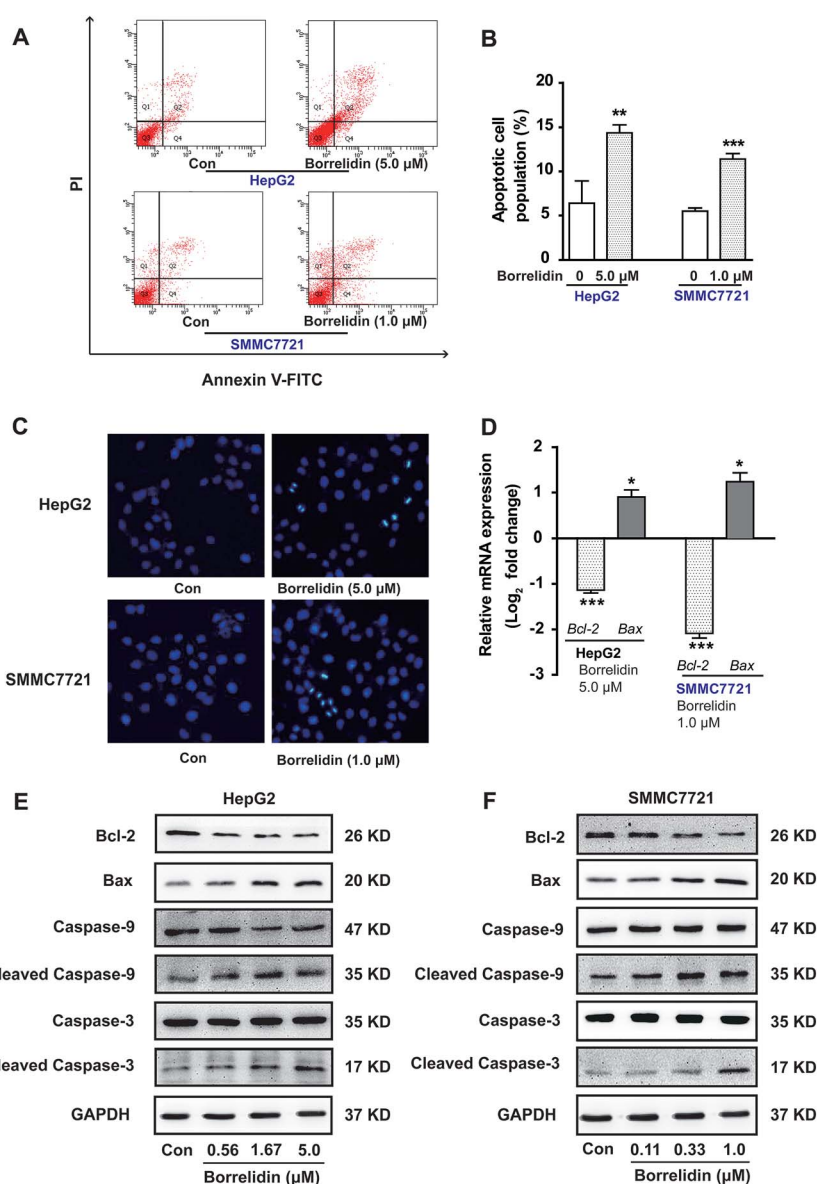


Fig. 4 Effect of borrelidin on inducing apoptosis of HCC cells. (A) Flow cytometric analysis on apoptosis of HepG2 and SMMC7721 cells treated with borrelidin. (B) The percentages of annexin V-positive cells of HepG2 and SMMC7721 cells treated with borrelidin. (C) Morphological alterations of HepG2 and SMMC7721 cells treated with borrelidin stained with Hoechst 33258. (D) The mRNA levels of *Bcl-2*, *Bax* of borrelidin treated HepG2 cells and SMMC7721 cells were quantified by real-time qPCR. The protein levels of Bcl-2, Bax, caspase-3, cleaved caspase-3, caspase-9 and cleaved caspase-9 of HepG2 cells (E) and SMMC7721 cells (F) treated with borrelidin were assessed by western blot. The vehicle-treated group was the control (Con). Data were represented as means \pm SD ($n = 3$). * $p < 0.05$ vs. control; ** $p < 0.01$ vs. control; *** $p < 0.001$ vs. control.



caspases cascade played a dominant role in the induction of cell apoptosis.¹⁶ Caspase-9, an initiator caspase, was cleaved into the active form and then activated caspases-3 by proteolytic cleavage, subsequently induced the digestion of structural proteins, DNA degradation and ultimately phagocytosis.^{16–18} To further explore the effects of borrelidin on the apoptosis-related molecules, the mRNA and protein expression of Bcl-2, Bax, caspase-3 and caspase-9 cleavages in two HCC cells were investigated by real-time qPCR or western blot assay. As shown, the expression of Bcl-2 was significantly down-regulated, while Bax up-regulated in HepG2 and SMMC7721 cells after borrelidin treatment (Fig. 4D–F). Western blot analysis demonstrated that the protein levels of cleaved caspase-9 and cleaved caspase-3 were significantly increased in a dose-dependent manner in the borrelidin treated group (Fig. 4E and F). We concluded that borrelidin could induce apoptosis through unbalance of Bcl-2 family and activation of caspase family in HCC cells.

