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New prenylxanthenes, polyketide hemiterpenoid pigments from the endophytic fungus *Emericella* sp. XL029 and their anti-agricultural pathogenic fungal and antibacterial activities†

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Four new prenylxanthenes (1–4) together with five known compounds (5–9) were isolated from the solid-substrate fermentation culture of *Emericella* sp. XL029 associated with the leaves of *Panax notoginseng*. Their structures and relative configurations were elucidated by extensive spectroscopic analysis and comparison of NMR data to those of known compounds. The anti-agricultural pathogenic fungal and antibacterial activities of all isolated compounds were evaluated against eight agricultural pathogenic fungi and nine human pathogenic bacteria. The anti-agricultural pathogenic fungal assay indicated that compounds 1, 2, 4, 8 and 9 displayed significant activity against *Drechslera maydis* with MIC value of 25 $\mu\text{g mL}^{-1}$. Moreover, compound 1 also exhibited significant activity against other three fungi including *Rhizoctonia cerealis*, *Fusarium oxysporum* and *Phylospora piricola*, and compounds 4 and 9 against *Rhizoctonia cerealis* with MIC value of 25 $\mu\text{g mL}^{-1}$. Compound 5 only demonstrated significant activity against *Phylospora piricola* with MIC value of 25 $\mu\text{g mL}^{-1}$. Further antibacterial activity tests revealed that compounds 1–9 showed significant antibacterial activity against all tested Gram-positive (except for drug resistant *Staphylococcus aureus*) and Gram-negative bacteria with MIC values ranging from 12.5 to 50 $\mu\text{g mL}^{-1}$. While only compounds 1, 4 and 9 displayed moderate activity against drug resistant *Staphylococcus aureus* with MIC value of 50 $\mu\text{g mL}^{-1}$.

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Introduction

Naturally occurring xanthenes, an important class of secondary metabolites featuring a common 9*H*-xanthen-9-one scaffold, are frequently found in plants, fungi, ferns, and lichens. Such secondary metabolites have been shown to display a range of pharmacological activities, such as antimicrobial, antioxidant and cytotoxic activities due to their tricyclic core as well as their highly diverse substituents.^{1–3} Naturally occurring prenylxanthenes, as an important type of secondary metabolites with a range of pharmacological activities, are particularly

common in plants (mainly in Clusiaceae, Gentianaceae, Moraceae, and Polygalaceae families) and fungi. Due to their unique structures and significant therapeutic potential, they have the promising potential applications in acting as lead compounds for new generations of anticancer, antibacterial, immunomodulatory, antidiabetic, and antiobesity agents, *etc.*⁴

Plant endophytic fungi inhabiting the normal tissues of host plants without causing apparent pathogenic symptoms are well known producers of novel secondary metabolites with outstanding potential for generating novel therapeutic leads.⁵ The fungal genus *Emericella*, as the major group of fungi, has been reported to be the promising resources for secreting novel bioactive prenylxanthenes, such as *E. rugulosa*, *E. varicolor*, *E. nidulans*, *E. sp.* SCSIO 05240, and *E. striata*.^{6–14} Furthermore, fungi of the *Emericella* species are the sexual states of *Aspergillus* species, the perfect states of *Aspergillus*, has been the source of various classes of bioactive compounds.^{8,23,24}

During our studies in searching for novel bioactive compounds from plant endophytic fungi, we have isolated the interesting pigment-producing fungal strain (XL029) associated with the leaves of *Panax notoginseng*, belonging to the genus *Emericella*. The present study was undertaken to search for novel pigments from the solid-substrate fermentation culture of

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† Electronic supplementary information (ESI) available: 1D and 2D NMR spectra, HRESIMS spectra, IR spectra, UV spectra of new compounds 1–4, and X-ray crystallographic data of 4. CCDC 1525724. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c7ra04762b



