



Cite this: *RSC Adv.*, 2017, 7, 41435

# Utilization of chromic polydiacetylene assemblies as a platform to probe specific binding between drug and RNA†

Anothai Kamphan,<sup>ab</sup> Changjun Gong,<sup>c</sup> Krishnagopal Maiti,<sup>c</sup> Souvik Sur,<sup>c</sup> Rakchart Traiphol<sup>\*abd</sup> and Dev P. Arya<sup>ib\* c</sup>

Recognition of nucleic acids remains an important endeavor in biology. Nucleic acids adopt shapes ranging from A-form (RNA and GC rich DNA) to B-form (AT rich DNA). We show, in this contribution, shape-specific recognition of A–U rich RNA duplex by a neomycin (Neo)–polydiacetylene (PDA) complex. PDA assemblies are fabricated by using a well-known diacetylene (DA) monomer, 10,12-pentacosadiynoic acid (PCDA). The response of poly(PCDA) assemblies is generated by mixing with a modified neomycin–PCDA monomer (Neo–PCDA). The functionalization by neomycin moiety provides specific binding with homopolyribonucleotide poly(rA)–poly(rU) stimulus. Various types of alcohols are utilized as additives to enhance the sensitivity of poly(PCDA)/Neo–PCDA assemblies. A change of absorption spectra is clearly observed when a relatively low concentration of poly(rA)–poly(rU) is added into the system. Furthermore, poly(PCDA)/Neo–PCDA shows a clear specificity for poly(rA)–poly(rU) over the corresponding DNA duplex. The variation of linker between neomycin moiety and conjugated PDA backbone is found to significantly affect its sensitivity. We also investigate other parameters including the concentration of Neo–PCDA and the DA monomer structure. Our results provide here preliminary data for an alternative approach to improve the sensitivity of PDA utilized in biosensing and diagnostic applications.

Received 28th June 2017  
Accepted 2nd August 2017

DOI: 10.1039/c7ra07178g

rsc.li/rsc-advances

## 1. Introduction

A number of antibiotics that stop the growth of foreign pathogens, such as bacteria and viruses, are known to bind nucleic acids such as DNA and RNA. One such class of antibiotics are aminoglycosides, well known to bind bacterial ribosomal RNA.<sup>1</sup> Neomycin is one such antibiotic, belonging to aminoglycosides containing the deoxystreptamine core. The binding of neomycin, and other aminoglycosides drugs, to bacterial RNA affects the production of proteins through miscoding in translation, ultimately leading to bacterial death.<sup>2–5</sup> In addition to binding to ribosomal RNA, aminoglycosides have been shown to bind numerous other RNA,<sup>6–10</sup> DNA,<sup>11–17</sup> hybrid duplex,<sup>18,19</sup>

triplex<sup>20–23</sup> and quadruplex<sup>24–26</sup> structures of therapeutic interest.<sup>27</sup>

Conventional methods for the study of neomycin drug–RNA interactions utilize established analytical tools. For example, circular dichroism (CD) spectroscopy is used to study conformational changes of RNA during ligand–RNA binding.<sup>11,28</sup> Isothermal titration calorimetry (ITC), differential scanning calorimetry (DSC) and UV-vis spectroscopy are used to elucidate the thermodynamic parameters of ligand–RNA interactions.<sup>11,28</sup> Fluorescence intercalator displacement (FID) assays can be used for the study of selectivity, affinity, stoichiometry and binding site size of RNA and ligand.<sup>11,28</sup> New and simple approaches that can help us study small molecule's interaction with biopolymers would be welcome addition to these established rigorous methods of ligand–nucleic acids interactions.

Polydiacetylene (PDA) is a conjugated polymer, which has received tremendous attention as a chromatic sensor. PDA can be prepared *via* 1,4-addition of highly organized assemblies of diacetylene monomers, normally initiated by UV light irradiation.<sup>29,30</sup> PDA possesses color transition, typically from blue to red, upon the exposure to various external stimuli such as heat,<sup>31–42</sup> acids and bases,<sup>40,43–46</sup> chemicals<sup>47–56</sup> and biomolecules.<sup>57–66</sup> The perturbation by these stimuli generally weakens segmental interactions within the PDA assemblies. This results in the reorientation of conjugated backbone and alkyl chain. The decrease of conjugation length of PDA provides the shift of

<sup>a</sup>Department of Chemistry, Faculty of Science, Naresuan University, Phitsanulok 65000, Thailand

<sup>b</sup>Laboratory of Advanced Polymers and Nanomaterials, School of Materials Science and Engineering, Center of Excellence for Innovation in Chemistry, Faculty of Science, Mahidol University at Salaya, Phuttamonthon 4 Road, Salaya, Nakhon Pathom 73170, Thailand

<sup>c</sup>Laboratories of Medicinal Chemistry, Department of Chemistry, Clemson University, Clemson, South Carolina 29634, USA. E-mail: dparya@clemson.edu

<sup>d</sup>NANOTEC-MU Excellence Center on Intelligent Materials and Systems, Faculty of Science, Mahidol University, Rama 6 Road, Ratchathewi, Bangkok 10400, Thailand. E-mail: rakchart.tra@mahidol.ac.th

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c7ra07178g



absorption spectrum to lower energy region and hence induces color transition.<sup>30,67–70</sup> Many prior reports demonstrate the ability of PDA-based biosensors for the detection of viral RNA with quick and reliable testing without requiring advanced instruments. Amano *et al.* reviewed the construction of PDA with desirable physical, optical and electrical properties that responds to the influenza virus's RNA. The exposure of influenza virus to modified-PDA solution causes blue shift in the absorption spectra, corresponding to blue-to-red color transition of PDA.<sup>71</sup> Park *et al.* demonstrates the detection of RNA by using PDA conjugated with DNA complementary to RNA of pathogenic bacteria. The hybridization of a specific DNA to RNA enhances fluorescent intensity of PDA-based sensor, leading to the application as a biochip in the detection of pathogenic microorganisms.<sup>72</sup> In this report, we demonstrate the use of PDA in a modified approach for the investigation of RNA–ligand interactions.

Here, PCDA modified neomycin head groups (Neo-PCDA) is synthesized to generate a specific binding site for RNA. Previous calorimetry and FID studies have shown that the binding of poly(rA)–poly(rU), a homopolymeric dsRNA, with neomycin has high  $K_a$  values ( $\sim 10^7 \text{ M}^{-1}$ ), and low  $AC_{50}$  ( $0.02 \pm 0.01 \mu\text{M}$ ) respectively, representing high affinities.<sup>11,14</sup> These studies are in concert with other data showing nanomolar binding of neomycin to RNA targets. Therefore, poly(rA)–poly(rU) was used here as a model RNA for investigating the neomycin–RNA interaction. We also demonstrate here that the sensitivity of poly(PCDA)/Neo-PCDA assemblies can be significantly improved by using alcohol additives. An addition of alcohols weakens the strength of interactions within the poly(PCDA)/Neo-PCDA assemblies resulting in the increase of colorimetric response compared to its pure system.<sup>73</sup> The developments of specific and rapid color transition of PDA to biomolecules could open wide-range of possibilities for biosensor, drug detection, and diagnostic applications.

