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Quantification of ionic organo(fluoro)phosphates in decomposed lithium battery electrolytes

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A method for the separation, identification and quantification of the ionic decomposition products of LiPF₆ based organic electrolytes was developed. It employed the simultaneous coupling of two-dimensional ion chromatography (2D IC) with electrospray-ionization mass spectrometry (ESI-MS) and with inductively coupled plasma-mass spectrometry (ICP-MS). The electrolyte under investigation was a commercially available electrolyte by BASF, consisting of 1 mol L⁻¹ LiPF₆ in a mixture of one linear and one cyclic organic carbonate (50 : 50 wt%). The quantification was based on the use of a structurally similar compound as calibration standard for the ICP-MS measurement, while the identification was carried out with ESI-MS. Several organo(fluoro)phosphate species were successfully separated and quantified. Since these species show a high structural similarity to the warfare agent sarin, their investigation is of special interest with regard to their potential toxicity. Additionally, a quantification study of an electrolyte extracted from a cycled lithium metal cell was carried out.

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1. Introduction

The Lithium Ion Battery (LIB) is a key energy storage technology of the 21st century. Not only is it widely used for consumer electronics and portable devices, but it also plays an important role in stationary energy storage and electric vehicles.^{1,2} Of all the rechargeable electrochemical energy storage technologies, the lithium ion battery is one of the most commonly applied due to its high specific energy and power performance.² Despite these advantages, there are still challenges to overcome regarding its safety and limited lifetime.^{3–6} The limited battery lifetime is attributed to aging effects due to reaction and decomposition of battery components. In order to improve the LIB lifetime, understanding the aging mechanisms and development of new components with increased aging stability are of great significance.

The electrolyte is an LIB component, which is strongly influenced by aging. The state of the art conducting salt is LiPF₆. It has a high conductivity and fulfills the manifold requirements better than any other conducting salt, however, LiPF₆ is thermally and chemically instable.^{7–11} The electrolyte solvent mixture is composed of linear and cyclic organic carbonates, as they provide the electrolyte with a number of properties, which contribute positively to the performance of the LIB, such as low viscosity and high dielectric constant.^{12–18} The aging of the

electrolyte is a major drawback for a long-term battery cell performance. Due to its hygroscopicity, LiPF₆ always contains small amounts of water, which is known to accelerate the decomposition process of LiPF₆. In the organic carbonate solvents, LiPF₆ stands in equilibrium with LiF and PF₅.^{19,20} In presence of protic impurities like water, PF₅ reacts to inorganic phosphates under release of HF.²¹ At elevated temperatures beginning already with 40 °C and at high positive electrode potentials >4.9 V Li/Li⁺, the phosphates undergo reactions with the carbonates.^{22–24} Through these reactions CO₂ gas and organophosphates dissolved in the electrolyte are formed. These compounds are known to impair the LIB performance²⁵ and furthermore some organophosphates have structural similarities with nerve agents like sarin²⁶ and are potentially toxic.^{27–31} This toxicity is due to the inhibition of the mammal enzyme acetylcholinesterase^{26,30,32}

Most of the previous analytical studies were focused on the identification of possible decomposition products. The first studies were carried out with DSC^{33,34} and TGA³⁵ in order to describe the thermal stability of the electrolyte. The organophosphates previously reported in literature were detected with numerous analytical methods. By using nuclear magnetic resonance (NMR),^{8,36–40} different ionic and non-ionic main compounds were identified. Yet, NMR due to the high limit of detection low concentrations of organophosphates were not detected. High resolution-electrospray ionization-mass spectrometry (HR-ESI-MS) in the positive mode,^{7,9,21,41–43} liquid chromatography-based methods^{30,44–46} and gas chromatography-mass spectrometry (GC-MS) were applied to find and identify neutral organophosphates^{7,9,41–43,47} while ion chromatography-

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electrospray ionization-mass spectroscopy with product ion scan (IC-ESI-MS/MS)^{22,48,49} proved to be highly useful for the identification of ionic compounds. The attempts mainly covered the quantification of HF by spectroscopic ellipsometry^{50,51} as well as the educt LiPF₆. Further fluoride quantification was done by capillary electrophoresis (CE) and IC with conductivity detection and a fluoride sensitive electrode.^{21,24,52–54} So far, the quantification of the organophosphates was difficult due to the absence of analytical standards. Only two substances, dimethyl fluorophosphate (DMFP) and diethyl fluorophosphate (DEFP) were successfully quantified by GC-MS⁴⁷ with synthesized calibration standards. However, recently, approaches with liquid chromatography-tandem mass spectrometry were carried out.^{52,55} For the ionic compounds, a method was published which used a relative quantification with an internal standard with IC-ESI-MS²⁴ in order to monitor the degradation process.