2.4. Borrelidin inhibited the migratory and invasive ability of HCC cells

The progression of metastasis exacerbated the probability of morbidity and mortality in cancer populations.¹⁹ Cell

migration and invasion were critical steps in the process of metastasis of cancer cells. In the current study, the effect of borrelidin on the migration and invasion of HCC cells were investigated. Firstly, a wound healing assay was conducted to detect cell mobility after borrelidin treatment. The results showed that borrelidin significantly decreased the movement ability of HepG2 and SMMC7721 cells in a dose-dependent manner (Fig. 5A). Then, the invasive and migratory behavior of borrelidin treated HepG2 and SMMC7721 cells were analyzed using Boyden chamber assays with or without Matrigel. The results demonstrated that inhibition occurred in a dose-dependent manner compared to control group (Fig. 5B). Matrix metalloproteinases (MMPs) exhibited the ability to degrade the extracellular matrix (ECM) and contributed to the invasion of cancer cells.²⁰ Among them, MMP-2 and MMP-9 were crucial for degrading the main component of ECM.^{21–23} In order to investigate the effect of borrelidin on MMP-2 and MMP-9 expressions in HCC cells, we detected the intracellular protein levels of MMP-2 and MMP-9 of HepG2 and SMMC7721 cells by western blot analysis. As shown, the protein expressions of MMP-2 and MMP-9 were down-regulated in a dose-dependent manner (Fig. 5C).

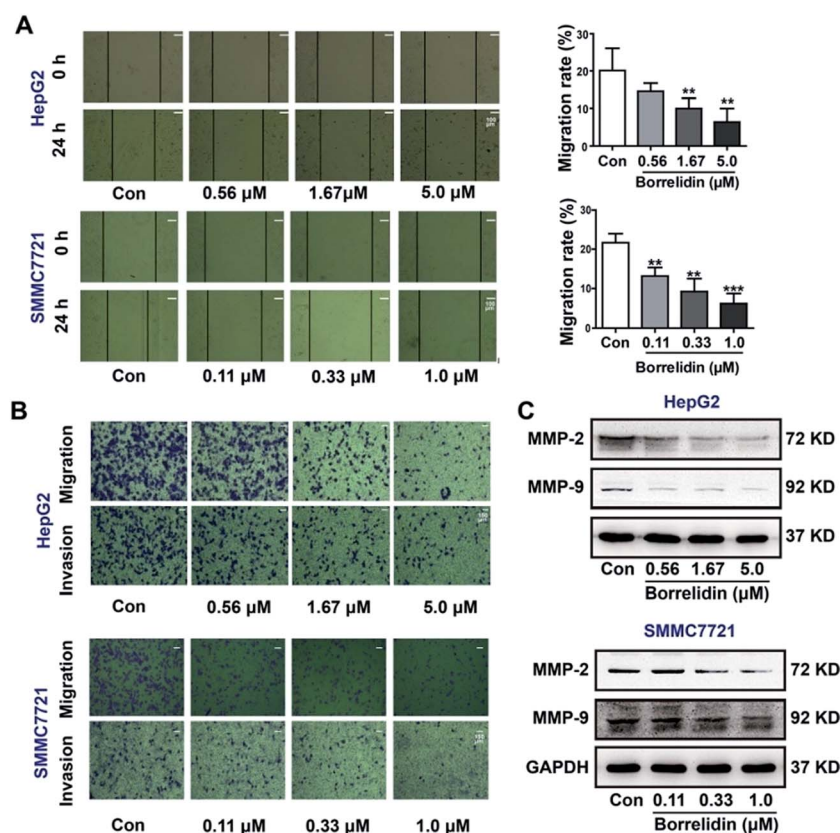


Fig. 5 Borrelidin inhibited the migration and invasion potential of HCC cells. (A) The wound healing assay for evaluating the effect of borrelidin on mobility of HepG2 cells and SMMC7721 cells. The average scraped width per well at 0 h and 24 h was measured and calculated as migration rate. (B) The transwell migration assay with or without Matrigel for assessing the effect of borrelidin on invasive or migratory potential of HepG2 cells and SMMC7721 cells. (C) The intracellular MMP-2 and MMP-9 protein expression levels of HepG2 and SMMC7721 cells treated with borrelidin were assessed by western blot. The vehicle-treated group was the control (Con). Data were represented as means \pm SD ($n = 3$). * $p < 0.05$ vs. control; ** $p < 0.01$ vs. control; *** $p < 0.001$ vs. control.



2.5. MAPKs signaling pathway was regulated by borrelidin in HCC cells

MAPKs family proteins, including the p38, extracellular signal-regulated kinases 1/2 (ERK1/2), and c-Jun amino (N)-terminal kinases 1/2 (JNK), played essential roles in anti-tumor agent-induced apoptosis and cell cycle arrest.^{24–27} JNK and p38 kinase could distort the balance of Bcl-2 family proteins, leading to cell apoptosis.²⁸ The activation of ERK signaling pathway was responsible for the cleavage of caspases.^{29,30} Besides, activation of ERK could increase levels of p21 and p27 as well as reduce cyclin D and cyclin E expressions to cause cell cycle arrest.^{31,32} To investigate the effect of borrelidin on MAPKs signaling pathway, we performed western blot analysis to investigate the expressions of p38, ERK1/2 and JNK in the borrelidin treated cells. As shown in Fig. 6, phosphorylation levels of p38, ERK1/2 and JNK were obviously increased in borrelidin-treated HCC cells in a dose-dependent manner. The results indicated that the anti-tumor effect of borrelidin on HCC cells was related to up-regulating MAPKs signaling pathway.

