

Cite this: *RSC Adv.*, 2017, 7, 33558

Fabrication and investigation of high-quality glass-ceramic (GC)–polymethyl methacrylate (PMMA) composite for regenerative medicine†

Ricardas Golubevas,^a Aleksej Zarkov,^a Laurynas Alinauskas,^a Zivile Stankeviciute,^a Giedrius Balciunas,^b  Editas Garskaite ^{*a} and Aivaras Kareiva^a

The preparation of a glass-ceramic (GC)–polymethyl methacrylate (PMMA) composite material is reported. Precursor GC powders were synthesised *via* sol–gel method at 600 °C. XRD analysis revealed the formation of sodium calcium silicate ($\text{Na}_6\text{Ca}_3\text{Si}_6\text{O}_{18}$) as a main constituent phase of GC powders. The cylindrical-shaped GC–PMMA composite samples with fractional GC content ranging from 75 to 95% were prepared *via* photopolymerization reaction. The largest compressive strength for the composites containing 90% of GC has been determined. The growth of ceramic layer on the surface of composites immersed into simulated body fluid (SBF) under static conditions was studied by ICP-OES, FE-SEM/EDX, FTIR and XRD techniques. The changes in the concentrations of Ca, Si and P after the dissolution indicate the degradation process of GC–PMMA composites. The structures of the resultant surface confirmed the formation of new ceramic layer on the surface of composite materials. FTIR and XRD analyses revealed the deposition of low-crystallinity calcium phosphate layer with the composition similar to that of the carbonated hydroxyapatite (cHAP) after the soaking in SBF.

Received 8th May 2017
Accepted 27th June 2017

DOI: 10.1039/c7ra05188c

rsc.li/rsc-advances

1. Introduction

Biomaterials for bone repair receive considerable attention worldwide. Currently, the autograft bone (*i.e.* bone from the patient) is the gold standard in the treatment of various bone defects and performing spinal fusion procedures.^{1,2} Bone allografts (*i.e.* taken from a donor) or xenografts (*i.e.* bone from an animal source) can be used as alternatives; however, they suffer from potential risks such as disease transmission, adverse host immune response, they are in short supply and are also expensive. The global bone grafts and substitutes market is projected to rise steadily and reach US\$2.7 billion by 2020 with a compound annual growth rate of 3.8%.³ Therefore, the scaffold engineering for bone regenerative medicine is an alternative strategy and has been an area of great focus.^{4–6}

Bioactive glasses (amorphous, Ca- and possibly P- containing silica based materials) are group of bioactive ceramics used as a substitute for the autologous bone. Implanted bioactive glasses dissolve and stimulate both osteoconduction (support bone growth and encourage the ingrowth of surrounding bone) and osteoinduction (capable of promoting the differentiation of progenitor cells down an osteoblastic lineage).^{1,4,7} Sol–gel derived bioactive glass has advantages over the dense bioactive glass produced by conventional solid melt method, because it possess nanoporosity and larger surface area that courses more rapid dissolution rate and subsequently enhance faster new bone formation.⁸ However, one of the biggest challenges for utilization of bioactive glasses is its brittleness.^{6,7}

Alternative approach is to use various polymers that offer advantage in terms of possibility to produce materials that possess toughness and plasticity. The most commonly used synthetic polymers are polylactic acid (PLA), poly(glycolic acid) (PGA), polycaprolactone (PCL) and bioinert polymers such as polyurethane (PLU) or polymethylmethacrylate (PMMA).^{9–13} Acrylic polymer PMMA is commonly used as bone cement or in oral/maxillofacial surgery.^{10,14} Although PMMA is regarded as highly biocompatible material, it is also associated with several biological drawbacks. For instance, it is not biodegradable, the surface properties of PMMA are unfavourable for tissue adhesion and the tissues damage around cement surface due to thermal necrosis caused by PMMA polymerization and residual liquid monomer may also occur.^{15–17}

^aInstitute of Chemistry, Vilnius University, Naugarduko 24, Vilnius LT-03225, Lithuania. E-mail: edita.garskaite@chf.vu.lt

^bScientific Institute of Thermal Insulation, Vilnius Gediminas Technical University, Linkmenu st. 28, Vilnius LT-08217, Lithuania

† Electronic supplementary information (ESI) available: Values of diameter, height, mass and compressive strength for GC–PMMA composites, FE-SEM micrographs showing the surface of the pellets: (a) neat PMMA, ((b) and (c)) GC(75%)–PMMA(25%) composites, and (d) GC(95%)–PMMA(5%) composite, FTIR spectra of GC (annealed at 600 °C) and grinded GC–PMMA composite pellets, and absorption FTIR spectra recorded from the surface of GC(85%)–PMMA(15%) composite pellets 1, 2 and 3 weeks after the dissolution in SBF. See DOI: 10.1039/c7ra05188c

Several PMMA–bioceramic composites have been studied as alternatives to PMMA with objective to improve bioactivity and accelerate bone restoration. For instance, to improve the filler/cement interface the β -tricalcium phosphate (β -TCP) has been encapsulated with poly(ethylene glycol) (PEG) and later incorporated to a poly(methyl methacrylate) bone cement.¹⁸ The preparation of nanocomposite composed of hydroxyapatite (HAP) and carbon-nanotube reinforced PMMA has been also reported.¹⁹ Furthermore, Chen *et al.* demonstrated enhanced bioactivity of silicate bioceramic–PMMA composite compare to that of neat PMMA,¹⁵ while results obtained by Ravarian *et al.* showed that PMMA–bioactive glass composite supports osteoblast growth and differentiation.²⁰

Currently, a research trend is to fabricate scaffolds from bioceramic and polymer component with architectures mimicking the three-dimensional interconnected porosity of natural bone and an ability to guide cellular attachment and promote bone growth is one of the critical issues.²¹ Here we present the synthesis of glass-ceramic (GC) powders and fabrication of GC–PMMA composite *via* polymerization induced by photoionization process. The composition, morphology and mechanical properties of prepared materials are reported. The growth of ceramic layer on the composite surfaces immersed into simulated body fluid (SBF) under static conditions was also studies and discussed.

2. Experimental

2.1. Sol–gel synthesis of bioglass powders

The starting components for the synthesis of GC were calcium nitrate tetrahydrate ($\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$, $\geq 98\%$, Fluka), sodium nitrate (NaNO_3 , 99.5%, Chempur), tetraethyl orthosilicate ($\text{Si}(\text{OC}_2\text{H}_5)_4$, TEOS, 98%, Aldrich) and triethyl phosphate (TEP, $\text{PO}(\text{C}_2\text{H}_5)_3$, $\geq 98\%$, Aldrich). To synthesize GC powders a modified synthesis procedure published elsewhere²² was used. Firstly, 5 mM of citric acid ($[(\text{C}_6\text{H}_8\text{O}_7)]$, CA, $\geq 99.5\%$, Sigma-Aldrich) solution was prepared. To 26 mL of this solution 1 mL of TEP and 11.6 mL of TEOS were added. The mixture was stirred for 1 h at 65–70 °C until clear sol was obtained. In the following step, $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ (7.15 g, 0.03 mol) and NaNO_3 (4.66 g, 0.05 mol) salts were added and the mixture was further stirred for 60 min at 70 °C until sol turned into clear and transparent gel (the molar ratios of Ca : P : Na : Si was 0.4 : 0.1 : 0.6 : 1). The obtained Si–Ca–Na–P–O precursor gel was then dried in the oven at 100 °C for 24 h and turned into fine powders. The prepared powders were then calcined at 600, 700 and 1000 °C (heating rate 1 °C min^{-1}) for 5 h in air with intermediate grinding in agate mortar. The GC material annealed at 600 °C was further used to prepare GC–PMMA composites.