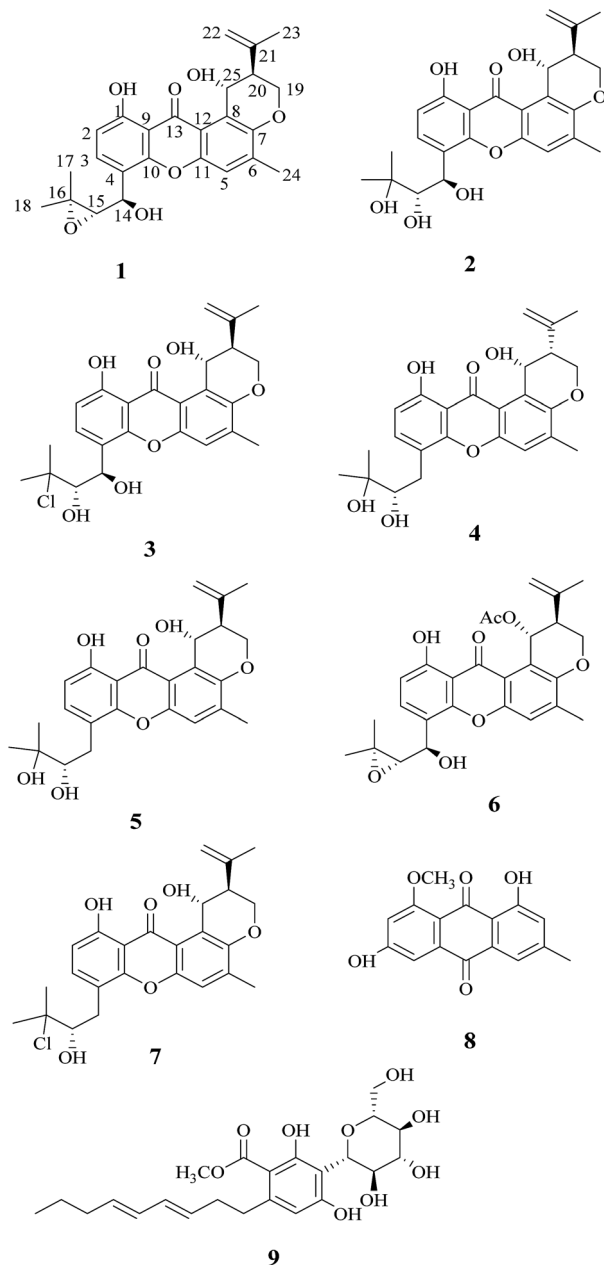


Fig. 1 Structures of compounds 1–9.

Emericella sp. XL029, which led to the isolation of nine compounds (1–9) including four new ones (1–4) (Fig. 1), seven of them belonging to prenylxanthenes. In this paper, the isolation, structure elucidation and biological activities of these compounds are described.

Results and discussion

Compound 1 was obtained as a yellow powder. The molecular formula was determined to be $C_{25}H_{26}O_7$ based on the HRESIMS at m/z 437.1586 ($[M - H]^-$, calcd 437.1606), implying the presence of thirteen degrees of unsaturation. The IR spectrum exhibited absorptions of hydroxyl (3446 cm^{-1}), aromatic ketone

(1643 cm^{-1}) functionalities. The ^1H NMR (Table 1) and HSQC spectra indicated the presence of three aromatic protons [δ_{H} 7.22 (s, H-5), 6.85 (d, $J = 8.6$ Hz, H-2), and 7.83 (d, $J = 8.6$ Hz, H-3)], four tertiary methyls [δ_{H} 1.31 (s, H-17), 1.50 (s, H-18), 1.86 (s, H-23), and 2.36 (s, H-24)], two methylenes including one oxygenated [δ_{H} 4.43 (dd, $J = 3.2, 10.9$, H-19a), 4.35 (dd, $J = 3.0, 10.9$, H-19b)] and one olefinic signal [δ_{H} 4.82 (s, H-22a), 4.60 (s, H-22b)], four methine signals [δ_{H} 5.14 (d, $J = 7.4$, H-14), 3.11 (d, $J = 7.6$, H-15), 2.74 (d, $J = 2.8$, H-20), and 5.43 (t, $J = 3.1, 6.2$, H-25)] as well as two exchangeable proton signals at δ_{H} 12.80 (s, 1-OH) and 4.97 (d, $J = 3.9$, 25-OH). The ^{13}C NMR and DEPT spectra (Table 1) revealed the presence of 25 carbons, which were assigned as four methyls, two methylenes (one oxygenated and one olefinic carbon), seven methines (including aromatic and oxygenated carbons), and twelve nonprotonated carbons (including one ketone carbonyl). The COSY correlations (H-19/H-20/H-25, corresponding to OCHCHCH₂O spin system) and the key HMBC correlations (Fig. 2) from H-19 to C-7, from H-20 to C-8, and from H-25 to C-7 and C-12 suggested the presence of dihydropyran ring, which was fused to an aromatic unit at C-7 and C-8. The isopropenyl moiety was deduced by the key HMBC correlations from H-22 to C-20, C-21 and C-23, which was connected to the pyran ring at C-20. A comprehensive analysis of 1D NMR data along with above evidences revealed the structure of 1 was mostly identical to 14-methoxytjixanthone,¹³ previously reported from the fungus *Emericella rugulosa*, except for the absence of methoxy group at C-14 and the presence of hydroxyl group, which was further confirmed by HMBC experiments and molecular weight. The relative configurations of all chiral centers in 1 were determined to be 14*R**, 15*S**, 20*S**, 25*R**, respectively, by comparison of the coupling constants, NMR data with those of reported tjixanthone derivatives,^{12–14} and the experimental optical rotation data [$[\alpha]_{\text{D}}^{25} -59^\circ$ (c 0.183, CHCl_3)] with those of related known compound 14-methoxytjixanthone-25-acetate [$[\alpha]_{\text{D}}^{20} -38^\circ$ (c 0.1, CHCl_3), which was further confirmed by the NOE correlations (Fig. 3) of H_a-19 with H_b-22, H-23, and H-25, H_b-19 with H-20 and OH-25, H-25 with H_b-22 and H-23. Thus, compound 1 was determined as 14-hydroxytjixanthone.

