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Introduction

Hydrogen bonding is the most important and widespread intermolecular interaction.¹ The scientific branches involving the importance of hydrogen bonding are very diverse, including inorganic and organic chemistry, supramolecular chemistry, biochemistry, and medicine.² It is operative in determining molecular conformation, molecular aggregation, and the function of a vast number of chemical systems ranging from inorganic to biological and is emerging as a powerful tool in the design of functional materials.³ In recent years, receptors using hydrogen bonding have been extensively developed, and a variety of functions have been investigated, including recognition, sensing, catalysis, and carrier-mediated transport across membranes.⁴

Perylene Diimide (PDI) is a valuable functional dye, which has numerous potential properties such as unique light-harvesting, redox properties, high chemical persistency, thermal durability, and outstanding optical and electronic properties. It has been applied as pigments, in fluorescent detectors, and semiconductors in organic electronics and photovoltaics.⁵

Anions are ubiquitous in environment and food samples and play an important role in numerous types of biological and medicinal processes.⁶ The design and development of synthetic receptors to recognize specific anions selectively had received more and more attention in recent years.⁷ Using PDI derivatives

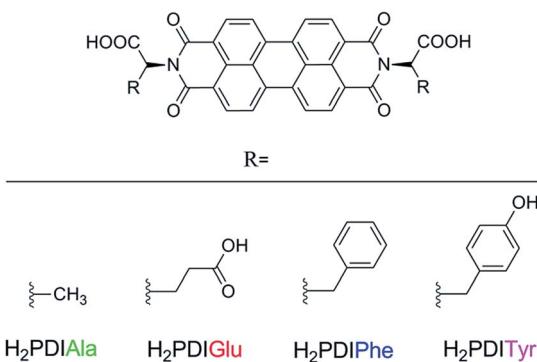
to detect anions became a developing trend of research. Sourav Saha and co-workers used aliphatic chain modified PDI to detect F^- and OH^- , which demonstrated anion- π electronic interactions from OH^- and F^- ions to PDI in aprotic solvents.⁸ On the other hand, a type of PDI derivative incorporated with functional groups of chloroacetamide ($-\text{NH}-(\text{C}=\text{O})-\text{CH}_2-\text{Cl}$) has been studied for fluoride ion detection.⁹ It possesses a specific selectivity and high sensitivity for fluoride ion (F^-) among the halogen anions. The recognition was attributed to the intermolecular proton transfer between a hydrogen atom on the amide of the sensor and the fluoride anion. Based on this result, a new colorimetric and ratiometric fluorescent sensor for naked-eye detection of fluoride ion based on PDI-amide derivatives has been designed and synthesized.¹⁰ In consideration of the important effect of hydrogen bonding, the PDI derivatives equipped with a $-\text{COOH}$ group, such as amino acid, should form a strong hydrogen bonding. However, there is no report of PDI-amino acid derivatives applied in detection of anions in literature. Amino acids are essential structural units and physiologically active substances of life and are environment friendly. The new material designed with amino acids has advantages of being green and safe, low price, easy to synthesize and so on.

Herein, we report the studies of PDI-amino acid derivatives (H_2PDIAAs) as fluorescent sensors to detect anions, particularly halogen ions, in organic solvents. Four types of H_2PDIAAs have been selected, which are perylene diimides functionalized with L-alanine (H_2PDIALa), L-glutamic acid (H_2PDIGlu), L-phenylalanine (H_2PDIPhe) and L-tyrosine (H_2PDITyr) (Scheme 1). A comparative study and discussion on their fluorescent properties have been provided explaining their interaction with halogen ions in DMF as H_2PDIAAs have good solubility and strong fluorescence in DMF only. This work applies the PDI-amino acid derivatives on the fluorescent sensor to detect anions based on hydrogen bonding for the first time.