Nevertheless, an absolute quantification of the organophosphates can be achieved with ICP-MS with a phosphorous containing calibration standard. Therefore, it is crucial that the individual species are well separated before they are transferred into the ICP-MS. In this work a newly developed method for the quantification of the organo(fluoro)phosphates is presented. An IC/IC system as presented by Kraft *et al.*⁴⁹ has been modified to fulfill the needs of the hyphenation to the ICP-MS and a simultaneous hyphenation to the ESI-MS was applied.

2. Material and methods

2.1 Chemicals

Battery grade electrolyte LP30 (50/50 wt% ethylene carbonate (EC)/dimethyl carbonate (DMC) and 1 M LiPF₆), and LP50 (50/50 wt% ethylene carbonate (EC)/ethyl methyl carbonate (EMC) and 1 M LiPF₆) SelectiLyte™ were purchased from BASF (Ludwigshafen, Germany). Deionized water from a Millipore Milli-Q water system with a LC-PAK cartridge (Bedford, USA) was used as contaminant and for the preparation of eluents and internal standard. Sodium carbonate (Certipur), sodium bicarbonate (for analysis) and sodium hydroxide solution (30%, Suprapur) were purchased from Merck (Darmstadt, Germany). Acetonitrile (HPLC gradient grade) for sample dilution and organic elution gradient was obtained from VWR (Bruchsal, Germany). As internal standard tellurium ICP-MS Standard in 2–3% HNO₃ (Certipur, Merck, Darmstadt, Germany) and for the calibration dibutyl phosphate (97.0% purity, Sigma-Aldrich, Steinheim, Germany) were applied.

2.2 IC/IC conditions

For the two-dimensional anion exchange chromatography, devices from Metrohm (Herisau, Switzerland) were used. The samples were introduced into the system with an 889 IC sample center. The measurements on the first dimension were carried out with an 850 Professional IC equipped with a conductivity detection unit. The second dimension was run with two 872 extension modules containing a pump, a chemical suppressor and a CO₂ suppressor. For the detection a second conductivity detector was used. All instruments were operated with the

software MagIC Net 3.0 (Metrohm, Herisau, Switzerland). On the first dimension a Na₂CO₃/NaHCO₃ eluent (1.8/1.7 mmol L^{−1}) was used with a gradient step. From 9 to 16 minutes 50% of acetonitrile was added while the carbonate concentration remained unchanged. The second dimension was run isocratically with an eluent out of 10 mM NaOH.

Further conditions can be found in Table 1.

2.3 ICP-MS conditions

All ICP-MS measurements were carried out with an Agilent 7700× Series ICP-MS (Agilent Technologies, Santa Clara, USA). The instrument was operated with the software Mass Hunter A.01.02 from Agilent Technologies (Santa Clara, USA). The internal standard (Te ICP-Standard (984 ± 4 mg kg^{−1}), diluted by factor 10 000 in deionized water) was pumped with a flow of 1.3 mL min^{−1} against counter pressure. All further conditions are given in Table 2.

2.4 ESI-MS conditions

For the ESI-MS measurements a triple quadrupole/linear ion trap instrument 3200QTrap (AB Sciex, Framingham, USA) was used. The instrument was controlled with the software Analyst 1.5.2 by AB Sciex (Framingham, USA). All other conditions can be found in Table 3.

2.5 Measurement setup

The two mass spectrometers were coupled simultaneously after the IC. The setup was used as shown in Fig. 1. Therefore, a split into both devices was applied after the second detector in the

Table 1 IC parameters

Instrument	Metrohm 850 Professional IC
Eluent 1	1.80 mM Na ₂ CO ₃ , 1.70 mM NaHCO ₃
Column 1	Metrosep A supp 4250 × 4.0
Guard column 1	Metrosep A supp 4/5 guard/4.0
Flow rate 1	1.0 mL min ^{−1}
Preconcentration column	Metrosep A PCC 1/4.0
Eluent 2	10 mM NaOH
Column 2	Metrosep A supp 10 250 × 4.0 mm
Guard column 2	Metrosep A supp 10 guard/4.0 mm
Flow rate 2	1.0 mL min ^{−1}
Column temperature	40 °C
Sample processor	Metrohm 889 sample center
Injection volume	10 µL

Table 2 ICP-MS parameters

Instrument	Agilent 7700 Series ICP-MS
RF power	1550 W
Carrier gas flow	1.05 L min ^{−1}
Torch	Quartz, 1.8 mm i.d.
Nebulizer	Micro mist
Spray chamber temperature	2 °C
Sampling depth	8.0 mm
Integration time	0.1 s
Split	1 : 1



Table 3 ESI-MS parameters

Instrument	AB Sciex 3200 QTrap
Mode	Negative, full scan
Mass range	m/z 50–250
Integration time	1 s
Ion spray voltage	−4250 V
Curtain gas	30 psi
Ion source gas 1	35 psi
Ion source gas 2	35 psi
Temperature	450 °C
Declustering potential	−20 V
Entrance potential	−8 V

IC. Due to tubing and pump speed, the ESI-MS was operated with a flow of 0.2 mL min^{-1} . The tube to the ICP-MS was then led into a four-way connector. The internal standard was added at this point with a flow rate of 1.3 mL min^{-1} (measured without counter pressure). Also connected to this four way connector were the sample introduction and the split tube, which were pumped by the peristaltic pump of the Agilent 7700 Series ICP-MS at a pump rate of 0.5 rps, resulting in a 1 : 1 split.