## 2. Experiment

### 2.1 The synthesis of Neo-PCDA1 and Neo-PCDA2

The chemical structures of neomycin Neo-PCDA1 and Neo-PCDA2 are shown in Fig. 1. Neo-PCDA1 and Neo-PCDA2 were synthesized as shown in Schemes 1 and 2. Boc-protected

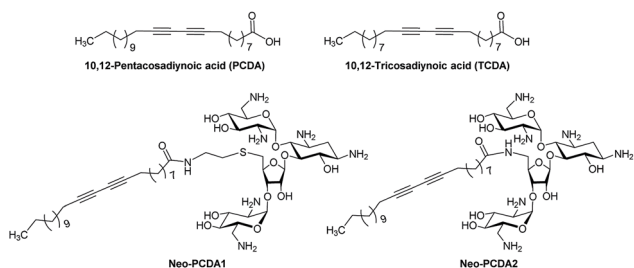
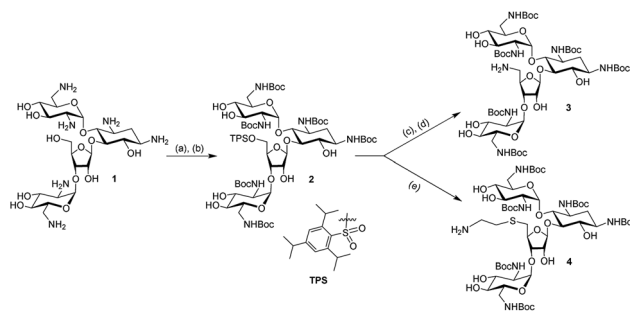
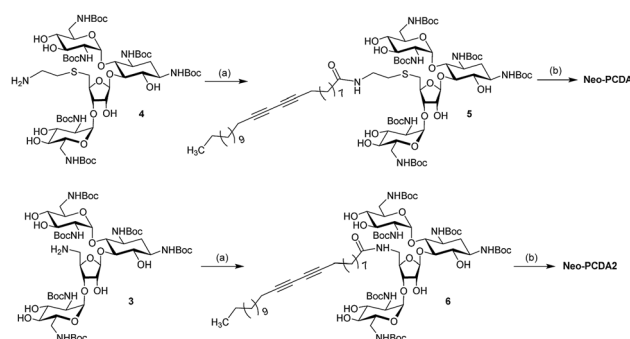


Fig. 1 Chemical structures of diacetylene monomer, 10,12-pentacosadiynoic acid (PCDA) and 10,12-tricosadiynoic acid (TCDA); and synthesized neomycin–PCDA conjugates, Neo-PCDA1 and Neo-PCDA2.



Scheme 1 Reagents and conditions: (a)  $(\text{Boc})_2\text{O}$ ,  $\text{H}_2\text{O}$ , methanol,  $\text{Na}_2\text{CO}_3$ , r.t., 18 h, 65%; (b) 2,4,6-triisopropylbenzenesulfonyl chloride, pyridine, r.t., 40 h, 50%; (c)  $\text{NaN}_3$ ,  $\text{DMF}/\text{H}_2\text{O}$ ,  $90^\circ\text{C}$ , 12 h, 95%; (d)  $\text{H}_2$ ,  $\text{Pd/C}$ , ethanol, r.t., 12 h, quantitative; (e)  $\text{HSCH}_2\text{CH}_2\text{NH}_2$ , ethanol,  $\text{Na}(\text{metal})$ , r.t., 12 h, 60%.



Scheme 2 Reagents and conditions: (a) 10,12-pentacosadiynoic acid (PCDA), TBTU, DIPEA,  $\text{DMF}$ , r.t., 18 h, 80% (for 5) and 74% (for 6); (b)  $\text{TFA}$ ,  $\text{CH}_2\text{Cl}_2$ , r.t., 3 h, 76% (for Neo-PCDA1) and 71% (for Neo-PCDA2).

neomycin amine derivatives 3 and 4 were prepared from commercially available neomycin B (1) according to published schemes (Scheme 1).<sup>11,14,22,74–76</sup> All the compounds were characterized and spectral data was matched with the reported values from the literatures. Amine functionalized neomycin derivatives 3 and 4 were coupled with 10,12-pentacosadiynoic acid (PCDA) monomer (Aldrich) in the presence of coupling reagent *O*-(Benzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium tetrafluoroborate (TBTU) in  $\text{DMF}$  solution, followed by deprotection with trifluoroacetic acid in dichloromethane, leading to neomycin-PCDA conjugates, Neo-PCDA1 and Neo-PCDA2 (Scheme 2). All the four new compounds 5, 6, Neo-PCDA1 and Neo-PCDA2 were characterized by nuclear magnetic resonance spectroscopy (NMR, Bruker Advance-500 and 300) and MALDI-TOF mass spectrometry (Bruker Microflex) and the spectra are provided in the ESI.† One of the  $\alpha$ -anomeric protons of two pyranoside units in 5, appeared as a singlet at 5.38 ppm whereas the  $\beta$ -anomeric proton of furanoside unit resonates at 5.15 ppm as a singlet, in  $^1\text{H}$  NMR spectrum of 5 (Fig. S2†). The peak at 99.09 and 100.52 ppm corresponded to  $\alpha$ -anomeric carbon of two pyranoside moieties whereas  $\beta$ -anomeric carbon of furanoside moiety appeared at 111.08 in  $^{13}\text{C}$  NMR spectrum of 5 (Fig. S3†). Further, the formation of the compound 5 was confirmed by the appearance of molecular ion peak at 1653.268



$[M + Na]^+$ , as the base peak in MALDI-TOF mass spectrum of **5** (Fig. S1†). In  $^1H$  NMR spectrum of **6** (Fig. S8†),  $\beta$ -anomeric proton of the furanoside moiety appeared as a singlet at 5.08 ppm whereas the corresponding anomeric carbon resonated at 111.48 ppm, in  $^{13}C$  NMR spectrum of **6** (Fig. S9†). The anomeric peaks of compound **6** were assigned by the help of HMQC spectrum (Fig. S10†). The formation of compound **6** was further confirmed by MALDI-TOF mass spectrum (Fig. S7†), where the molecular ion peak appeared as the base peak at 1593.065  $[M + Na]^+$ .

The details of the synthetic procedures for compounds **5**, **6**, Neo-PCDA1 and Neo-PCDA2 are given below.

**2.1.1. Compound 5.** To a solution of compound **4** (0.1 g, 0.078 mmol) in DMF (1 mL), the mixture of 10,12-pentacosadiynoic acid (0.146 g, 0.39 mmol) and TBTU (0.126 g, 0.39 mmol) in DMF (0.5 mL) was added. DIPEA (0.136 mL, 0.78 mmol) was added to the reaction mixture and it was stirred at room temperature for 18 h. The solvent was removed and the crude reaction mixture was purified by column chromatography (silica gel) to furnish compound **5** (0.102 g, 80%);  $^1H$  NMR ( $CD_3OD$ , 300 MHz)  $\delta$  5.38 (s, 1H), 5.15 (s, 1H), 4.26 (s, 2H), 3.91 (s, 1H), 3.77–3.71 (m, 2H), 3.60–3.39 (m, 13H), 3.22–3.16 (m, 3H), 2.78–2.74 (m, 2H), 2.36 (t,  $J = 7.9$  Hz, 2H), 2.27–2.20 (m, 6H), 1.99–1.94 (m, 1H), 1.65–1.61 (m, 3H), 1.47–1.44 (m, 57H), 1.34–1.30 (m, 27H), 0.90 (t,  $J = 6.6$  Hz, 3H);  $^{13}C$  NMR ( $CD_3OD$ , 75 MHz)  $\delta$  176.36, 159.08, 158.88, 158.48, 158.21, 158.18, 157.92, 111.08, 100.52, 99.09, 87.02, 82.67, 80.73, 80.59, 80.40, 80.01, 77.86, 75.71, 75.65, 74.54, 74.48, 73.25, 72.87, 71.57, 68.89, 66.44, 58.09, 53.60, 50.81, 41.69, 40.09, 37.18, 36.95, 33.06, 31.68, 31.64, 30.74, 30.66, 30.58, 30.45, 30.31, 30.26, 30.12, 30.05, 29.84, 29.78, 29.52, 29.02, 28.88, 28.85, 28.76, 26.99, 23.72, 19.69, 18.53, 14.45; MALDI-TOF  $m/z$  calcd for  $C_{80}H_{139}N_7O_{25}SNa [M + Na]^+$ : 1652.94, found 1653.268.