2.6. Borrelidin inhibited SMMC7721 cells xenograft growth *in vivo*

To investigate effect of borrelidin against HCC cells *in vivo*, a human hepatoma SMMC7721 xenograft model was established. As shown, borrelidin significantly inhibited the tumor growth compared to the control group (Fig. 7A and B). Besides, histological studies on tumor tissue sections with H&E staining showed that borrelidin changed the histology of human tumor xenografts in nude mice (Fig. 7C). Tumor tissues in the control group grew actively with deep-dyed big nucleolus. While tumor in the borrelidin treatment group showed distinct pathological features including fewer karyokinetic events, loss of cell-cell contacts and cell necrosis.

Based on the study *in vitro*, borrelidin inhibited the growth of SMMC7721 cells through causing cell cycle arrest and inducing apoptosis. Therefore, we performed western blot to verify the mechanism *in vivo*. As expected, the protein expressions of

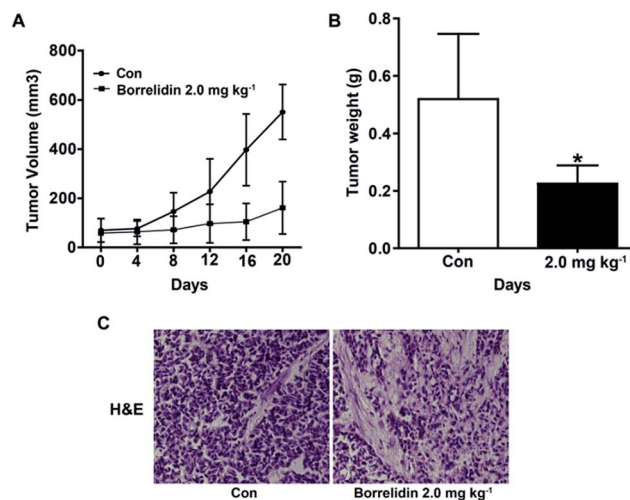


Fig. 7 The inhibitory effect of borrelidin on tumor growth in SMMC7721 cell xenograft model. (A) Tumor volumes were measured at the indicated days. (B) Tumor weight were recorded after mice were sacrificed. (C) Representative micrographs of tumor section of H&E staining. The vehicle-treated group was the control (Con). Data were represented as means \pm SD ($n = 5$). Statistical differences were considered significantly at the level of $*p < 0.05$, vs. control.

cyclin D3, cyclin E1 were reduced and p21 was increased, and cleavage of caspase-3 was activated in the tissues treated with borrelidin (Fig. 8A). Furthermore, TUNEL assay was performed

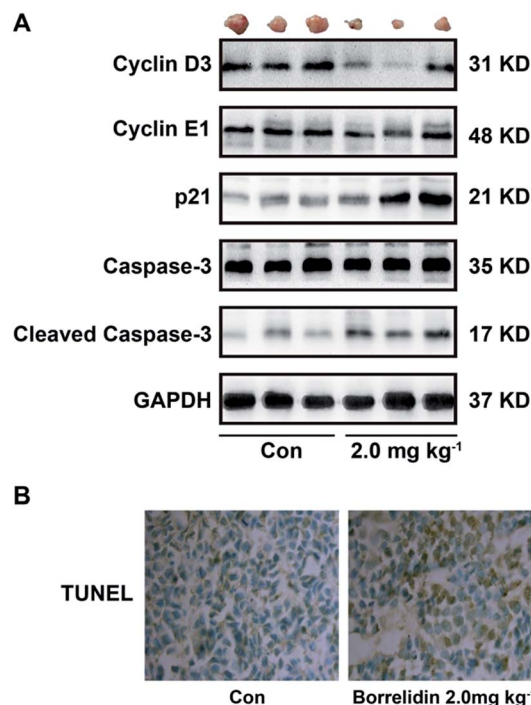


Fig. 8 Borrelidin caused cell cycle arrest and induced apoptosis in SMMC7721 cell xenograft model. (A) Protein expressions of cyclin D3, cyclin E1, p21, caspase-3 and cleaved caspase-3 of tumor tissues treated with borrelidin (0 and 2 mg kg⁻¹) were assessed by western blot. (B) TUNEL staining of representative micrographs of tumor section treated with borrelidin (0 and 2 mg kg⁻¹). The vehicle-treated group was the control (Con).

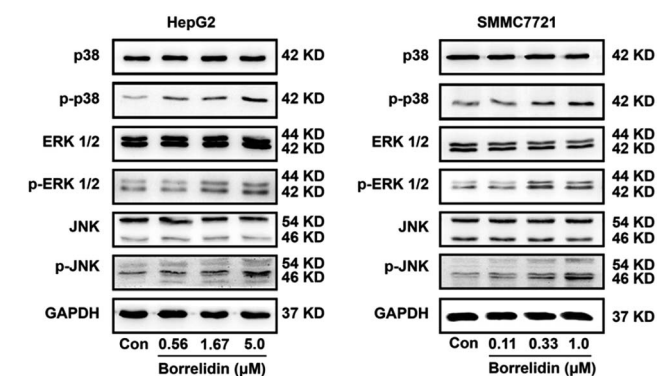


Fig. 6 Up-regulations of MAPKs pathway modulated cell cycle and apoptosis of HCC cells treated with borrelidin. The protein expression of p38, p-p38, ERK1/2, p-ERK1/2, JNK and p-JNK of HepG2 cells (A) and SMMC7721 cells (B) treated with borrelidin were assessed by western blot. The vehicle-treated group was the control (Con).