2.6 Sample preparation

In order to quantitatively study the influence of moisture on the aging of LiPF_6 based electrolytes, the samples aging was carried out after addition of water. For simplicity reasons, the electrolyte LP30 was chosen as it produces only organo(fluoro)phosphates with methyl groups. These species can be fully separated

while the aging products of LP50 which also contains ethyl side chains undergo group separation.⁴⁹ The formation reactions for the homologous species with methyl and ethyl side chains follow the same pathways and hence do not play an influential role during the reaction. Therefore, LP30 can be considered as a simplified system for LP50 which was applied during the initial method development.

The aging of LiPF_6 based electrolyte is initiated by traces of water in the electrolyte. The water reacts with the penta-fluorophosphate which is in a constant equilibrium with lithium fluoride and hexafluorophosphate. A temperature of 95 °C was chosen to age the samples within short times. In the quantification part the samples were prepared with different amounts of water to gain information about the kinetics of the organo(fluoro)phosphate formation.

Different amounts of water were added to the electrolyte to a maximum of 2.5 vol%. A minimum of three identical samples per amount of water was prepared. The samples were prepared in a dry room (dew point: −50 °C). The electrolyte (1 mL each) was stored in gas-tight stainless steel containers (containers: workshop, University of Münster, cap and perfluoralkoxy (PFA) sealing by Swagelok, Solon, USA) at 95 °C for 24 h.

After aging, the samples were diluted in acetonitrile by a factor of 100.

For method development, samples with 0.8 vol% of water were aged at 95 °C for 29 days.

The electrolyte from the cycled cells was extracted by centrifugation of the separator for 15 minutes at 8.5 rpm. The

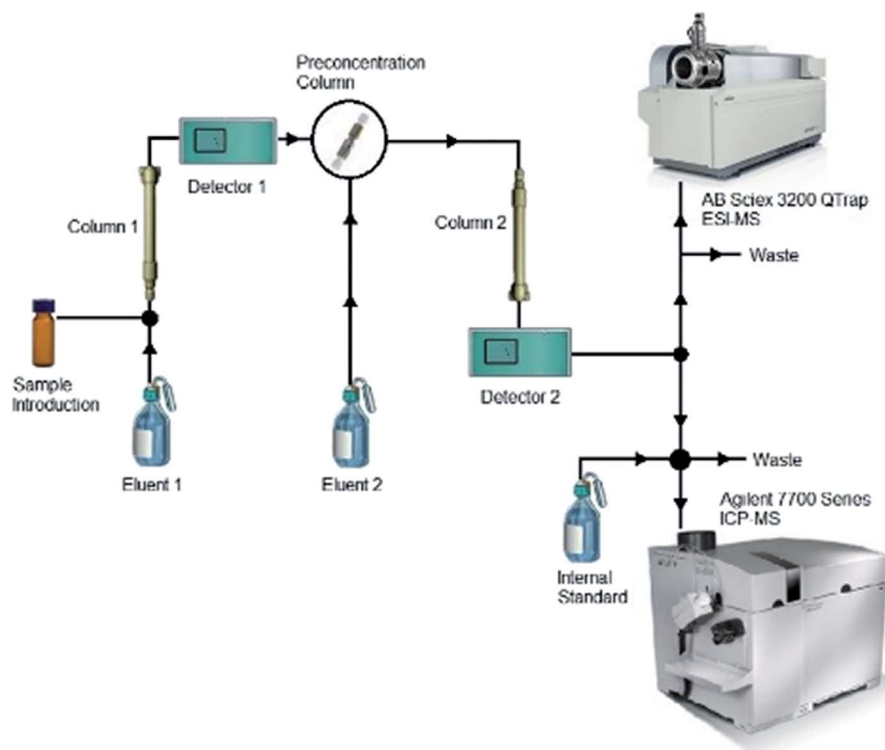


Fig. 1 Experimental setup. The two-dimensional IC is hyphenated to the ESI-MS and the ICP-MS with a three way split. The internal standard is added to the ICP-MS after the initial split.



samples were subsequently diluted in acetonitrile by a factor of 10.

The calibration standards were prepared by dilution of dibutyl phosphate in acetonitrile in the range from 0.5 mg kg⁻¹ to 100 mg kg⁻¹.