2.2. Preparation of GC–PMMA composite pellets

The starting components of the GC–PMMA composite were methyl 2-methylprop-2-enoate (methylmethacrylate, MMA, $\text{C}_5\text{H}_8\text{O}_2$, Evonik-Degussa GmbH) (5.00 g, 0.05 mol), (diphenylphosphinyl)-((2,4,6-trimethylphenyl)methanone) (Genocure TPO (defined as TPO), $\text{C}_{22}\text{H}_{21}\text{O}_2\text{P}$, Rahn USA Corp.) (1 g, 0.003

mol) and (2-benzyl-2-(dimethylamino)-4'-morpholinobutyrophenone) (Irgacure 396 (defined as BDMP), $\text{C}_{23}\text{H}_{30}\text{N}_2\text{O}_2$, BASF Corp.) (1 g, 0.003 mol). Firstly, these components were mechanically mixed in a beaker (the weight ratio of MMA : TPO : BDMP was 71.4 : 14.3 : 14.3). Photoinitiators TPO and BDMP were used for the bulk and surface cure, respectively. The GC powders heated at 600 °C were then added to the prepared MMA–TPO–BDMP mixture (the weight ratio of GC : MMA–TPO–BDMP was 75 : 25) and further mixed until the homogeneous GC–PMMA composite precursor was obtained. In the following step, 0.5 g of composite powders were pressed mechanically to cylindrical-shaped specimens of 8 mm in diameter and 5.1 mm in height (average value reported) using VLP hydraulic press (Enerpac) (10 kN) and left under natural light conditions at room temperature for 5 days to enable photopolymerization of the photoactive compounds.

The pellets of neat PMMA and GC–PMMA composites with fractional GC content of 85, 90 and 95% were additionally fabricated using a procedure described above. It is noteworthy that the preparation of pellets was constricted due to the liquidised form of the MMA–TPO–BDMP–GC mixture at the concentration of MMA–TPO–BDMP mixture above 25%.

2.3. Preparation of a simulated body fluid (SBF) solution and composite dissolution

1 L of the simulated body fluid (SBF) electrolyte solution was prepared by dissolving 7.996 g (0.138 mol) of sodium chloride (NaCl , $\geq 99\%$, Aldrich), 0.350 g (0.004 mol) of sodium hydrogen carbonate (NaHCO_3 , premium, Sigma), 0.224 g (0.003 mol) of potassium chloride (KCl , $\geq 99.0\%$, Sigma), 0.228 g (0.001 mol) of potassium phosphate dibasic trihydrate ($\text{K}_2\text{HPO}_4 \cdot 3\text{H}_2\text{O}$, $\geq 99.0\%$, Sigma-Aldrich), 0.305 g (0.001 mol) of magnesium chloride hexahydrate ($\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$, BioXtra $\geq 99.0\%$, Sigma-Aldrich), 0.244 g (0.002 mol) of calcium chloride (CaCl_2 , anhydrous, Bio-Reagent, $\geq 96.0\%$, Sigma) and 0.071 g (0.005 mol) of sodium sulphate (Na_2SO_4 , BioXtra, $\geq 99.0\%$, Sigma-Aldrich) into the distilled water. Prepared solution was buffered at pH of 7.25 with 50 mmol L^{-1} of (6.057 g, 0.05 mol) tris(hydroxymethyl)amino-methane ($[(\text{CH}_2\text{OH})_3\text{CNH}_2]$, ACS Reagent, $\geq 99.8\%$, Sigma-Aldrich) and 40 mL of 1 kmol L^{-1} hydrochloric acid (HCl , ACS, 36.6–38%, Alfa Aesar) solution. The SBF solution (ion concentrations are shown in Table 1) was prepared according to Kokubo *et al.*²³

GC(85%)–PMMA(15%), GC(90%)–PMMA(10%) and GC(95%)–PMMA(5%) pellets were soaked in 20 mL of SBF solution and kept in closed fluoroplastic PFA (Roth) tins (placing one sample into each tin) at a starting pH of 7.25 at 37 °C for 1, 2 and 3 weeks.

2.4. Characterization

Thermogravimetric analysis (TG) and differential scanning calorimetry (DSC) of the Si–Ca–Na–P–O gels were performed using Perkin Elmer STA 6000 Simultaneous Thermal Analyzer. Dried samples of ~5–10 mg were heated from 25 to 900 °C at a heating rate of 10 °C min^{-1} in a dry flowing air (20 mL min^{-1}).

The phase composition of annealed GC powders was studied by X-ray diffraction (XRD, Rigaku, MiniFlex II, Cu-K α radiation,



Table 1 The concentrations of ions in the SBF

Ion	Na ⁺	K ⁺	Mg ²⁺	Ca ²⁺	Cl ⁻	HCO ₃ ⁻	HPO ₄ ²⁻	SO ₄ ²⁻
Concentration, mmol L ⁻¹	142.0	5.0	1.5	2.2	147.8	4.2	1.0	0.5

$\lambda = 0.1542$ nm, 40 kV, 100 mA, $2\theta = 10$ – 60°) analysis. Identification of crystalline phases was achieved by comparing the experimentally obtained XRD patterns to the database provided by JCPDS (Joint Committee on Powder Diffraction Standards).

Morphological features of the samples and elemental distribution were evaluated using field emission scanning electron microscopy (FE-SEM, SU70, Hitachi) equipped with the energy dispersive X-ray spectrometer (EDS) and using INCA software (Oxford Instruments). The samples were examined using secondary electron (SE) imaging mode. The electron beam acceleration voltage was 5 kV and 20 kV for SEM and EDS analysis, respectively. X-ray accusation time of 60 seconds was used to obtain the EDS spectra (9 spectra were obtained over different regions for each sample).

Infrared spectra of the ceramic glass and composite powder samples were recorded using Fourier transform infrared (FT-IR) spectrometer (Frontier FT-IR), Perkin Elmer, BaF₂ plate (ZnSe ATR crystal, liquid-nitrogen-cooled mercury cadmium telluride (MCT) detector), 4000–600 cm⁻¹. FTIR absorbance spectra of the surface of composite pellets before and after the dissolution were recorded in reflectance geometry using Thermo scientific Nicolet iZ10 spectrometer.

The compressive strength tests were performed with a universal testing machine “Haunsfield H10KS” (Hounsfield Test Equipment Ltd, Redhill, UK) and software “Qmat Professional”. The cell load was of 10 kN with a resolution of 0.1 N (load measurement accuracy: $\pm 0.5\%$ of indicated load from 2% to 100% capacity). Load speed was of 20 mm min⁻¹ (position measurement accuracy: 0.001 mm, speed accuracy: $\pm 0.005\%$ of set speed). To calculate density (ρ), the weight, height and diameter of prepared specimens were estimated using precision balance Mettler toledo MS1003S ($e = 0.01$ g, $d = 0.001$ g) and DIGI-MET indicator ($e = 0.01$ mm, $d = 0.001$ mm) from Helios Preisser.

To eliminate the impact of varying densities of specimens (which forms during its preparation) on compressive strength, the estimated strength σ_{estim} , corresponding to mean density of specimens, was calculated according to eqn (1):

$$\sigma_{\text{estim}} = \sigma_i \frac{\rho_{\text{mean}}}{\rho_i} \quad (1)$$

where σ_i is the actual strength of sample, ρ_{mean} – mean density of tested samples (1905 kg m⁻³) and ρ_i – density of the tested sample.