**2.1.2. Compound 6.** To a solution of compound **3** (0.1 g, 0.082 mmol) in DMF (1 mL), the mixture of 10,12-pentacosadiynoic acid (0.154 g, 0.41 mmol) and TBTU (0.132 g, 0.41 mmol) in DMF (0.5 mL) was added. DIPEA (0.142 mL, 0.82 mmol) was added to the reaction mixture and it was stirred at room temperature for 18 h. The solvent was removed and the crude reaction mixture was purified by column chromatography (silica gel) to furnish compound **6** (0.96 g, 74%);  $^1H$  NMR ( $CD_3OD$ , 500 MHz)  $\delta$  5.48 (s, 1H), 5.08 (s, 1H), 4.30 (s, 1H), 4.06 (s, 1H), 3.93–3.86 (m, 4H), 3.75 (s, 2H), 3.56–3.50 (m, 11H), 3.25–3.11 (m, 3H), 2.34–2.30 (m, 3H), 2.24 (t,  $J = 6.8$  Hz, 3H), 1.95–1.93 (m, 1H), 1.66–1.63 (m, 2H), 1.47–1.42 (m, 65H), 1.34–1.30 (m, 20H), 0.90 (t,  $J = 7.0$  Hz, 3H);  $^{13}C$  NMR ( $CD_3OD$ , 125 MHz)  $\delta$  176.58, 159.06, 158.87, 158.55, 158.49, 158.25, 157.89, 111.48, 100.42, 98.80, 88.47, 80.94, 80.74, 80.66, 80.36, 80.31, 79.47, 77.83, 76.20, 75.51, 74.42, 73.35, 72.69, 72.61, 72.55, 71.65, 68.99, 66.46, 66.44, 56.80, 53.53, 52.50, 51.30, 49.85, 43.91, 42.64, 41.95, 36.96, 35.69, 33.05, 30.73, 30.66, 30.57, 30.44, 30.39, 30.35, 30.12, 29.86, 29.83, 29.54, 29.49, 29.02, 28.89, 28.84, 28.80, 28.74, 27.19, 23.72, 19.68, 14.46; MALDI-TOF  $m/z$  calcd for  $C_{78}H_{135}N_7O_{25}Na [M + Na]^+$ : 1592.94, found 1593.065.

**2.1.3. Neo-PCDA1.** To a solution of compound **5** (0.07 g, 0.043 mmol) in dichloromethane (0.5 mL) trifluoroacetic acid (0.5 mL) was added and stirred at room temperature for 3 h. The

solvent was removed under vacuum and the residue was dissolved in deionized water (2 mL) and washed with diethylether ( $3 \times 10$  mL). Lyophilization of the aqueous solution yielded a pale yellow powder of Neo-PCDA1 (0.034 g, 76%);  $^1H$  NMR (500 MHz,  $D_2O$ )  $\delta$  5.93 (d,  $J = 4.0$  Hz, 1H), 5.32 (d,  $J = 1.8$  Hz, 1H), 5.19 (s, 1H), 4.45–4.37 (m, 1H), 4.31 (m, 1H), 4.21 (t,  $J = 4.9$  Hz, 1H), 4.13 (m, 2H), 4.00 (t,  $J = 9.7$  Hz, 1H), 3.94–3.76 (m, 5H), 3.72 (s, 1H), 3.69–3.55 (m, 5H), 3.52–3.41 (m, 3H), 3.40–3.21 (m, 8H), 3.18 (m, 1H), 3.08 (d,  $J = 7.4$  Hz, 2H), 2.42–2.37 (m, 1H), 2.16 (s, 2H), 1.79 (m, 1H), 1.44 (s, 2H), 1.21 (m, 30H);  $^{13}C$  NMR (75 MHz,  $D_2O$ )  $\delta$  163.55, 163.09, 162.61, 162.14, 122.10, 118.24, 114.37, 110.51, 110.21, 96.05, 95.46, 95.33, 84.79, 81.50, 77.71, 75.21, 73.57, 72.46, 70.57, 70.07, 69.41, 69.17, 68.09, 67.93, 67.60, 67.33, 60.18, 54.34, 53.45, 50.83, 49.75, 49.64, 48.38, 42.52, 40.42, 40.08, 32.04, 29.85, 29.16, 28.09, 22.72, 17.68, 16.21, 13.96, 12.09; MALDI-TOF  $m/z$  calculated for  $C_{50}H_{91}N_7O_{13}S [M]^+$  1029.64, found 1030.75.

**2.1.4. Neo-PCDA2.** To a solution of compound **6** (0.07 g, 0.045 mmol) in dichloromethane (0.5 mL) trifluoroacetic acid (0.5 mL) was added and stirred at room temperature for 3 h. The solvent was removed under vacuum and the residue was dissolved in deionized water (2 mL) and washed with diethylether ( $3 \times 10$  mL). Lyophilization of the aqueous solution yielded a pale yellow powder of Neo-PCDA2 (0.031 g, 71%);  $^1H$  NMR (500 MHz,  $D_2O$ )  $\delta$  5.93 (d,  $J = 3.9$  Hz, 1H), 5.33 (s, 1H), 5.19 (s, 1H), 4.46–4.37 (m, 1H), 4.32 (s, 1H), 4.22 (t,  $J = 4.9$  Hz, 1H), 4.13 (m, 2H), 4.00 (t,  $J = 9.7$  Hz, 1H), 3.93–3.80 (m, 4H), 3.73 (s, 1H), 3.68–3.55 (m, 5H), 3.50–3.44 (m, 2H), 3.42–3.23 (m, 7H), 3.19 (m, 1H), 3.08 (m, 2H), 2.42–2.37 (m, 1H), 2.14 (s, 2H), 1.85–1.77 (m, 1H), 1.39 (s, 2H), 1.22 (m, 30H);  $^{13}C$  NMR (126 MHz,  $D_2O$ )  $\delta$  163.30, 163.02, 162.74, 162.45, 119.82, 117.50, 115.18, 112.86, 110.20, 96.06, 95.47, 95.33, 84.78, 81.49, 77.72, 75.21, 75.15, 73.56, 72.46, 70.61, 70.56, 70.08, 69.40, 69.17, 68.19, 67.99, 67.61, 67.33, 62.50, 60.17, 54.34, 53.44, 50.83, 49.76, 49.64, 48.38, 42.52, 40.43, 40.06, 32.01, 29.81, 28.23, 27.97, 22.71, 17.69, 16.22, 13.97, 12.10; MALDI-TOF  $m/z$  calculated for  $C_{48}H_{87}N_7O_{13} [M]^+$  970.27, found 970.71.

## 2.2 The preparation of PDA/Neo-PCDA assemblies

The assemblies of PDA/Neo-PCDA were prepared by using two DA monomers, including 10,12-pentacosadiynoic acid (PCDA) and 10,12-tricosadiynoic acid (TCDA). DA monomer and Neo-PCDA powder were dissolved in ethanol and then filtered by using a 0.45  $\mu m$  nylon filter to remove polymerized impurities. The ratios of Neo-PCDA were 0, 5, 10, 15 and 20 mol%. The DA/Neo-PCDA solutions in ethanol were slowly dried at 60  $^\circ C$  in a water bath. Phosphate-buffered saline solutions (PBS buffer) at various pH values (6, 7, and 8) were added into the DA/Neo-PCDA films to provide 0.25 mM aqueous suspension. The pH of PBS buffer was adjusted by using HCl and NaOH solutions. The samples were sonicated at 70  $^\circ C$  for 90 min to disperse DA/Neo-PCDA films into aqueous medium. The suspensions were allowed to equilibrate at room temperature and then stored at 4  $^\circ C$  overnight to induce self-assembly process. The cloudy suspensions of DA/Neo-PCDA were irradiated under UV light (10 W,  $\lambda \sim 254$  nm) for 5 min, resulting in blue PDA/Neo-PCDA





vesicles. Effect of Neo-PCDA structure was studied by using Neo-PCDA1 and Neo-PCDA2 prepared by the same method.