2.7 Cell assembly and cycling procedure

For constant current cycling, coin cells with lithium metal as anode and LiNi_{0.5}Mn_{1.5}O₄ (LNMO) as cathode were assembled in a dry room (dew point: -50 °C). The LNMO cathodes were prepared in-house. They consist out of 86 wt% of LNMO (Customcells), of 9 wt% conductive additive Super C65 (TIMCAL) and of 5 wt% polyvinylidene difluoride (Solvay). The components were solved in anhydrous *N*-methyl-2-pyrrolidone (Sigma Aldrich) before coating on an aluminum foil. They were then dried at 70 °C, pressed and afterwards dried again under vacuum at 120 °C. The mass loading of the active material was 2 mg cm⁻² and the diameter of the electrodes was 12 mm. The diameter of the lithium metal anode was 15 mm. The separator was prepared out of 5 layers of Freudenberg 2190 and one layer of Celgard 2500 (Celgard LLC, Charlotte, NC), the diameter was 17 mm. The separator was orientated in a way that the Celgard side faced the cathode. Here, as electrolyte LP50 (150 µL) was applied.

The cells were cycled in a Maccor series 4000 battery test system (Maccor Inc.). Three formation cycles were carried out with a charge and discharge current of 0.0678 mA between an upper cut-off voltage of 4.9 V and a lower cut-off voltage of 3.5 V. Then the cells were charged with 0.339 mA to 5.5 V and held there for 10 h. Finally, the cells were again discharged to 3.5 V.

3. Results and discussion

3.1 Separation

In the first dimension of the IC, the organo(fluoro)phosphates were insufficiently separated from each other and appeared as a shoulder on the fluoride signal as shown in Fig. 2. A separation could be possible after a long elution time on a column with hydrophobic character, enabling the interaction with alkyl groups of the organo(fluoro)phosphates. The maximum content of organic solvents for the hydrophobic column was limited to 10%, preventing the elution of the hexafluorophosphate. PF₆⁻ was only eluted after the application of an organic gradient with high contents. However, the addition of organic compounds would further make the plasma unstable or extinguish it. Therefore, the method of choice was the application of a two-dimensional chromatography using the cutting of the fraction with the organo(fluoro)phosphates from the first dimension and their transfer onto the second column.

In the second dimension, the fluoride signal still overlapped with three organo(fluoro)phosphate signals in the conductivity detector (Fig. 3). In the phosphorous selective detection in the ICP-MS (Fig. 4) it is shown that there were five baseline separated signals. Three of these signals were eluted together with the fluoride and could therefore not be detected in the conductivity detector. The other two signals were also visible in

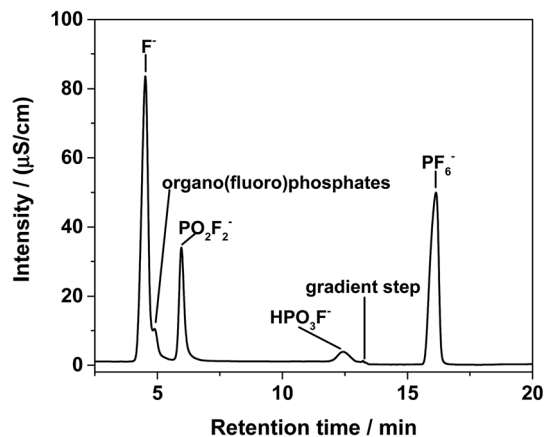


Fig. 2 Ion chromatogram by conductivity detection of LP30 after addition of 0.8 vol% of water, aged at 95 °C for 29 days, 1st dimension. The precursor hexafluorophosphate and its decomposition products including organo(fluoro)phosphates were detected. The organo (fluoro)phosphates were insufficiently separated from the fluoride signal.

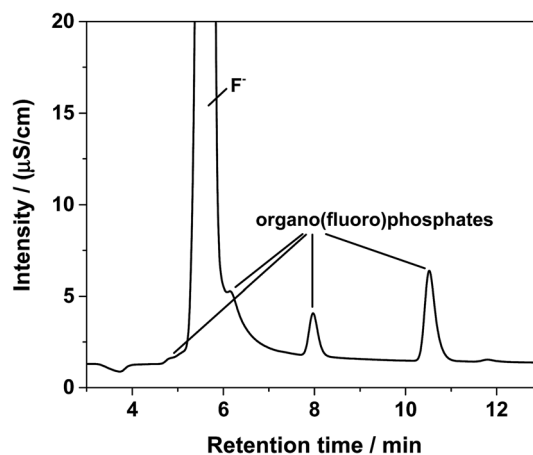


Fig. 3 Ion chromatogram by conductivity detection of LP30 after addition of 0.8 vol% of water, aged at 95 °C for 29 days, 2nd dimension.

the conductivity measurement. They were therefore phosphorous containing species.