Compound 2 was obtained as a yellow powder. The molecular formula was determined to be $C_{25}H_{28}O_8$ by the HRESIMS at m/z 479.1684 ($[M + \text{Na}]^+$, calcd 479.1676), suggesting twelve degrees of unsaturation. Analysis of its 1D NMR spectroscopic data (Table 1) revealed the presence of 25 carbons, which were assigned as four methyls, two methylenes, seven methines, and twelve nonprotonated carbons. By comparison of its NMR spectra with those of 1 revealed that compound 2 has the same tjixanthone type skeleton,^{14,17,18} and its NMR data were almost identical to those of 1, obviously differing only in the chemical shifts at C-15 and C-16. The carbon chemical shifts at C-15 (δ_{C} 77.2) and C-16 (δ_{C} 73.9) were shifted significantly downfield in 2 compared to 1 [C-15 (δ_{C} 67.6) and C-16 (δ_{C} 60.5)], indicating that the 14,15-epoxy ring in 1 was opened to form 14,15-diol moiety in 2, which was further supported by evidence deduced from its molecular formula that has one more oxygen and two hydrogen atoms (18 mass units difference) than 1. It suggested that compound 2 is the epoxide ring-opening product of 1. The



Table 1 ^1H and ^{13}C NMR data for compounds for 1–4

No.	1^a		2^a		3^a		4^a	
	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	δ_{C} , type	δ_{H} (<i>J</i> in Hz)
1	161.7, C		160.5, C		161.2, C		160.4, C	
2	110.6, CH	6.85, d (8.6)	109.9, CH	6.79, d (8.5)	110.2, CH	6.83, d (8.4)	109.9, CH	6.76, d (8.3)
3	135.1, CH	7.83, d (8.6)	135.1, CH	7.84, d (8.5)	134.5, CH	7.81, d (8.5)	138.1, CH	7.52, d (8.3)
4	117.8, C		119.4, C		119.7, C		116.1, C	
5	118.9, CH	7.22, s	118.9, CH	7.15, s	119.0, CH	7.23, s	119.2, CH	7.23, s
6	138.8, C		138.5, C		138.7, C		138.7, C	
7	149.8, C		149.6, C		149.7, C		149.4, C	
8	121.3, C		120.9, C		121.2, C		121.6, C	
9	109.0, C		108.6, C		108.8, C		109.2, C	
10	152.1, C		151.0, C		151.2, C		153.1, C	
11	151.8, C		151.7, C		151.9, C		151.7, C	
12	116.9, C		116.7, C		116.9, C		116.7, C	
13	184.2, C		184.1, C		184.2, C		184.2, C	
14	66.5, CH	5.14, d (7.4)	66.4, CH	5.62, s	65.3, CH	5.68, brs	31.9, CH ₂	3.19, brd (14.0); 2.68, dd (10.5, 14.0)
15	67.6, CH	3.11, d (7.6)	77.2, CH	3.44, s	79.6, CH	3.73, brd (4.4)	77.7, CH	3.72, brd (10.2)
16	60.5, C		73.9, C		74.3, C		72.9, C	
17	24.8, CH ₃	1.31, s	27.0, CH ₃	1.32, s	29.4, CH ₃	1.79, s	23.5, CH ₃	1.36, s
18	19.7, CH ₃	1.50, s	26.0, CH ₃	1.50, s	29.0, CH ₃	1.78, s	26.6, CH ₃	1.41, s
19	64.7, CH ₂	4.43, dd (3.2, 10.9); 4.35, dd (3.0, 10.9)	64.5, CH ₂	4.39, dd (3.0, 10.9); 4.30, dd (2.7, 10.9)	64.6, CH ₂	4.43, dd (2.6, 10.9); 4.35, dd (3.0, 10.9)	64.0, CH ₂	4.47, dd (2.1, 10.1); 4.33, dd (10.4, 11.9)