Dissolution of fabricated composite pellets was monitored by measuring the concentrations of Si, Ca and P in soaking solutions. Quantitative determination of the elements was performed using Perkin Elmer Optima 7000 DV inductively coupled plasma optical emission spectrometer (ICP-OES).

3. Results and discussion

3.1. Crystallization and composition evaluation of glass-ceramic powders

The results of TG and DSC analyses of Si-Ca-Na-P-O gel powders were used to evaluate thermal behaviour of GC. TG/DTG and DSC curves of representative Ca-P-Na-Si-O gel powders are shown in Fig. 1. Six main steps of weight loss are clearly seen in the DTG curve. The first very small weight loss of $\sim 3\%$ without significant events in the DTG curve was observed by heating the sample up to 200 °C and assigned to the removal of adsorbed water. The second step of weight loss (6%) is observed in the range of 220–270 °C and assigned to the decomposition of precursor gel and evolution of NO_x gasses and organic fragments. This is also evident from the exothermic signals in the DSC curve (maxima at 222 °C and 234 °C). The next small weight loss of about 2% observed at 334 °C (DTG curve) was further assigned to the decomposition of organics and evolution of absorbed water. Then, the significant mass loss at around 580–710 °C and simultaneous broad endothermic peak (DSC curve) with three maxima at 605, 662 and 702 °C (DTG curve) takes place. This step is related to the nucleation of primary crystals and crystallization processes of glass ceramic. Similar results were reported by Boccaccini *et al.* where sintering and crystallization behaviour of bioglass powders was investigated.²⁴

One shall note that in thermal treatment of Ca-P-Na-Si-O gel powders more than one crystalline phase may form depending on the temperature.^{25,26} The last thermal effect with weight loss of $\sim 5\%$ is observed at 764 °C (DTG curve) followed by endothermic reaction (DSC curve, maxima at 759 °C) and can be further assigned to the increased crystallization of glass-ceramic powders as well as decomposition of nitrate into nitrite²⁷ and evolution of gasses.

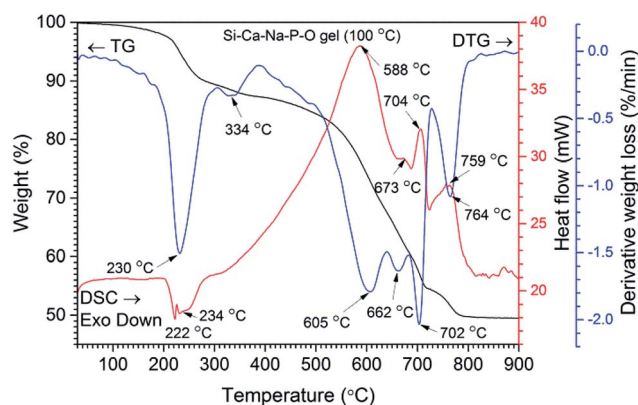


Fig. 1 TG/DTG and DSC curves of Si-Ca-Na-P-O gel dried at 100 °C.



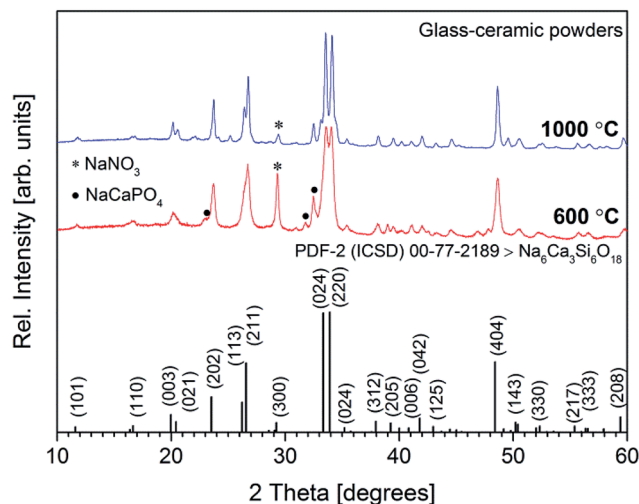


Fig. 2 X-ray diffraction patterns of glass-ceramic powders annealed at 600 and 1000 °C.

XRD patterns of Ca–P–Na–Si–O gel powders annealed at 600 and 1000 °C are presented in Fig. 2.

Significant reflections in the XRD pattern were observed at $2\theta = 11.68^\circ, 16.73^\circ, 20.13^\circ, 23.64^\circ, 26.65^\circ, 33.56^\circ, 34.07^\circ, 35.33^\circ, 38.03^\circ, 39.46^\circ, 41.04^\circ, 41.95^\circ, 43.18^\circ, 48.60^\circ, 50.48^\circ, 52.18^\circ, 55.69^\circ, 56.50^\circ$ and 59.69° confirming the formation of crystalline solid at around 600 °C. These reflections were assigned to the (101), (110), (033), (202), (211), (024), (220), (024), (312), (205), (006), (042), (125), (404), (143), (330), (217), (333) and (208) diffraction peaks of sodium calcium silicate ($\text{Na}_6\text{Ca}_3\text{Si}_6\text{O}_{18}$) [JCPDS no. 77-2189] phase. The formation of other sodium calcium silicates such as $\text{Na}_2\text{Ca}_2\text{Si}_3\text{O}_9$ [JCPDS no. 22-1455] and $\text{Na}_4\text{Ca}_4\text{Si}_6\text{O}_{18}$ [JCPDS no. 79-1089] might also be observed, since different sodium calcium silicates exhibit very similar reference patterns and often consist of a solid solution.^{22,28,29} The reflections observed at $2\theta = 23.12^\circ, 31.04^\circ, 31.75^\circ$ and 32.50° were assigned to the buchwaldite (NaCaPO_4) [JCPDS no. 29-1193] phase, while the intensive reflection at $2\theta = 29.37^\circ$ was assigned to the sodium nitrate (NaNO_3 phase [JCPDS no. 36-1474]). Similar results were reported by Pirayesh and Nychka where NaNO_3 phase was observed at the temperatures up to 600 °C.³⁰ Further annealing at 700 °C (data not presented) and 1000 °C increased crystallinity, whereas the sharp peak of NaNO_3 phase in the XRD pattern slightly diminished. This is probably due to the change of material phase, considering that the nitrates at higher temperatures decompose to the respective nitrites and oxygen (designated NaNO_2 [JCPDS no. 83-2254] phase).²⁷

3.2. Morphology evaluation of GC powders and GC–PMMA composites

To see distribution of glass-ceramic within the PMMA matrix the surface morphological examination was performed. The FE-SEM micrographs of GC powders annealed at 600 °C and selected pellets of GC–PMMA (wt ratios of 75/25 and 95/5) composite are presented in Fig. 3.

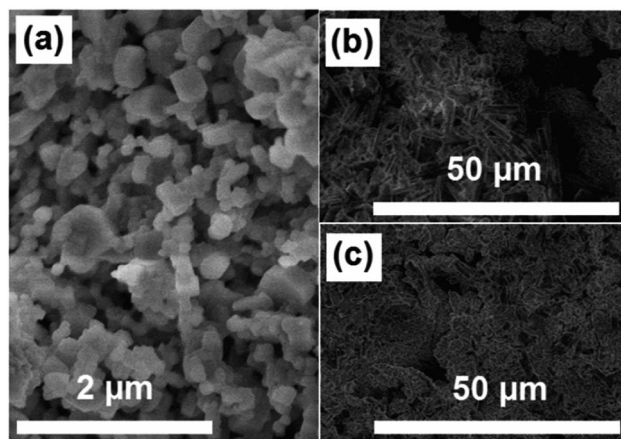


Fig. 3 FE-SEM micrographs showing surface morphology of (a) GC powders synthesised at 600 °C, and (b) GC(75%)–PMMA(25%) and (c) GC(95%)–PMMA(5%) composites (FE-SEM micrographs showing larger surface regions of composites are presented in ESI Fig. S1†).

