


PAPER

[View Article Online](#)
[View Journal](#) | [View Issue](#)Cite this: *RSC Adv.*, 2017, 7, 37478Received 19th June 2017
Accepted 25th July 2017

DOI: 10.1039/c7ra06813a

rsc.li/rsc-advancesPhthalide derivatives from *Ligusticum chuanxiong*†Xu Zhang, Bing Han, Zi-Ming Feng, Ya-Nan Yang,  Jian-Shuang Jiang
and Pei-Cheng Zhang*

Eleven new phthalide derivatives (1–11) have been isolated from the rhizome of *Ligusticum chuanxiong*. In particular, 1 and 2 contain a mercaptopropionic acid moiety in the phthalide derivatives. All the structures, including their absolute configurations, were determined by UV, IR, HRESIMS, 1D and 2D NMR spectroscopic measurements, and by a comparison of the experimental and calculated electronic circular dichroism (ECD) spectra. Results of a bioassay showed that compound 4 has a moderate neuroprotective activity on human neuroblastoma SH-SY5Y cell injury induced by H₂O₂ and oxygen glucose deprivation (OGD).

Introduction

The rhizome of *Ligusticum chuanxiong* Hort. (syn. *Ligusticum striatum* DC.) (Apiaceae), named Chuanxiong in traditional Chinese medicine, is commonly used for treating atherosclerosis, cardiovascular disease, anemia and hypertension.¹ This plant is mainly distributed in the Sichuan province but also in Yunnan, Guizhou, and Guangxi provinces in China. So far, the phenolic acids, alkaloids, and phthalides^{2–16} contained in it have been investigated. Among these compounds, tetramethylpyrazine and ferulic acid were considered to be the active constituents.^{17–24} The phthalides from *L. chuanxiong* have attracted particular attention due to their vasodilative,^{17,25,26} neuroprotective,²⁷ antiproliferative,^{28,29} anti-inflammatory²⁴ and antibacterial effects.³⁰ In particular, Z-ligustilide, a phthalide isolated from *L. chuanxiong*, displays various pharmacological activities including neuroprotective, anti-inflammatory, anti-proliferative and vasorelaxation effects.^{31–34} It is considered a major bioactive compound relevant to the therapeutic effects and has attracted great interest in recent years. Overall, more than 40 phthalide lactones have been obtained from the rhizome of *L. chuanxiong*,^{35,36} but most of them were isolated from the liposoluble fraction. In consideration of the bioavailability *in vivo*, the phthalide derivatives from the water-soluble extract of this plant deserve to be the primary focus of further studies. In the course of our study, eleven new phthalide derivatives (1–11) have been isolated and their structures were elucidated by various spectroscopic methods (Fig. 1). Subsequently, their neuroprotective activities on SH-SY5Y cell injury

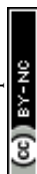
induced by H₂O₂ and OGD were evaluated at a concentration of 10 μM.

Results and discussion

Compound 1 was obtained as a white amorphous powder, and its molecular formula was established as C₁₅H₂₀O₇S by HRESIMS at *m/z* 345.0999 [M + H]⁺ (calcd 345.1003). The IR spectrum peaks at 3387 cm^{−1}, 1741 cm^{−1} and 1682 cm^{−1} suggest the presence of hydroxy, γ-lactone and diene groups, respectively. In the ¹H NMR spectra (Table 1), a double bond at δ_H 5.48 (1H, t, *J* = 7.5 Hz, H-8), six methylene resonances at δ_H 2.43 (2H, m, H-4), 1.81 (2H, m, H-5), 2.31 (2H, q, *J* = 7.5 Hz, H-9), 1.56 (2H, m, H-10), 3.41 (2H, t, *J* = 6.0 Hz, H-11), 2.89 (1H, dd, *J* = 7.0, 13.5 Hz, H-1'a), 3.02 (1H, dd, *J* = 4.5, 13.5 Hz, H-1'b), one methine resonance at δ_H 3.50 (1H, brs, H-7), and two oxygen-methine resonances at δ_H 4.10 (1H, brs, H-6) and 4.17 (1H, dd, *J* = 4.5, 7.0 Hz, H-2') were presented. The ¹³C NMR spectrum (Table 1) shows 15 carbon resonances, including two carboxyl carbons at δ_C 173.9 (C-3') and 168.1 (C-1), four olefinic carbons, five methylene carbons, a methine carbon resonance at δ_C 41.8 (C-7), an oxygen-methylene carbon resonance at δ_C 60.2 (C-11), and two oxygen-methine resonances at δ_C 67.5 (C-6) and 70.4 (C-2'). In the HMBC experiment (Fig. 2), the correlations from H-8 (δ_H 5.48) to C-3 (δ_C 147.7) and C-3a (δ_C 151.6) suggest the existence of a 1,3-butadiene moiety. Furthermore, it was confirmed to be connected to a 3-hydroxypropyl on the basis of the correlations of the ¹H–¹H COSY experiment (Fig. 2), including those between H-8 and H-9, H-9 and H-10, and H-10 and H-11. In addition, the ¹H–¹H COSY correlations from H-4 to H-5, from H-5 to H-6, and from H-6 to H-7 together with the HMBC correlations from H-6 (δ_H 4.10) to C-7a, from H-7 (δ_H 3.50) to C-7a and C-3a, from H-5 (δ_H 1.81) to C-3a, and from H-4 (δ_H 2.43) to C-3a and C-7a indicate the existence of a cyclohexene moiety. The 2-carboxyl-2-hydroxy-ethylthio moiety on position C-7 was confirmed by the ¹H–¹H COSY correlations

State Key Laboratory of Bioactive Substance and Function of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences, Peking Union Medical College, Beijing 100050, People's Republic of China. E-mail: pczhang@imm.ac.cn

† Electronic supplementary information (ESI) available: 1D NMR, 2D NMR HRMS, IR, and ECD spectra. See DOI: 10.1039/c7ra06813a



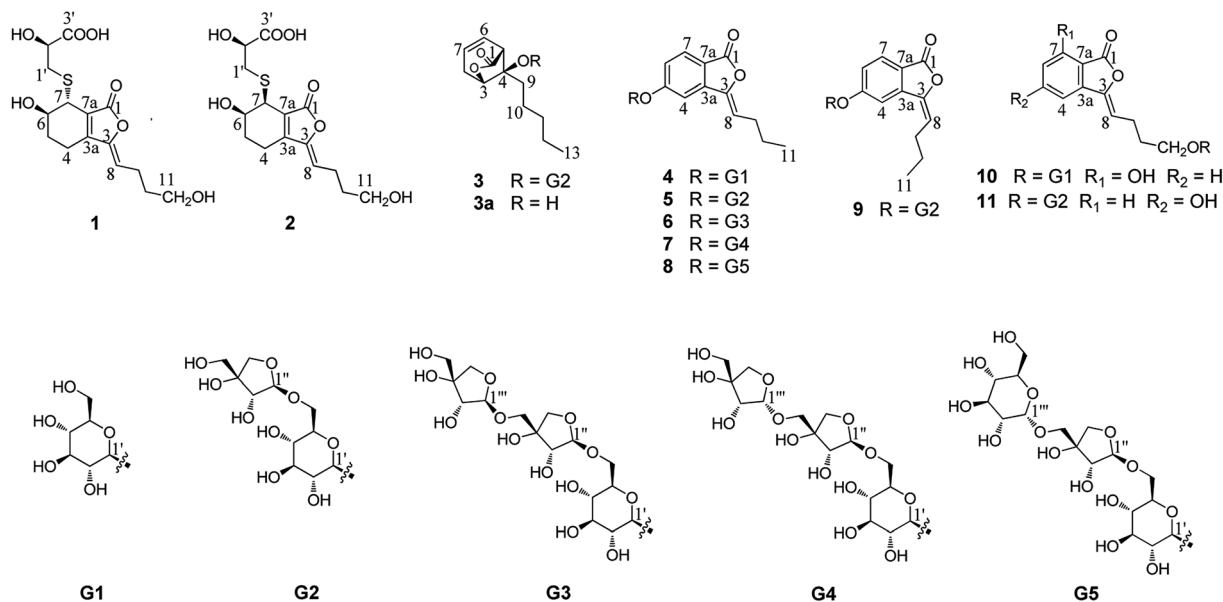


Fig. 1 Chemical structures of 1–11.