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† Electronic supplementary information (ESI) available: Spectra (IR, UV-vis, ^1H NMR, ^{13}C NMR, ESI-MS, fluorescence life time), or other electronic format. See DOI: [10.1039/c7ra08005k](https://doi.org/10.1039/c7ra08005k)





Scheme 1 Structural representations of PDI functionalized with L-alanine (H_2PDIAla), L-glutamic acid (H_2PDIGlu), L-phenylalanine (H_2PDIPhe) and L-tyrosine (H_2PDITyr).

Experimental

Materials and measurements

All materials were reagent grade and obtained from commercial sources and used without further purification. IR spectra were recorded on a Nicolet-360 FT-IR spectrophotometer using KBr pellets. Absorbance spectra were obtained with a Persee TU-1950 spectrophotometer; the luminescent spectra were recorded at room temperature on Hitachi F-7000 FL spectrophotometer with a xenon arc lamp ashen light source spectra, with the pass width of $E_x = 2.5$ nm, $E_m = 2.5$ nm. NMR spectra were recorded on a Bruker 400 MHz, and DMSO- d_6 was used as the solvent, with TMS as the internal standard.

General procedure for H_2PDIAAs

The four types of H_2PDIAAs have been synthesized according to the method reported in literature.¹¹ Furthermore, 3,4,9,10-perylene tetracarboxylic acid dianhydride (0.500 g, 1.28 mmol) in DMSO (50 mL) was heated at 100 °C with stirring. The amino acids, L-alanine (H_2PDIAla) (0.341 g, 3.83 mmol), L-glutamic acid (H_2PDIGlu) (0.563 g, 3.83 mmol), L-phenylalanine (H_2PDIPhe) (0.632 g, 3.83 mmol) or L-tyrosine (H_2PDITyr) (0.594 g, 3.83 mmol) was dissolved in 2 M KOH aq. (2.5 mL). The solution of potassium amino acid salt was added to the mixture. The mixture was stirred for 3 hours at 100 °C before cooling to the room temperature with the formation of a precipitate, which was recovered by filtration and recrystallized from hot water and acetone to yield the dipotassium salt K_2PDIAAs . Moreover, 2 M HCl aq. (5 mL) was added, recovered by filtration, and washed extensively with water before being dried *in vacuo* (NMR, ESI-MS, IR for H_2PDIAAs in Fig. S1–S16†). *N,N'*-Bis-(L-alanine)-3,4,9,10-perylene tetracarboxylic diimide (H_2PDIAla). Yields: H_2PDIAla (0.512 g, 75%). ^1H NMR (400 MHz, DMSO- d_6): δ 7.88 (d, $J = 7.7$ Hz, 4H), 7.75 (d, $J = 8.1$ Hz, 4H), 5.52 (dd, $J = 13.9$, 6.8 Hz, 2H), 1.67 (d, $J = 7.0$ Hz, 6H). ^{13}C NMR (100 MHz, DMSO- d_6): δ 172.2, 162.2, 133.2, 130.8, 127.9, 124.4, 123.5, 122.0, 49.5, 15.4. IR (film): $\nu = 1747, 1695, 1653, 1592, 1437, 1343, 1256, 974, 809, 747$ cm⁻¹. ESI-MS: $m/z = 535$ [M + H]⁺. *N,N'*-Bis-(L-glutamic acid)-3,4,9,10-perylene tetracarboxylic diimide (H_2PDIGlu). Yields: H_2PDIGlu (0.611 g, 73%). ^1H NMR (400 MHz, DMSO- d_6):

