

Cite this: *RSC Adv.*, 2017, 7, 42357

Incensfuran: isolation, X-ray crystal structure and absolute configuration by means of chiroptical studies in solution and solid state†‡

Najeeb Ur Rehman,^a Hidayat Hussain,^b *^{ab} Sulaiman Al-Shidhani,^a Satya Kumar Avula,^a Ghulam Abbas,^{ac} Muhammad Usman Anwar,^d ^a Marcin Górecki,^{de} Gennaro Pescitelli *^d and Ahmed Al-Harrasi*^aReceived 3rd July 2017
Accepted 21st August 2017

DOI: 10.1039/c7ra07351h

rsc.li/rsc-advances

A new cembrane diterpene named incensfuran (**1**), biogenetically derived from incensole (**2**), was isolated from crude extracts of the *Boswellia papyrifera* Hochst. The structure of incensfuran (**1**) was elucidated by 1D and 2D NMR study, and its absolute configuration was determined by single crystal X-ray diffraction analysis and electronic and vibrational circular dichroism spectroscopy (ECD, VCD). Moreover semisynthesis of incensfuran (**1**) was achieved after treatment of incensole (**2**) with $\text{BF}_3 \cdot \text{OEt}_2$. In addition epoxidation of incensfuran (**1**) provided expoxide analog **3** with 80% yield.

Introduction

Diterpenes are an important class of natural products based on twenty carbon atoms which can be found both in marine organisms and terrestrial plants. They are biogenetically derived from condensation of four isoprene units *via* deoxyxylulose phosphate or mevalonate pathways. Diterpenoids are generally found in Nature as linear, bicyclic, tricyclic or tetracyclic, pentacyclic, and macrocyclic diterpenes.¹ Cembranoids are 14-membered macrocyclic diterpene displaying a C-1 isopropyl group along with three methyl groups at C-4, C-8 and C-12. Cembranes have been reported from various natural sources including marine invertebrates (*viz.*, soft corals and sponges), plants (*viz.*, frankincense, resins, and tobacco), and insects.² The chemical diversity can be generated among cembranoids through polyoxygenation, yielding hydroxyl group (which are normally esterified by small-sized aliphatic or aromatic acids), keto groups, furan ring formation or γ -lactone group.³ However, the oxygenation and stereochemical

diversity of cembranoids are quite complex, for instance incensole (**2**) contains a 14-membered carbocyclic ring which incorporates the tetrahydrofuran ring. Incensole (**2**) and its acetate demonstrated a wide range of therapeutic applications such as: anti-proliferative, anti-inflammatory, neuroprotective, antidepressive-like and also cytotoxic properties.⁴

Due to their wide range of biological effects connected with unique structural variations, incensole analogs have received special interest among scientists with respect to their isolation and synthesis.² As a part of our ongoing research we have investigated various *Boswellia* species. Here, we reported the isolation from *Boswellia papyrifera* Hochst. of a new cembrane diterpene named incensfuran (**1**) along with known compound *viz.*, incensole (**2**). Its structure and relative configuration was elucidated by NMR and X-ray studies, and the absolute configuration was confirmed by means of electronic and vibrational circular dichroism (ECD, VCD) in solution, and in the case of ECD also in the solid state. In particular, we tested the applicability of the solid-state ECD/TDDFT method⁵ to the target compound which is fairly challenging, since it possesses a weak solid-state ECD spectrum with a band <200 nm, due to the presence of only one alkene chromophore in its structure.

Results and discussion

Phytochemical investigation of *B. papyrifera* resulted in the isolation and characterization of a new cembrane analog, incensfuran (**1**) (Scheme 1).

Compound **1** was obtained as colorless crystal and its molecular formula was confirmed as $\text{C}_{20}\text{H}_{34}\text{O}_2$ by ESIMS and NMR data. Moreover, the IR spectrum contained the absorption band of an olefinic (1670 cm^{-1}) functionality. This was further supported from ¹H-NMR spectrum peak at δ 6.01 (br. s, 1H, H-13)

^aUoN Chair of Oman's Medicinal Plants and Marine Natural Products, University of Nizwa, PO Box 33, 616 Birkat Al Mauz, Nizwa, Oman. E-mail: hussainchem3@gmail.com; aharrasi@unizwa.edu.om; Fax: +968-25446289; Tel: +968-25446328

^bCenter for Marine Biotechnology and Biomedicine, Scripps Institution of Oceanography, University of California-San Diego, La Jolla, CA 92093-0204, USA

^cDepartment of Biological Sciences and Chemistry, University of Nizwa, PO Box 33, 616 Birkat Al Mauz, Nizwa, Oman