Table 1 ¹H NMR (500 MHz) and ¹³C NMR (125 MHz) data of compounds 1–3 in DMSO-*d*₆

Position	1		2		3	
	δ_{H} (f in Hz)	δ_{C}	δ_{H} (f in Hz)	δ_{C}	δ_{H} (f in Hz)	δ_{C}
1		168.1		167.1		174.7
3		147.7		147.4	4.58, brs	78.9
3a		151.6		150.9		
4	2.43, m	16.1	2.41, m	19.6		84.2
5	1.81, m	22.9	1.74, m	25.8	3.21, dd (1.5, 7.5)	47.7
6	4.10, brs	67.5	3.95, dd (3.5, 10.5)	68.2	5.87, m	123.9
7	3.50, brs	41.8	3.81, d (3.5)	43.6	5.77, m	127.9
7a		124.2		126.1		
8	5.48, t (7.5)	112.7	5.42, t (7.5)	112.5	2.46, m	31.3
9	2.31, q (7.5)	22.4	2.29, q (7.5)	22.4	1.67, m	26.1
10	1.56, m	31.8	1.55, m	31.7	1.38, m	21.7
11	3.41, t (6.0)	60.2	3.40, t (6.5)	60.2	1.19, m	31.3
12					1.25, m	21.7
13					0.85, t (7.5)	13.7
1'	2.89, dd (7.0, 13.5)	36.9	2.93, dd (7.0, 13.5)	37.9	4.38, d (8.0)	97.2
	3.02, dd (4.5, 13.5)		3.07, dd (4.5, 13.5)			
2'	4.17, dd (4.5, 7.0)	70.4	4.12, dd (4.5, 7.0)	70.7	2.94, m	73.2
3'		173.9		174.1	3.28, m	75.2
4'					2.99, m	70.1
5'					3.14, m	76.9
6'					3.35, m	67.9
1''					3.81, dd (1.5, 10.5)	
2''					4.84, d (3.0)	109.4
3''					3.74, brs	75.9
4''						78.6
					3.58, d (9.5)	73.4
					3.85, d (9.5)	
5''					3.35, m	63.4

from H-1' (δ_{H} 2.89, 3.02) to H-2' (δ_{H} 4.17) and the HMBC correlations from H-1' and H-2' to the carbonyl carbon at δ_{C} 173.9 and from H-7 to C-1' at δ_{C} 36.9 in consideration of the

changes of the chemical shifts of C-7 and C-1'. The HMBC correlation from H-7 to the carbonyl at δ_{C} 168.1 indicates that C-7a connected with the carbonyl. Thus, the planar structure was



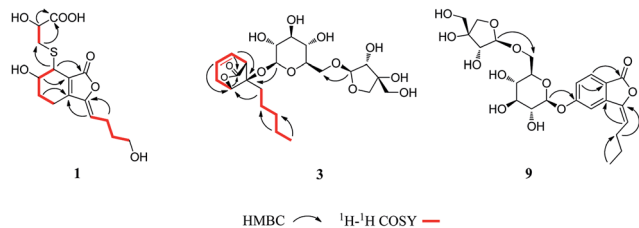


Fig. 2 Key HMBC and ^1H - ^1H COSY correlations of **1**, **3** and **9**.

determined to be 6-hydroxy-7-(2-carboxyl-2-hydroxy-ethylthio)-3-(4-hydroxybutylidene)-4,5,6,7-tetrahydrophthalide. The absolute configuration of C-2' in **1** was established by a dimolybdenum tetraacetate $[\text{Mo}_2(\text{AcO})_4]$ -induced circular dichroism procedure.³⁷ The diagnostic Cotton effect approximately 344.5 nm was negative, so the absolute configuration of C-2' in **1** was assigned as *S*. In the ROESY spectra (Fig. 3), the correlation between H-8 and H-4 proved the *Z*-configuration of the double bond on position C-3, and the correlation between H-6 and H-1' suggested that their relative configuration was *trans*. Their absolute configuration was determined by comparing the experimental and calculated ECD data based on the time-dependent density functional theory (TD-DFT) method at the B3LYP/6-31+G (d,p) level. Considering that the variations in the side chain resulted in too many conformations and had little effect on the CD spectrum of the tetrahydrophthalide core in **1**, the selected conformers **1Ja** (6*S*,7*S*) and **1Jb** (6*R*,7*R*) were calculated after optimization (Fig. 4). A comparison of the theoretically calculated and experimental ECD curves (Fig. 5) permitted the assignment of the absolute configuration of **1** as 6*R*,7*R*. Finally, the structure of **1** was determined to be (3*Z*,3*aE*)-(6*R*,7*R*,2'*S*)-6-hydroxy-7-(2-carboxyl-2-hydroxyethylthio)-3-(4-hydroxybutylidene)-4,5,6,7-tetrahydrophthalide, named thio-senkyunolide A.

The HRESIMS of **2** gave the same molecular formula as **1**, $\text{C}_{15}\text{H}_{20}\text{O}_7\text{S}$, based on the $[\text{M} + \text{Na}]^+$ ion peak at m/z 367.0827 (calcd 367.0822). The ^1H and ^{13}C NMR spectra of **2** (Table 1) were closely comparable to those of **1**, with the minor differences occurring in the signals in the cyclohexene moiety, that is, the ^{13}C NMR data were shifted downfield, except that of C-3a (Table 1). Further 2D NMR analysis, including HSQC, HMBC, and ^1H - ^1H COSY experiments, confirmed that compound **2** was the optical isomer of compound **1**. Using the same method, the absolute configuration of C-2' in **2** was established as *S* by a dimolybdenum tetraacetate $[\text{Mo}_2(\text{AcO})_4]$ -induced circular

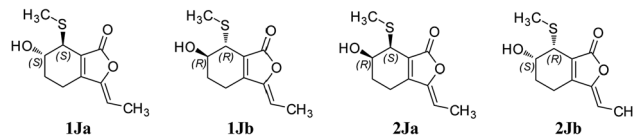


Fig. 4 The structures of **1Ja**, **1Jb**, **2Ja** and **2Jb**.

dichroism procedure,³⁷ whose diagnostic Cotton effect at approximately 345 nm was also negative. In the ROESY spectra, the correlation between H-8 and H-4 proved the *Z*-configuration of the double bond at position C-3, but the correlation between H-6 and H-1' was not observed in **2** compared with **1**, which suggested that the relative configuration between them was *cis*. The absolute configurations of 6*R*,7*S* were further confirmed by a comparison of the experimental ECD spectrum and calculated ECD data using the aforementioned methods (Fig. 5). Thus, the structure of **2** was determined to be (3*Z*,3*aE*)-(6*R*,7*S*,2'*S*)-6-hydroxy-7-(2-carboxyl-2-hydroxyethylthio)-3-(4-hydroxybutylidene)-4,5,6,7-tetrahydrophthalide, named thio-senkyunolide B.