δ 12.5 (br s, 4H), 8.43–7.94 (m, 8H), 5.66–5.53 (m, 2H), 2.61–2.47 (m, 4H), 2.46–2.25 (m, 4H). ^{13}C NMR (100 MHz, DMSO- d_6): δ 174.0, 170.9, 162.3, 133.1, 130.7, 128.0, 124.6, 123.1, 121.8, 52.7, 30.8, 23.9. IR (film): $\nu = 1751, 1697, 1642, 1591, 1363, 1255, 1168, 809, 747$ cm⁻¹. ESI-MS: $m/z = 651$ [M + H]⁺. *N,N'*-Bis-(L-phenylalanine)-3,4,9,10-perylene tetracarboxylic diimide (H_2PDIPhe). Yields: H_2PDIPhe (0.652 g, 74%). ^1H NMR (400 MHz, DMSO- d_6): δ 8.32–7.98 (m, 8H), 7.34–7.01 (m, 10H), 6.03–5.90 (m, 2H), 3.95–3.18 (m, 4H). ^{13}C NMR (100 MHz, DMSO- d_6): δ 170.6, 162.18, 137.9, 133.8, 131.1, 129.1, 128.2, 128.1, 126.4, 125.2, 123.6, 121.7, 54.0, 34.3. IR (film): $\nu = 1736, 1698, 1659, 1592, 1435, 1365, 1254, 1168, 810, 747$ cm⁻¹. ESI-MS: $m/z = 687$ [M + H]⁺. *N,N'*-Bis-(L-tyrosine)-3,4,9,10-perylene tetracarboxylic diimide (H_2PDITyr). Yields: H_2PDITyr (0.593 g, 70%). ^1H NMR (400 MHz, DMSO- d_6): δ 8.29–7.75 (m, 8H), 7.15–7.97 (m, 4H), 6.64–6.51 (m, 4H), 5.99–5.85 (m, 2H), 3.58–3.46 (m, 2H), 3.44–3.31 (m, 2H). ^{13}C NMR (100 MHz, DMSO- d_6): δ 170.8, 162.1, 155.9, 133.4, 130.9, 130.1, 127.9, 127.8, 124.9, 123.2, 121.6, 115.0, 54.1, 33.4. IR (film): $\nu = 1692, 1649, 1592, 14434, 1402, 1367, 1253, 1009, 825, 745$ cm⁻¹. ESI-MS: $m/z = 719$ [M + H]⁺.

Preparation of solutions for absorbance and fluorescence detections

The stock solution of H_2PDIAAs was prepared in DMF (1×10^{-5} M), and each inorganic salt (Na₂HPO₄, Na₂HPO₄, Na₃PO₄, Na₄P₂O₇ (PPI), NaNO₃, NaHCO₃, Na₂CO₃, NaHSO₄, Na₂SO₄, tetrabutylammonium fluoride (TBAF), tetrabutylammonium chloride (TBACl), tetrabutylammonium bromide (TBABr), tetrabutylammonium iodide (TBAI), tetrabutylammonium hydroxide (TBAOH)) was dissolved in DMF (1.5×10^{-2} M).

Test-paper measurement

The silica gel plate (2×1.5 cm²) was immersed in the DMF solution of H_2PDIAla (1×10^{-5} M) by soaking half of it and then dried using the blower. The letters (BIT) were written both in the upper and lower panels with the TBAF solution and then put under the visible light and UV light ($\lambda = 365$ nm).

Fluorescence lifetime decay

Fluorescence decay lifetime was measured using a time-correlated single photon counting instrument. Only freshly prepared solutions were used for the spectroscopic study, and all experiments were carried out at room temperature (298 K). Fluorescence lifetime was measured using Fluorolog-Tau-3 picosecond level fluorescence spectroscopy.

Results and discussion

UV-vis spectra

The UV-vis spectra of H_2PDIAAs show the typical features of perylene absorption with three peaks.¹² The interactions of H_2PDIAAs with different anions were studied by UV-vis absorption spectroscopy, in which there was found to be an obvious change in the presence of F⁻ and OH⁻ ions among Cl⁻, Br⁻, I⁻, H₂PO₄⁻, HPO₄²⁻, PO₄³⁻, PPI, NO₃⁻, HCO₃⁻, CO₃²⁻, HSO₄⁻, and SO₄²⁻ ions (Fig. S17†). Fig. 1 exhibits the results of