3.2 Identification

The signals were identified with ESI-MS. The matching mass chromatogram is shown in Fig. 5. The obtained masses for the five signals were assigned to the organo(fluoro)phosphate species that were already identified by Kraft *et al.*⁴⁹ The substances which were found in amounts > LOQ (limit of quantification) were methyl fluorophosphate (MFP) 1, ethylene phosphate (EP) 2, dimethyl phosphate (DMP) 3, methoxyethyl fluorophosphate (MEFP) 4 and methoxyethyl methyl phosphate (MEMP) 5. The structures are given in Table 4.

LP50 was used as electrolyte in the electrochemical investigations for the following reasons: the wetting behavior of LP30 with separators, which enable a good electrolyte recovery by centrifugation, was very poor, which resulted in inefficient



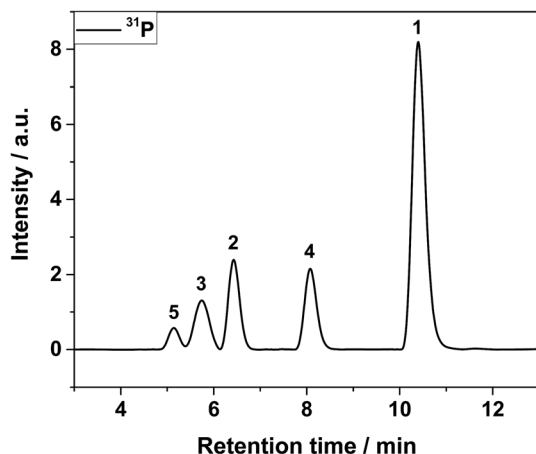


Fig. 4 ICP-MS chromatogram after 2nd dimension IC of LP30 after addition of 0.8 vol% of water, aged at 95 °C for 29 day.

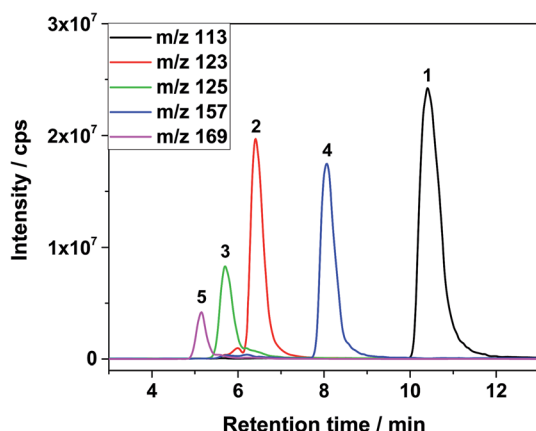


Fig. 5 ESI-MS mass chromatogram after 2nd dimension IC of LP30 after addition of 0.8 vol% of water, aged at 95 °C for 29 days.

charge/discharge cycling of the cell. LP50 contains ethyl methyl carbonate (EMC) instead of DMC. Therefore, the decomposition products will have both, methyl and ethyl side chains instead of methyl side chains only. An overall overview about the possible decomposition products can be achieved. The individual species with different side chains were not separable by the applied method. The organo(fluoro)phosphates hence underwent a group separation.⁴⁹ The 2D-IC-ICP chromatogram alongside with the ESI-chromatogram are shown in Fig. 6.

The additional species that differed from LP30 decomposition products are given in Table 5. MEFP and MEMP as well as their homologues with ethyl side chain have not been detected. The retention times were higher compared to the chromatogram in Fig. 4 because for this measurement a new A sup 10 column was applied which still had a higher capacity.

3.3 Development of a method for the quantification of organo(fluoro)phosphates

For the quantification of the organo(fluoro)phosphates it was necessary to have a standard which was treatable with the same

separation method as the analytes. Therefore, it was essential to find species that eluted from the first separation column without organic modifier with a short retention time. The standard also needed to be eluted from the second column with the same aqueous sodium hydroxide solution as the analytes with a short retention time and in a sharp signal. Therefore, three phosphorus containing substances were investigated. The hexafluorophosphate salt, *i.e.*, the precursor for the organo (fluoro)phosphates was not among the candidates for the standards because it eluted on the first column only under application of an organic gradient. This was already known from the first dimension separation of the electrolyte (Fig. 2). Phosphate was investigated as the first candidate for the calibration standard. It showed the same behavior as hexafluorophosphate and only eluted after application of an organic gradient. The other candidates, monofluorophosphate and dibutyl phosphate showed similar retention behavior on both columns. Dibutyl phosphate was chosen as calibration standard due to the higher structural similarity to the organo(fluoro) phosphates. Further, there was also a fluoride signal visible in the chromatograms of monofluorophosphate. This was attributed to hydrolysis of the substance.