Compound **3** was obtained as a white amorphous powder. Its molecular formula was identified as $\text{C}_{23}\text{H}_{36}\text{O}_{12}$ (m/z 503.2148 $[\text{M} - \text{H}]^-$) by negative HRESIMS. The IR absorptions at 3336 cm^{-1} , 1774 cm^{-1} and 1049 cm^{-1} represented a hydroxyl group, carbonyl group and alkenyl group, respectively. The ^1H NMR and ^{13}C NMR spectra (Table 1) of **3** displayed an amyl side chain that produced a methyl resonances [δ_{H} 0.85 (3H, t, $J = 7.5\text{ Hz}$, H-13); δ_{C} 13.7] and four methylene resonances [δ_{H} 1.25 (2H, m, H-12), 1.19 (2H, m, H-11), 1.38 (2H, m, H-10), 1.67 (2H, m, H-9); δ_{C} 21.7, 31.3, 21.7, 26.1, respectively]; two olefinic [δ_{H} 5.77 (1H, m, H-7), 5.87 (1H, m, H-6); δ_{C} 127.9, 123.9]; an oxymethine [δ_{H} 4.58 (1H, brs, H-3); δ_{C} 78.9] and a methine [δ_{H} 3.21 (1H, dd, $J = 1.5$, 7.5 Hz, H-5); δ_{C} 47.7]; and one methylene [2.46 (2H, m, H-8); δ_{C} 31.3]. Furthermore, a carbonyl carbon at δ_{C} 174.7 (C-1) and a quaternary carbon δ_{C} 84.2 (C-4) were observed. The presence of two anomeric protons at δ_{H} 4.38 (1H, d, $J = 8.0\text{ Hz}$, H-1') and 4.84 (1H, d, $J = 3.0\text{ Hz}$, H-1'') and eleven carbons proved that compound **3** contained two sugar moieties. Comparison of the NMR data of **3** with those of the known compound ligusticoside A¹⁶ suggested that these two compounds shared a similar skeleton except sugar moieties. In the HMBC spectrum, the correlations H-1'/C-4 and H-1''/C-6' proved that the glucose moiety was attached at C-4 and the pentose was attached at the C-6' of glucose (Fig. 2). The configurations of the apiose and glucose were determined to be *D*-configurations by GC analysis after acidic hydrolysis and chiral derivatization (retention times at 18.26, 29.32 min). The β -anomeric configurations were deduced on the basis of their coupling constants (Glc: $J = 8.0\text{ Hz}$; Api: $J = 3.0\text{ Hz}$). In the ROESY spectrum of **3**, the correlations of H-8/H-9, H-8/H-10, H-5/H-9, H-5/H-10 and H-3/H-9 indicated that H-3, H-5 and amyl chain were located on the same side (Fig. 3).

The absolute configuration of **3** was determined by comparing the experimental and calculated ECD data based on the TD-DFT method at the B3LYP/6-31+G (d,p) level. Considering the numerous conformations of sugar, we used the aglycone **3a**

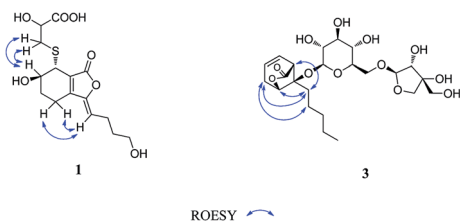


Fig. 3 Key ROESY correlations of **1** and **3**.



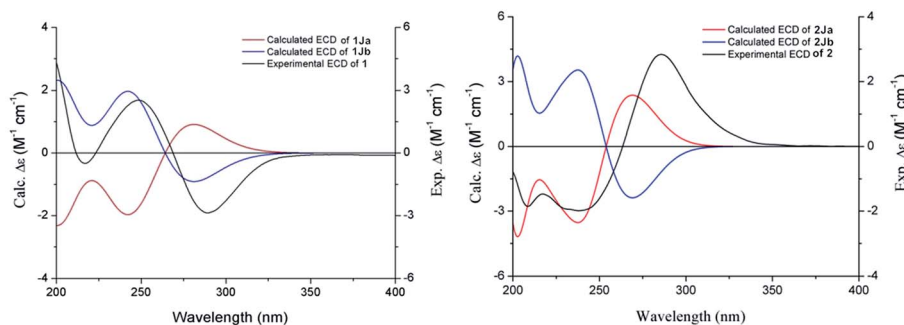


Fig. 5 Experimental ECD and calculated ECD spectrum of **1** and **2** in MeOH.

which was obtained from the acidic hydrolysis for ECD calculations. Through all wavelengths, the absolute configuration of **3S,4R,5R** was confirmed by matching the calculated spectrum with the experimental ECD data (Fig. 6). Thus, the structure of **3** was confirmed and named ligusticoside B.

Compound **4** was obtained as a white amorphous powder. The positive HRESIMS gave the $[M + H]^+$ ion peak at m/z 367.1399, in accordance with an empirical molecular formula of $C_{18}H_{22}O_8$. The 1H NMR spectrum (Table 2) of **4** presented a methyl resonance at δ_H 0.95 (3H, t, $J = 7.5$ Hz, H-11), two methylene resonances at δ_H 1.51 (2H, m, H-10) and 2.36 (2H, q, $J = 7.5$ Hz, H-9) and an olefinic proton at δ_H 5.96 (1H, t, $J = 8.0$ Hz, H-8). An ABX system at δ_H 7.80 (1H, d, $J = 8.5$ Hz, H-7), 7.20 (1H, dd, $J = 8.5, 2.0$ Hz, H-6) and 7.59 (1H, d, $J = 2.0$ Hz, H-4) was presented. Additionally, the presence of multiple protons between δ_H 3.20 and 3.70 and the presence of a doublet at δ_H 5.12 (1H, d, $J = 6.5$ Hz, H-1') suggested the occurrence of a glycoside moiety. The ^{13}C NMR spectrum (Table 2) of **4** showed 18 carbons, including a carbonyl carbon at δ_C 165.9 (C-1), a tertiary olefinic carbon at δ_C 109.5 (C-8), a quaternary olefinic carbon at δ_C 145.0 (C-3), six aromatic carbons at δ_C 141.4 (C-3a), 106.4 (C-4), 162.8 (C-5), 119.4 (C-6), 126.6 (C-7), and 116.9 (C-7a), and two methylene carbon resonances at δ_C 27.4 (C-9) and 22.0 (C-10). In addition, six oxygenated carbons that contributed to a glycoside moiety were also observed at δ_C 99.9 (C-1'), 73.1 (C-2'), 76.6 (C-3'), 69.5 (C-4'), 77.2 (C-5'), and 60.5 (C-6').

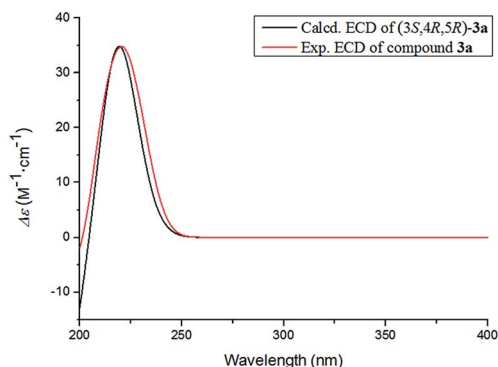


Fig. 6 Experimental ECD and calculated ECD spectrum of **3a** in MeOH.