Table 1 Physiological and biochemical index of xenograft nude mice

	Body weight (g)	Liver index (mg g ⁻¹)	Spleen index (mg g ⁻¹)	AST (karU)	ALT (karU)
Con	25.1 ± 1.0	53.7 ± 3.0	7.5 ± 1.7	97.2 ± 4.7	76.2 ± 3.6
Borrelidin (2 mg kg ⁻¹)	22.7 ± 2.5	54.9 ± 1.8	7.2 ± 1.8	97.9 ± 4.2	75.9 ± 2.9

in the xenografted tissues. Compared to vehicle treatment, the borrelidin treated tissues exhibited more TUNEL-positive cells, which demonstrated that borrelidin caused cell apoptosis accompanied by DNA breaks (Fig. 8B). Besides, there were no significant changes in body weight, liver index, spleen index, serum activities of AST and ALT between vehicle and borrelidin-treated group, which indicated that borrelidin did not cause severe toxicity in mice (Table 1).

3. Conclusions

Taken together, borrelidin caused G0/G1 cell cycle arrest, induced caspase-dependent apoptosis and inhibited the growth of HCC cells through up-regulating MAPKs signaling pathway. Moreover, it could attenuate migration and metastasis behavior of HCC cells. Borrelidin may serve as a potential lead for development of anti-tumor agent for HCC treatment.

4. Experimental

4.1. General experimental procedures

NMR spectra were collected on a Bruker AV-600 spectrometer, δ in ppm rel. to TMS, J in Hz. ESIMS were recorded on an Agilent 1290-6420 Triple Quadrupole LC-MS spectrometer. Silica gel (100–200 mesh, 200–300 mesh, Qingdao Marine Chemical Ltd., Qingdao, China), Sephadex LH-20 (GE Healthcare Bio-sciences AB, Uppsala, Sweden), YMC*GEL ODS-A (S-50 μ m, 12 nm) (YMC Co., Ltd, Kyoto, Japan) were used for column chromatography. Flow cytometric analysis was used a LSR-Fortessa flow cytometer (BD, Franklin Lakes, NJ, USA). MTT, protein quantification and serum analysis were analyzed using a microplate reader (BioTek Synergy H1, BioTek Instruments, Inc., Vermont, USA). Human hepatocellular carcinoma cell lines HepG2 and SMMC7721 were purchased from Chinese Academy of Sciences (Shanghai, China). Fetal bovine serum (FBS), L-glutamine and penicillin-streptomycin were purchased from Gibco BRL Co., Ltd. (Gaithersburg, MD, USA). Antibodies of cyclin D1, cyclin D3, cyclin E1, CDK2, CDK4, CDK6, p21, Bax, Bcl-2, caspase-3, caspase-9, MMP-2, MMP-9, p38, p-p38 (Thr180/Tyr182), ERK1/2, p-ERK1/2 (Thr202/Tyr204), JNK, p-JNK (Thr183/Tyr185) and GAPDH were purchased from Cell Signaling Technology (Dancers, MA, USA).

4.2. Microbial material

The organism for producing borrelidin was isolated from fresh fecal samples excreted by healthy adult *Hylobates hooleck* living in Yunnan Wild Animal Park, Kunming, Yunnan province, in October 2013. The strain was identified as *Streptomyces*

vinaceusdrappus by Dr Xiu Chen based on morphological characteristics and 16S rRNA gene sequences. The BLAST result showed that the sequence was most similar (100.0%) to the sequence of *S. vinaceusdrappus* (strain: NBRC 13099^T, GenBank accession no. AB184311). The strain (no. YIM 100880) was deposited at the Yunnan Institute of Microbiology, Yunnan University, China.

4.3. Fermentation for *Streptomyces vinaceusdrappus*

A slant culture of the strain was used to inoculate 500 mL Erlenmeyer flasks containing 100 mL of seed medium composed of 4 g L⁻¹ yeast extract, 4 g L⁻¹ glucose, 5 g L⁻¹ malt extract, 1.0 mL multiple vitamin solution, and 1.0 mL trace element solution at a pH of 7.2 with no adjustment. The flasks were cultured for 2 days at 28 °C on a rotary shaker at 180 rpm. This seed culture was used to inoculate fermentation medium with a 10% volume. The fermentation was carried out in a 500 mL Erlenmeyer flask containing 100 mL of fermentation medium composed of 10 g L⁻¹ soybean meal, 2 g L⁻¹ peptone, 5 g L⁻¹ glucose, and 5 g L⁻¹ soluble starch at a pH of 7.0 with no adjustment, and it was incubated for 7 days at 28 °C on a rotary shaker at 180 rpm.