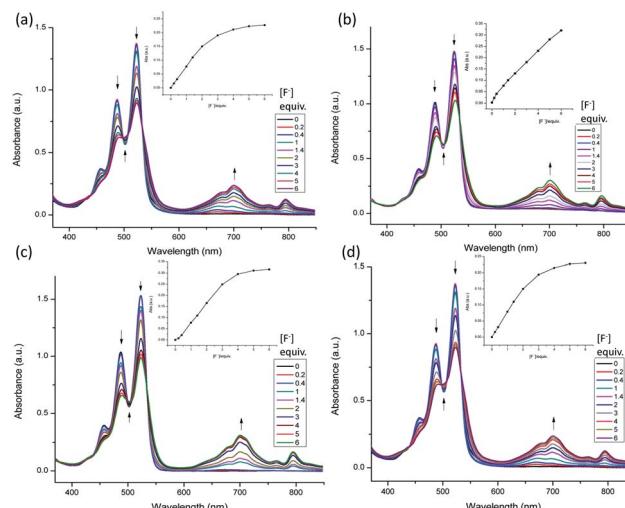


Fig. 1 UV-vis titration of H₂PDIAAs with TBAF in DMF (1×10^{-5} M), (a) H₂PDIALa, (b) H₂PDIGlu, (c) H₂PDIPhe, (d) H₂PDITyr. Inset: UV absorbance intensity at 700 nm vs. concentration of F⁻ ion.

UV-vis titration by adding TBAF into H₂PDIAAs in DMF solution. Clearly, the typical absorption peaks red-shifted, and a new peak appeared around 700 nm. With the same condition, the UV-vis spectrometric titrations of H₂PDIAAs with other anions did not show any significant change. Moreover, the inset pictures in Fig. 1 suggest that it has a good linearity between the absorbance at 700 nm with the concentrations of F⁻ from 0.2 eq. to 6 eq., which indicates that H₂PDIALa can detect F⁻ quantitatively. The linear equation was found to be $y = 0.0152x + 0.0008$ ($R = 0.9991$), where "y" is the absorbance at 700 nm measured at a given F⁻ concentration, and "x" represents the equivalent of F⁻ added. When the concentration of F⁻ was less than 1 equivalent of PDI, there was no large absorbance around 700 nm, and the absorbance appeared after the concentration of F⁻ was more than 1 equivalent of PDI. The same trend in the detection was found using H₂PDIPhe and H₂PDITyr (Fig. 1c and d inset), but H₂PDIGlu showed a different tendency in detection of F⁻ because it contains four carboxyl groups and has a capacity to accept more F⁻ ions (Fig. 1b inset, S18†). The OH⁻ ion shows similar phenomenon with F⁻ ion when it interacts with H₂PDIAAs (Fig. S19†).

Fluorescent properties

The interactions between H₂PDIAAs and various anions were studied by fluorescence spectra. There was an obvious change in fluorescence spectra of H₂PDIAAs in the presence of F⁻ and OH⁻ ions as shown in Fig. S20.† Furthermore, fluorescence titration by adding standard solution of TBAF in DMF has been applied in investigation of H₂PDIAAs interacting with F⁻ and OH⁻ ions (Fig. 2 and S21†). The four compounds have different fluorescence intensity from the strongest 7500 (a.u., H₂PDIPhe) to the lowest 1500 (a.u., H₂PDITyr), which may be related to the different substitutes on the branched chain of the four compounds. In order to understand the fluorescence behavior of H₂PDIAAs in solution better, we measured the fluorescence lifetime of these four compounds in DMF and calculated the