Irregular micro-voids within some areas of composite surface were observed, while samples of neat PMMA appeared to be dense (ESI Fig. S1†). The content and distribution of voids varies within different surface regions examined and increases with an increase of PMMA content. This can be attributed to the inorganic–organic composite preparation procedure and non-uniform distribution of glass ceramic material within polymer matrix. The size of the irregular voids was estimated to be in the range of $20 \times 10 \pm 5 \mu\text{m}$ and $1 \times 0.5 \pm 0.25 \mu\text{m}$ for the GC(75%)–PMMA(25%) and GC(95%)–PMMA(5%) composites, respectively (FE-SEM micrographs). One might find that the formed voids can lead to some porosity which is one of the most important characteristics for scaffolding materials, since it allow migration and proliferation of osteoblasts and mesenchymal cells, effective vascularisation, bone ingrowth as well as improves mechanical interlocking between artificial implant biomaterial and surrounding natural bone providing mechanical stability at the interface.³¹

3.3. Spectroscopic characterization of GC–PMMA composites

Fig. 4 shows absorption FTIR spectra of PMMA and GC(75%)–PMMA(25%) composite. FTIR spectrum of neat PMMA in the region of $1700\text{--}1100 \text{ cm}^{-1}$ exhibits characteristic bands located at $1722, 1656, 1595, 1436, 1386, 1228, 1192$ and 1143 cm^{-1} . Similar absorption spectrum of PMMA was presented by Gonzalez-Benito and Gonzalez-Gaitano.³² FTIR spectrum of GC(75%)–PMMA(25%) composite showed broad and distinctive bands in the region of $1100\text{--}850 \text{ cm}^{-1}$. Bands centered at 1018 and 922 cm^{-1} were assigned to the Si–O–Si vibrational modes of orthosilicate (SiO_4^{4-}) tetrahedral group, whereas the phosphate vibrations are not very well resolved.^{33,34} The shoulder at 881 cm^{-1} and a couple bands in the $600\text{--}550 \text{ cm}^{-1}$ region were related to the P–O vibrations which overlap with mixed stretching and bending vibrations of Si–O–Si bond.^{33,34} In the $1500\text{--}1350 \text{ cm}^{-1}$ region broad bands were obtained and



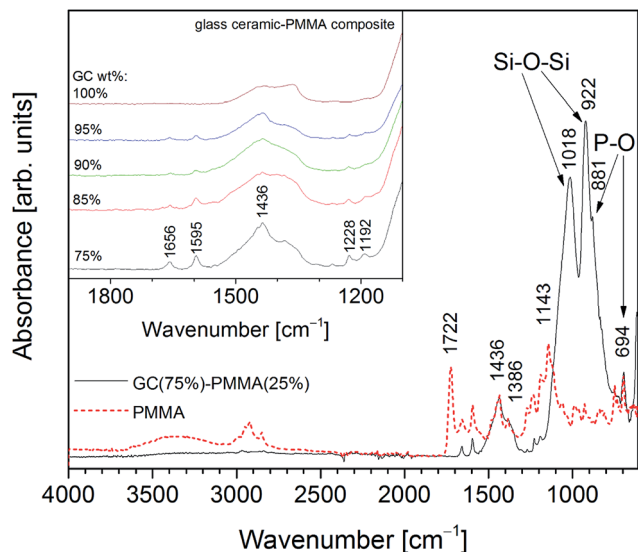


Fig. 4 Absorption FTIR spectra of GC(75%)–PMMA(25%) composite and PMMA (inset: absorption FTIR spectra of GC (synthesised at 600 °C) and GC–PMMA composites in the range of 1100–1900 cm^{-1}).

assigned to the PMMA matrix overlapping with C–O vibrations of carbonate group. The enlarged FTIR spectra of glass ceramic powders and GC–PMMA composites in the 1900–1100 cm^{-1} region are presented in Fig. 4 inset. The results clearly point out that with increase of PMMA content the intensity of vibrational modes of PMMA (bands located at 1656, 1595, 1228 and 1192 cm^{-1}) also increases. Furthermore, the intense PMMA band centered at 1722 cm^{-1} assigned to the C=O stretching vibrations^{17,35} disappears in the spectra of GC–PMMA composites.

Moreover, to see if spectral features of the composite surface vary from the bulk material the FTIR spectra from the powdered composites (pellets were grinded in an agate mortar) were additionally recorded. Spectral features of the powdered materials are in a good agreement with spectra obtained from the surface of the composite pellets (ESI Fig. S2†).

3.4. Mechanical properties of GC–PMMA composites

The mechanical properties of GC–PMMA composites were investigated by the uniaxial compressive strength test (Fig. 5). In this system, the compressive strength of composites (σ_i) was in the range of 42–50 MPa (the compressive strength of neat PMMA pellets was measured to be 126 MPa) and decreased with the increase of glass ceramic–polymer ratio. The composites containing 75, 85 and 90% of glass ceramic exhibited very similar compressive strength values. The compressive strength of bioactive glass scaffolds spans from 0.2 to 150 MPa and is composition, microstructure and/or fabrication method dependent,^{7,36,37} although macroporous bioactive glass scaffolds can display compressive strength values even below 0.2 MPa.³⁸ Material crystallinity also plays an important role in mechanical strength and generally increases when the crystalline phase is increased. For example, Daguno *et al.* reported that the amount of crystal phase formed in the bioactive glass improves

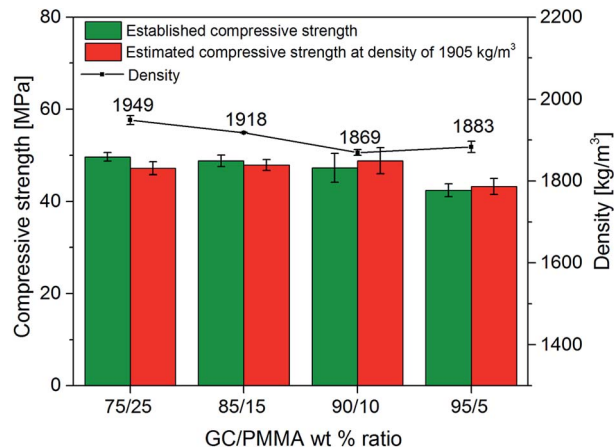


Fig. 5 Density of samples, measured compressive strength, and estimated compressive strength at density of 1905 kg m^{-3} of GC–PMMA composite materials.

the hardness, fracture toughness, bending strength and elastic modulus of the material depending on the heat treatment temperature.³⁹ Similar results were obtained in the work presented by Sabree *et al.* showing that ceramic scaffolds made of amorphous material exhibit brittleness compared to those produced of purely crystalline material.⁴⁰ The compressive or diametral tensile strength of the PMMA can also be altered by incorporating different additives into the bone cement.^{10,15,41,42} The density (ρ) values (Fig. 5) showed that composites containing 90% and 95% of GC exhibit slightly lower density in comparison to those containing 75% and 85%, while calculated compressive strength at $\rho = 1900 \text{ kg m}^{-3}$ shows that the highest value was obtained of composite containing 90% of GC. One can also observe that composite containing 95% of GC decreased in compressive strength by 11.4% and this is attributed mostly to the small amount of supporting PMMA matrix as well as decreased density of material. The results imply that GC(90%)–PMMA(10%) composite yields the highest strength and lowest density compared to all fabricated materials.