A calibration with dibutyl phosphate using six data points resulted in a straight line graph with the slope of 6.54 ± 0.08 and the intercept of -0.8 ± 0.2 . The R^2 value was 0.9992 which proved a strong correlation between the signal and the concentration. The limits of quantification and detection were determined graphically from the peak height and noise width. The LOQ was 0.4 mg L^{-1} (0.01 mmol L^{-1}) while the LOD was 0.12 mg L^{-1} ($3.8 \times 10^{-3} \text{ mmol L}^{-1}$) both relating to phosphorus. These limits were still low considering that the quantification was applied after a separation instead of measuring the total phosphorous amount.

An internal standard of 100 ppb tellurium which was constantly injected into the system was used in order to monitor the plasma stability.

3.4 Quantification

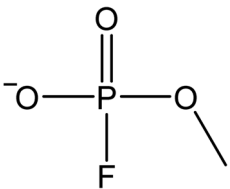
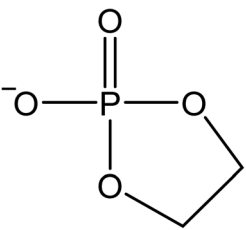
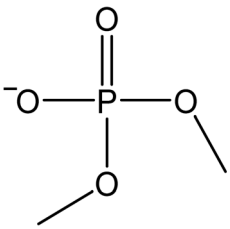
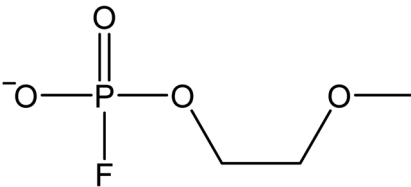
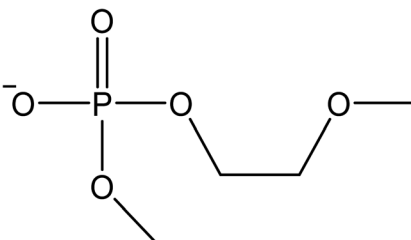
For quantification, samples of LP30 with different amounts of water, which had been aged under the same conditions, have been analyzed with the previously described method. The quantification results are shown in Fig. 7. The errors marked with error bars were the standard deviations from the three samples, which were used for each data point. The error for the single measurements was 1.5% and thus negligible compared to the standard deviations, which resulted from non-uniform aging of the samples. The standard deviations could go up to 10% of the measured mean value.

From Fig. 7, it can be seen that without water addition there were no decomposition products observed after one day of thermal aging. In order to gain information about the reaction mechanisms in the electrolyte without water addition, the electrolyte from a cycled cell at elevated cut-off voltage was investigated.

The chromatogram is shown in Fig. 6 and the corresponding quantification results are given in Table 6. The results show that



Table 4 Structures of the organo(fluoro)phosphates from the thermal aging of LP30

Number	Name	<i>m/z</i>	Chemical structure
1	Methyl fluorophosphate (MFP)	113	
2	Ethylene phosphate (EP)	123	
3	Dimethyl phosphate (DMP)	125	
4	Methoxyethyl fluorophosphate (MEFP)	157	
5	Methoxyethyl methyl phosphate (MEMP)	169	

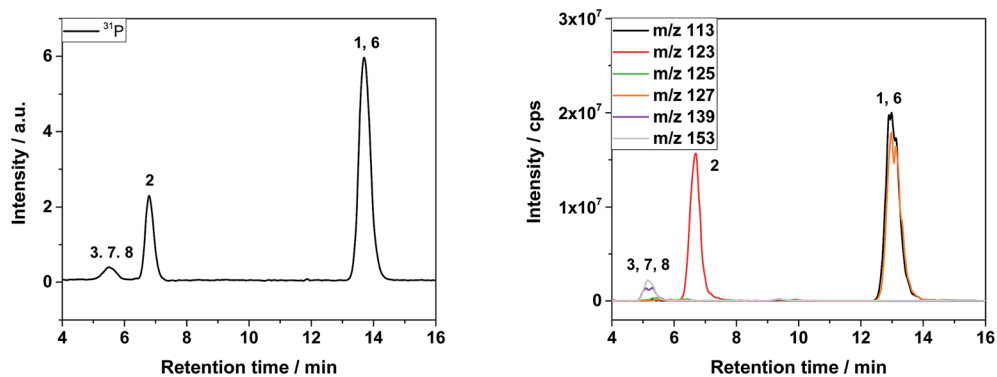
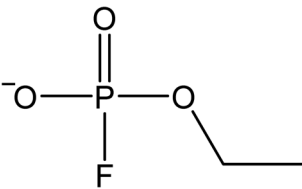
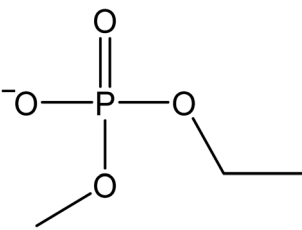
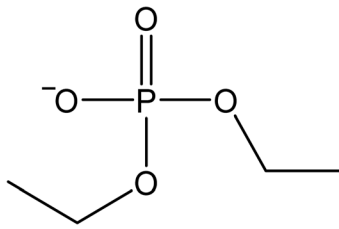
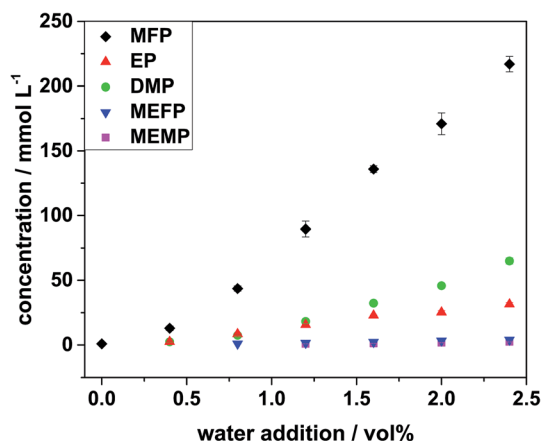
**Fig. 6** ICP-MS chromatogram and ESI-chromatogram of LP50 from aged Li metal/LNMO coin cells.