In the HMBC spectrum of **4**, the correlations H-7/C-1, H-6/C-7a, H-4/C-3, H-8/C-3a, H-9/C-3, H-10/C-8 and H-11/C-9 confirmed the presence of a butenylphthalide moiety. The glucose moiety was attached at C-5 by the correlation of H-1'/C-5. In the NOE experiment, when irradiating H-4, H-8 generated a gain, and when irradiating the H-8, H-4 also generated gain signal (ESI[†]). Thus the geometric configuration of the double bond was confirmed as *Z*. The glucose was confirmed as having a *D*-configuration through the same method as used for **3**. The relatively large coupling constant ($J = 6.5$ Hz) of the anomeric proton suggested that the glucose moiety was β -configured. Therefore, the structure of **4** was established as (*Z*)-3-butyldiene-5-*O*- β -*D*-glucopyranosyl phthalide, named ligusticoside A.

The molecular formula of compound **5** was confirmed as $C_{23}H_{30}O_{12}$ by the $[M - H]^-$ ion peak at m/z 497.1664 in HRESIMS. The 1H NMR spectrum (Table 2) of **5** presented a methyl resonance at δ_H 0.95 (3H, t, $J = 7.5$ Hz, H-11), two methylene resonances at δ_H 1.52 (2H, m, H-10) and 2.35 (2H, m, H-9) and an olefinic proton at δ_H 5.95 (1H, t, $J = 8.0$ Hz, H-8). An ABX system at δ_H 7.83 (1H, d, $J = 8.5$ Hz, H-7), 7.23 (1H, dd, $J = 8.5, 2.0$ Hz, H-6) and 7.54 (1H, d, $J = 2.0$ Hz, H-4) was presented. According to the above 1H NMR spectroscopic data, compound **5** had a similar skeleton to **4** except for the glycoside moiety. The presence of multiple protons between δ_H 3.10 and δ_H 3.89 and the presence of two doublets at δ_H 5.10 (1H, d, $J = 7.0$ Hz, H-1') and 4.80 (1H, d, $J = 3.0$ Hz, H-1'') suggested the occurrence of two sugar moieties. Compared with the ^{13}C NMR spectroscopic data of **4**, compound **5** had an additional five carbons whose resonances at δ_C 109.2 (C-1''), 75.9 (C-2''), 78.8 (C-3''), 73.3 (C-4''), and 63.1 (C-5'') were observed, and combined with the coupling constant ($J = 3.0$ Hz), the pentose was confirmed to be β -apiose. The apiose and glucose moieties were further determined to have *D*-configurations by the above method. The β -anomeric configuration was deduced on the basis of the coupling constant (Glc: $J = 7.0$ Hz). The connective position of apiose was identified by the correlation of H-6'/C-1'', which confirmed that the C-1'' of the apiofuranosyl was located at the C-6' of the glucopyranosyl in the HMBC spectrum. The geometric configuration of the double bond was confirmed as *Z* by comparison with the data of 1H NMR and ^{13}C NMR in **4**. Thus, compound **5** was established as (*Z*)-3-butyldiene-5-*O*- β -*D*-apiofuranosyl-(1 \rightarrow 6)- β -*D*-glucopyranosyl phthalide, named ligusticoside B.



Table 2 ^1H and ^{13}C NMR data of compounds 4–7 in $\text{DMSO}-d_6$

Position	4^a		5^a		6^b		7^b	
	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}
1		165.9		165.9		165.8		165.8
3		145.0		144.9		144.9		144.9
3a		141.4		141.3		141.2		141.2
4	7.59, d (2.0)	106.4	7.54, d (2.0)	106.7	7.54, d (1.8)	106.6	7.54, d (1.8)	106.6
5		162.8		162.7		162.6		162.6
6	7.20, dd (2.0, 8.5)	119.4	7.23, dd (2.0, 8.5)	119.1	7.23, dd (1.8, 9.0)	119.0	7.23, dd (1.8, 8.4)	119.0
7	7.80, d (8.5)	126.6	7.83, d (8.5)	126.7	7.82, d (9.0)	126.6	7.82, d (8.4)	126.6
7a		116.9		117.0		117.0		117.0
8	5.96, t (8.0)	109.5	5.95, t (8.0)	109.6	5.95, t (7.8)	109.4	5.95, t (7.8)	109.4
9	2.36, q (7.5)	27.4	2.35, m	27.4	2.36, m	27.4	2.35, m	27.3
10	1.51, m	22.0	1.52, m	22.0	1.52, m	21.9	1.52, m	21.9
11	0.95, t (7.5)	13.8	0.95, t (7.5)	13.8	0.95, t (7.8)	13.7	0.95, t (7.2)	13.7
Glc								
1'	5.12, d (6.5)	99.9	5.10, d (7.0)	99.9	5.12, d (7.2)	99.8	5.11, d (7.2)	99.8
2'	3.20, m	73.1	3.13, m	73.1	3.29, m	73.0	3.29, m	73.0
3'	3.29, m	76.6	3.30, m	75.6	3.30, m	75.6	3.28, m	75.6
4'	3.29, m	69.5	3.29, m	69.8	3.16, m	69.6	3.14, m	69.6
5'	3.47, m	77.2	3.63, m	76.5	3.63, m	76.4	3.62, m	76.4
6'	3.47, m	60.5	3.43, m	67.4	3.45, dd (6.6, 10.8)	67.3	3.45, dd (6.6, 10.8)	67.2
	3.70, m		3.88, m		3.88, d (10.8)		3.88, m	
Api-1								
1''			4.80, d (3.0)	109.2	4.80, d (3.0)	108.7	4.80, d (3.0)	108.9
2''			3.73, dd (3.0, 6.5)	75.9	3.73, dd (3.0, 6.6)	76.1	3.69, dd (3.0, 6.6)	76.8
3''				78.8		78.7		77.6
4''			3.59, d (9.0)	73.3	3.58, d (9.6)	73.4	3.64, m	73.6
			3.86, m		3.84, d (9.0)		3.86, m	
5''			3.33, m	63.1	3.33, m	69.1	3.36, d (9.6)	69.8
					3.50, d (9.6)		3.62, m	
Api-2								
1'''					4.81, d (3.6)	109.0	4.85, d (4.8)	102.1
2'''					3.76, dd (3.6, 6.0)	75.6	3.76, dd (4.8, 9.6)	71.4
3'''						77.4		76.1
4'''					3.60, m	73.3	3.65, m	73.8
					3.84, d (9.0)		3.90, d (9.6)	
5'''					3.30, m	63.1	3.23, dd (5.4, 9.6)	63.0
					3.33, m		3.28, m	

^a 500 MHz for ^1H NMR, 125 MHz for ^{13}C NMR. ^b 600 MHz for ^1H NMR, 150 MHz for ^{13}C NMR.

The molecular formula of **6** was assigned as $\text{C}_{28}\text{H}_{38}\text{O}_{16}$ based on the $[\text{M} + \text{Na}]^+$ ion peak at m/z 653.2046 (calcd 653.2052) in the HRESIMS. By carefully comparing the related data, **6** had similar structural characteristics to **4** and **5**, with the main difference that **6** had a three-monosaccharide unit. In the ^1H NMR spectrum (Table 2) of **6**, the presence of multiple protons between δ_{H} 3.15 and δ_{H} 3.89 and the presence of three anomeric protons at δ_{H} 5.12 (1H, d, $J = 7.2$ Hz, H-1'), 4.80 (1H, d, $J = 3.0$ Hz, H-1'') and 4.81 (1H, d, $J = 3.6$ Hz, H-1''') suggested the occurrence of three monosaccharide units. The ^{13}C NMR resonances (Table 2) at δ_{C} 99.8, 108.7, and 109.0 indicated that **6** had a pyranose and two furanoses. The apiose and glucose were determined to have D -configurations through the same method as used above. The β -anomeric configurations were confirmed on the basis of their coupling constants (Glc: $J = 7.2$ Hz; Api-inner: $J = 3.0$ Hz; Api-outer: $J = 3.6$ Hz). In the HMBC spectrum, the glucose moiety

was also attached at C-5 by the correlation of H-1'/C-5. The correlation H-6'/C-1'' confirmed that the apiofuranosyl was located at the glucopyranosyl. The correlation H-5''/C-1''' confirmed that the outer apiofuranosyl was located at the inner apiofuranosyl. Thus, compound **6** was established as (Z)-3-butylidene-5-O- β -D-apiofuranosyl-(1 \rightarrow 5)- β -D-apiofuranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl phthalide, named ligusticumside C.