4.4. Extraction, isolation and identification of borrelidin

The completed fermentation broth (80 L) was separated into filtrate and mycelium by centrifugation. The filtrate was extracted with EtOAc three times. The combined EtOAc extract was subjected to open silica gel (100–200 mesh) column chromatography eluting with a CH₂Cl₂–MeOH solvent system (from 100 : 1 to 1 : 1, and MeOH at last) to yield eight fractions, Fr.1–Fr.8. Fraction 4 (4.5 g) was subjected to Sephadex LH-20 chromatography (MeOH) to produce 9 subfractions (Fr.4.1–Fr.4.9). Fr.4.4 (0.3 g) and Fr.4.5 (2.0 g) containing borrelidin were collected and combined on the basis of their TLC profile (CH₂Cl₂–MeOH = 15 : 1, R_f = 0.48) and further separated by ODS column chromatography, eluting with water–methanol (25 : 75) to give a purified borrelidin (about 1.25 g).

Borrelidin: white amorphous power, $[\alpha]_D^{25} = -12.53^\circ$ (c 0.76, MeOH); ESI-MS m/z : 490 $[M + H]^+$, 512 $[M + Na]^+$. ¹H-NMR (600 MHz, DMSO- d_6) δ (ppm): 6.95 (1H, d, J = 11.4 Hz), 6.46 (1H, brt, J = 13.0 Hz), 6.31 (1H, ddd, J = 14.7, 10.5, 4.3 Hz), 4.88 (1H, dt, J = 10.7, 3.6 Hz), 4.06 (1H, d, J = 9.6 Hz), 3.75 (1H, dt, J = 9.8, 3.0 Hz), 2.47–2.54 (2H, m), 2.37 (1H, ddd, J = 14.9, 10.7, 4.1 Hz), 2.29 (1H, q, J = 7.9 Hz), 2.25 (1H, dd, J = 15.4, 3.2 Hz), 2.07 (1H, dd, J = 15.4, 9.8 Hz), 1.84–1.90 (2H, m), 1.80 (1H, m), 1.60–1.73 (5H, m), 1.55 (1H, m), 1.35 (1H, m), 1.10 (2H, m), 1.00 (1H, brt, J = 13.0 Hz), 0.93 (1H, m), 0.92 (3H, d, J = 6.4 Hz, CH₃), 0.86 (1H, brt, J = 11.1 Hz), 0.78 (3H, d, J = 6.4 Hz, CH₃), 0.77 (3H, d, J =



6.4 Hz, CH₃), 0.74 (3H, d, $J = 7.2$ Hz, CH₃), 0.60 (1H, brt, $J = 11.5$ Hz); ¹³C-NMR (150 MHz, DMSO-*d*₆) δ (ppm): 177.6, 171.3, 143.6, 139.2, 127.8, 119.5, 116.8, 75.3, 71.0, 70.6, 48.5, 48.1, 45.9, 43.4, 37.6, 37.4, 35.9, 35.7, 34.8, 31.2, 29.3, 27.3, 26.4, 25.2, 20.8, 18.8, 18.6, 15.4. The spectral properties were identical with those reported in the literature.^{33,34}

4.5. Cell culture

Human hepatocellular carcinoma cell lines HepG2 and SMMC7721 were cultured in DMEM and RPMI-1640 culture medium, respectively, with 10% fetal bovine serum (FBS), 1% L-glutamine, 100 units per mL penicillin and 100 $\mu\text{g mL}^{-1}$ streptomycin at 37 °C in a humidified 5% CO₂ atmosphere incubator (Thermo Fisher Scientific, Waltham, MA, USA).

4.6. MTT assay

The cytotoxicity and proliferation inhibition were quantified by the MTT assay. HepG2 and SMMC7721 cells (7×10^3 cells per well) were seeded in 96-well plates respectively and exposed to borrelidin at six concentrations (0.33, 1.0, 3.3, 10.0, 33.0 and 100.0 μM). After 48 h incubation at 37 °C, 20 μL MTT was added into each well and incubated another 4 h. And then the liquid in the wells was removed. After that, 150 μL DMSO was added into each well. Use a microplate reader at 570 nm to measure the absorbance of each well. The results were represented as the mean \pm SD from three independent experiments. The inhibition rate (%) = $[(A_{570}(\text{control}) - A_{570}(\text{borrelidin})) / A_{570}(\text{control})] \times 100\%$ and the IC₅₀ values were calculated by a non-linear regression analysis using SPSS 13.0 (SPSS Inc., Chicago, IL, USA).

4.7. Measurement of cell cycle arrest

HepG2 and SMMC7721 cells (5×10^5 cells per well) were seeded in 6-well plates with 5.0 μM and 1.0 μM of borrelidin treatment for 24 h, respectively, then trypsinized, centrifuged and washed with cold PBS, and fixed with 75% ethanol left overnight at -20 °C. Following adequate washing with PBS, the fixed cells were incubated with PI working fluid (Dingguo Changsheng Biotechnology Co. Ltd., Beijing, China) for 30 min in the dark at room temperature. Samples were immediately run on a LSR-Fortessa flow cytometer with a total of 20 000 cells collected and analyzed with the PI-based cell cycle phase. The results were analyzed using ModFit LT 4.1 (Verity Software House, Topsham, ME, USA) and represented as the mean \pm SD from three independent experiments.

4.8. Hoechst 33258 staining

Cells stained Hoechst 33258 (Beyotime Biotechnology Co. Ltd, Shanghai, China) was used to evaluate the morphological changes. HepG2 and SMMC7721 cells (10^5 cells per well) were seeded in 6-well plates and treated with 5.0 μM and 1.0 μM of borrelidin for 24 h, respectively. Then cells were fixed with paraformaldehyde for 5 min, washed three times with PBS and stained with 500 μL Hoechst 33258 for 5 min in the dark at room temperature. Morphological changes were evaluated and

photographed by fluorescence microscope (DMi8, Leica, Heidelberg, Germany).