20	44.9, CH	2.74, d (2.8)	44.7, CH	2.68, brs	44.9, CH	2.74, d (2.4)	43.9, CH	2.55, brd (11.8)
21	142.5, C		142.3, C		142.5, C		142.1, C	
22	112.4, CH ₂	4.82, s; 4.60, s	112.3, CH ₂	4.78, s; 4.55, s	112.3, CH ₂	4.82, s; 4.59, s	111.6, CH ₂	5.07, s; 4.78, s
23	22.5, CH ₃	1.86, s	22.4, CH ₃	1.82, s	22.5, CH ₃	1.86, s	22.5, CH ₃	1.99, s
24	17.5, CH ₃	2.36, s	17.3, CH ₃	2.32, s	17.5, CH ₃	2.36, s	17.5, CH ₃	2.37, s
25	63.3, CH	5.43, t (3.1, 6.2)	63.1, CH	5.38, brs	63.2, CH	5.41, brs	61.2, CH	5.48, t (3.2, 5.8)
OH-1		12.80, s		12.59, s		12.66, s		12.60, s
OH-14				2.68, brs		3.07, brs		
OH-15				2.68, brs		3.07, brs		2.21, brs
OH-16								1.65, brs
OH-25		4.97, d (3.9)		5.13, d (3.3)		4.98, d (3.8)		4.54, d (3.3)

^a ^1H (400 MHz) and ^{13}C (100 MHz) NMR data in CDCl_3 .

relative configurations of **2** were assigned as 14*R**, 15*S**, 20*S**, 25*R**, respectively, the same as those of **1**, by comparison of the ^1H and ^{13}C NMR spectroscopic data and optical rotation value with those of **1**, and further confirmed by the NOESY experiment (Fig. 3). Thus, compound **2** was determined as 14-hydroxyltjixanthone hydrate.

Compound **3** was obtained as a yellow powder. The molecular formula was determined to be $\text{C}_{25}\text{H}_{27}\text{ClO}_7$ by the HRESIMS at m/z 473.1364 ($[\text{M} - \text{H}]^-$, calcd 473.1373), implying the presence of twelve degrees of unsaturation. Analysis of its ^1H and ^{13}C NMR spectroscopic data (Table 1) suggested that compound **3** also has the tjixanthone type skeleton, and its NMR data were almost identical to those of co-isolated known compound 15-chlorotjixanthone hydrate (**7**),¹² differing only in the chemical shifts at C-14. The carbon chemical shifts at C-14 [δ_{C} 65.3 (d)] was shifted significantly downfield in **3** compared to **7** [δ_{C} 31.9 (t)], indicating that the hydroxyl group was substituted at C-14 in **3**, which was confirmed by HMBC correlations (Fig. 2) and

its molecular formula that has one more oxygen atom than **7**. The relative configurations of all chiral centers in **3** are the same as those of **2**, by comparison of the coupling constants, 1D NMR, NOESY data and the experimental optical rotation data [$[\alpha]_{\text{D}}^{24} -53^\circ$ (c 0.167, CHCl_3)] with those of compound **2** [$[\alpha]_{\text{D}}^{24} -40^\circ$ (c 0.1, CHCl_3)]. Finally, compound **3** was established as 14-hydroxyl-15-chlorotjixanthone hydrate.