Compound **7** had the same molecular formula ($\text{C}_{28}\text{H}_{38}\text{O}_{16}$) as **6** through the HRESIMS at m/z 653.2066 $[\text{M} + \text{Na}]^+$. A careful comparison of the IR, UV and NMR data of **6** and **7** suggested that they had the same planar structure. The main difference was the chemical shifts of C-1''' and C-2''' of the outer apiose at δ_{C} 102.1 and 71.4 in **7**, instead of δ_{C} 109.0 and 75.6 in **6**, respectively. This was confirmed by the coupling constant of H-1''' ($J = 4.8$ Hz in **7** vs. $J = 3.6$ Hz in **6**). Clearly, the outer sugar unit had an α configuration instead of the β configuration in **6**.



The sugar moieties were determined to be D-configurations of apiose and glucose through the same method as used above. Therefore, the structure of **7** was confirmed as (Z)-3-butyldiene-5-O- α -D-apiofuranosyl-(1 \rightarrow 5)- β -D-apiofuranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl phthalide, named ligusticumside D.

According to the $[M + Na]^+$ ion peak at m/z 683.2164 in the HRESIMS, the molecular formula of **8** was assigned as $C_{29}H_{40}O_{17}$. Comparing its NMR data with those of **7**, both of them are trisaccharides of butyldienephthalide with the major difference between them being the type of the outer sugar. The presence of an anomeric proton at δ_H 4.64 (1H, d, $J = 3.5$ Hz, H-1'') and carbons at δ_C 99.2, 72.8, 72.1, 70.0, 73.3, and 60.8 suggested that the outer sugar of **8** was α -glucose instead of α -apiose in **7**. The apiose and glucose were determined to have D-configurations by the same method as used above. Thus, the structure of **8** was confirmed as (Z)-3-butyldiene-5-O- α -D-glucopyranosyl-(1 \rightarrow 5)- β -D-apiofuranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl phthalide, named ligusticumside E.

The molecular formula of compound **9** was determined to be $C_{18}H_{22}O_8$ through HRESIMS ($[M + Na]^+$ 521.1629). The identical data of HRESIMS, IR, and UV suggested that **5** and **9** were isomers, with the difference being the geometric configuration of the double bond. In the 1H NMR spectrum of **9** (Table 3), an olefinic proton was presented at δ_H 5.90 (1H, t, $J = 7.8$ Hz, H-8). In the NOE experiment, when irradiating H-4, H-8 did not generate a gain, and when irradiating H-8, H-4 also did not generate a gain signal, which was entirely different from the results for **5** (ESI $^+$). Additionally, the C-4 (δ_C 110.1) and C-8 (δ_C 114.2) were shifted significantly downfield in the ^{13}C NMR spectrum when compared to those of **5** (C-4 at δ_C 106.7 and C-8 at δ_C 109.6). Thus, the double bond was proven to have an *E* configuration, and compound **9** was confirmed as (*E*)-3-butyldiene-5-O- β -D-apiofuranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl phthalide, named ligusticumside F.

Compound **10** was obtained as a white amorphous powder. The HRESIMS showed a positive ion at m/z 405.1156 $[M + Na]^+$ that matched the molecular formula $C_{18}H_{22}O_9$. The IR spectrum presented hydroxyl (3395 cm^{-1}) and γ -lactone (1757 cm^{-1}) peaks. The 1H NMR spectrum of **10** (Table 3) showed three aromatic resonances at δ_H 7.27 (1H, d, $J = 8.0$ Hz, H-4), 7.54 (1H, t, $J = 8.0$ Hz, H-5), and 6.89 (1H, d, $J = 8.0$ Hz, H-6), which were assigned to a 1,2,3-trisubstituted benzene ring, an olefinic resonance at δ_H 5.86 (1H, t, $J = 7.5$ Hz, H-8), two methylene resonances at δ_H 2.40 (2H, q, $J = 7.5$ Hz, H-9) and 1.71 (2H, m, H-10), and a pair of oxygen-methylene resonances at δ_H 3.50 (1H, m, H-11a) and 3.83 (1H, m, H-11b), as well as the glucoside resonances. The ^{13}C NMR experiment (Table 3) presented 18 carbon resonances, including one carbonyl carbon at δ_C 164.4 (C-1), six aromatic carbons, two olefinic carbons at δ_C 145.0 (C-3) and 107.9 (C-8), two methylene carbon resonances at δ_C 22.3 (C-9) and 28.9 (C-10), one oxygen-methylene resonance at δ_C 68.1 (C-11), and a set of glucose carbon resonances, implying that **10** was still a phthalide derivative with a glucose unit. The correlation of H-1'/C-11 in the HMBC spectrum suggested that the glucose moiety was located at C-11. The glucose was confirmed as having a D-configuration by the same method as used above. The β -configuration was deduced based on the coupling

constant ($J = 8.0$ Hz) of H-1'. Thus, compound **10** was determined to be (Z)-3-(4-O- β -D-glucopyranosyl-butyldiene)-7-hydroxyphthalide, named ligusticumside G.

Compound **11** had the molecular formula $C_{23}H_{30}O_{13}$ by the analysis of its HRESIMS, which gave a positive ion at m/z 537.1587 $[M + Na]^+$ (calcd 537.1579). An ABX system at δ_H 7.17 (1H, d, $J = 2.0$ Hz, H-4), 6.97 (1H, dd, $J = 2.0, 8.0$ Hz, H-6) and 7.68 (1H, d, $J = 8.0$ Hz, H-7) (Table 3) in the 1H NMR spectrum suggested the presence of the same trisubstituted aromatic ring as in **4** instead of that in **10**. In the HMBC experiment, H-7 (δ_H 7.68) correlated with C-5 (δ_C 163.8) and C-3a (δ_C 141.8), H-6 (δ_H 6.97) correlated with C-4 (δ_C 114.4) and C-7a (δ_C 105.6), and H-4 (δ_H 7.17) correlated with C-6 (δ_C 118.5), C-7a (δ_C 105.6), and C-3 (δ_C 144.9), indicating that the aromatic hydroxyl was located at position C-5. The additional resonances at δ_H 4.84 (1H, d, $J = 3.0$ Hz, H-1'') and from δ_H 3.84 to 3.30 were attributed to one pentose in **11**. The ^{13}C NMR spectrum showed 23 carbon resonances (Table 3), and the corresponding pentose resonances at δ_C 109.2 (C-1''), 75.8 (C-2''), 78.8 (C-3''), 73.4 (C-4''), and 63.1 (C-5'') were observed. The apiofuranosyl and glucopyranosyl were determined to have D-configurations by the same method as used above (retention times at 18.26, 29.32 min). The coupling constants of the anomeric protons indicated the β -configuration (Glc: $J = 8.0$ Hz; Api: $J = 3.0$ Hz). Moreover, the correlation between H-1'' (δ_H 4.84) and C-6' (δ_C 68.2) confirmed that the C-1'' of the apiofuranosyl was located at the C-6' of the glucopyranosyl. Thus, compound **11** was confirmed as (Z)-3-[4-O- β -D-apiofuranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl-butyldiene]-7-hydroxyphthalide, named ligusticumside H.