3.5. GC–PMMA composite dissolution in SBF solution

Fig. 6 shows the concentrations of Si, P and Ca in a soaking solutions as a function of dissolution time for GC(85%)–PMMA(15%), GC(90%)–PMMA(10%) and GC(95%)–PMMA(5%) composite materials. The concentration of Si in dissolution medium gradually increased over the immersion time and after 3 weeks reached 115 ppm, 135 ppm and 165 ppm for the GC(95%)–PMMA(5%), GC(85%)–PMMA(15%) and GC(90%)–PMMA(10%) composites, respectively, with most significant alteration obtained after the first week (70–90 ppm). This sudden increase in Si concentration shows and indicates the fast dissolution rate of the glass in the SBF. The highest concentrations of Si were obtained for the GC(90%)–PMMA(10%) material and this could be attributed to the higher porosity of composite and faster surface chemistry. The concentration of Ca in the dissolution medium, on the contrary, falls gradually (it ranges from 90 ppm to 10–20 ppm). However, different variation in the



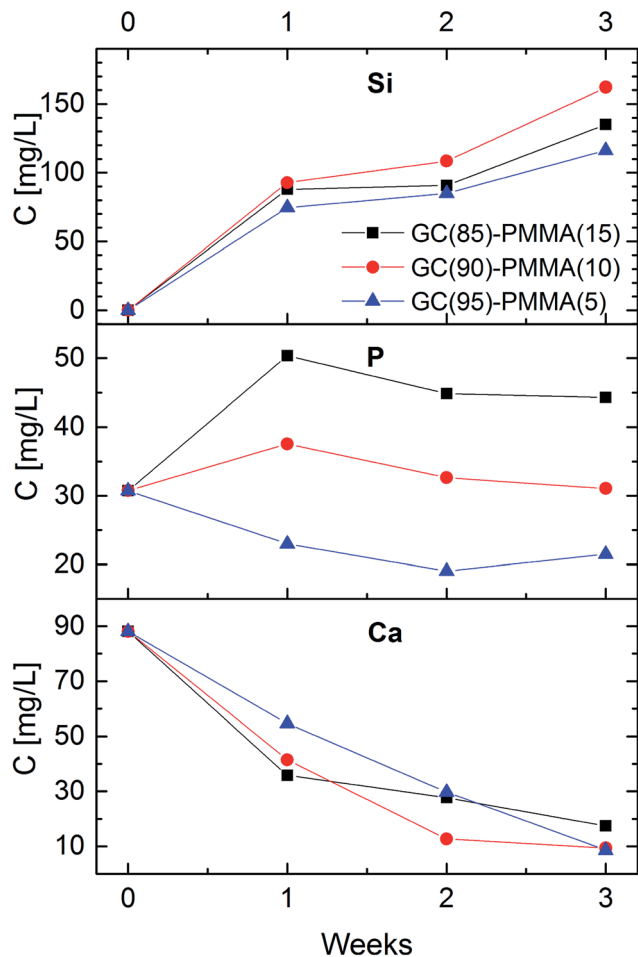


Fig. 6 Concentrations of Ca, P and Si in the soaking solution as a function of immersion time.

content of phosphorus was obtained. An increase in P concentration was observed first week after the dissolution for the GC(85%)–PMMA(15%) and GC(90%)–PMMA(10%) composites, following with small decrease after the second and third week of dissolution. The concentrations of P for the GC(95%)–PMMA(5%) composite initially decreases and for the last week of dissolution slightly increase. Generally, these processes are due to the dissolution–precipitation reactions that occur at glass ceramic surface in contact with SBF. A rapid ion exchange between glass ceramic surface and dissolution medium induces the formation of hydrated silica-rich layer. Subsequently Ca^{2+} and HPO_4^{2-} ions diffuse towards the composite surface and as a result a calcium phosphate ceramic is formed *via* heterogeneous nucleation. The mechanism and sequence of reactions taking place on the bioactive glass surface when in contact with physiological fluids and involved in the apatite layer formation were reported elsewhere.^{21,43,44} The dissolution of glass ceramic is important since it relates to the bioactivity. Nevertheless, it is solution parameters, glass composition, sintering temperature and topography dependent.^{8,44}

3.6. Morphological and structural characterization of GC–PMMA composites after the dissolution in SBF

Fig. 7 depicts FE-SEM micrographs of the different morphological structures formed on the surface of GC–PMMA composite materials soaked in the SBF solution for 1, 2 and 3 weeks. The morphological features differ significantly compared to those obtained from as-prepared samples. These changes again indicate the leaching and diffusion processes of ions towards the composite surface, with subsequent formation of ceramic material on the surfaces of all the samples. Microscopic surface cracking was also observed after samples were soaked in SBF solution (micrographs not presented). Some cracks were slightly less pronounced when samples were soaked for 3 weeks. This indicates the formation of thicker ceramic component layer on the surface.

EDS analysis showed the elemental composition of newly formed ceramic material on the composite surface after the dissolution. The representative FE-SEM micrographs with EDS-based elemental mapping and EDS spectra of GC(95%)–PMMA(5%) composite before and three weeks after the dissolution are presented in Fig. 8. Elements such as C, O, Na, Mg, Si, P, Cl and Ca were detected and the average atomic percent (%) ratios of Mg : Si : P : Ca were 2.6 : 4.6 : 11.9 : 19.6. The ceramic composition was similar to that of the HAP with an average Ca : P ratio of 1.65 ($n = 9$). Nonstoichiometric nature of deposited material generally can be explained by the existence of substitutions and vacancies in the crystal lattice of HAP (stoichiometry for HAP Ca/P is 1.67).⁴⁵ Furthermore, the presence of different amorphous apatitic phases might also be present. Raynaud *et al.* demonstrated that HAP with a Ca/P atomic ratio of 1.65 dissociated into a mixture of HAP and β -TCP after heating at 1000 °C.⁴⁶

The surface of GC(95%)–PMMA(5%) composite before dissolution contained C, O, Na, Si, P and Ca and the average atomic% ratios of Si : P : Ca were 6.3 : 1.0 : 5.1 (10.95 : 1.2 : 7.8 and 10.88 : 8.62 : 1.09 : 5.72 for GC(90%)–PMMA(10%) and GC(85%)–PMMA(15%) composites, respectively). Based on these observations, we can conclude that the layer rich in calcium and phosphorus has been deposited on the surface of composite materials. The EDS-based mapping were observed over entire samples and showed homogeneous distribution of the P and Ca elements on the surface of composites after the dissolution.