4.9. Measurement of apoptosis

HepG2 and SMMC7721 cells (5×10^5 cells per well) were seeded in 6-well plates and treated with 5.0 μM and 1.0 μM of borrelidin for 24 h, respectively. Then trypsinized, centrifuged and washed with cold PBS, and re-suspended in 490 μL of the provided binding buffer. 5 μL FITC-labeled Annexin V and 5 μL PI (BD, Franklin Lakes, NJ, USA) were added to the samples and mixed gently. After 15 min incubation in the dark at room temperature, samples were immediately run on a LSR-Fortessa flow cytometer with a total of 20 000 cells collected and analyzed with the Annexin V-FITC/PI apoptosis method. Cytographs were performed using BD FACSDiva Software (BD, Franklin Lakes, NJ, USA). The results were represented as the mean \pm SD from three independent experiments.

4.10. Wound healing assay

HepG2 and SMMC7721 cells (10^6 cells per well) were seeded in 6-well plates. After cells were confluent, two vertical linear "wounds" were carefully scratched by a 200 μL plastic pipette tip. Then washed with PBS and added into 2 mL serum-free medium with 0, 0.56, 1.67 and 5.0 μM of borrelidin for HepG2 cell, and 0, 0.11, 0.33 and 1.0 μM of SMMC7721 cell, respectively, then incubated at 37 °C and photographed every 12 h by inverted microscope.

4.11. The transwell assay and tumor invasion assay

A Boyden chamber (Thermo Fisher Scientific, Roskilde, Denmark) was used to examine the cell migration and invasion. For the invasion assay, BD Matrigel™ Basement Membrane Matrix (BD, Franklin Lakes, NJ, USA) was thawed overnight at 4 °C and diluted with cold serum-free medium (1 : 3). 200 μL of DMEM (for HepG2 cells) or RPMI-1640 (for SMMC7721 cells) medium with 20% FBS was added to the lower chamber of the transwell plate. The upper chamber of transwell was coated with 100 μL Matrigel. After Matrigel solidified, 200 μL of DMEM or RPMI-1640 medium with 1% FBS and 5×10^5 cells and different concentrations of borrelidin (0, 0.56, 1.67 and 5.0 μM for HepG2 cells; 0, 0.11, 0.33 and 1.0 μM for SMMC7721 cells) were added to the upper chamber of the transwell plates, respectively. After 24 h incubation at 37 °C, removed non-invaded cells with a cotton swab and fixed invaded cells in methanol and stained with crystal violet. For migration, the procedure was the same as the invasion assay except absence of Matrigel. The pictures were photographed by inverted microscope. The results were represented as the mean \pm SD from three independent experiments.

4.12. RNA extraction and real-time qPCR

The effects of borrelidin on the gene expression were examined in HepG2 and SMMC7721 cells by SYBR Green real-time quantitative PCR (Applied Biosystems, Waltham, MA, USA). HepG2 and SMMC7721 cells (8×10^5 cells per well) were seeded in 6-well plates and treated with 5.0 μM and 1.0 μM of borrelidin



for 24 h, respectively. The total RNA from cells was extracted using TRIzol Reagent (Ambion, Foster City, CA, USA) according to the manufacturer's instructions. The cDNA was synthesized from the extracted total RNA using a reverse transcriptase kit (Promega, Madison, WI, USA). The forward and reverse primers for *Cyclin D1* were 5-CTGTGCTGCGAAGTGGAACCAT-3 and 5-TTCATGGCCAGCGGAAGACCTC-3, respectively; *Cyclin D3* were 5-AGCGCCTTTCCCAACTCTA-3 and 5-CCAGGGTTACCACCATTGT-3; *Cyclin E1* were 5-GCTCGGGCTTTGTCCAG-3 and 5-CTTCCTGGACACGCTGGT-3; *CDK2* were 5-CCTTGTTTGTCCCTTCTAC-3 and 5-CAAATCCACCCACTATGA-3; *CDK4* were 5-CCTCTTTGGCAGCTGGTCAC-3 and 5-ACCGACACCAATTTTCAGCCA-3; *CDK6* were 5-CTGAATGCTCTTGCTCCTT-3 and 5-AAAGTTTGGTGGTCCTTGA-3; *p21* were 5-AGGCACCGAGGCACTCAGAG-3 and 5-TGACAGGTC-CACATGGTCTTCC-3; *Bax* were 5-CTGAGCAGATCATGAAGACA-3 and 5-AGTTTGCTGGCAAAGTAGAA-3; *Bcl-2* were 5-CTTTGAGTTC-GGTGGGGTCA-3 and 5-GGGCCGTACAGTTCCACAAA-3; *GAPDH* were 5-GGCAAGTTCAATGGCAGT-3 and 5-TGGTGAAGACGCCA-GTAGACTC-3. Real-time qPCR was performed using at 95 °C for 10 s, 60 °C for 20 s, and 72 °C for 20 s for total 35 cycles. *GAPDH* was used as an internal control. The $2^{-\Delta\Delta C_T}$ method was used to analyze relative changes in gene expressions for statistical analysis.³⁵