Compounds **1–11** were tested for their neuroprotective effects on SH-SY-5Y cell injury induced by H_2O_2 and OGD with L-NBP (L-3-*n*-butylphthalide) as a positive control. The results showed that compound **5** exhibited a weak neuroprotective effect with the increase cell viability rate of 15.60% and compound **4** exhibited a moderate neuroprotective effect with the increase cell viability rate of 24.35%, compared with the positive control L-NBP with increase cell viability rate of 18.26% on H_2O_2 -induced neurotoxicity. And compound **4** exhibited a moderate neuroprotective effect with the increase cell viability rate of 13.72%, compared with the positive control L-NBP with increase cell viability rate of 4.03% on OGD-induced neurotoxicity.

Experimental

General experimental procedures

The optical rotations, UV spectra and ECD spectra were recorded with JASCO P-2000, V650 and J-815 spectrometer (JASCO, Easton, MD, USA), respectively. The Infrared spectra were measured on Nicolet 5700 spectrometer (Thermo Scientific, FL, USA). The NMR spectra were recorded with Varian 500 MHz (Bruker-Biospin, Billerica, MA, USA) and 600 MHz NMR spectrometers (Varian, Inc., Palo Alto, CA, USA). HRESIMS reports were obtained from Agilent 6520 HPLC-Q-TOF (Agilent Technologies, Waldbronn, Germany) and LCMS-IT-TOF system (Shimadzu Scientific Instruments Inc., Kyoto, Japan). Preparative HPLC was performed using a Shimadzu LC-10AT with



Table 3 ^1H and ^{13}C NMR data of compounds **8–11** in $\text{DMSO}-d_6$

Position	8^a		9^b		10^a		11^a	
	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}
1		165.9		165.6		164.4		166.1
3		144.9		144.7		145.0		144.9
3a		141.3		139.4		141.0		141.8
4	7.54, d (2.0)	106.6	7.45, d (1.8)	110.1	7.27, d (8.0)	110.3	7.17, d (2.0)	114.4
5		162.7		162.5	7.54, t (8.0)	136.6		163.8
6	7.23, dd (2.0, 8.5)	119.0	7.32, dd (1.8, 9.0)	118.7	6.89, d (8.0)	116.4	6.97, dd (2.0, 8.0)	118.5
7	7.82, d (8.5)	126.8	7.89, d (9.0)	127.0		157.2	7.68, d (8.0)	126.8
7a		117.0		118.7		109.4		105.6
8	5.96, t (8.0)	109.6	5.90, t (7.8)	114.2	5.86, t (7.5)	107.9	5.89, t (7.5)	108.7
9	2.35, m	27.4	2.52, m	27.2	2.40, q (7.5)	22.3	2.41, q (7.5)	22.3
10	1.51, m	22.0	1.57, m	22.3	1.71, m	28.9	1.72, m	28.9
11	0.95, t (7.5)	13.8	0.98, t (7.8)	13.6	3.50, m	68.1	3.51, m	67.7
					3.83, m		3.80, m	
Glc-1								
1'	5.12, d (7.5)	99.8	5.11, d (7.2)	100.2	4.12, d (8.0)	102.9	4.14, d (8.0)	102.8
2'	3.29, m	73.1	3.30, m	73.0	2.94, t (8.0)	73.4	2.95, overlap	73.2
3'	3.29, m	75.6	3.31, m	75.7	3.06, m	76.9	3.12, t (8.5)	76.6
4'	3.16, m	69.5	3.14, m	69.8	3.03, q (9.0)	70.1	2.98, overlap	70.2
5'	3.64, m	76.4	3.89, m	76.2	3.11, m	76.8	3.25, m	75.5
6'	3.46, m	67.4	3.44, dd (6.6, 10.8)	67.5	3.42, dd (5.0, 11.0)	61.1	3.39, dd (7.0, 11.0)	68.2
	3.89, m		3.87, m		4.12 d (8.0)		3.82 m	
Api								
1''	4.83, d (4.0)	108.8	4.80, d (3.0)	109.2			4.84, d (3.0)	109.2
2''	3.71, dd (4.0, 6.5)	76.8	3.75, dd (3.0, 6.6)	75.8			3.74, d (3.0)	75.8
3''		77.7		78.7				78.8
4''	3.65, m	73.7	3.57, d (9.0)	73.2			3.56, d (9.5)	73.4
	3.91, m		3.86, d (9.0)				3.84, d (9.5)	
5''	3.25, m	69.6	3.31, m	62.9			3.30, t (11.0)	63.1
	3.64, m						3.32, t (11.0)	
Glc-2								
1'''	4.64, d (3.5)	99.2						
2'''	3.29, m	72.8						
3'''	3.16, m	72.1						
4'''	3.07, m	70.0						
5'''	3.40, m	73.3						
6'''	3.45, m	60.8						
	3.58, m							

^a 500 MHz for ^1H NMR, 125 MHz for ^{13}C NMR. ^b 600 MHz for ^1H NMR, 150 MHz for ^{13}C NMR.

a ODS-A column (250 mm \times 20 mm, 5 μm ; YMC Corp., Kyoto, Japan). The Agilent 1200 series system was used to carry on the HPLC-DAD analysis with an Apollo C₁₈ column (250 mm \times 4.6 mm, 5 μm ; Alltech Corp., Lexington, KY, USA). The Agilent 7890A was used to carry on the GC analysis with a capillary column, HP-5 (60 m \times 0.32 mm, with a 1 μm film; Agilent Technologies Inc., CA, USA). Macroporous resin Diaion HP-20 (Mitsubishi Chemical Corp., Tokyo, Japan), RP-C₁₈ (50 μm , YMC Corp.), and Sephadex LH-20 (Pharmacia Fine Chemicals, Uppsala, Sweden) were used to column chromatograph.

Plant material

The roots of *Ligusticum chuanxiong* Hort. were collected from Pengzhou Town, Sichuan Province in PRC, in Jun 2013 and

identified by professor L. Ma. A voucher specimen (ID-S-2594) was deposited at the Institute of Materia Medica, Peking Union Medical College and Chinese Academy of Medical Sciences, Beijing, China.

Extraction and isolation

The powdered rhizome of *L. chuanxiong* Hort. (100.0 kg) was exhaustively extracted with 80% EtOH under reflux condition. The solvent was evaporated by reduced pressure and then the residue (23.1 kg) was partitioned successively with EtOAc and *n*-BuOH. The *n*-BuOH-soluble portion (1300 g) was applied on a HP-20 column to give five fractions A–E through gradient elution with H₂O, 15% ethanol, 30% ethanol, 50% ethanol, and 95% ethanol, respectively. Fraction C (103.0 g) was



chromatographed over an RP-C₁₈ column, eluting with H₂O/MeOH (from 100 : 0 to 0 : 100) to give 16 fractions (C1–C16) on the basis of HPLC analyses. Fraction C5 was subjected to a Sephadex LH-20 with a gradient of increasing MeOH (0–100%) in H₂O and then separated by preparative HPLC (MeOH/H₂O, 30 : 70, V/V, HOAc, 0.01%) to give **2** (4 mg). Fraction C13 was separated by column chromatography over a Sephadex LH-20 using H₂O as the eluent and was further purified by preparative HPLC (MeOH/H₂O, 45 : 55, v/v, HOAc, 0.01%) to give **1** (64 mg). Compounds **10** (4 mg) and **11** (4 mg) were obtained from fraction C14 with the above method. Fraction D (48.0 g) was chromatographed over an RP-C₁₈ column, eluting with H₂O/MeOH (from 95 : 5 to 0 : 100) to give 24 fractions (D1–D24) on the basis of HPLC and TLC analyses. Fraction D9 was purified by a Sephadex LH-20 with a gradient of increasing MeOH (0–100%) in H₂O and then separated by preparative HPLC (MeCN/H₂O, 28 : 72, V/V, HOAc, 0.02%) to give **4** (70 mg), **5** (50 mg), **6** (3 mg), **7** (4 mg), **8** (3 mg), and **9** (10 mg). Fraction D8 was chromatographed over a Sephadex LH-20 with a gradient of increasing MeOH (0–100%) in H₂O to give 19 fractions (D8-1–D8-19) on the basis of HPLC and TLC analyses. Fraction D8-2 was chromatographed over silica gel (EtOAc/MeOH/H₂O, from 120 : 1:0.5 to 1 : 1:0.5) to give **3** (120 mg).