The presence of newly formed ceramic material on the surface of composites was further examined by XRD analysis and FTIR spectroscopy. XRD patterns of GC(95%)–PMMA(5%) composite before and after the dissolution are presented in Fig. 9. Generally, the poorly crystalline nature of deposited ceramic material is indicated by the broad peaks and low resolution. One can observe that the sharp diffraction peaks of sodium calcium silicate ($\text{Na}_6\text{Ca}_3\text{Si}_6\text{O}_{18}$) [JCPDS no. 77-2189] phase in the XRD pattern are less expressive and broader in comparison to those obtained from composites before dissolution. It should be noted, that intensity of reflections at $2\theta \sim 31.00^\circ$ and 32.50° increased compare to the main reflections of $\text{Na}_6\text{Ca}_3\text{Si}_6\text{O}_{18}$ phase observed at $2\theta = 33\text{--}34^\circ$. This result



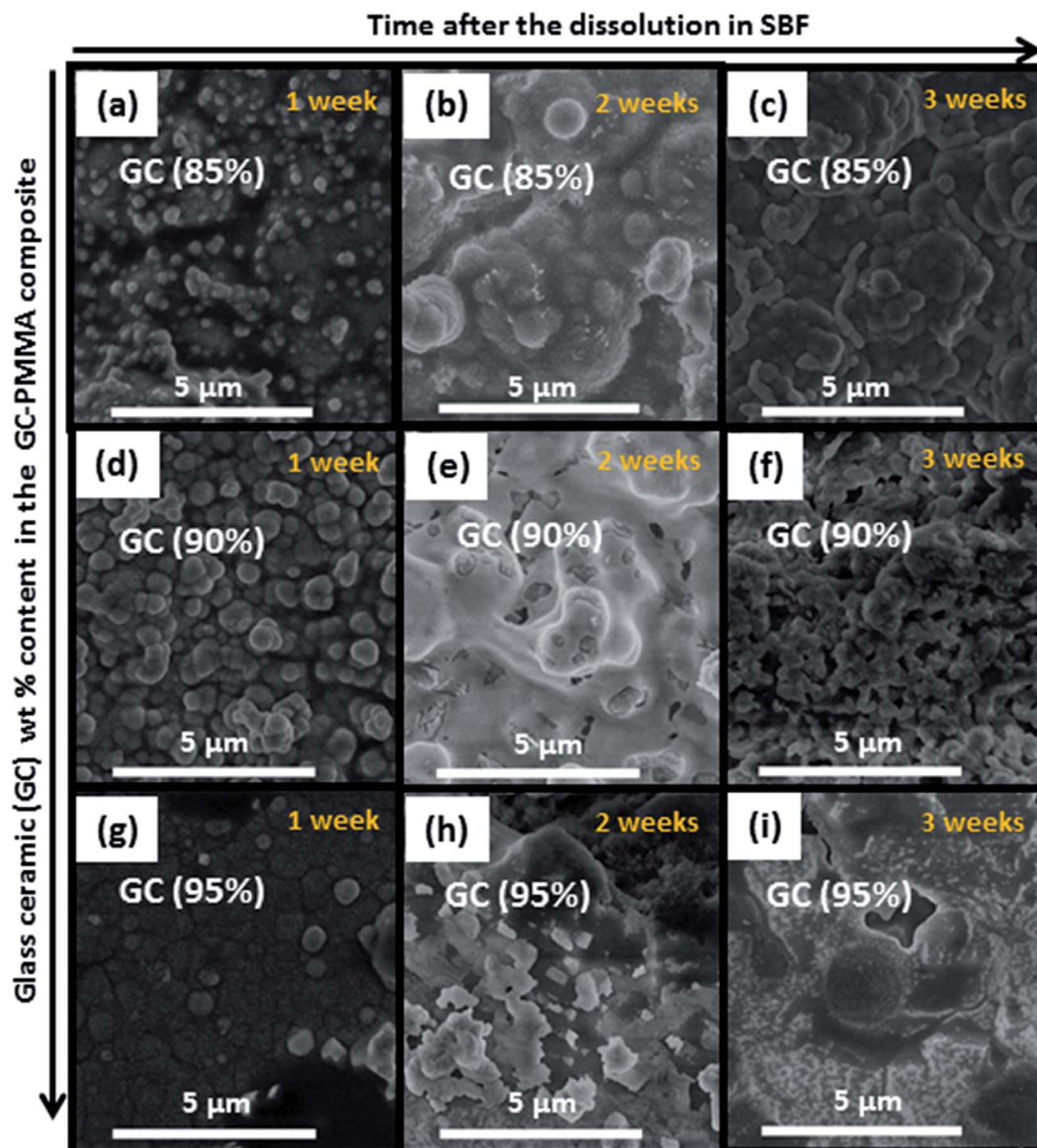


Fig. 7 FE-SEM micrographs of the surface of the ((a)–(c)) GC(85%)–PMMA(15%), ((d)–(f)) GC(90%)–PMMA(10%) and ((g)–(i)) GC(95%)–PMMA(5%) composites after 1, 2 and 3 weeks of soaking in SBF.

indicate the possible formation of poorly crystalline calcium phosphate after the composites were soaked in SBF solution (the main Bragg peaks characteristic of crystalline HAP arise in the region from 31 to 33° degrees [JCPDS no. 09-0432]⁴⁷). The rate of cHAP layer deposition was studied by Pereira *et al.*⁴⁴ Furthermore, in the XRD pattern of composite materials after the dissolution of three weeks, the sharp reflections at $2\theta = 31.68^\circ$, 45.42° and 56.50° were obtained and assigned to the NaCl [JCPDS no. 96-900-6375] phase. This result is attributed to the different post-dissolution treatment of the composite samples. It has been reported that to prevent aging of mineralized ceramic, materials are treated in acetone. However, we previously observed (data not reported) that such post-dissolution treatment induces slight demineralization of deposited ceramic and solvent becomes turbid. To assess actual composition and structure of the surface layer the XRD patterns

from the as-deposited materials after the dissolution for 3 weeks were recorded. The composites containing 85% and 95% of GC showed similar diffraction patterns (data not presented).

Absorption FTIR spectra recorded from the surface of GC(95%)–PMMA(5%) composite material after the dissolution for 1, 2 and 3 weeks are presented in Fig. 10. FTIR spectra show characteristic bands of the phosphate (PO_4^{3-}) and carbonate (CO_3^{2-}) groups and the deposited material was identified as carbonate containing HAP. In the $1100\text{--}900\text{ cm}^{-1}$ region, a broad band with a maximum at 995 cm^{-1} (shoulder at 936 cm^{-1}) was obtained and assigned to the triply degenerate asymmetric stretching mode, ν_3 , of the apatitic PO_4^{3-} groups. It has been previously reported that band obtained at around 600 cm^{-1} indicates a formation of amorphous calcium phosphate phase, while splitting of this band shows the presence of the crystalline phase.^{33,34,45,48} The bands observed in the low



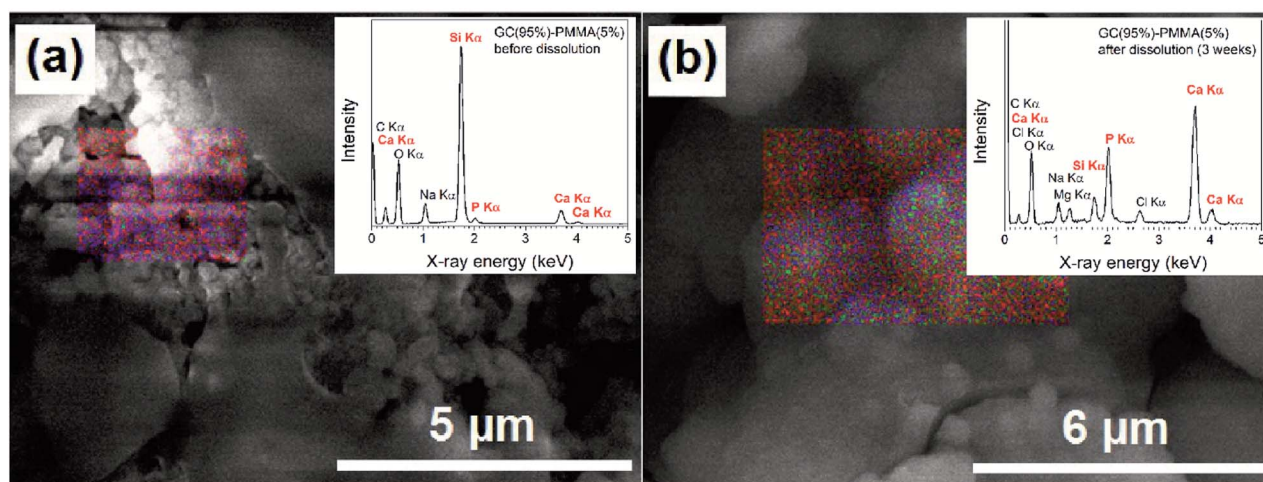


Fig. 8 FE-SEM micrographs showing EDS-based elemental mapping and EDS spectra (insets) of GC(95%)-PMMA(5%) composites (a) before dissolution and (b) three weeks after the dissolution in SBF (designated EDS mapping colours: Ca – red, P – green, O – blue).