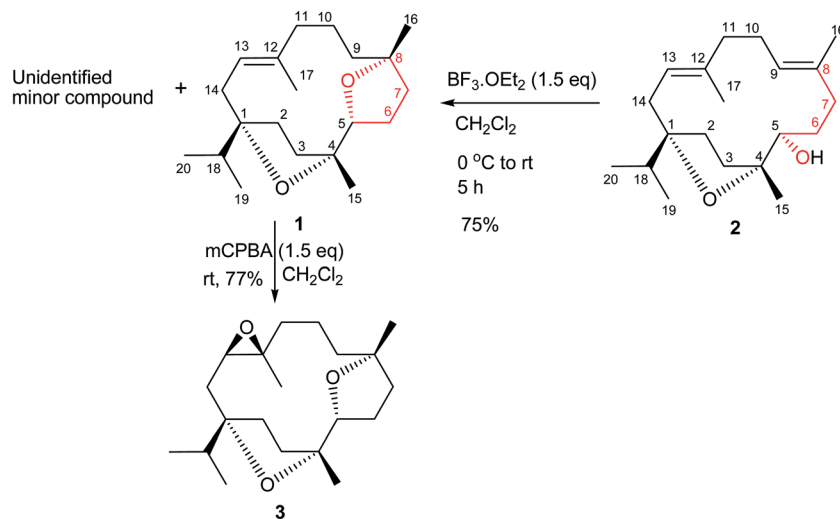
^dDipartimento di Chimica e Chimica Industriale, Università di Pisa, via Moruzzi 13, 56124 Pisa, Italy. E-mail: gennaro.pescitelli@unipi.it

^eInstitute of Organic Chemistry, Polish Academy of Sciences, ul. Kasprzaka 44/52, 01-224 Warsaw, Poland

† Dedicated to Professor Atta-ur-Rahman on the occasion of his 75th birthday.

‡ Electronic supplementary information (ESI) available. CCDC 1555949. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c7ra07351h





Scheme 1 Structure of incensuran (1) and incensole (2).

and ^{13}C -NMR peaks at δ 132.2 (C-12) and 121.2 (C-13). Furthermore, NMR displayed typical signals for isopropyl group of cembrane skeleton δ_{H} : 0.87 (d, $J = 6.6$ Hz, 3H, H-19); δ_{C} : 17.7; δ_{H} : 1.01 (d, $J = 6.6$ Hz, 3H, H-20); δ_{C} : 17.1 and cembrane skeleton was further confirmed from other methyl signals in NMR spectra δ_{H} : 1.55 (s, 3H, 17-Me); δ_{C} : 18.8; δ_{H} : 1.10 (s, 3H, 15-Me); δ_{C} : 24.0; δ_{H} : 1.04 (s, 3H, 16-Me); δ_{C} : 25.6). In addition, ^1H -NMR spectrum also possessed well-resolved oxymethine signal at 4.14 (dd $J = 9.6, 3.0$ Hz, 1H, H-5) supported by ^{13}C -NMR peak at 83.4.⁶

The DEPT NMR spectrum demonstrated the presence of 20 carbons divided into five methyls, eight methylenes, three methines (including one oxygenated C-5 methine at δ 83.4), and four quaternary carbons including three oxygenated tertiary carbons at δ 88.7, 86.5, and 83.7 for C-1, C-4, and C-8 respectively. Moreover, the complete structure of compound **1** was accomplished by COSY and HMBC experiments (Fig. 1) which illustrated oxygenation at C-1, C-4, C-5, and C-8 in the form of two furan rings. The cembrane skeleton as well two furan units were supported by key HMBC correlations between Me-20 and Me-19 with C-1 and C-18; Me-15 with C-3, C-4, and C-5; Me-16 with C-7, C-8, and C-9; Me-17 and C-11, C-12, and C-13; H-2 with C-1, C-3, C-4, and C-14; H-3 with C-1, C-2, C-4, and C-5; H-5 with C-3, C-4, C-6, and C-7; H-6 with C-4, C-5, C-7, and C-8; H-7 with C-5, C-6, C-8, and C-9; H-10 with C-8, C-9, C-11, and C-12; H-13 with C-1, C-11, C-12, and C-14, suggested the C-5 and C-8 furano-incensole.^{4,7}

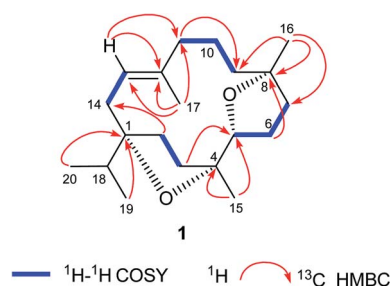


Fig. 1 Key COSY and HMBC correlations of incensuran **1**.

The relative stereochemistry at C-1, C-4, C-5, and C-8 was confirmed by the NOESY correlations of H-5 with Me-16 and Me-15, and of Me-19/20 with Me-15. Moreover, the structure of **1** was also confirmed by single-crystal X-ray diffraction analysis (Fig. 2). Moreover incensuran (**1**) was prepared (75%) from incensole (**2**) via Lewis acid mediated cyclization with $\text{BF}_3 \cdot \text{OEt}_2$ along with unidentified minor compound. Incensuran (**1**) was also epoxidized with *m*CPBA in CH_2Cl_2 and yielded epoxide analog **3** with 80% yield.