Structure characterization

Thiosenkyunolide A, 1. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 201 (4.01), 277 (4.09) nm; $[\alpha]_{\text{D}}^{20}$ –49 (c 0.1 MeOH); HRESIMS m/z 345.0999 $[M + H]^+$ (calcd 345.1003), m/z 367.0823 $[M + Na]^+$ (calcd 367.0822); IR (KBr) ν_{\max} : 3387, 2938, 1741, 1682, 1633, 1514, 1414, 1230, 1184, 1093, 1043, 954 cm^{–1}; ¹H NMR and ¹³C NMR see Table 1.

Thiosenkyunolide B, 2. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 201 (3.84), 277 (4.06) nm; $[\alpha]_{\text{D}}^{20}$ +30 (c 0.1 MeOH); HRESIMS m/z 345.1008 $[M + H]^+$ (calcd 345.1003), m/z 367.0827 $[M + Na]^+$ (calcd 367.0822); IR (KBr) ν_{\max} : 3347, 2932, 2882, 1756, 1680, 1632, 1412, 1323, 1239, 1090, 1048, 1026, 997, 966 cm^{–1}; ¹H NMR and ¹³C NMR see Table 1.

Ligisticoside B, 3. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 203 (4.18), 216 (3.90) nm; $[\alpha]_{\text{D}}^{20}$ +4 (c 0.1 MeOH); HRESIMS m/z 503.2148 $[M - H]^-$ (calcd 503.2134); IR (KBr) ν_{\max} : 3336, 2924, 1774, 1275, 1350, 1049 cm^{–1}; ¹H NMR and ¹³C NMR see Table 1.

Ligisticumside A, 4. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 253 (4.42), 273 (4.16) nm; $[\alpha]_{\text{D}}^{20}$ –73 (c 0.1 MeOH); HRESIMS m/z 367.1399 $[M + H]^+$ (calcd 367.1387); IR (KBr) ν_{\max} : 3377, 2929, 1759, 1689, 1613, 1484, 1297, 1078, 991 cm^{–1}; ¹H NMR and ¹³C NMR see Table 2.

Ligisticumside B, 5. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 253 (4.35), 274 (4.07) nm; $[\alpha]_{\text{D}}^{20}$ –96 (c 0.1 MeOH); HRESIMS m/z 497.1664 $[M - H]^-$ (calcd 497.1665); IR (KBr) ν_{\max} : 3406, 2931, 1763, 1611, 1482, 1293, 1066, 1011 cm^{–1}; ¹H NMR and ¹³C NMR see Table 2.

Ligisticumside C, 6. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 252 (4.19), 275 (3.92) nm; $[\alpha]_{\text{D}}^{20}$ –78 (c 0.1 MeOH); HRESIMS m/z 653.2046 $[M + Na]^+$ (calcd 653.2052); IR

(KBr) ν_{\max} : 3372, 2930, 1762, 1681, 1611, 1387, 1293, 1066, 930 cm^{–1}; ¹H NMR and ¹³C NMR see Table 2.

Ligisticumside D, 7. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 253 (4.27), 272 (4.03) nm; $[\alpha]_{\text{D}}^{20}$ –30 (c 0.1 MeOH); HRESIMS m/z 653.2066 $[M + Na]^+$ (calcd 653.2052); IR (KBr) ν_{\max} : 3398, 2932, 1765, 1687, 1613, 1295, 1072, 930 cm^{–1}; ¹H NMR and ¹³C NMR see Table 2.

Ligisticumside E, 8. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 253 (4.38), 270 (4.16) nm; $[\alpha]_{\text{D}}^{20}$ –29 (c 0.1 MeOH); HRESIMS m/z 683.2164 $[M + Na]^+$ (calcd 683.2158); IR (KBr) ν_{\max} : 3384, 2926, 1762, 1688, 1614, 1294, 1073, 928 cm^{–1}; ¹H NMR and ¹³C NMR see Table 3.

Ligisticumside F, 9. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 254 (4.08), 272 (3.91) nm; $[\alpha]_{\text{D}}^{20}$ –21 (c 0.1 MeOH); HRESIMS m/z 521.1629 $[M + Na]^+$ (calcd 521.1629); IR (KBr) ν_{\max} : 3399, 2931, 1765, 1689, 1612, 1483, 1294, 1072, 953 cm^{–1}; ¹H NMR and ¹³C NMR see Table 3.

Ligisticumside G, 10. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 198 (3.96), 225 (4.14), 265 (3.95), 330 (3.66), 339 (3.70) nm; $[\alpha]_{\text{D}}^{20}$ –10 (c 0.1 MeOH); HRESIMS m/z 405.1156 $[M + Na]^+$ (calcd 405.1156); IR (KBr) ν_{\max} : 3395, 2928, 2880, 1757, 1687, 1606, 1473, 1372, 1297, 1199, 1163, 1079, 1017, 895 cm^{–1}; ¹H NMR and ¹³C NMR see Table 3.

Ligisticumside H, 11. White amorphous powder; UV λ_{\max} (MeOH) (log ϵ): 254 (4.54) nm; $[\alpha]_{\text{D}}^{20}$ –30 (c 0.1 MeOH); HRESIMS m/z 537.1587 $[M + Na]^+$ (calcd 537.1579); IR (KBr) ν_{\max} : 3375, 2932, 2884, 1753, 1689, 1610, 1481, 1467, 1381, 1295, 1165, 1057, 999, 932 cm^{–1}; ¹H NMR and ¹³C NMR see Table 3.

Determination of the absolute configuration of sugar

Compounds **5** (2 mg) was dissolved in 1 mol L^{–1} CF₃COOH (14 mL) and then the mixture was heated in 70 °C for 1 h. The mixture was then extracted three times with EtOAc, and the aqueous layer was freeze-dried to obtain residue. Using the same method with the literature,³⁸ the residue was dissolved in anhydrous pyridine (2 mL), L-cysteine methyl ester hydrochloride (4 mg) was added, and then the mixture was heated in a water bath (60 °C) for 1 h. After the reaction solution was dried under vacuum, N-trimethylsilylimidazole (1 mL) was added, and the solution was heated in a water bath (60 °C) for 1 h and extracted three times with H₂O/*n*-hexane. Then, the *n*-hexane layer was analyzed using GC under conditions as follows: injection temperature, 300 °C; detector temperature (FID), 300 °C; capillary column, HP-5 (30 m × 0.32 mm, Dikma); start temperature, 200 °C, raised to 260 °C at a rate of 10 °C min^{–1}, and the final temperature maintained for 30 min; and N₂ used as the carrier gas.

Neuroprotective activities of compounds 1–11

The assay method of neuroprotective effects about compounds refer to the procedures of literature.³⁹

Conclusions

In the course of a search for neuroprotective compounds from *Ligisticum chuanxiong* Hort., eleven new phthalide derivatives



were obtained. Among them, two are substituted with the rare mercaptopropionic acid. All compounds were tested for their neuroprotective effects, and it was observed that compound 4 had moderate effect against H₂O₂ and OGD-induced neurotoxicity in SH-SY5Y cells at 10 μ M. These results could contribute to the better understanding of the therapeutic usage of *Ligusticum chuanxiong* Hort.