### 2.3 The study of colorimetric response of PDA/Neo-PCDA to poly(rA)-poly(rU)

Polyadenylic-uridylic acid (Poly(rA)-poly(rU)) was purchased from Midland Certified Company. Tuning colorimetric response of PDA/Neo-PCDA to poly(rA)-poly(rU) was induced by using two alcohols, butanol and hexanol. The alcohols were added into PDA/Neo-PCDA solution at various concentrations by using a micropipette. After the addition of alcohols, the solution was shaken by using vortex for a few seconds. Poly(rA)-poly(rU) solution in PBS buffer (pH 8) was added into PDA/Neo-PCDA solution at various concentrations by using a micropipette. The mixture was shaken for a few seconds prior to each addition. The color transition of PDA/Neo-PCDA assemblies was followed by measuring the absorption spectra (UV-vis spectrophotometer, Agilent technologies, Cary series). The concentration of PDA/Neo-PCDA assemblies in each experiment was controlled to be the same by adjusting the absorbance value. To quantify the extent of blue-to-red color transition of PDA/Neo-PCDA assemblies, the colorimetric response (% CR) was defined and calculated as follows: % CR =  $[(PB_0 - PB)/PB_0] \times 100$ . The PB is the percent blue calculated from  $A_{640}/(A_{540} + A_{640})$ , where  $A_{540}$  and  $A_{640}$  are the absorbance of red ( $\lambda = 540$ ) and blue ( $\lambda = 640$ ) phases of vesicles, respectively. The initial  $PB_0$  value was determined before the addition of poly(rA)-poly(rU). Morphology of these solutions was also studied by using transmission electron microscope (TEM, Hitachi H7600). The samples were prepared by dropping diluted PDA/Neo-PCDA solutions onto copper grids, followed by air-drying at room temperature.

The specificity of PDA/Neo-PCDA was studied by testing with a control DNA sequence containing only adenine and thymine bases. The poly(PCDA)/Neo-PCDA1 containing 0.5% v/v hexanol was used. The solution of control DNA in PBS buffer (pH 8) was added into poly(PCDA)/Neo-PCDA1 solution by using the same testing process as poly(rA)-poly(rU) system. The colorimetric response of poly(PCDA)/Neo-PCDA1 vesicles was followed by measuring the absorption spectra.

## 3. Results and discussion

### 3.1 Morphologies of poly(PCDA)/Neo-PCDA assemblies and the optimization

Pure poly(PCDA), poly(PCDA)/Neo-PCDA1 and poly(PCDA)/Neo-PCDA2 solutions show a blue color, indicating proper packing of PCDA monomers required for the topolymerization process. Morphologies of resultant poly(PCDA)/Neo-PCDA assemblies revealed by TEM are shown in Fig. 2 (bottom). A model of the Neo-PCDA compounds bound to RNA is also included in Fig. 2 (top). It has been observed that pure poly(PCDA) shows spherical shape with featureless internal pattern.<sup>21</sup> The TEM image of poly(PCDA)/Neo-PCDA1 also shows spherical particles with diameter of about 100 nm (see Fig. 2a). In contrast, poly(PCDA)/Neo-PCDA2 assemblies

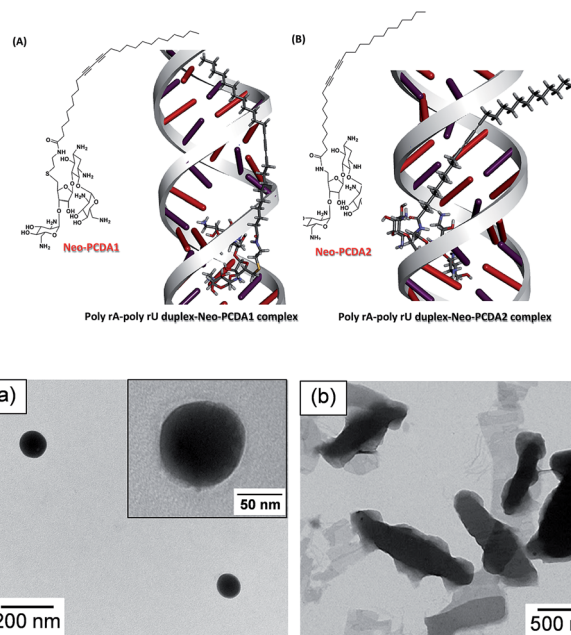


Fig. 2 In silico models (above) and TEM images (below) of poly(PCDA) prepared by mixing with 10 mol% of (a) Neo-PCDA1 and (b) Neo-PCDA2.

exhibit relatively large rod-like and sheet-like shapes as shown in Fig. 2b. This is attributed to the difference of chemical linker between PCDA and neomycin head group (see Fig. 1). The Neo-PCDA2 constitutes much shorter linker compared to that of the Neo-PCDA1, which in turn significantly affects their packing structure. Neo-PCDA2 has a shorter linker compared to Neo-PCDA1 (see Fig. 1), resulting in a larger head group repulsion. This provides the increase of head group area and the decrease in the packing parameter, leading to the differences in molecular organization and ultimately different shape and size of poly(PCDA)/Neo-PCDA assembly.<sup>39,77,78</sup> Jung,<sup>66,79</sup> Cheng<sup>80</sup> and Ye<sup>81</sup> also demonstrate that the variation of head group structure provides different shape and size of PDA assemblies. Flower-like morphology, rectangular shape, fiber, ribbon with various size in the range of nanometer to micrometer are observed. It is important to note that the differences of shape and size of PDA significantly affect its colorimetric response as well. This will be discussed in the following section.

To achieve the highest response of poly(PCDA)/Neo-PCDA assemblies to poly(rA)-poly(rU), concentration of Neo-PCDA and pH of PBS buffer were optimized. We observed that the poly(PCDA)/Neo-PCDA1 assemblies containing 20 mol% of Neo-PCDA1 at pH 8 provided the highest response to poly(rA)-poly(rU) (see Fig. S14 in ESI†). RNA-binding affinity of an aminoglycoside is known to vary with the change in pH, as the protonated states of the amino groups are affected.<sup>82</sup> However, this system exhibits large precipitation, which is clearly undesirable for sensing applications. Therefore, the highest possible concentration of Neo-PCDA, 10 mol%, providing clear blue solution and no precipitation of the assemblies, was used for the entire study.



### 3.2 Colorimetric response of poly(PCDA)/Neo-PCDA1 to poly(rA)-poly(rU)

Poly(PCDA)/Neo-PCDA1 assemblies were prepared by mixing of PCDA and 10 mol% Neo-PCDA1 in PBS buffer at pH 8. Initial absorption spectrum of poly(PCDA)/Neo-PCDA1 exhibits  $\lambda_{\text{max}}$  at  $\sim 640$  nm. The spectrum hardly changes after the addition of poly(rA)-poly(rU) up to  $\sim 13 \mu\text{M}$  (see Fig. 3a). Interestingly, the colorimetric response of poly(PCDA)/Neo-PCDA1 drastically increases when a small amount of hexanol, 0.5% v/v, is added into the system. Absorption peak at  $\sim 540$  nm grows significantly upon the addition of poly(rA)-poly(rU) (see Fig. 3b), indicating the formation of red poly(PCDA)/Neo-PCDA1. A blue-to-purple color transition of poly(PCDA)/Neo-PCDA1 can be observed as shown in Fig. 3d. According to our previous studies, the added hexanol can penetrate into the poly(PCDA)/Neo-PCDA1 layers which in turn weakens inter-/intrachain interactions within the assemblies.<sup>51,52,73</sup> This perturbation enhances colorimetric response of poly(PCDA)/Neo-PCDA1 to poly(rA)-poly(rU).