Determination of the absolute configuration

Similar to the previous report on 5-*epi*-incensole,⁸ in order to assign the absolute configuration of incensuran **1**, we performed chiroptical studies based on the tandem use of ECD and

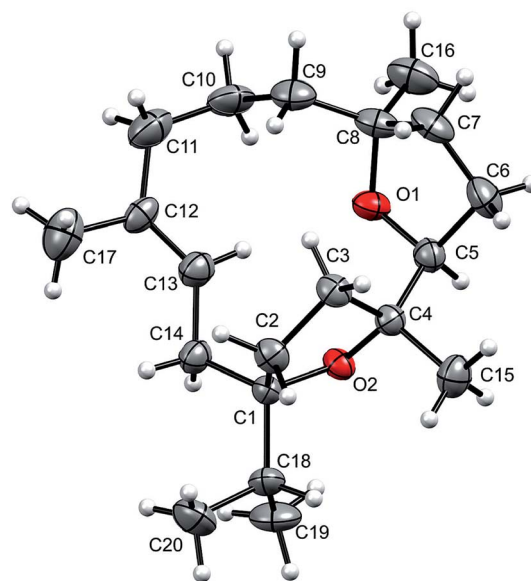


Fig. 2 Molecular structure of incensuran (**1**). Thermal ellipsoids are presented with 50% probability.



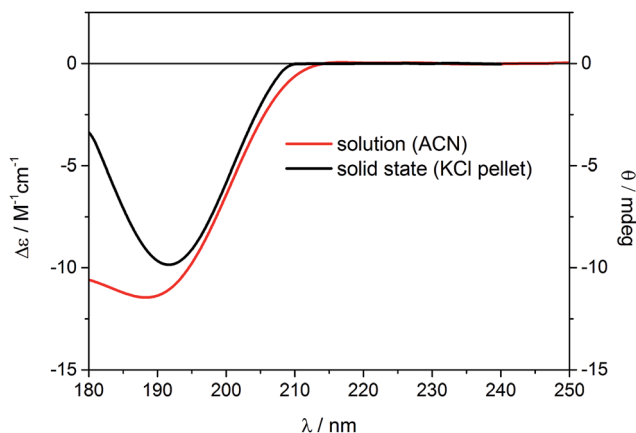


Fig. 3 Experimental ECD spectra of (1*S*,4*R*,5*R*)-incensuran (**1**) measured in acetonitrile solution (red line, left scale) and the solid state as KCl pellet (black line, right scale).

VCD spectroscopy both in solution and solid state. First, we applied the so-called solid-state ECD/TDDFT methodology,^{5,9} which is based on the comparison between the measured ECD spectrum in the solid state as a microcrystalline KBr or KCl pellet, and the spectrum computed by TDDFT method using the X-ray geometry as input structure (after optimization of all H-atoms). Consequently, all uncertainties related to the conformational search are avoided. Over the last decade, it has been shown that this methodology is particular beneficial for flexible medium-size natural products,^{5,9–16} and can be also applied to differentiate the conformational polymorphic forms of active pharmaceutical ingredients.¹⁷ In Fig. 3, the measured ECD spectra of incensuran **1** in acetonitrile solution (red line) and in the solid-state as KCl pellet (black line) are presented. As can be seen, both spectra display a negative ECD band with a minimum centered at ~ 190 nm.

In Fig. 4, the ECD spectrum calculated at CAM-B3LYP/TZVP level using the X-ray geometry with the (1*S*,4*R*,5*R*)-configuration is shown (black line). The agreement with the experimental solid-state ECD spectrum (Fig. 3) allowed us to establish the absolute configuration of incensuran **1** to be 1*S*,4*R*,5*R*. At the same time, we also confirmed that the absolute configuration at C-1 and C-4 is the same as incensole.^{6,8}

Having in mind that **1** possesses only one alkene chromophore in its structure, it is not surprising that ECD spectra in both media showed only a single negative band below 200 nm. Therefore, we decided to support the assignment by computing solution ECD and VCD spectra. In the first step of the computational procedure,^{18,19} a thorough conformational analysis was run with molecular mechanics (Merck Molecular Force Field, MMFF) within an energy range of 10 kcal mol⁻¹. Next, all the obtained structures (31) were submitted to DFT optimization at the B3LYP/6-31G(d) level of theory to reduce the total number of conformers found. Then, the remaining conformers (20) within 3 kcal mol⁻¹ were further re-optimized *in vacuo* using a higher level of theory, *i.e.* B3LYP/6-311+G(d,p). Finally, a set of 6 stable conformers with Boltzmann population >1% at 298 K were selected (see Table S1, ESI[†]). The most abundant DFT structure