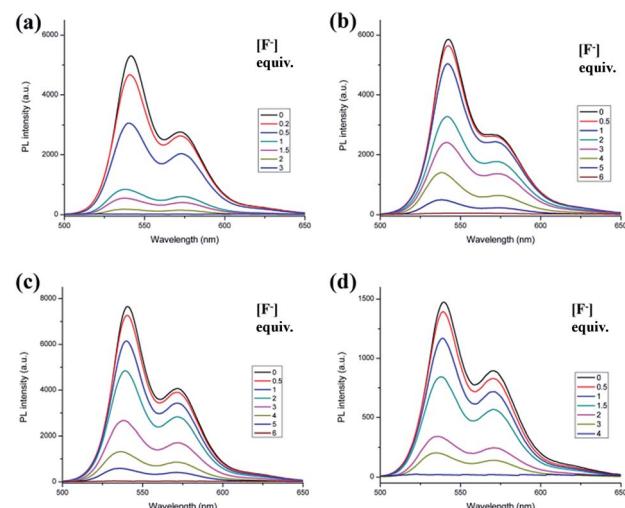


Fig. 2 Fluorescence emission of H₂PDIAAs with TBAF titration in DMF (1×10^{-5} M) ($\lambda_{\text{ex}} = 525$ nm), (a) H₂PDIALa, (b) H₂PDIGlu, (c) H₂PDIPhe and (d) H₂PDITyr.

theoretical quantum yield, shown in Table 1, in which the quantum yield of H₂PDIPhe in the four compounds is the largest one, and H₂PDITyr is the lowest. As a general rule, the fluorescence intensity is proportional to the fluorescence quantum yield. Therefore, this result is consistent with the fluorescence intensity measured with the same conditions, as shown in Fig. 2. The fluorescence lifetime of H₂PDITyr is longer than that of the other three H₂PDIAAs.

As shown in Fig. 2, the H₂PDIALa has been found to have sensitivity higher than that of the other three compounds. The OH⁻ ion also has similar phenomenon with the interaction of H₂PDIAAs (Fig. S21†), and fluorescence quenching can also be observed.

Fluorescent detectors for anions

Besides the UV-vis and fluorescent spectra, the selectivity of H₂PDIAAs to F⁻ and OH⁻ ions can also be well identified by the naked eye with colorimetric change (Fig. 3). The interactions of H₂PDIALa probe with various anions (H₂PO₄⁻, HPO₄²⁻, PO₄³⁻, PPI, NO₃⁻, HCO₃⁻, CO₃²⁻, HSO₄⁻, SO₄²⁻, Cl⁻, Br⁻, I⁻) compared with F⁻ were studied. Upon adding various anions to the blank solution of H₂PDIALa, no appreciable color change was noticed under the visible light and UV light (Fig. 3a). Therefore, it was inferred that there were no interactions between H₂PDIALa and various anions mentioned.

Table 1 The quantum yield (Φ_F) and lifetime of H₂PDIAAs in DMF solution (1×10^{-5} M)

	H ₂ PDIALa	H ₂ PDIGlu	H ₂ PDIPhe	H ₂ PDITyr
Quantum yield (Φ_F) ^a	0.27	0.36	0.66	0.11
τ (ns) ^b	4.51	4.65	4.54	5.06

^a The quantum yield (Φ_F) was measured relative to the fluorescein solution in 0.1 M NaOH ($\Phi_F = 0.79$).¹³ ^b Fluorescence lifetime values ($\lambda_{\text{em}} = 541, 572$ nm).