The study of its morphology by TEM supports our hypothesis. The morphology of poly(PCDA)/Neo-PCDA1 changes from spherical shape to sheet-like structure after the addition of 0.5% v/v hexanol as shown in Fig. 4a. This observation suggests that the presence of hexanol induces segmental reorganization within the assemblies, enhancing the colorimetric response to

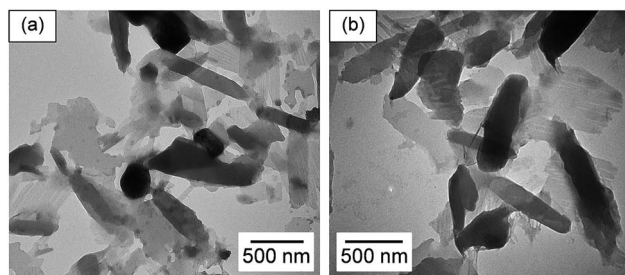


Fig. 4 TEM images of poly(PCDA)/Neo-PCDA1 (a) with the addition of 0.5% v/v hexanol and then (b) was disturbed by the addition of 9.52  $\mu\text{M}$  poly(rA)-poly(rU).

poly(rA)-poly(rU). We also observe that the perturbation by poly(rA)-poly(rU) further induces the agglomeration of poly(PCDA)/Neo-PCDA1 (see Fig. 4b). It is worthwhile to note that the morphologies revealed by TEM could be different from those of PDA assemblies in the solution state due to the sample preparation process. However, the change of morphology in dried state still reflects the properties of PDA assemblies in solution.

The plots of % CR clearly show that the increase of hexanol concentration systematically enhances poly(PCDA)/Neo-PCDA1 response to poly(rA)-poly(rU) (see Fig. 3c). The higher amounts of hexanol causes increased perturbation of poly(PCDA)/Neo-PCDA1 assemblies,<sup>51,52,73</sup> resulting in more blue-to-purple color transition as shown in Fig. 3d. However, the further increase of hexanol concentration results in a cloudy suspension arising from its limited solubility in water.<sup>83</sup> A control set of poly(PCDA)/Neo-PCDA1 assemblies containing 0.5% v/v hexanol was also investigated. The absorption spectra of this solution were measured at the same time interval with the measurement of other solutions without the addition of poly(rA)-poly(rU) solution. This system shows a slight increase of CR value to  $\sim 8\%$  while the addition of poly(rA)-poly(rU) provides the CR value of  $\sim 20\%$  as shown in Fig. 3c. The difference of these two sets indicates the color transition of poly(PCDA)/Neo-PCDA1 induced by interaction between Neo-PCDA1 and poly(rA)-poly(rU).

Schematic illustration in Fig. 5a concludes our study in this first part. The self-assembly of PCDA and Neo-PCDA monomers takes place in a solution state. A blue phase of poly(PCDA)/Neo-PCDA assemblies is obtained *via* photopolymerization process. The insertion of alcohols into poly(PCDA)/Neo-PCDA layers weakens the interactions within the system, which in turn enhances the sensitivity. The specific binding between neomycin moiety and poly(rA)-poly(rU) induces the color transition. In this study, butanol was also used as an additive in order to investigate the effect of alcohol alkyl tail on the colorimetric response of poly(PCDA)/Neo-PCDA1. The highest concentration of butanol, 5% v/v, was added into poly(PCDA)/Neo-PCDA1 while the suspension still remained blue. We observe that the addition of butanol provides lower response of poly(PCDA)/Neo-PCDA1 compared to the use of hexanol (see Fig. 5b). Although the added concentration of butanol is 10 times higher than that of hexanol, the % CR reaches plateau at

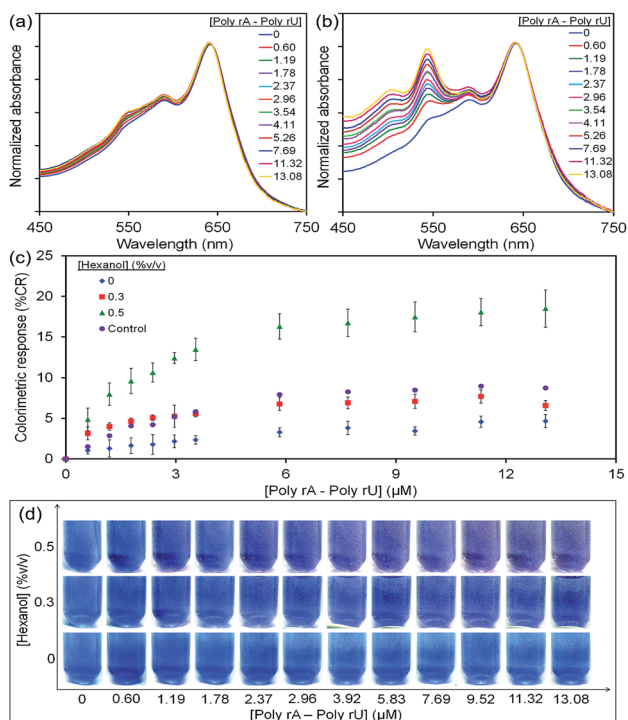


Fig. 3 Absorption spectra of (a) pure poly(PCDA)/Neo-PCDA1 and (b) with the addition of 0.5% v/v hexanol upon testing with poly(rA)-poly(rU). (c) The plots of % CR and (d) color photographs show color transition of poly(PCDA)/Neo-PCDA1 upon increasing poly(rA)-poly(rU) concentration. Error bars are obtained from 3 experiments (control: poly(PCDA)/Neo-PCDA1 with the addition of only 0.5% v/v hexanol, no addition of poly(rA)-poly(rU)).



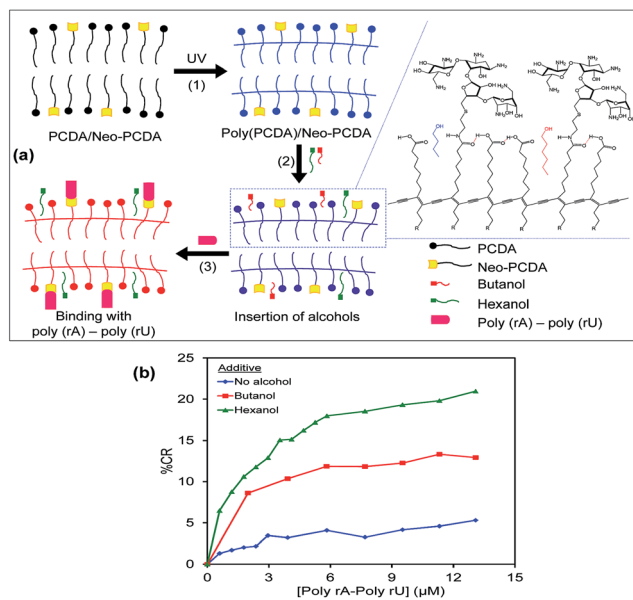


Fig. 5 (a) Model illustrates the formation of poly(PCDA)/Neo-PCDA, the insertion of alcohols and the binding between neomycin moiety with poly(rA)-poly(rU). (b) The plot shows the effect of alcohol additives on sensitivity of poly(PCDA)/Neo-PCDA1 to poly(rA)-poly(rU). Concentrations of the added alcohols are 5% v/v for butanol and 0.5% v/v for hexanol.

about 12% upon increasing concentration of poly(rA)-poly(rU) to  $\sim 13 \mu\text{M}$ . Our results suggest that hexanol with relatively long chain can penetrate into deeper region of poly(PCDA) layer. This induces larger extent of perturbation (see model in Fig. 5a), providing higher colorimetric response.<sup>51,73</sup> It is possible to use other types of alcohols as additives to enhance the sensitivity of PDA. The effects of alcohol structures on colorimetric response of PDAs are previously reported by our group.<sup>51,73</sup> It is worthwhile to note that a phospholipid (dimyristoylphosphatidylcholine, DMPC) is normally used for improving sensitivity of PDA assemblies to biomolecules in previous studies.<sup>53,60,61</sup> We demonstrate in this study that alcohol is an interesting additive for enhancing the sensitivity of PDA to biomolecules. The alcohol is also much cheaper compared to the DMPC.