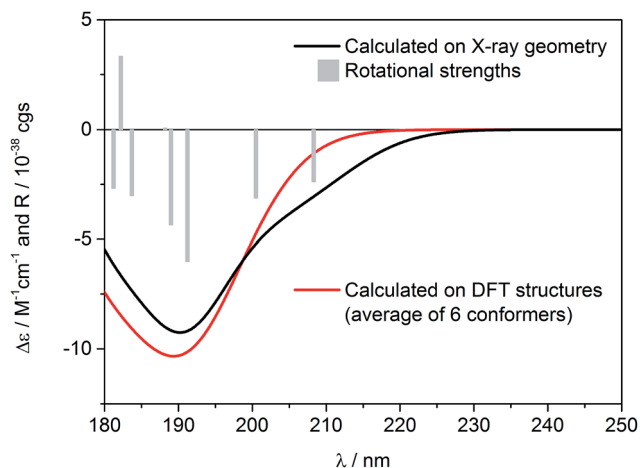


Fig. 4 Calculated ECD spectra for (1*S*,4*R*,5*R*)-incensuran (**1**) at CAM-B3LYP/TZVP level of theory. Black line: calculated on the X-ray geometry. Red line: calculated as Boltzmann average over 6 DFT-optimized structures. Vertical bars represent calculated rotational strengths on the X-ray geometry (for better clarity these values are divided by 4). Spectra red-shifted by 12 nm, Gaussian band-width 0.35 eV.

is almost coincident with the X-ray geometry (Fig. 5, and S1, ESI[†]). The remaining low-energy conformers (Fig. S1, ESI[†]) show moderate variations in the conformation of the macrocycle ring, especially in the region ranging from C-8 to C-11, while the region from C-1 to C-8 is more rigid because of the presence of the oxygen bridges. Furthermore, some differences in the conformation of tetrahydrofuran rings are also observed.

Solution ECD spectrum was simulated at the TDDFT level of theory using DFT-optimized geometries. The calculations were performed *in vacuo* for the first 50 excited electronic states using two different levels, *i.e.* CAM-B3LYP/aug-cc-pVDZ and CAM-B3LYP/TZVP. The obtained results were consistent, however a slightly better match between experimental and computed spectra was obtained using CAM-B3LYP/TZVP level, so these results are shown here (Fig. 4). As expected, there is also a good agreement between the solution and calculated ECD spectrum, which further confirms the configurational assignment of (1*S*,4*R*,5*R*)-incensuran (**1**).

One may argue that a single ECD band, though consistently reproduced by calculations, may not suffice to assign the

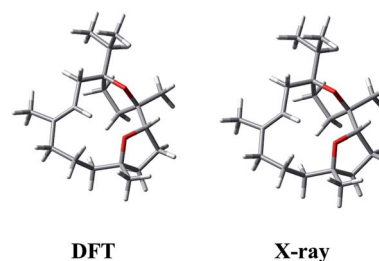


Fig. 5 Comparison between the lowest-energy DFT structure calculated *in vacuo* at B3LYP/6-311+G(d,p) level and the X-ray structure of (1*S*,4*R*,5*R*)-incensuran (**1**).



absolute configuration of a new compound confidently. This is true in general, however we wish to observe that the experimental ECD of incensfuran (**1**) is not weak; its g -factor ($\Delta\epsilon/\epsilon$) is -0.001 , both experimental and calculated. The absolute value is therefore well above the assumed sensitivity of ECD spectra (10^{-5}). Still, it is a good practice to use independent chiroptical methods to assign the absolute configuration of new natural products.¹⁹ Therefore, we confirmed ECD results using VCD too.

Theoretical VCD and IR spectra were calculated at the B3LYP/6-311+G(d,p) level applying the PCM model for CHCl_3 ($\epsilon = 4.9$) in order to reflect the impact of solvent not only on conformational equilibrium, but also on vibrational transitions, by reoptimizing the previously found set conformers (Table S2, ESI†). The simulated VCD/IR spectra are shown in Fig. 6. As can be seen, the Boltzmann averaged spectra (red curves) are in good agreement with the experimental data recorded in CDCl_3 (black curves). As it would be expected, similar results were obtained by carrying out *in vacuo* calculations (Fig. S2, ESI†). Some discordances in the relative intensities of calculated VCD bands may be observed in the range of $1300\text{--}1100\text{ cm}^{-1}$. Nevertheless, the experimental and calculated IR spectra correspond perfectly in the range $1600\text{--}950\text{ cm}^{-1}$, resulting indeed in a good agreement of VCD signs too, which allowed us to confirm independently the previous assignment obtained using ECD spectroscopy. Note that at the concentration of 0.19 M , the $\text{C}=\text{C}$ stretching ($1600\text{--}1700\text{ cm}^{-1}$) is almost silent in the VCD spectrum and faintly visible in the IR one.