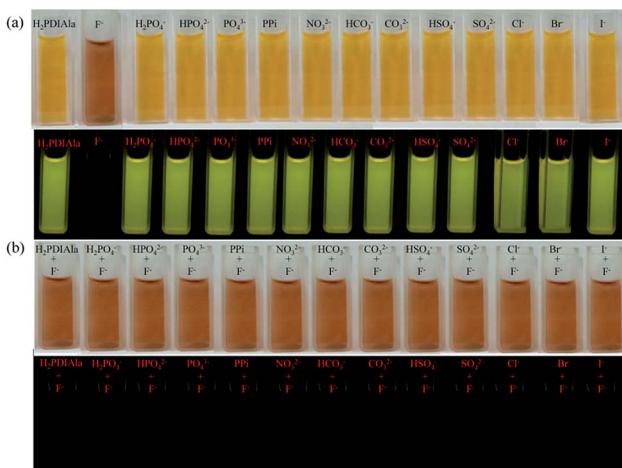


Fig. 3 (a) The photographs of colorimetric identification by naked eye and fluorescence emission ($\lambda = 365$ nm) of H₂PDIAla with various ions (6 eq., each). (b) The photographs of colorimetric identification by naked eye and fluorescence emission ($\lambda = 365$ nm) of H₂PDIAla–F⁻ ion with various ions (6 eq., each).

Furthermore, with the addition of F⁻ ion to the above solution, the color of both blank solution and that containing various anion's solution changed from yellow to brown as observed by the naked eye and fluorescence quenching under UV light (Fig. 3b). Clearly, the H₂PDIAla can be a fluorescent probe for F⁻ ion even with the existence of various other anions. Moreover, the detections by H₂PDIGlu, H₂PDIpHe and H₂PDTyr were almost the same except for the difference of color changes with H₂PDIGlu from yellow to purple, H₂PDIpHe from yellow to brick red, and H₂PDTyr from jacinth to brown (Fig. S22–S24†). The detection for OH⁻ ion by the fluorescent probe H₂PDIAAs has been processed, and a similar phenomenon was observed with fluorescence quenching and a color change could be observed by the naked eye (Fig. S25–S28†).

In order to improve the convenience and rapidity of F⁻ ion detection out of the laboratory, we made a simple test-paper, which used the existing H₂PDIAAs solution with silica gel plate. By soaking half of the silica gel plate in the solution and drying, the letters "BIT" were written in both the upper and lower panels with the TBAF solution. There was a significant difference between the soaked part and the control part. The letters "BIT" could clearly be observed under the visible light and UV light, but there was nothing on the control part (Fig. 4). It suggests that this method can be used to detect F⁻ ion conveniently and practically.

Mechanism of the fluorescent detectors

The NMR titration experiment is very useful in studying the mechanism of H₂PDIAAs to detect anions in solutions (Fig. 5). According to our NMR titration experiment, when the addition of F⁻ was equal or less than 1 equivalent of PDI, the signal of core structure protons always existed, whereas the signal of carboxylic acid moved to a low magnetic field and became broader. On the other hand, the signal of core structure protons gradually disappeared with the increasing of F⁻ concentration

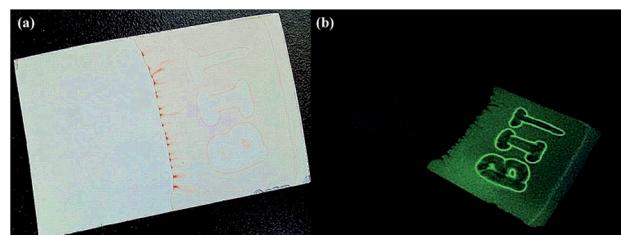


Fig. 4 Photographs of test-paper of H₂PDIAla solution in DMF (1×10^{-5} M) of TBAF. (a) Visible light of the test-paper, (b) UV light ($\lambda = 365$ nm) of the test-paper.

after more than 1 equivalent of PDI. The results indicated that when the concentration of F⁻ was equal or less than 1 equivalent of PDI, the association between H₂PDIAAs and F⁻ could be through the hydrogen bonding between the H⁺ of carboxylic acid ($-\text{COOH}$ proton) and F⁻(COO–H···F⁻). The formation of hydrogen bonding followed by deprotonation of the $-\text{COOH}$ fragment could reduce the electron density on the carboxylic moiety, which decreases the electron density of the core benzene ring of PDI, deshielding the corresponding protons and moving them to a low magnetic field.¹⁴ Clearly, when the concentration of F⁻ is more than 1 equivalent of PDI, there is an interaction between core structure and F⁻. Moreover, UV-vis titration experiments indicated that the PDI anion radicals were generated at high concentration of F⁻. These results indicated that anion–π interaction may play the leading role in the association between H₂PDIAAs and F⁻ at high concentrations of F⁻. A similar phenomenon for OH⁻ ion was observed (Fig. S32†). Based on the above observations and inferences, the synergistic effect of hydrogen bonding and anion–π interaction in the fluorescent detection of H₂PDIAAs could be proposed