### 3.3 The specificity of poly(PCDA)/Neo-PCDA1 to poly(rA)-poly(rU)

High specificity of PDA to its target biomolecules is necessary for the utilization as a biosensor. Here, we explore the specificity of poly(PCDA)/Neo-PCDA1 to poly(rA)-poly(rU) by comparing the colorimetric response to an AT rich DNA (5'-ATATATATA-TATATAT-3'). The results are illustrated in Fig. 6. As discussed earlier, the addition of  $13.06 \mu\text{M}$  of poly(rA)-poly(rU) induces the increase of % CR to  $\sim 22\%$ . In contrast, the addition of DNA hardly induces any colorimetric response. The plot of % CR is similar to that of the control set. No significantly change is observed in this system. The further increase of DNA concentration to  $800 \mu\text{M}$  still shows similar behavior. This result indicates that the poly(PCDA)/Neo-PCDA1 has high specificity to poly(rA)-poly(rU) attributed to the specific binding with

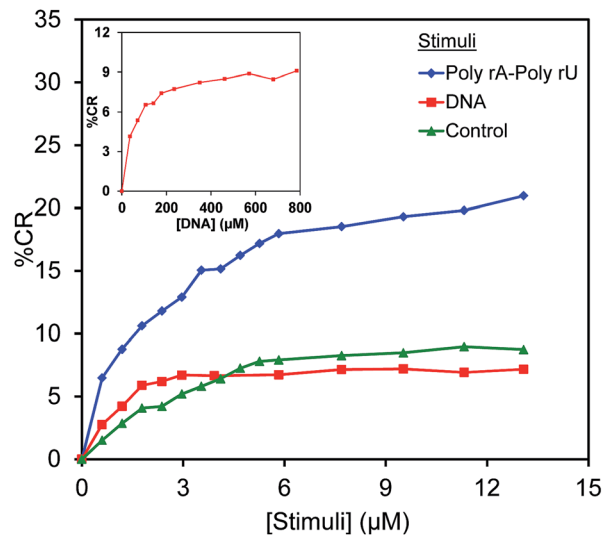


Fig. 6 The % CR curves of poly(PCDA)/Neo-PCDA1 containing 0.5% v/v hexanol upon testing with each stimulus. The inset shows the % CR of this system upon the addition of DNA at extremely high concentration (control: poly(PCDA)/Neo-PCDA1 with the addition of 0.5% v/v hexanol, no addition of poly(rA)-poly(rU)).

neomycin moiety. This corresponds to low  $AC_{50}$  for the binding of neomycin moiety and poly(rA)-poly(rU).<sup>11,28</sup>

### 3.4 The effect of Neo-PCDA structure on colorimetric response

Specific binding site of Neo-PCDA is an important key for the recognition of poly(rA)-poly(rU). Here, we investigate the effect of Neo-PCDA structure on the binding ability of poly(PCDA)/Neo-PCDA to poly(rA)-poly(rU). Our result shows that the change of linker between neomycin moiety and PCDA significantly influences the colorimetric response of their assemblies. The Neo-PCDA2 constitutes a shorter linker compared to that of the Neo-PCDA1 (see Fig. 1). Fig. 7a shows that the addition of poly(rA)-poly(rU) into poly(PCDA)/Neo-PCDA2 assemblies causes slight growth of absorption peak at  $\sim 540 \text{ nm}$ , corresponding to the formation of some red phase. The plot of % CR clearly

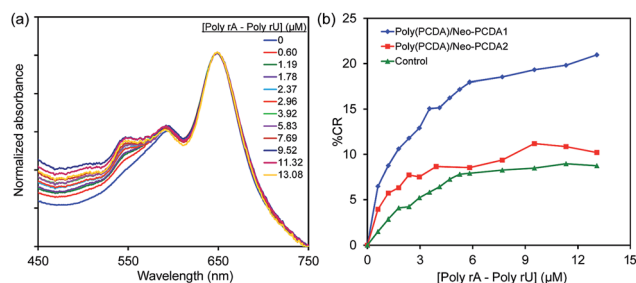


Fig. 7 (a) The absorption spectra of poly(PCDA)/Neo-PCDA2 containing 0.5% v/v hexanol upon testing with poly(rA)-poly(rU). (b) The plot of % CR shows different responses of poly(PCDA)/Neo-PCDA1 and poly(PCDA)/Neo-PCDA2 to poly(rA)-poly(rU) (control: poly(PCDA)/Neo-PCDA1 with the addition of 0.5% v/v hexanol, no addition of poly(rA)-poly(rU)).





shows that the color response of poly(PCDA)/Neo-PCDA2 is much lower than that of poly(PCDA)/Neo-PCDA1 (see Fig. 7b). The CR value of poly(PCDA)/Neo-PCDA2 system is slightly higher than that of the control set of poly(PCDA)/Neo-PCDA1 system. The study of their morphologies by TEM (see Fig. 2) reveals that the poly(PCDA)/Neo-PCDA1 and poly(PCDA)/Neo-PCDA2 assemblies exhibit a different shape and size. The poly(PCDA)/Neo-PCDA2 assemblies exhibit much larger size compared to the system of poly(PCDA)/Neo-PCDA1. It has been known that the smaller PDA has higher response to external stimuli.<sup>84</sup> Therefore, the larger size of poly(PCDA)/Neo-PCDA2 assemblies likely causes the lower response to poly(rA)-poly(rU).

We further investigate the color-transition behaviors of poly(PCDA)/Neo-PCDA1 and poly(PCDA)/Neo-PCDA2 upon increasing temperature (see ESI Fig. S15<sup>†</sup>). Significant drop of absorbance in the system of poly(PCDA)/Neo-PCDA2 is observed upon increasing temperature attributed to the formation of large aggregates and then precipitation. In addition, this system also shows lower % CR compared to pure poly(PCDA) and poly(PCDA)/Neo-PCDA1, corresponding to higher stability. This result is consistent with the lower response of poly(PCDA)/Neo-PCDA2 to poly(rA)-poly(rU).

### 3.5 The effect of DA monomer structure on colorimetric response

The variation of DA monomer structure is an approach for enhancing colorimetric response of PDA to its stimuli. In this part, the PDA/Neo-PCDA1 assemblies were fabricated by using TCDA monomer which has slightly shorter alkyl tail than PCDA monomer. Fig. 8a shows the absorption spectra of poly(TCDA)/Neo-PCDA1 containing 0.5% v/v hexanol upon the addition of poly(rA)-poly(rU) at various concentrations. We observe significant growth of absorption peak at ~540 nm. The plot of % CR clearly shows that poly(TCDA)/Neo-PCDA1 has higher CR values compared to the system of poly(PCDA)/Neo-PCDA1 (see Fig. 8b) attributed to the shorter alkyl tail of poly(TCDA). The decrease of alkyl chain length results in weaker dispersion interaction along the PDA layers.<sup>51,52,73</sup> Therefore, we observe higher response of poly(TCDA)/Neo-PCDA1 to poly(rA)-poly(rU) compared to poly(PCDA)/Neo-PCDA1.

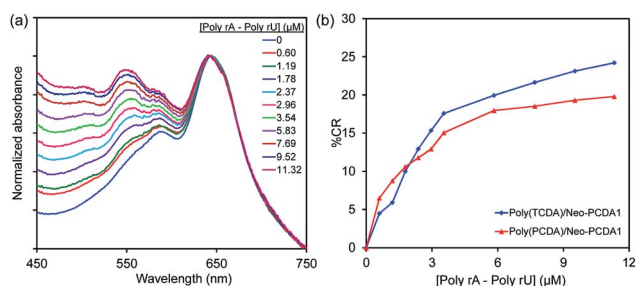


Fig. 8 Absorption spectra of (a) poly(TCDA)/Neo-PCDA1 and (b) the plot of % CR shows the effect of DA monomer structure on the response to poly(rA)-poly(rU).