Table 5 Structure of the organo(fluoro)phosphates from the cyclic aging of LP50

Number	Name	<i>m/z</i>	Chemical structure
6	Ethyl fluorophosphate (DFP)	127	
7	Ethyl methyl phosphate (MEP)	139	
8	Diethyl phosphate (DEP)	153	

**Fig. 7** Quantification results for the five organo(fluoro)phosphates after aging for 24 h at 95 °C with addition of different amounts of water.

the main aging product in the cycling aging was MFP, just as it was in the aging with water addition. Yet, with $3.7 \pm 0.2 \text{ mmol L}^{-1}$ the total amount was low compared to the yields in the aging with water. Upon addition of only 0.4 vol% of water

Table 6 Quantification results for the organo(fluoro) phosphates from the electrolyte of the cycled cells

Species	MFP + DFP	EP	DMP + EMP + DEP
Amount/mmol L ⁻¹	3.7 ± 0.2	0.93 ± 0.06	0.26 ± 0.04

the amount of MFP was at $13 \pm 1 \text{ mmol L}^{-1}$ after 24 h of thermal aging. This shows that the thermal aging at 95 °C with water was highly aggressive compared to cyclic aging.

The second most abundant species was EP. Comparison with the thermally aged samples shows that this was not in accordance with the findings in the samples with water addition. In this case DMP occurred in higher concentrations than EP did. This phenomenon was attributed to the decomposition of PF_6^- to less fluorinated phosphates. They are considered precursors for the formation of the organo(fluoro)phosphates. The more complex organo(fluoro)phosphates MEFP and MEMP were only formed after high amounts of water addition. They were formed as a concurrence reaction to the ring closure of the EP formation since both reactions have fluoroethyl phosphate as a precursor. The results showed that the reaction of fluoroethyl phosphate with water, resulting in EP formation was faster than the reaction with DMC.

These results lead to the conclusion that the organo(fluoro)phosphate formation kinetics depend on the aging pathway. Hence, different pathways were favored in thermal and cyclic aging. Yet, the investigation of the mechanism was not feasible by IC based measurements alone and require theoretical calculations of the influence of thermal and electrochemical influences of the reaction constants.

Regarding the influences of the water addition the amount of decomposition products increased with the amount of added water. The increase of the amounts of the organo(fluoro)phosphates was almost linear with the higher water addition. Only the first two measurement points for each organo(fluoro)phosphate did not match this trend. This was also observed regarding the accumulated amounts of the organo(fluoro)



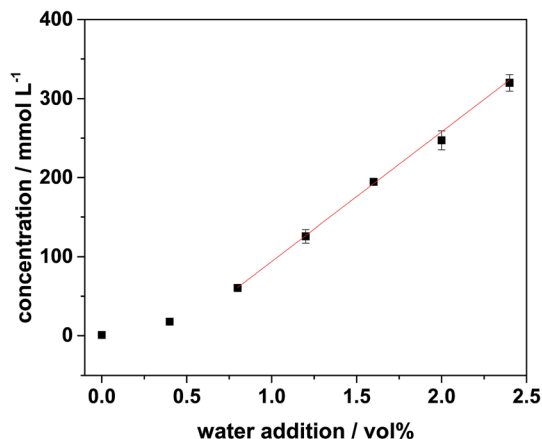


Fig. 8 Accumulated quantification for the organo(fluoro)phosphates after aging for 24 h at 95 °C with addition of different amounts of water.