Compound **4** was isolated as a yellow needle crystal with the same molecular formula $\text{C}_{25}\text{H}_{28}\text{O}_7$ as that of co-isolated known compound **5**,¹² on the basis of HRESIMS peak at m/z 463.1720 ($[\text{M} + \text{Na}]^+$, calcd 463.1727), suggesting twelve degrees of unsaturation. The ^{13}C NMR spectrum of **4**, which displayed 25 carbon signals analyzing by DEPT and HSQC data, indicated the presence of four methyls, three methylenes, six methines, and twelve quaternary carbons. Comparison of the NMR spectroscopic data of **4** with compound **5** revealed that both have the same planar structures, which further confirmed by HMBC correlations. It is suggested that the relative configurations of **4**



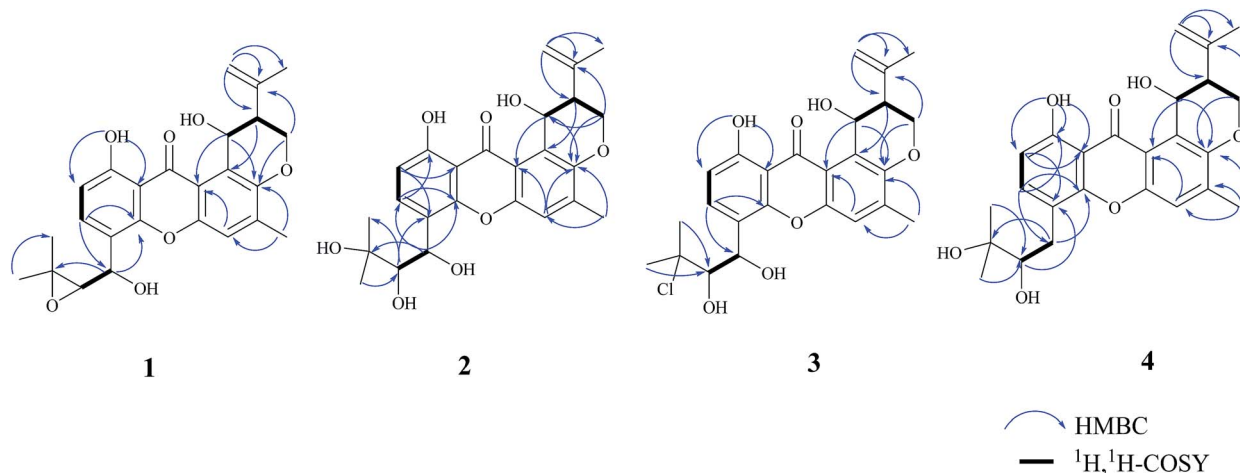


Fig. 2 Key HMBC and ^1H , ^1H -COSY correlations of compounds 1–4.

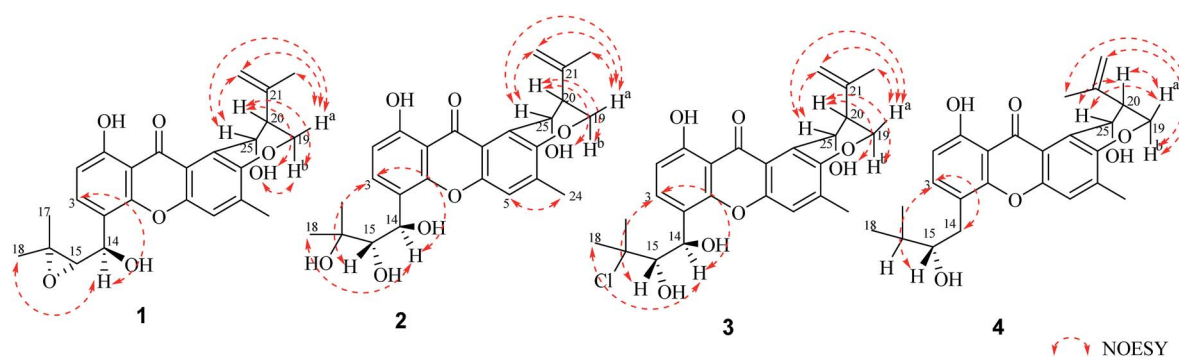


Fig. 3 Key NOESY correlations of compounds 1–4.