Summarizing, here we demonstrated that the solid-state ECD/TDDFT method confirmed its strength even in the presence of a single alkene moiety as the only chromophore unit in the molecule. Previously, we had applied the same technique to compounds containing other weak chromophores such as two non-conjugated double bonds,⁸ two non-conjugated ester moieties,²⁰ and two anhydride groups.²¹ Furthermore, we showed that the stereochemical assignment can be performed using very low amount of sample ($\sim 0.2\text{ mg}$ in this case) required

to record a significant solid-state ECD spectrum $<200\text{ nm}$. In contrast, to measure solution VCD spectrum one need a much larger amount of sample ($\sim 11\text{ mg}$ in the present case).

Conclusion

A new cembrane diterpene, incensfuran (**1**), was isolated from the crude extracts of the *B. papyrifera* Hochst and its structure elucidated by means of spectroscopic data analysis including 1D (^1H and ^{13}C NMR spectra) and 2D (COSY, HMBC, NOESY) spectra. The relative and absolute configuration was established as $(1S,4R,5R)\text{-1}$ by single crystal X-ray analysis in tandem with electronic and vibrational circular dichroism. In the course of our work we also confirmed that the solid-state ECD/TDDFT method can be reliably applied to compounds having one substituted olefin as the only chromophore in their structure.

Experimental section

General

Optical rotations were measured on a KRUSS P P3000 polarimeter (A. Kruss Optronik, Germany). IR spectra were recorded on a Bruker, ATR-Tensor 37 spectrophotometer. ESI-MS were recorded on a Waters Quattro Premier XE Mass Spectrometer (Waters, Milford, MA). The ^1H and ^{13}C NMR spectra were recorded on Bruker NMR spectrometers operating at 600 MHz (150 MHz for ^{13}C). The chemical shift values are reported in ppm (δ) units and the coupling constants (J) are given in Hz. For TLC, pre-coated aluminum sheets (silica gel 60 F-254, E. Merck) were used. Visualization of the TLC plates was achieved under UV light at 254 and 366 nm and by spraying with ceric sulfate reagent.

ECD spectra were measured with a Jasco (Tokyo, Japan) J-715 spectropolarimeter at room temperature, using 100 nm min^{-1} scanning speed, a step size of 0.2 nm , a bandwidth of 1 nm , a response time of 0.5 s , and an accumulation of 8 scans. The

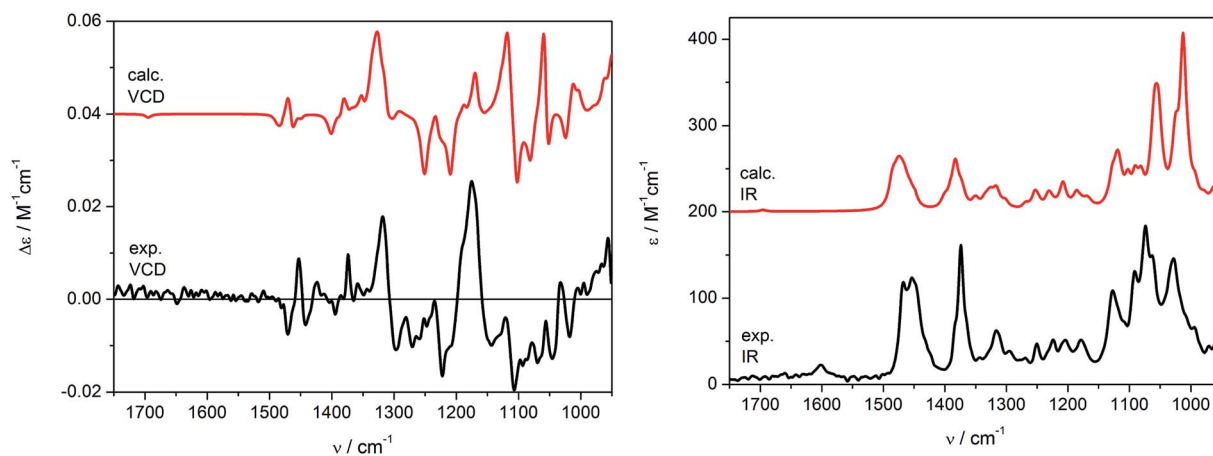


Fig. 6 Calculated (red lines) and experimental (black lines) VCD (left) and IR (right) spectra for $(1S,4R,5R)$ -incensfuran (**1**). Calculated spectra are shown with a vertical offset for clarity. Calculations run at B3LYP/6-311+G(d,p) level using PCM for CHCl_3 as Boltzmann average over 6 conformers; for better comparison the calculated VCD spectrum was multiplied by 2; frequencies scaled by a factor 0.985; band-width $\sigma = 6\text{ cm}^{-1}$.



solution spectrum of **1** was measured on a 3.3 mM CH₃CN solution in a quartz cell with a path length of 0.02 cm. The ECD solid-state spectrum was obtained using the KCl pellet technique described previously.⁹ In particular, ~0.1 mg of **1** was mixed and ground with 200 mg of KCl, and then ~100 mg of this mixture was used to prepare the pellet. IR/VCD spectra were recorded at room temperature using a Jasco (Tokyo, Japan) FVS-6000 VCD spectrometer between 2000–850 cm⁻¹ with 4 cm⁻¹ resolution using 4000 scans, on 0.19 M CDCl₃ solution in a BaF₂ cell with a path length of 100 μm. The final spectrum was obtained by subtracting the CDCl₃ spectrum recorded in the same conditions.