4.13. Western blot analysis

In vitro experiment, HepG2 and SMMC7721 cells (8×10^5 cells per well) were seeded in 6-well plates and treated with different concentrations (0, 0.56, 1.67 and 5.0 μ M for HepG2 cell, and 0, 0.11, 0.33 and 1.0 μ M for SMMC7721 cell) of borrelidin for 24 h, then washed with cold PBS and lysed with RIPA lysis buffer (Beyotime Biotechnology Co. Ltd, Shanghai, China) containing 2% protease inhibitor cocktail (Promega, Madison, WI, USA). *In vivo* experiment, tumor tissue homogenate was lysed for western blot analysis. The protein concentration of the lysates was determined by a BCA Protein Assay Kit (Beyotime Biotechnology Co. Ltd, Shanghai, China). Equal quantities of protein (40–60 μ g) were separated by SDS-PAGE, transferred to PVDF membranes (Millipore, Billerica, MA, USA), and blocked with 5% non-fat milk for 90 min at room temperature. The membranes were incubated with specific primary antibodies overnight at 4 °C and subsequently probed with corresponding secondary antibodies for 1 h at room temperature. Then, protein bands were detected with ECL select western Blot detection reagent (Millipore, Billerica, MA, USA) by chemiluminescence (Tanon 5500, Shang Hai, China). The relative amounts of protein loaded was normalized to the respective GAPDH bands.

4.14. Animal and tumor xenograft studies

BALB/c nude mice (weighing 18–20 g) were used for building a xenograft model. Mice were administrated with 100 μ L SMMC7721 cell suspension (5×10^6 cells) by subcutaneous injection, and when tumor volumes reaching 50 mm³, mice was randomly allocated to two groups ($n = 5$ per group). Animals of borrelidin treated group was administered borrelidin (2 mg kg⁻¹, dissolved in 60% 1,2-propanediol) once every other day by intraperitoneal injection. Control group were treated

with vehicle in the same way. Body weight and tumor size (tumor volume = length (mm) \times width (mm) \times depth (mm) \times $\pi/6$) were measured every other day. After 3 weeks of treatment, whole blood of each mouse was collected from the orbit for serum analysis. Then, the mice were sacrificed, and the tumor was excised. The wet weight of internal organs and tumor of each mouse were recorded. A part of tumor was prepared for western blot and other part for paraffin block. The study was performed in strict accordance with the Guide for the Care and Use of Laboratory Animals (Ministry of Science and Technology of China, 2006), and all experimental protocols were approved by the Laboratory Ethics Committees of College of Life and Health Sciences, Northeastern University (Shenyang, China).

4.15. Hematoxylin and eosin (H&E) staining

Tumor tissues were fixed in 4% paraformaldehyde, paraffin-embedded and section at 5 μ m. Afterwards, the tissue section was deparaffinized in xylene and rehydrated in a series of graded alcohols. To visualize histology, the tissue section was stained with hematoxylin and eosin and photographed using a microscope (Olympus, Model BX40F4, Tokyo, Japan).

4.16. Detection of apoptotic cells *in situ*

The number of apoptosis cells was identified by Tdt-mediated dUTP nick end labelling (TUNEL) method with an apoptosis detection kit (Millipore, Billerica, MA, USA) according to the manufacturer's instructions.³⁶

4.17. Serum analysis

Serum from each mouse was analyzed to observe changes of aspartate aminotransferase (AST) and L-alanine 2-oxoglutarate aminotransferase (ALT) levels. Serum AST and ALT analysis were conducted in 96-well plates using Aspartate aminotransferase Assay Kit and Alanine aminotransferase Assay Kit (Jiancheng Bioengineering Ins., Nanjing, China) respectively according to the manufacture's instruction. The enzyme activity was monitored by measuring the optical density (510 nm) of the plates.

5. Data analysis

All results are presented graphically as the group mean \pm standard deviation (SD) from at least three independent experiments. Student's *t*-test was used to determine differences between two groups using SPSS 13.0. The threshold value for acceptance of differences was 5% ($p \leq 0.05$).

Conflicts of interest

The authors declare no competing financial interest.