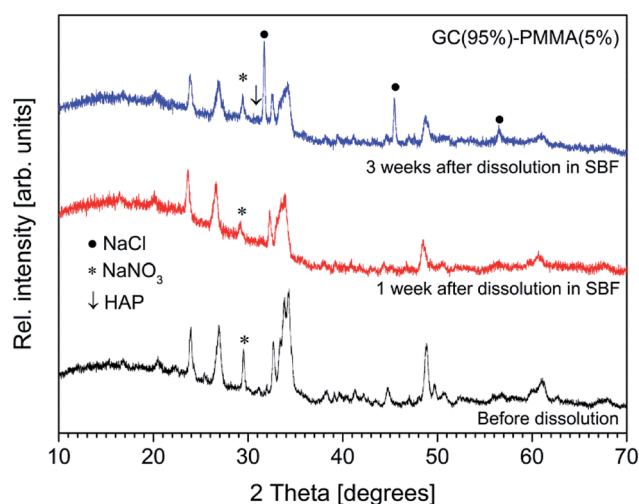


Fig. 9 XRD diffractograms of GC(95%)-PMMA(5%) composite surface before and after the dissolution in SBF.

wavenumber region of $650\text{--}600\text{ cm}^{-1}$ (maxima at 608 cm^{-1}) were attributed to the triply degenerated bending mode, ν_4 , of the O–P–O bonds.⁴⁷ An insignificant splitting of this band indicates the deposition of poorly crystalline calcium phosphate – amorphous material as a major phase with a small presence of crystalline phase.^{8,34} Furthermore, the characteristic bands corresponding to the CO_3^{2-} group were obtained in the $1600\text{--}1350\text{ cm}^{-1}$ interval.⁴⁷ This suggests that carbonate group is being incorporated into the deposited ceramic to form cHAP. The peaks appear for substitutions both in phosphate (B-type carbonate) and OH (A-type carbonate) sites: a band located at 1494 cm^{-1} was assigned to the stretching modes, ν_3 , while the bands at 1454 cm^{-1} and 1436 cm^{-1} to the bending modes, ν_4 or ν_3 , of C–O bond.^{9,45} In the lower wavenumber region a band arising from the bending mode, ν_2 , of C–O bond (CO_3^{2-} group) was observed at $\sim 865\text{ cm}^{-1}$ and assigned to the A-type substitution, while broad band $\sim 700\text{ cm}^{-1}$ was designated to the in-

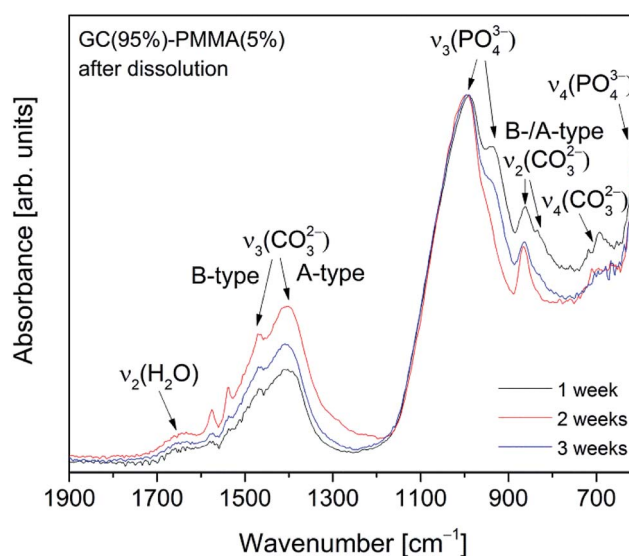


Fig. 10 Absorption FTIR spectra recorded from the surface of GC(95%)-PMMA(5%) composites 1, 2 and 3 weeks after the dissolution in SBF.

plane band, ν_4 , CO_3^{2-} group. In another work by Pereira *et al.* the formation of cHAP on the surface of sol-gel derived bioglass surfaces *in vitro* has been demonstrated.³⁴

It is known that HAP is thermodynamically the most stable phase in physiological conditions and has the ability for direct chemical bonding to the bone. However, previous studies have shown that mechanism of bone apatite formation involves metastable intermediate products including an amorphous calcium phosphate.⁴⁹ This indicates the favourable transformation of deposited low crystallinity calcium phosphate into the crystalline hydroxyapatite. GC(90%)-PMMA(10%) and GC(85%)-PMMA(15%) composites exhibited very similar spectral features showing the formation of A- and B-substituted



carbonated apatite under identical experimental conditions (ESI Fig. S3†).

Although we can conclude that in the current work the inorganic–organic composites were successfully prepared and the growth of low crystallinity carbonated HAP on the surface of composites immersed into SBF under static conditions was obtained, the studies examining the effect of the observed surface morphology on cell behaviour *in vitro* and *in vivo* still has to be performed and will be a subject of further studies.

4. Conclusions

In this work glass-ceramic powders were synthesised *via* sol–gel method at 600 °C. XRD analysis showed that during sintering the crystallization takes place and sodium calcium silicate ($\text{Na}_6\text{Ca}_3\text{Si}_6\text{O}_{18}$) was observed as a main phase. Homogeneous GC–PMMA composites with fractional GC content ranging from 75 to 95% were successfully prepared *via* photopolymerization reaction. The mechanical compressive strength of GC–PMMA composite materials ranged from 42.5 to 49.6 MPa. The GC(90%)–PMMA(10%) composites exhibited the highest strength and lowest density. ICP-OES results indicated the degradation process of the glass ceramic in simulated body fluid. A combination of XRD and SEM/EDS analyses revealed that the alteration products are mostly amorphous material with the composition similar to that of the HAP (average atomic% ratio of Ca/P = 1.65). According to FTIR analysis the ceramic layer formed after the dissolution was low crystallinity calcium phosphate with crystalline phase attributed to carbonate A- and B-substituted HAP. The findings indicate that fabricated composites could be potential materials for bone scaffold engineering when extreme strength is not required.

Acknowledgements

This research was funded by a grant KALFOS (No. LJB-2/2015) from the Research Council of Lithuania.