For X-ray measurements, single crystals of **1** were mounted on a glass fiber and examined at 296 K on a Bruker D8 Venture APEX diffractometer equipped with Photon 100 CCD area detector using graphite-monochromated Mo-Kα radiation (λ = 0.71073 Å).[‡]

Plant material

The *B. papyrifera* frankincense was donated by Dr Saifeldin Elnegrabi from Sudan. All of these samples were authenticated by Dr Mustafa Mansi (botanist), Department of Biological Sciences and Chemistry, University of Nizwa, the Sultanate of Oman and voucher specimen (no. BSHR-01/2012) was deposited with the Herbarium of the Chair of Oman's Medicinal Plants and Marine Natural Products.

Extraction and purification

The air-dried powdered resin (1.5 kg) of *B. papyrifera* was finely extracted with MeOH (10 l) at r.t. (3 × 15 days) and evaporated under reduced pressure to yield a crude MeOH extract (1.3 kg). The crude MeOH extract was subjected to Vacuum Liquid Chromatography (VLC) using 10, 20, 30, 50% *n*-hexane/EtOAc, and washed with EtOAc to afford five fractions (SF₁–SF₅). Fraction (SF₁), 10% *n*-hexane/EtOAc, was further subjected to column chromatography using *n*-hexane, 2%, 5%, and 10% *n*-hexane/EtOAc as a mobile phase to get four sub-fractions (SSF₁–SSF₄). After taking TLC, fraction two (SSF₂) was further chromatographed and gave semi-pure compound which was loaded directly on recycling preparative HPLC, using CHCl₃ solvent system stabilized with 0.6% EtOH. Incensfuran (**1**, 55.5 mg) was eluted as a UV-inactive compound at a retention time of 51.7 min with 3.5 ml min⁻¹ flow rate. The compound was recrystallized in 30% CHCl₃/MeOH to make a white needle like crystals.

Incensfuran (**1**): white crystals; [α]_D²⁵ = -15.2 (c 0.26, MeOH); IR (KBr): 1670, 1460, 1382, 1053, 978 and 841 cm⁻¹; IR (CDCl₃): 1467, 1452, 1374, 1315, 1250, 1224, 1203, 1177, 1127, 1074, 1026 cm⁻¹; VCD (CDCl₃, sign): 1470 (-), 1454 (+), 1442 (-), 1423 (+), 1395 (-), 1373 (+), 1317 (+), 1297 (-), 1270 (-), 1222 (-), 1175 (+), 1143 (-), 1107 (-), 1068 (-), 1046 (+) and 1018 cm⁻¹ (-); ECD (CH₃CN, Δε): 188.3 nm (-11.4); ECD (KCl, mdeg): 191.6 nm (-9.9); ¹H-NMR (600 MHz, CDCl₃): 6.01 (br. s, 1H, H-13), 4.14 (dd, *J* = 9.6, 3.0 Hz, 1H, H-5), 2.20 (m, 2H, H-11), 2.27 (m, 1H, H-2b), 2.13 (m, 1H, H-14a), 2.03 (m, 1H, H-14b), 2.01 (m, 1H, H-3a), 1.97 (m, 1H, H-9a), 1.80 (m, 1H, H-18), 1.78 (m, 1H, H-2a), 1.64 (m, 1H, H-6a), 1.71 (m, 2H, H-7a, H-3b), 1.67 (m, 1H,

H-6b), 1.53 (m, 2H, H-7b), 1.55 (s, 3H, 17-Me), 1.48 (m, 1H, H-9b), 1.42 (m, 1H, H-10a), 1.33 (m, 1H, H-10b), 1.10 (s, 3H, 15-Me), 1.04 (s, 3H, 16-Me), 1.01 (d, *J* = 6.6 Hz, 3H, H-20), 0.87 (d, *J* = 6.6 Hz, 3H, H-19); ¹³C NMR (150 MHz, CDCl₃): δ 132.2 (C-12), 121.2 (C-13), 88.7 (C-1), 86.5 (C-4), 83.4 (C-5), 83.7 (C-8), 41.1 (C-18), 40.9 (C-11), 38.5 (C-3), 38.4 (C-7), 35.2 (C-6), 31.3 (C-14), 31.1 (C-11), 28.1 (C-2), 25.6 (C-16), 24.0 (C-15), 19.8 (C-10), 18.8 (C-17), 17.7 (C-19), 17.1 (C-20); ESIMS: *m/z* (rel. int.): 329.02 [M + Na]⁺ (100); (calcd for C₂₀H₃₄O₂ + Na, 329.2); 635.2 [2M + Na] (50).