Acknowledgements

This work was supported by the CAMS Innovation Fund for Medical Sciences (CIFMS) (No. 2016-I2M-1-010).

References

- 1 Editing Group for The Compilation of Chinese Herbal Medicines, *The Compilation of Chinese Herbal Medicines*, People's Medical Publishing House, Beijing, 1996, pp. 133–135.
- 2 J. Huang, X. Q. Lu, C. Zhang, J. Lu, G. Y. Li, R. C. Lin and J. H. Wang, *Fitoterapia*, 2013, **91**, 21–27.
- 3 C. P. Miao, S. H. Wu, B. Z. Luo, J. Wang and Y. W. Chen, *Fitoterapia*, 2010, **81**, 1088–1090.
- 4 M. Kim, S. O. Kim, M. Lee, J. H. Lee, W. S. Jung, S. K. Moon, Y. S. Kim, K. H. Cho, C. N. Ko and E. H. Lee, *Eur. J. Pharmacol.*, 2014, **740**, 504–511.
- 5 S. L. Li, S. S. K. Chan, G. Lin, L. Ling, R. Yan, H. S. Chung and Y. K. Tam, *Planta Med.*, 2003, **69**, 445–451.
- 6 N. Y. Yang, D. C. Ren, J. A. Duan, X. H. Xu, N. Xie and L. J. Tian, *Helv. Chim. Acta*, 2009, **92**, 291–297.
- 7 L. S. Lim, P. Shen, Y. H. Gong and E. L. Yong, *Phytochemistry*, 2006, **67**, 728–734.
- 8 J. Huang, X. Q. Lu, J. Lu, G. Y. Li, H. Y. Wang, L. H. Li, R. C. Lin and J. H. Wang, *J. Asian Nat. Prod. Res.*, 2013, **15**, 1237–1242.
- 9 Y. H. Li, S. L. Peng, Y. Zhou, K. B. Yu and L. S. Ding, *Planta Med.*, 2006, **72**, 652–656.
- 10 T. Naito, T. Katsuhara, K. Niitsu, Y. Ikeya, M. Okada and H. Mitsuhashi, *Phytochemistry*, 1992, **31**, 639–642.
- 11 T. Naito, K. Niitsu, Y. Ikeya, M. Okada and H. Mitsuhashi, *Phytochemistry*, 1992, **31**, 1787–1789.
- 12 T. Naito, Y. Ikeya, M. Okada, H. Mitsuhashi and M. Maruno, *Phytochemistry*, 1996, **41**, 233–236.
- 13 J. Yang, X. L. Feng, Y. Yu, Q. Wang, J. Zou, C. X. Wang, Z. Q. Mu, X. S. Yao and H. Gao, *China's Med.*, 2016, **11**, 1–7.
- 14 W. Wei, W. Xu and X. W. Yang, *J. Asian Nat. Prod. Res.*, 2017, **19**, 704–711.
- 15 P. O. Donkor, Y. Chen, L. Q. Ding and F. Qiu, *J. Ethnopharmacol.*, 2016, **194**, 530–548.
- 16 X. L. Chang, Z. Y. Jiang, Y. B. Ma, X. M. Zhang, K. W. K. Tsim and J. J. Chen, *J. Asian Nat. Prod. Res.*, 2009, **11**, 805–810.
- 17 M. J. Liang, L. C. He and G. D. Yang, *Life Sci.*, 2005, **78**, 128–133.
- 18 E. Y. Kim, J. H. Kim and M. R. Rhyu, *Biol. Pharm. Bull.*, 2010, **33**, 1360–1363.
- 19 K. N. Nam, K. P. Kim, K. H. Cho, W. S. Jung, J. M. Park, S. Y. Cho, S. K. Park, T. H. Park, Y. S. Kim and E. H. Lee, *Cell Biochem. Funct.*, 2013, **31**, 707–712.
- 20 X. Yang, Y. C. Wang, L. L. Li, Y. C. Jin, L. Sironi, Y. Wang and Y. Wang, *Fitoterapia*, 2014, **95**, 240–246.
- 21 W. Li, S. Zhang, Q. H. Gao, J. W. Hou and T. T. Wei, *Res. Chem. Intermed.*, 2004, **30**, 605–613.
- 22 T. F. Lee, Y. L. Lin and Y. T. Huang, *Planta Med.*, 2007, **73**, 527–534.
- 23 H. Y. Qi, S. O. Siu, Y. Chen, Y. F. Han, I. K. Chu, Y. Tong, A. S. Y. Lau and J. H. Rong, *Chem.-Biol. Interact.*, 2010, **183**, 380–389.
- 24 L. Liu, Z. Q. Ning, S. Shan, K. Zhang, T. Deng, X. P. Lu and Y. Y. Cheng, *Planta Med.*, 2005, **71**, 808–813.
- 25 W. C. Ko, C. C. Liao, C. H. Shih, C. B. Lei and C. M. Chen, *Planta Med.*, 2002, **68**, 1004–1009.
- 26 J. W. Tian, F. H. Fu, W. L. Jiang, C. Y. Wang, F. Sun and T. P. Zhang, *China J. Chin. Mater. Med.*, 2005, **30**, 466–468.
- 27 W. X. Gong, Y. Z. Zhou, X. Li, X. X. Gao, J. S. Tian, X. M. Qin and G. H. Du, *Molecules*, 2016, **21**, 549.
- 28 T. F. Lee, Y. L. Lin and Y. T. Huang, *Planta Med.*, 2007, **73**, 527–534.
- 29 Y. L. Lin, T. F. Lee, Y. J. Huang and Y. T. Huang, *J. Gastroenterol. Hepatol.*, 2006, **21**, 1257–1265.
- 30 Y. Sim and S. Shin, *Arch. Pharmacol. Res.*, 2008, **31**, 497–502.
- 31 X. Kuang, J. R. Du, Y. X. Liu, G. Y. Zhang and H. Y. Peng, *Pharmacol., Biochem. Behav.*, 2008, **88**, 213–221.
- 32 J. Wang, J. R. Du, Y. Wang and C. Y. Wang, *Acta Pharmacol. Sin.*, 2010, **31**, 791–797.
- 33 J. R. Du, B. Bai, X. Kuang, Y. Yu, C. Y. Wang, Y. Ke, Y. J. Xu, A. H. C. Tzang and Z. M. Qian, *J. Ethnopharmacol.*, 2006, **108**, 54–58.
- 34 Q. Lu, T. Q. Qiu and H. Yang, *Eur. J. Pharmacol.*, 2006, **542**, 136–140.
- 35 X. Ran, L. Ma, C. Peng, H. Zhang and L. P. Qin, *Pharm. Biol.*, 2011, **49**, 1180–1189.
- 36 W. Wei, X. W. Wu and X. W. Yang, *RSC Adv.*, 2016, **6**, 61037–61046.
- 37 G. Snatzke, U. Wagner and H. P. Wolff, *Tetrahedron*, 1981, **37**, 349–361.
- 38 M. L. Gan, M. T. Liu, L. S. Gan, S. Lin, B. Liu, Y. L. Zhang, J. C. Zi, W. X. Song and J. G. Shi, *J. Nat. Prod.*, 2012, **75**, 1373–1382.
- 39 S. W. Huang, J. W. Qiao, X. Sun, P. Y. Gao, L. Z. Li, Q. B. Liu, B. Sun, D. L. Wu and S. J. Song, *J. Funct. Foods*, 2016, **24**, 183–195.