References

- 1 M. M. Stevens, *Mater. Today*, 2008, **11**, 18–25.
- 2 P. V. Giannoudis, H. Dinopoulos and E. Tsiridis, *Injury*, 2005, **36**, S20–S27.
- 3 Global Data, MediPoint: Bone Grafts and Substitutes – Global Analysis and Market Forecast, Executive Summary, 2014, January, Retrieved from: <https://www.marketresearch.com/>, Reference code: GDME0192MAR.
- 4 J. R. Jones, E. Gentleman and J. Polak, *Elements*, 2007, **3**, 393–399.
- 5 K. Jahan and M. Tabrizian, *Biomater. Sci.*, 2016, **4**, 25–39.
- 6 J. J. Li, D. L. Kaplan and H. Zreiqat, *J. Mater. Chem. B*, 2014, **2**, 7272–7306.
- 7 Q. Fu, E. Saiz, M. N. Rahaman and A. P. Tomsia, *Mater. Sci. Eng., C*, 2011, **31**, 1245–1256.
- 8 P. Sepulveda, J. R. Jones and L. L. Hench, *J. Biomed. Mater. Res.*, 2002, **61**, 301–311.
- 9 E. Garskaite, L. Alinauskas, M. Drienovsky, J. Krajcovic, R. Cicka, M. Palcut, L. Jonusauskas, M. Malinauskas, Z. Stankeviciute and A. Kareiva, *RSC Adv.*, 2016, **6**, 72733–72743.
- 10 M. Arora, E. K. S. Chan, S. Gupta and A. D. Diwan, *World J. Orthoped.*, 2013, **4**, 67–74.
- 11 J. M. Moran, D. Pazzano and L. J. Bonassar, *Tissue Eng.*, 2004, **9**, 63–70.
- 12 D. M. Yunos, Z. Ahmad and A. R. Boccaccini, *J. Chem. Technol. Biotechnol.*, 2010, **85**, 768–774.
- 13 Z. Li, G. Lang, X. Chen, H. Sacks, C. Mantzur, U. Tropp, K. T. Mader, T. C. Smallwood, C. Sammon, R. G. Richards, M. Alini and S. Grad, *Biomaterials*, 2016, **84**, 196–209.
- 14 G. Lewis, *J. Biomed. Mater. Res.*, 1997, **38**, 155–182.
- 15 L. Chen, D. Zhai, Z. Huan, N. Ma, H. Zhu, C. Wu and J. Chang, *RSC Adv.*, 2015, **5**, 37314–37322.
- 16 G. Lewis, *J. Biomed. Mater. Res., Part B*, 2006, **76**, 456–468.
- 17 A. K. Riau, D. Mondal, G. H. F. Yam, M. Setiawan, B. Liedberg, S. S. Venkatraman and J. S. Mehta, *ACS Appl. Mater. Interfaces*, 2015, **7**, 21690–21702.
- 18 B. Vázquez, M. P. Ginebra, X. Gil, J. A. Planell and J. San Román, *Biomaterials*, 2005, **26**, 4309–4316.
- 19 M. K. Singh, T. Shokuhfar, J. J. d. A. Gracio, A. C. M. de Sousa, J. M. D. F. Ferreira, H. Garmestani and S. Ahzi, *Adv. Funct. Mater.*, 2008, **18**, 694–700.
- 20 R. Ravarian, C. M. Murphy, A. Schindeler, A. Rawal, J. M. Hook and F. Dehghani, *RSC Adv.*, 2015, **5**, 60681–60690.
- 21 A. J. Salinas and M. Vallet-Regi, *RSC Adv.*, 2013, **3**, 11116–11131.
- 22 J. Faure, R. Drevet, A. Lemelle, N. Ben Jaber, A. Tara, H. El Btaouri and H. Benhayoune, *Mater. Sci. Eng., C*, 2015, **47**, 407–412.
- 23 T. Kokubo and H. Takadama, *Biomaterials*, 2006, **27**, 2907–2915.
- 24 A. R. Boccaccini, Q. Chen, L. Lefebvre, L. Gremillard and J. Chevalier, *Faraday Discuss.*, 2007, **136**, 27–44.
- 25 J. Massera, S. Fagerlund, L. Hupa and M. Hupa, *J. Am. Ceram. Soc.*, 2012, **95**, 607–613.
- 26 R. Golovchak, P. Thapar, A. Ingram, D. Savytskii and H. Jain, *Acta Biomater.*, 2014, **10**, 4878–4886.
- 27 C. M. Kramer, Z. A. Munir and J. V. Volponi, *Thermochim. Acta*, 1982, **55**, 11–17.
- 28 R. L. Siqueira, O. Peitl and E. D. Zanotto, *Mater. Sci. Eng., C*, 2011, **31**, 983–991.
- 29 I. Cacciotti, M. Lombardi, A. Bianco, A. Ravaglioli and L. Montanaro, *J. Mater. Sci.: Mater. Med.*, 2012, **23**, 1849–1866.
- 30 H. Pirayesh and J. A. Nychka, *J. Am. Ceram. Soc.*, 2013, **96**, 1643–1650.
- 31 G. Hannink and J. J. C. Arts, *Injury*, 2011, **42**, S22–S25.
- 32 J. González-Benito and G. González-Gaitano, *Macromolecules*, 2008, **41**, 4777–4785.
- 33 A. J. Salinas, M. Vallet-Regi and I. Izquierdo-Barba, *J. Sol-Gel Sci. Technol.*, 2001, **21**, 13–25.
- 34 M. M. Pereira, A. E. Clark and L. L. Hench, *J. Biomed. Mater. Res.*, 1994, **28**, 693–698.



- 35 T. Serizawa, K.-i. Hamada, T. Kitayama, N. Fujimoto, K. Hatada and M. Akashi, *J. Am. Chem. Soc.*, 2000, **122**, 1891–1899.
- 36 B. Thavornyutikarn, P. Tesavibul, K. Sitthiseripratip, N. Chatarapanich, B. Feltis, P. F. A. Wright and T. W. Turney, *Mater. Sci. Eng., C*, 2017, **75**, 1281–1288.
- 37 X. Wang, L. Zhang, X. Ke, J. Wang, G. Yang, X. Yang, D. He, H. Shao, Y. He, J. Fu, S. Xu and Z. Gou, *RSC Adv.*, 2015, **5**, 102727–102735.
- 38 Y. Minaberry and M. Jobbágy, *Chem. Mater.*, 2011, **23**, 2327–2332.
- 39 J. K. M. F. Daguano, K. Strecker, E. C. Ziemath, S. O. Rogero, M. H. V. Fernandes and C. Santos, *J. Mech. Behav. Biomed. Mater.*, 2012, **14**, 78–88.
- 40 I. Sabree, J. E. Gough and B. Derby, *Ceram. Int.*, 2015, **41**, 8425–8432.
- 41 S. Shinzato, T. Nakamura, T. Kokubo and Y. Kitamura, *J. Biomed. Mater. Res.*, 2001, **54**, 491–500.
- 42 G. Lewis, *J. Biomed. Mater. Res., Part B*, 2008, **84**, 301–319.
- 43 L. L. Hench, *J. Am. Ceram. Soc.*, 1991, **74**, 1487–1510.
- 44 M. M. Pereira, A. E. Clark and L. L. Hench, *J. Am. Ceram. Soc.*, 1995, **78**, 2463–2468.
- 45 W. Querido, L. G. Abraçado, A. L. Rossi, A. P. C. Campos, A. M. Rossi, R. A. S. San Gil, R. Borojevic, A. Balduino and M. Farina, *Calcif. Tissue Int.*, 2011, **89**, 358.
- 46 S. Raynaud, E. Champion, D. Bernache-Assollant and P. Thomas, *Biomaterials*, 2002, **23**, 1065–1072.
- 47 E. Garskaite, K.-A. Gross, S.-W. Yang, T. C.-K. Yang, J.-C. Yang and A. Kareiva, *CrystEngComm*, 2014, **16**, 3950–3959.
- 48 S. Radin, P. Ducheyne, B. Rothman and A. Conti, *J. Biomed. Mater. Res.*, 1997, **37**, 363–375.
- 49 M. Vallet-Regi and D. A. Navarette, Biological Apatites in Bone and Teeth, in *Nanoceramics in Clinical Use: From Materials to Applications*, 2nd edn, 2015, ch. 1, pp. 1–29.