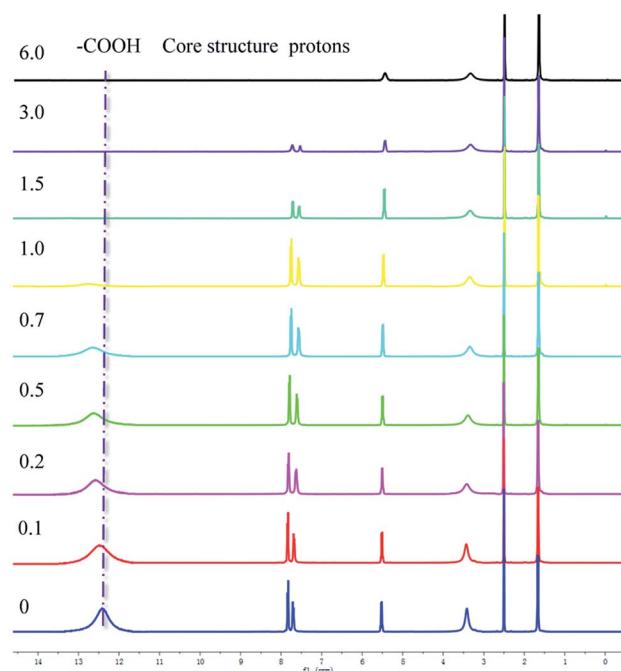
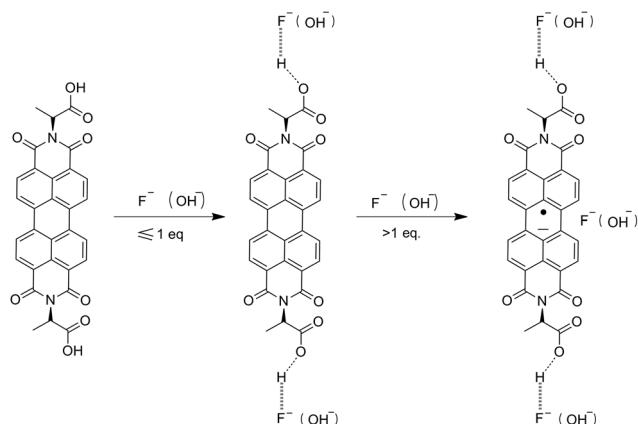


Fig. 5 The ¹H NMR titrations of H₂PDIAla with TBAF in DMSO-*d*₆.





Scheme 2 Proposed mechanism of H_2PDIAA fluorescent probe to detect fluoride and hydroxide ions.

and, the mechanism of fluorescent detection of H_2PDIAAs towards anions can be inferred to be a synergistic effect (Scheme 2). To the best of our knowledge, this is the first report of PDI-amino acid derivatives as fluorescent probes to detect anions.

Conclusion

In summary, we have explored and discussed the fluorescent detection of anions using four types of PDI-amino acids derivatives. The result indicates that they are sensitive to F^- and OH^- ions in DMF solution with an obvious color change and fluorescence quenching in a short time. The detecting mechanism has been proposed according to the molecular structures and NMR titrations. In addition, we made a simple test-paper to develop its potential applications. These results constitute a promising platform for the exploitation and applications of H_2PDIAAs as fluorescent probes, which are induced by the synergistic effect of hydrogen bonding and anion- π interaction, and laid a solid foundation for the PDI's development for fluorescent detectors.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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