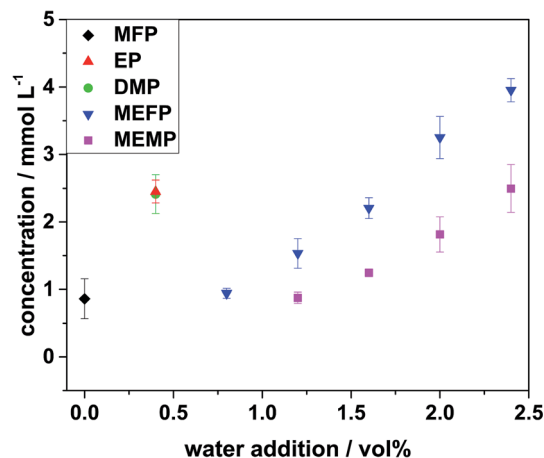


Fig. 9 Zoom-in from Fig. 6 to the lowest amount organo(fluoro)phosphates MEFP and MEMP.

phosphates. They are shown in Fig. 8. This effect could be caused by reactions that could not be monitored in this method. These reactions are hydrolysis steps of hexafluorophosphate, which lead to the formation of difluorophosphate and monofluorophosphate. These species were also precursors for the organo(fluoro)phosphate formation. They consumed small amounts of water and only with an excess of water the organo(fluoro)phosphate formation took place. Due to the two-dimensional approach, they were not visible in the ICP-MS chromatogram, but they have always been observed in the first dimension after conductivity detection. They were thus still present after the formation of the organo(fluoro)phosphates and lead to a consumption of water which could not be explained by organo(fluoro)phosphate formation alone. Furthermore, there existed also reaction pathways which lead to the formation of nonionic organo(fluoro)phosphates which were not accessible with ion chromatography.

It was clearly visible that MFP was formed in a high excess compared to the other organo(fluoro)phosphates. At an addition of 2.0 vol% of water 171 mmol L⁻¹ MFP were detected while the amount of EP (25 mmol L⁻¹) was only 15% of it. MEFP and MEMP were even less with only 1 and 2% compared with MFP. DMP was the second most abundant organo(fluoro)phosphate with 27% of the amount of MFP after addition of 2.0 vol% of water. According to Kraft *et al.*⁴⁹ DMP was formed from an alkoxylation reaction of MFP. Therefore, it was only formed if MFP has been formed before. The significantly higher amount of MFP shows that the formation of DMP from MFP was slower than the formation of MFP.

Furthermore, the more complex organo(fluoro)phosphates like MEFP and MEMP were only formed upon high amounts of water addition. In order to make their quantification more visible, Fig. 9 gives a zoom into the amounts to which they are formed. They have structural similarities to EP and it was suggested by Kraft *et al.*⁴⁹ that they were formed as a concurrence reaction. The current results show that the formation of EP was favored compared to the formation of MEFP and MEMP. In order to form EP, the precursor fluoroethyl phosphate reacted

with water while MEFP and MEMP were formed by a reaction with DMP. From the results it can be concluded that the reaction with water was faster than the reaction with DMP so that MEFP and MEMP were only formed if fluoroethyl phosphate existed in large quantities.

4. Conclusion

A new method for the quantification of ionic organo(fluoro)phosphates in LiPF₆-based LIB electrolytes was developed. The organo(fluoro)phosphates were generated by thermal aging at 95 °C for 24 h with addition of 0 to 2.5 vol% of water. The method uses a simultaneous hyphenation of a two dimensional heartcut ion chromatography with an ESI-MS and an ICP-MS. This enables the simultaneous identification and quantification of the different organo(fluoro)phosphate species. The quantification was realized by using dibutylphosphate as a structurally similar phosphorous containing calibration standard. The quantification of these substances enables the investigation of influences on the degradation of LiPF₆ in the LIB electrolyte. It is the first method which is able to compare the amounts of different ionic organo(fluoro)phosphate species by means of quantification.

In the current study the influence on the amount of water in the degradation process was investigated. The water amount was found to have a strong effect on the formation of the organo(fluoro)phosphates. The comparison of the amounts of the individual organo(fluoro)phosphates gives an insight into the relative formation rate of the substances. It shows that MFP is formed in the highest amount followed by DMP and EP. The rise of their amount shows a linear behavior. Yet, this is not true for low water amounts. This phenomenon is attributed to the decomposition of PF₆⁻ to less fluorinated phosphates. They are precursors for the formation of the organo(fluoro)phosphates. The more complex organo(fluoro)phosphates MEFP and MEMP are only formed after high amounts of water addition as they compete with EP for a common precursor.



As a first attempt of quantification of ionic organo(fluoro) phosphates in battery systems, a quantification from cycled Li-metal cells was carried out. It shows that MEFP and MEMP are not formed. MFP is still formed in the highest amount while in this case EP is formed in higher amounts than DMP. This points out that in thermal and cyclic aging different decomposition pathways are favored. Additionally the total amounts of organo(fluoro)phosphates after cyclic aging is lower than after thermal aging at 95 °C which shows that the addition of water and high temperature cause a more severe aging than cycling does.

In order to get the full picture, measurements with GC-ICP/MS should be carried out. In the current method, only ionic substances are accessible. A GC method is able to separate the volatile nonionic substances and is therefore complementary to the IC method.

Conflicts of interest

There are no conflicts of interest to declare.

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