is different from those of 5, supported by evidence from their opposite optical rotation data [compound 4, $[\alpha]_{\text{D}}^{25} +61^\circ$ (c 0.1, CHCl_3); compound 5, $[\alpha]_{\text{D}}^{25} -74^\circ$ (c 0.1, CHCl_3)]. The relative configuration of C-15 was assigned as S^* , the same as that of 5, deducing from the similar carbon and proton chemical shifts, and protons coupling constants. Further comparison of the ^{13}C NMR data with those of 5 indicated the chemical shifts at C-8, C-19, C-20, C-22 and C-25 are obviously different from those of 5, suggesting that the relative configurations at C-20 and C-25 are different in these two compounds. Finally, the relative configurations at C-20 and C-25 were determined to be R^* , R^* , by the NOESY correlations of H_b -19 with H_b -22, H-23, H_a -19 with H-20, H-25 with H-20 (Fig. 3), the same as related compound episamixanthone reported from *Aspergillus rugulosus*,¹⁹ and the configurations were further confirmed by a single-crystal X-ray diffraction analysis (Fig. 4). Thus, compound 4 was epitajixanthone hydrate.

Five known compounds (5–9) including three prenylxanthones derivatives (5–7) were also isolated and identified as tajixanthone hydrate (5),¹² 14-methoxytjixanthone-25-acetate (6),¹⁸ and 15-chlorotajixanthone hydrate (7),¹² questin (8),²⁰ and carnemycin B (9),²¹ respectively, based on the comparison of their spectroscopic data and optical rotation data with those reported in the literature. Moreover, the melting

points of 5 (174–175 °C), 6 (180–181 °C), 7 (174–175 °C), 8 (298–300 °C), and 9 (102–112 °C) were almost agreement with those of the known compounds.^{12,14,20,21}

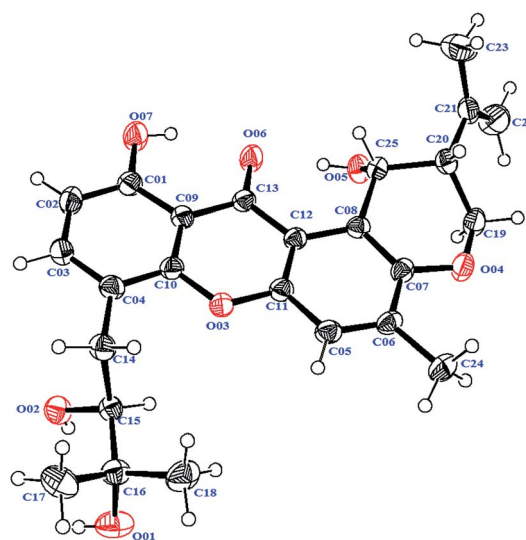
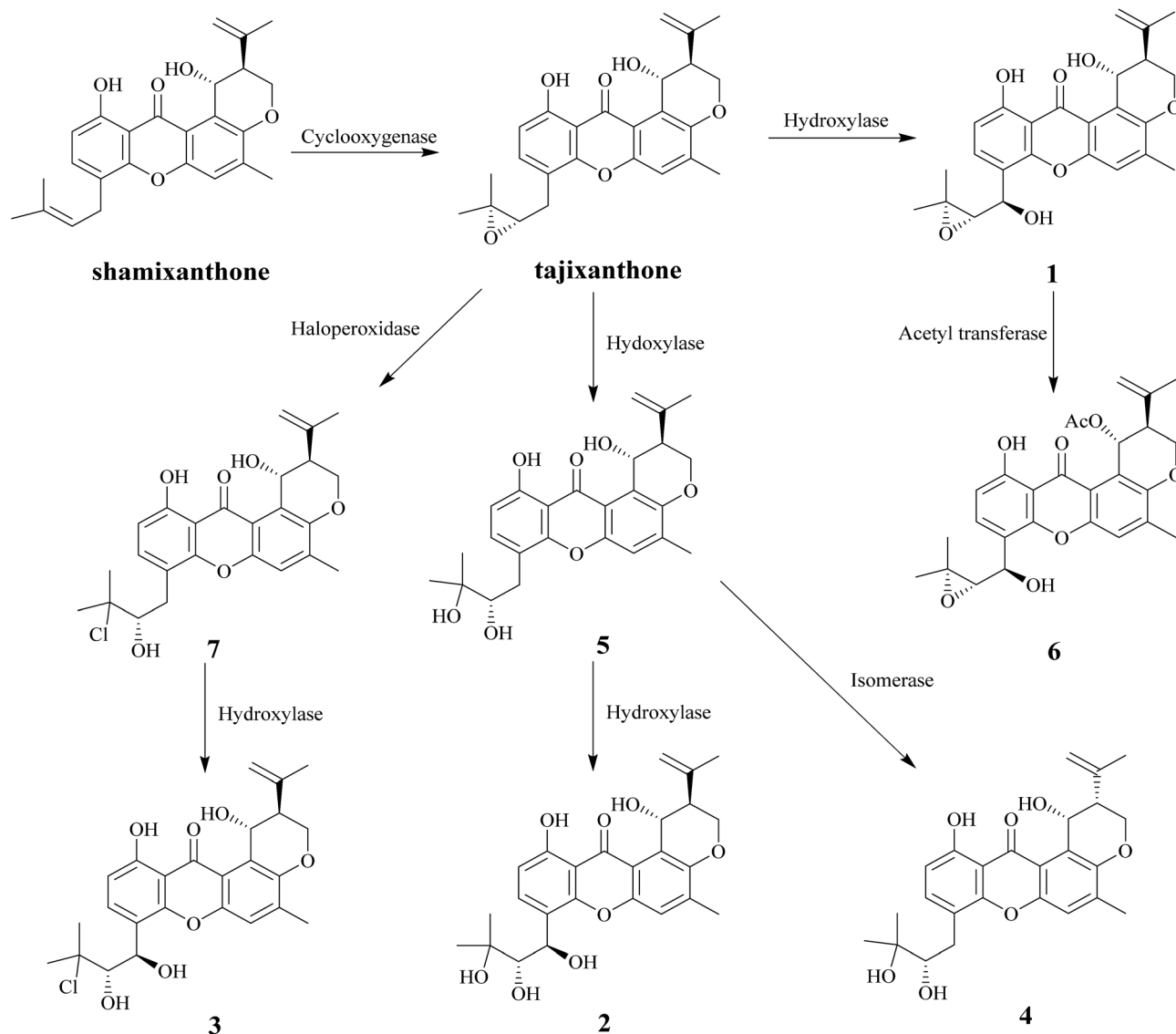