Compound 3

To a cooled (0°) soln. of incensfuran (**1**) (20 mg) in CH₂Cl₂ (5 ml) was added mCPBA (1.5 eq.) and the mixture was stirred at r.t. for 6 h. After taking TLC the reaction was quenched by addition of saturated Na₂SO₃ solution. The mixture was extracted with CH₂Cl₂ followed by washed with sat. aq. NaHCO₃ solution and dried with Na₂SO₄. Finally the residue was loaded directly on recycling preparative HPLC, using CHCl₃ solvent system stabilized with 0.6% EtOH to give compound **3** with 77% yield. Colourless solid; [α]_D²⁵ = +21.7 (c 0.04, MeOH); ¹H-NMR (600 MHz, CDCl₃): 4.02 (dd, *J* = 9.6, 3.6 Hz, 1H, H-5), 3.66 (dd, *J* = 10.2, 2.4, 1H, H-13), 2.19 (dd, *J* = 10.2, 3.6 Hz, 1H, H-9), 1.20 (m, 1H, H-9), 1.90 (m, 1H, H-7), 1.57 (m, 1H, C-7), 1.67 (m, 1H, H-11a), 1.52 (m, 1H, H-11b), 2.07 (m, 1H, H-2b), 2.19 (dd, *J* = 14.4, 2.4, 1H, H-14a), 1.47 (m, 1H, H-14b), 1.72 (m, 1H, H-3a), 1.87 (m, 1H, H-9a), 1.80 (m, 1H, H-18), 1.46 (m, 1H, H-2a), 1.85 (m, 1H, H-6a), 1.52 (m, 2H, H-7a, H-3b), 2.01 (m, 1H, H-6b), 1.16 (s, 3H, 16-Me), 1.53 (m, 2H, H-6b, H-7b), 1.21 (s, 3H, 17-Me), 1.65 (m, 1H, H-9b), 1.62 (m, 1H, H-10a), 1.78 (m, 1H, H-10b), 1.05 (s, 3H, 15-Me), 1.01 (d, *J* = 6.6 Hz, 3H, H-20), 0.81 (d, *J* = 6.6 Hz, 3H, H-19); ¹³C NMR (150 MHz, CDCl₃): δ 88.3 (C-1), 86.4 (C-4), 84.5 (C-5), 82.5 (C-8), 65.4 (C-13), 61.2 (C-12), 36.6 (C-18), 39.9 (C-11), 38.5 (C-3), 36.5 (C-7), 33.5 (C-6), 30.2 (C-14), 31.6 (C-9), 27.3 (C-2), 27.4 (C-16), 21.6 (C-15), 20.5 (C-10), 18.9 (C-17), 16.4 (C-19), 18.9 (C-20); ESIMS: *m/z* (rel. int.): 345.0 [M + Na]⁺; (calcd for C₂₀H₃₄O₃ + Na, 345.2).

Computational section

Conformational searches and preliminary DFT calculations were run with Spartan'14 (Wavefunction, Irvine CA), with standard parameters and convergence criteria. DFT and TDDFT calculations run with Gaussian'09 (Revision D.01. Gaussian, Inc: Wallingford, CT; 2013), with default grids and convergence criteria. The conformational search was run with the Monte Carlo algorithm implemented in Spartan'14 and using Merck Molecular Force Field (MMFF). The geometries were then optimized by DFT calculations first at the B3LYP/6-31G(d,p) level and then at the B3LYP/6-311G+(d,p) level of theory, first *in vacuo* and then using the PCM solvent model for CHCl₃. Next, all conformers with Boltzmann population of >1% at 298 K were considered for ECD (*in vacuo*) and VCD (*in vacuo* and CHCl₃) calculations. Frequency calculations were run at the B3LYP/6-311G+(d,p) level of theory using PCM for CHCl₃. IR and VCD spectra were obtained as sums of Lorentzian functions with a half-width at half height of 6 cm⁻¹, and a frequency scale factor of 0.985, and Boltzmann-averaged at 298 K using internal



energies. ECD calculations were run at the CAM-B3LYP/aug-cc-pVDZ and CAM-B3LYP/TZVP levels of theory *in vacuo*. ECD spectra were obtained as sums of Gaussian functions with an exponential half-width of 0.35 eV, using rotational strengths calculated in the dipole-length gauge, and Boltzmann averaged at 298 K using internal energies. Average spectra were generated with the software SpecDis, v. 1.70.^{22,23}

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors would like to thank The Oman Research Council (TRC) for the generous support through the project ORG/HSS/14/004. This work was also supported by the Wrocław Centre for Networking and Supercomputing (WCSS). M. G. gives thanks for the support from the Polish Ministry of Science and Higher Education ('Mobilnosc Plus' grant no. 1286/MOB/IV/2015/0).