## 4. Conclusions

Sensitive and specific PDA-based biosensor for the detection of homopolyribonucleotide poly(rA)-poly(rU) has been developed in this study. The poly(PCDA)/Neo-PCDA1 assemblies provide blue-to-purple color transition while pure poly(PCDA) vesicles show no change after the addition of poly(rA)-poly(rU). The addition of control DNA to poly(PCDA)/Neo-PCDA1 assemblies does not induce the color transition, indicating high specificity of poly(PCDA)/Neo-PCDA1 to poly(rA)-poly(rU). Sensitivity of poly(PCDA)/Neo-PCDA1 to poly(rA)-poly(rU) is enhanced by using alcohols as additives and the variation of DA monomer structure. The addition of small amount of hexanol results in significant increase of poly(PCDA)/Neo-PCDA1 response to poly(rA)-poly(rU). The penetration of alcohol molecules weakens interactions within the poly(PCDA)/Neo-PCDA1 assemblies, leading to the increase of colorimetric response. The variation of Neo-PCDA structure also affects its sensitivity. The use of Neo-PCDA1 with longer linker provides a higher response compared to that of the Neo-PCDA2. The PDA/Neo-PCDA assemblies prepared by using TCDA monomer exhibit a larger change compared to that of PCDA system. The development of sensitive and specific PDA to target biomolecules expands our toolkit for the utilization of PDA in biosensing, drug detection, and diagnostic applications.

## Conflicts of interest

There are no conflicts to declare.

## List of abbreviations

AC <sub>50</sub>	Concentration at 50% of maximum activity
CD	Circular dichroism spectroscopy
DA	Diacetylene (monomer)
DMF	<i>N,N</i> -Dimethylformamide
DMPC	Dimyristoylphosphatidylcholine
DNA	Deoxyribonucleic acid
FID	Fluorescence intercalator displacement assays
ITC	Isothermal titration calorimetry
MALDI	Matrix assisted laser desorption/ionization-time of flight mass spectrometry
Neo	Neomycin
Neo-PCDA	Conjugate of neomycin and PCDA
NMR	Nuclear magnetic resonance spectrophotometer
PBS buffer	Phosphate-buffered saline solutions
PCDA	10,12-Pentacosadiynoic acid
PDA	Polydiacetylene
Poly(rA)-poly(rU):	Polyadenylic-uridylic acid
RNA	Ribonucleic acid
TBTU	<i>O</i> -(Benzotriazol-1-yl)- <i>N,N,N',N'</i> -tetramethyluronium tetrafluoroborate
TCDA	10,12-Tricosadiynoic acid
TEM	Transmission electron microscope
TFA	Trifluoroacetic acid
TPS	2,4,6-Triisopropylbenzenesulfonyl chloride
UV	Ultraviolet



## Acknowledgements

We thank National Institutes for Health for financial support (grants R42GM097917, AI114114) to DPA. The research was also supported by the Thailand Research Fund (Grant BRG6080008). A. Kamphan thanks the Royal Golden Jubilee Ph.D. Program for supporting her Ph.D. scholarship. This work has partially been supported by the Nanotechnology Center (NANOTEC), Ministry of Science and Technology, Thailand, through its program of Center of Excellence Network.

## Notes and references

- 1 D. P. Arya, *Aminoglycoside antibiotics: from chemical biology to drug discovery*, John Wiley & Sons, 2007.
- 2 M. A. Nowak, S. Bonhoeffer, A. M. Hill, R. Boehme, H. C. Thomas and H. McDade, *Proc. Natl. Acad. Sci. U. S. A.*, 1996, **93**, 4398–4402.
- 3 J. L. Seyec, P. Chouteau, I. Cannie, C. Guguen-Guillouzo and P. Gripon, *J. Virol.*, 1999, **73**, 2052–2057.
- 4 J. Zhong, P. Gastaminza, G. Cheng, S. Kapadia, T. Kato, D. R. Burton, S. F. Wieland, S. L. Uprichard, T. Wakita and F. V. Chisari, *Proc. Natl. Acad. Sci. U. S. A.*, 2005, **102**, 9294–9299, DOI: 0503596102 [pii].
- 5 S. A. Waksman, H. A. Lechevalier and D. A. Harris, *J. Clin. Invest.*, 1949, **28**, 934–939, DOI: 10.1172/jci102182 [doi].
- 6 S. Kumar, P. Kellish, W. E. Robinson Jr, D. Wang, D. H. Appella and D. P. Arya, *Biochemistry*, 2012, **51**, 2331–2347.
- 7 S. Kumar and D. P. Arya, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 4788–4792.
- 8 N. Ranjan, S. Kumar, D. Watkins, D. Wang, D. H. Appella and D. P. Arya, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 5689–5693.
- 9 S. Nahar, N. Ranjan, A. Ray, D. P. Arya and S. Maiti, *Chem. Sci.*, 2015, **6**, 5837–5846.
- 10 S. Kumar, N. Ranjan, P. Kellish, C. Gong, D. Watkins and D. P. Arya, *Org. Biomol. Chem.*, 2016, **14**, 2052–2056.
- 11 S. Kumar, L. Xue and D. P. Arya, *J. Am. Chem. Soc.*, 2011, **133**, 7361–7375.
- 12 B. Willis and D. P. Arya, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 4974–4979.
- 13 B. Willis and D. P. Arya, *Biochemistry*, 2009, **49**, 452–469.
- 14 D. P. Arya, L. Xue and B. Willis, *J. Am. Chem. Soc.*, 2003, **125**, 10148–10149.
- 15 B. Willis and D. P. Arya, *Adv. Carbohydr. Chem. Biochem.*, 2006, **60**, 251–302.
- 16 D. P. Arya and B. Willis, *J. Am. Chem. Soc.*, 2003, **125**, 12398–12399.
- 17 B. Willis and D. P. Arya, *Biochemistry*, 2006, **45**, 10217–10232.
- 18 N. N. Shaw, H. Xi and D. P. Arya, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 4142–4145.
- 19 I. Charles, E. Davis and D. P. Arya, *Biochemistry*, 2012, **51**, 5496–5505.
- 20 H. Xi, S. Kumar, L. Dosen-Micovic and D. P. Arya, *Biochimie*, 2010, **92**, 514–529.
- 21 D. P. Arya, *Acc. Chem. Res.*, 2010, **44**, 134–146.
- 22 L. Xue, I. Charles and D. P. Arya, *Chem. Commun.*, 2002, 70–71.
- 23 L. Xue, N. Ranjan and D. P. Arya, *Biochemistry*, 2011, **50**, 2838–2849.
- 24 N. Ranjan, K. F. Andreasen, S. Kumar, D. Hyde-Volpe and D. P. Arya, *Biochemistry*, 2010, **49**, 9891–9903.
- 25 D. Watkins, N. Ranjan, S. Kumar, C. Gong and D. P. Arya, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 6695–6699.
- 26 N. Ranjan and D. P. Arya, *Molecules*, 2013, **18**, 14228–14240.
- 27 D. P. Arya, *DNA Binders and Related Subjects*, 2005, pp. 149–178.
- 28 H. Xi, E. Davis, N. Ranjan, L. Xue, D. Hyde-Volpe and D. P. Arya, *Biochemistry*, 2011, **50**, 9088–9113.
- 29 S. Okada, S. Peng, W. Spevak and D. Charych, *Acc. Chem. Res.*, 1998, **31**, 229–239.
- 30 R. W. Carpick, D. Y. Sasaki, M. S. Marcus, M. Eriksson and A. R. Burns, *J. Phys.: Condens. Matter*, 2004, **16**, R679.
- 31 B. Yoon, S. Lee and J. Kim, *Chem. Soc. Rev.*, 2009, **38**, 1958–1968.
- 32 M. van den Heuvel, D. W. Löwik and J. C. van Hest, *Biomacromolecules*, 2008, **9**, 2727–2734.
- 33 L. Rougeau, D. Picq, M. Rastello and Y. Frantz, *Tetrahedron*, 2008, **64**, 9430–9436.
- 34 I. S. Park, H. J. Park and J. Kim, *ACS Appl. Mater. Interfaces*, 2013, **5**, 8805–8812.
- 35 S. Wacharasindhu, S. Montha, J. Boonyiseng, A. Potisatituyenyong, C. Phollookin, G. Tumcharern and M. Sukwattanasinitt, *Macromolecules*, 2010, **43**, 716–724.
- 36 M. Gou, G. Guo, J. Zhang, K. Men, J. Song, F. Luo, X. Zhao, Z. Qian and Y. Wei, *Sens. Actuators, B*, 2010, **150**, 406–411.
- 37 J. Kim, J. Lee, H. Choi, D. Sohn and D. J. Ahn, *Macromolecules*, 2005, **38**, 9366–9376.
- 38 J. Pang, L. Yang, B. F. McCaughey, H. Peng, H. S. Ashbaugh, C. J. Brinker and Y. Lu, *J. Phys. Chem. B*, 2006, **110**, 7221–7225.
- 39 T. Pattanatornchai, N. Charoenthai and R. Traiphol, *J. Colloid Interface Sci.*, 2014, **432**, 176–181.
- 40 Q. Cheng and R. C. Stevens, *Langmuir*, 1998, **14**, 1974–1976.
- 41 A. Kamphan, C. Khanantong, N. Traiphol and R. Traiphol, *J. Ind. Eng. Chem.*, 2017, **46**, 130–138.
- 42 N. Traiphol, A. Chanakul, A. Kamphan and R. Traiphol, *Thin Solid Films*, 2017, **622**, 122–129.
- 43 Z. Yuan and T. W. Hanks, *Polymer*, 2008, **49**, 5023–5026.
- 44 S. J. Kew and E. A. Hall, *Anal. Chem.*, 2006, **78**, 2231–2238.
- 45 H. Jeon, J. Lee, M. H. Kim and J. Yoon, *Macromol. Rapid Commun.*, 2012, **33**, 972–976.
- 46 D. Seo and J. Kim, *Adv. Funct. Mater.*, 2010, **20**, 1397–1403.
- 47 J. Lee and J. Kim, *Chem. Mater.*, 2012, **24**, 2817–2822.
- 48 S. Lu, F. Luo, X. Duan, C. Jia, Y. Han and H. Huang, *J. Appl. Polym. Sci.*, 2014, **131**, 40634.
- 49 S. Lu, C. Jia, X. Duan, X. Zhang, F. Luo, Y. Han and H. Huang, *Colloids Surf., A*, 2014, **443**, 488–491.
- 50 F. Delbecq and T. Kawai, *Colloids Surf., A*, 2013, **430**, 85–90.
- 51 T. Pattanatornchai, N. Charoenthai, S. Wacharasindhu, M. Sukwattanasinitt and R. Traiphol, *J. Colloid Interface Sci.*, 2013, **391**, 45–53.