Fig. 4 ORTEP drawing of the compound 4.





Scheme 1 The formation of compounds 1–7 derived from tajixanthone and shamixanthone.

Table 2 Antifungal activity of compounds 1–9 (MIC, $\mu\text{g mL}^{-1}$)^a

Compounds	<i>R. solani</i>	<i>R. cerealis</i>	<i>D. maydis</i>	<i>F. oxysporum</i>	<i>S. sclerotiorum</i>	<i>C. fimbriata</i>	<i>F. oxysporum</i> f. sp. <i>vasinfectum</i>	<i>P. piricola</i>
1	50	25	25	25	50	50	50	25
2	100	50	25	50	100	100	100	50
3	>100	50	100	>100	100	>100	>100	50
4	>100	25	25	>100	100	>100	>100	50
5	100	50	100	100	>100	100	100	25
6	100	50	50	50	100	100	100	50
7	100	50	100	100	>100	>100	>100	50
8	>100	50	25	50	>100	>100	>100	50
9	50	25	25	50	50	100	50	50
Ketoconazole	6.25	3.12	1.56	3.12	3.12	12.50	6.25	3.12

^a MIC – minimum inhibitory concentrations.

Table 3 Antibacterial activity of compounds 1–9 (MIC, $\mu\text{g mL}^{-1}$)^a

Compounds	Gram-positive bacteria					Gram-negative bacteria			
	<i>M. luteus</i>	<i>S. aureus</i>	<i>B. megaterium</i>	<i>B. anthracis</i>	<i>B. paratyphosum B</i>	Drug resistant <i>S. aureus</i>	<i>P. aeruginosa</i>	<i>E. coli</i>	<i>E. aerogenes</i>
1	25	25	12.5	12.5	12.5	50	50	50	25
2	25	25	25	25	25	>100	50	50	50
3	25	25	25	25	25	>100	50	50	50
4	25	25	25	50	25	50	50	50	50
5	50	25	12.5	50	25	>100	50	50	25
6	25	25	12.5	25	25	>100	50	50	50
7	50	25	12.5	50	25	>100	50	50	50
8	25	25	12.5	12.5	25	>100	50	50	50
9	25	25	12.5	25	25	50	50	50	50
Ciprofloxacin	3.12	3.12	1.56	1.56	3.12	6.25	6.25	6.25	3.12

^a MIC – minimum inhibitory concentrations.