References

- V. Lanzotti, Diterpenes for therapeutic use, in *Natural products, Phytochemistry, botany, and mechanism of alkaloids, phenols and terpenes*, ed. K. G. Ramawati and J. M. Merilon, Springer-Verlag, Berlin, Heidelberg, 2013, pp. 3171–3191.
- F. Pollastro, S. Golin, G. Chianese, M. Y. Putra, A. S. Moriello, L. D. Petrocellis, V. García, E. Munoz, O. Tagliatalata-Scafati and G. Appendino, *J. Nat. Prod.*, 2016, **79**, 1762–1768.
- L. F. Liang and Y. W. Guo, *Chem. Biodiversity*, 2013, **10**, 2161–2196.
- A. A. Zaki, *Biosens. Anal. Biotechnol.*, 2014, **1**, 1–3.
- G. Pescitelli, T. Kurtán and K. Krohn, Assignment of the Absolute Configurations of Natural Products by Means of Solid-State Electronic Circular Dichroism and Quantum Mechanical Calculations, in *Comprehensive Chiroptical Spectroscopy*, ed. N. Berova, R. W. Woody, P. Polavarapu and K. Nakanishi, Wiley, New York, 2012, pp. 145–176.
- J. Ren, Y. G. Wang, A. G. Wang, L. Q. Wu, H. J. Zhang, W. J. Wang, Y. L. Su and H. L. Qin, *J. Nat. Prod.*, 2015, **78**, 2322–2331.
- T. E. Alemika, G. O. Onawunmi and T. A. Olugbade, *Journal of Pharmacy and Bioresources*, 2004, **1**, 7–11.
- S. K. Avula, H. Hussain, R. Csuk, S. P. Sommerwerk, P. Liebing, M. Górecki, G. Pescitelli, A. Al-Rawahi, N. Ur Rehman, I. R. Green and A. Al-Harrasi, *Tetrahedron: Asymmetry*, 2016, **27**, 829–833.
- G. Pescitelli, T. Kurtán, U. Flörke and K. Krohn, *Chirality*, 2009, **21**, E181–E201.
- T. Kurtán, R. Jia, Y. Li, G. Pescitelli and Y.-W. Guo, *Eur. J. Org. Chem.*, 2012, 6722–6728.
- A. Cimmino, V. Mathieu, M. Masi, R. Baroncelli, A. Boari, G. Pescitelli, M. Ferderin, R. Lisy, M. Evidente, A. Tuzi, M. C. Zonno, A. Kornienko, R. Kiss and A. Evidente, *J. Nat. Prod.*, 2016, **79**, 116–125.
- P. Zhang, L.-H. Meng, A. Mandi, X.-M. Li, T. Kurtán and B.-G. Wang, *RSC Adv.*, 2015, **5**, 39870–39877.
- P. Sun, D. X. Xu, A. Mandi, T. Kurtán, T. J. Li, B. Schulz and W. Zhang, *J. Org. Chem.*, 2013, **78**, 7030–7047.
- S. Lu, T. Kurtán, G. J. Yang, P. Sun, A. Mandi, K. Krohn, S. Draeger, B. Schulz, Y. H. Yi, L. Li and W. Zhang, *Eur. J. Org. Chem.*, 2011, **28**, 5452–5459.
- S. Lu, P. Sun, T. Li, T. Kurtán, A. Mandi, S. Antuš, K. Krohn, S. Draeger, B. Schulz, Y. Yi, L. Li and W. Zhang, *J. Org. Chem.*, 2011, **76**, 9699–9710.
- H.-W. Liu, X.-Z. Yu, D. Padula, G. Pescitelli, Z.-W. Lin, F. Wang, K. Ding, M. Lei and J.-M. Gao, *Eur. J. Med. Chem.*, 2013, **59**, 265–273.
- J. Frelek, M. Górecki, M. Łaszcz, A. Suszczyńska, E. Vass and W. J. Szczepek, *Chem. Commun.*, 2012, 5295–5297.
- G. Pescitelli and T. Bruhn, *Chirality*, 2016, **28**, 466–474.
- S. Superchi, P. Scafato, M. Górecki and G. Pescitelli, *Curr. Med. Chem.*, 2017, **24**, DOI: 10.2174/0929867324666170310112009.
- K. Krohn, U. Farooq, U. Flörke, B. Schulz, S. Draeger, G. Pescitelli, P. Salvadori, S. Antuš and T. Kurtán, *Eur. J. Org. Chem.*, 2007, 3206–3211.
- M. Saleem, H. Hussain, I. Ahmed, S. Draeger, B. Schulz, K. Meier, M. Steinert, G. Pescitelli, T. Kurtán, U. Flörke and K. Krohn, *Eur. J. Org. Chem.*, 2011, 808–812.
- T. Bruhn, A. Schaumlöffel, Y. Hemberger and G. Bringmann, *Chirality*, 2013, **25**, 243–249.
- T. Bruhn, A. Schaumlöffel, Y. Hemberger and G. Pescitelli, *SpecDis version 1.70*, Berlin, Germany, 2017. <https://specdis-software.jimdo.com/>.