- 52 N. Charoenthai, T. Pattanatornchai, S. Wacharasindhu, M. Sukwattanasinitt and R. Traiphol, *J. Colloid Interface Sci.*, 2011, **360**, 565–573.
- 53 K. Parambath Kootery, H. Jiang, S. Kolusheva, T. Vinod, M. Ritenberg, L. Zeiri, R. Volinsky, D. Malferrari, P. Galletti and E. Tagliavini, *ACS Appl. Mater. Interfaces*, 2014, **6**, 8613–8620.
- 54 S. Scoville and W. Shirley, *J. Appl. Polym. Sci.*, 2011, **120**, 2809–2820.
- 55 Y. Su, *React. Funct. Polym.*, 2006, **66**, 967–973.
- 56 Y. Su, J. Li and L. Jiang, *Colloids Surf., B*, 2004, **39**, 113–118.
- 57 R. Jelinek, *Drug Dev. Res.*, 2000, **50**, 497–501.
- 58 Y. Zhang, B. Ma, Y. Li and J. Li, *Colloids Surf., B*, 2004, **35**, 41–44.
- 59 G. Ma and Q. Cheng, *Talanta*, 2005, **67**, 514–519.
- 60 C. Wang, Z. Ma and Z. Su, *Sens. Actuators, B*, 2006, **113**, 510–515.
- 61 K. Kim, H. Choi, G. S. Lee, D. J. Ahn and M. Oh, *Colloids Surf., B*, 2008, **66**, 213–217.
- 62 M. van den Heuvel, D. W. Löwik and J. C. van Hest, *Biomacromolecules*, 2010, **11**, 1676–1683.
- 63 Y. K. Jung, T. W. Kim, J. Kim, J. Kim and H. G. Park, *Adv. Funct. Mater.*, 2008, **18**, 701–708.
- 64 Y. Xia, J. Deng and L. Jiang, *Sens. Actuators, B*, 2010, **145**, 713–719.
- 65 D. H. Kang, H. Jung, J. Lee, S. Seo, J. Kim, K. Kim and K. Suh, *Langmuir*, 2012, **28**, 7551–7556.
- 66 Y. K. Jung and H. G. Park, *Biosens. Bioelectron.*, 2015, **72**, 127–132.
- 67 D. Lee, S. K. Sahoo, A. L. Cholli and D. J. Sandman, *Macromolecules*, 2002, **35**, 4347–4355.
- 68 Y. Lifshitz, Y. Golan, O. Konovalov and A. Berman, *Langmuir*, 2009, **25**, 4469–4477.
- 69 X. Huang, S. Jiang and M. Liu, *J. Phys. Chem. B*, 2005, **109**, 114–119.
- 70 X. Wang, J. E. Whitten and D. J. Sandman, *J. Chem. Phys.*, 2007, **126**, 184905.
- 71 Y. Amano and Q. Cheng, *Anal. Bioanal. Chem.*, 2005, **381**, 156–164.
- 72 M. Park, K. Kim, D. J. Ahn and M. Oh, *Biosens. Bioelectron.*, 2012, **35**, 44–49.
- 73 A. Kamphan, N. Charoenthai and R. Traiphol, *Colloids Surf., A*, 2016, **489**, 103–112.
- 74 D. P. Arya, L. Xue and P. Tennant, *J. Am. Chem. Soc.*, 2003, **125**, 8070–8071.
- 75 I. Charles, L. Xue and D. P. Arya, *Bioorg. Med. Chem. Lett.*, 2002, **12**, 1259–1262.
- 76 K. Michael, H. Wang and Y. Tor, *Bioorg. Med. Chem.*, 1999, **7**, 1361–1371.
- 77 H. Y. Erbil, *Surface chemistry of solid and liquid interfaces*, Wiley Online Library, 2006.
- 78 R. Nagarajan, *Langmuir*, 2002, **18**, 31–38.
- 79 Y. K. Jung, H. G. Park and J. Kim, *Biosens. Bioelectron.*, 2006, **21**, 1536–1544.
- 80 Q. Cheng, M. Yamamoto and R. C. Stevens, *Langmuir*, 2000, **16**, 5333–5342.
- 81 Q. Ye, G. Zou, X. You, X. Yu and Q. Zhang, *Mater. Lett.*, 2008, **62**, 4025–4027.
- 82 F. Zhao, Q. Zhao, K. F. Blount, Q. Han, Y. Tor and T. Hermann, *Angew. Chem., Int. Ed.*, 2005, **44**, 5329–5334.
- 83 W. M. Haynes and D. R. Lide, *Handbook of Chemistry and Physics; National Institute for Standards and Technology*, CRC Press, New York, NY, 2010.
- 84 C. X. Guo, P. Boullanger, T. Liu and L. Jiang, *J. Phys. Chem. B*, 2005, **109**, 18765–18771.