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References

- 1 J. L. Petrick, M. Braunlin, M. Laversanne, P. C. Valery, F. Bray and K. A. McGlynn, *Int. J. Cancer*, 2016, **139**, 1534–1545.
- 2 J. U. Marquardt, P. R. Galle and A. Teufel, *J. Hepatol.*, 2012, **56**, 267–275.
- 3 M. A. Avila, C. Berasain, B. Sangro and J. Prieto, *Oncogene*, 2006, **25**, 3866–3884.
- 4 J. M. Llovet and J. Bruix, *J. Hepatol.*, 2008, **48**(suppl 1), 20–37.
- 5 D. J. Trader and E. E. Carlson, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 4767–4769.
- 6 T. Wakabayashi, R. Kageyama, N. Naruse, N. Tsukahara, Y. Funahashi, K. Kitoh and Y. Watanabe, *J. Antibiot.*, 1997, **50**, 671–676.
- 7 D. Habibi, N. Ogloff, R. B. Jalili, A. Yost, A. P. Weng, A. Ghahary and C. J. Ong, *Invest. New Drugs*, 2012, **30**, 1361–1370.
- 8 A. Sidhu, J. R. Miller, A. Tripathi, D. M. Garshott, A. L. Brownell, D. J. Chiego, C. Arevang, Q. Zeng, L. C. Jackson, S. A. Bechler, M. U. Callaghan, G. H. Yoo, S. Sethi, H. S. Lin, J. H. Callaghan, G. Tamayo-Castillo, D. H. Sherman, R. J. Kaufman and A. M. Fribley, *ACS Med. Chem. Lett.*, 2015, **6**, 1122–1127.
- 9 T. Kawamura, D. Liu, M. J. Towle, R. Kageyama, N. Tsukahara, T. Wakabayashi and B. A. Littlefield, *J. Antibiot.*, 2003, **56**, 709–715.
- 10 M. Malumbres and M. Barbacid, *Nat. Rev. Cancer*, 2009, **9**, 153–166.
- 11 H. Gali-Muhtasib and N. Bakkar, *Curr. Cancer Drug Targets*, 2002, **2**, 309–336.
- 12 S. K. Mantena, S. D. Sharma and S. K. Katiyar, *Carcinogenesis*, 2006, **27**, 2018–2027.
- 13 A. Deshpande, P. Sicinski and P. W. Hinds, *Oncogene*, 2005, **24**, 2909–2915.
- 14 D. Gafu, M. Weiß, I. Mylonas and A. Brüning, *J. Appl. Toxicol.*, 2014, **34**, 1109–1113.
- 15 A. Gross, J. M. McDonnell and S. J. Korsmeyer, *Genes Dev.*, 1999, **13**, 1899–1911.
- 16 I. Budihardjo, H. Oliver, M. Lutter, X. Luo and X. Wang, *Annu. Rev. Cell Dev. Biol.*, 1999, **15**, 269–290.
- 17 S. K. Mantena, S. D. Sharma and S. K. Katiyar, *Mol. Cancer Ther.*, 2006, **5**, 296–308.
- 18 E. M. Creagh, H. Conroy and S. J. Martin, *Immunol. Rev.*, 2003, **193**, 10–21.
- 19 P. Friedl and K. Wolf, *Nat. Rev. Cancer*, 2003, **3**, 362–374.
- 20 K. Kessenbrock, V. Plaks and Z. Werb, *Cell*, 2010, **141**, 52–67.
- 21 N. Ramos-DeSimone, E. Hahn-Dantona, J. Siple, H. Nagase, D. L. French and J. P. Quigley, *J. Biol. Chem.*, 1999, **274**, 13066–13076.
- 22 H. Zheng, H. Takahashi, Y. Murai, Z. Cui, K. Nomoto, H. Niwa, K. Tsuneyama and Y. Takano, *Anticancer Res.*, 2006, **26**, 3579–3583.
- 23 A. Moroz, F. K. Delella, R. Almeida, L. M. Lacorte, W. J. Fávoro, E. Deffune and S. L. Felisbino, *PLoS One*, 2013, **8**, e84757.
- 24 J. Wen, Y. Zhang, X. Chen, L. Shen, G. C. Li and M. Xu, *Biochem. Pharmacol.*, 2004, **68**, 323–331.
- 25 H. J. Kim, N. Chakravarti, N. Oridate, C. Choe, F. X. Claret and R. Lotan, *Oncogene*, 2006, **25**, 2785–2794.
- 26 H. S. Park, K. I. Park, D. H. Lee, S. R. Kang, A. Nagappan, J. A. Kim, E. H. Kim, W. S. Lee, S. C. Shin, Y. S. Hah and G. S. Kim, *Food Chem. Toxicol.*, 2012, **50**, 2407–2416.
- 27 C. W. Phang, S. A. Karsani, G. Sethi, S. N. A. Malek and S. N. Abd Malek, *PLoS One*, 2016, **11**, e0148775.
- 28 X. Sui, N. Kong, L. Ye, W. Han, J. Zhou, Q. Zhang, C. He and H. Pan, *Cancer Lett.*, 2014, **344**, 174–179.
- 29 S. J. Kim, N. W. Ju, C. D. Oh, Y. M. Yoon, W. K. Song, J. H. Kim, Y. J. Yoo, O. S. Bang, S. S. Kang and J. S. Chun, *J. Biol. Chem.*, 2002, **277**, 1332–1339.
- 30 C. Stefanelli, B. Tanti, M. Fattori, I. Stanic, C. Pignatti, C. Clo, C. Guarnieri, C. M. Calderera, C. A. Mackintosh, A. E. Pegg and F. Flamigni, *FEBS Lett.*, 2002, **527**, 223–228.
- 31 L. Brown and S. Benchimol, *J. Biol. Chem.*, 2006, **281**, 3832–3840.
- 32 H. Zhang and I. Rosdahl, *Int. J. Oncol.*, 2004, **25**, 1241–1248.
- 33 M. S. Kuo, D. A. Yurek and D. A. Kloosterman, *J. Antibiot.*, 1989, **42**, 1006–1007.
- 34 Z. Cao, G. Khodakaramian, K. Arakawa and H. Kinashi, *Biosci., Biotechnol., Biochem.*, 2012, **76**, 353–357.
- 35 M. W. Pfaffl, *Nucleic Acids Res.*, 2001, **29**, e45.
- 36 X. Jiang, X. Zhu, N. Liu, H. Xu, Z. Zhao, S. Li, J. Cai and J. Cao, *Int. J. Biol. Sci.*, 2017, **13**, 167–178.