In this study, nine compounds including four new ones were isolated from *Emericella* sp. XL029, eight of them (1–8) are pigments. Among these compounds, seven pigments (1–7) have tajixanthone type skeleton, the same as previously reported compounds tajixanthone and shamixanthone which has been confirmed to be derived through ring cleavage of an octaketide-derived anthraquinone with introduction of two dimethylallyl moieties by isotopic labelling experiments.²² It is suggested that the formation of compounds 1–7 were derived from the tajixanthone biosynthetic pathway through various modifications, such as hydroxylation, oxidation, chlorination, acetylation, and isomerization (Scheme 1).

All isolates (1–9) were tested for their anti-agricultural pathogenic fungal activity on eight agricultural pathogenic fungi, namely, *R. solani*, *R. cerealis*, *D. maydis*, *F. oxysporum*, *S. sclerotiorum*, *C. fimbriata*, *F. oxysporum* f. sp. *vasinfectum*, and *P. piricola*. The results are summarized in Table 2, indicating that compounds 1, 2, 4, 8 and 9 displayed significant activity against *D. maydis* with MIC value of 25 $\mu\text{g mL}^{-1}$. Moreover, compound 1 also exhibited significant activity against other three fungi including *R. cerealis*, *F. oxysporum* and *P. piricola*, and compounds 4 and 9 against *R. cerealis* with MIC value of 25 $\mu\text{g mL}^{-1}$, respectively. Compound 5 only demonstrated significant activity against *P. piricola* with MIC value of 25 $\mu\text{g mL}^{-1}$. Compounds 3, 6 and 7 showed moderate or weak activity against selective strains. Besides, all isolates (1–9) were evaluated for their antibacterial activity against Gram-positive (*M. luteus*, *S. aureus*, *B. megaterium*, *B. anthracis*, *B. paratyphosum B* and drug resistant *S. aureus*) and Gram-negative (*P. aeruginosa*, *E. coli* and *E. aerogenes*) bacteria (Table 3). Compounds 1–3, 6, 8 and 9 exhibited significant activity against *M. luteus*, *S. aureus*, *B. megaterium*, *B. anthracis*, and *B. paratyphosum B* with MIC values of 25 and 12.5 $\mu\text{g mL}^{-1}$, respectively. Compound 4 displayed significant activity against four Gram-positive bacteria including *M. luteus*, *S. aureus*, *B. megaterium*, and *B. paratyphosum B* with MIC value of 25 $\mu\text{g mL}^{-1}$, respectively. While compounds 5 and 7 exhibited antibacterial activity against *S. aureus*, *B. megaterium*, and *B. paratyphosum B* with MIC values of 25 and 12.5 $\mu\text{g mL}^{-1}$, respectively. In addition, only compounds

1, 4 and 9 showed moderate activity against drug resistant *S. aureus* with MIC value of 50 $\mu\text{g mL}^{-1}$. All isolated compounds displayed moderate activity against three Gram-negative (*P. aeruginosa*, *E. coli*, and *E. aerogenes*) bacteria with MIC value of 50 $\mu\text{g mL}^{-1}$.

Conclusions

In summary, we have reported the isolation, identification and biological study of nine compounds (1–9) including four new ones (1–4), seven of them belonging to prenylxanthenes from the solid-substrate fermentation culture of *Emericella* sp. XL029. The occurrence of prenylxanthone derivatives from *Emericella* sp. XL029 is in agreement with the previous findings,^{8–14} indicating that the isolation of these compounds might be a useful chemotaxonomic marker of the *Emericella* genus. The results of preliminary anti-agricultural pathogenic fungal and antibacterial assays suggested that these isolated prenylxanthone derivatives showed moderate activity. Moreover, previous findings predicted that the fungal metabolites produced by endophytic fungi may be useful for the growth of the plant host. In other words, those fungal metabolites that may protect the fungal and the host from parasites, as well as other predators such as bacteria, fungi, and animals.^{25–30} Thus, these newly discovered fungal pigments might be presumed to act as phytoalexins occurring in endophytic fungi.

Experimental section

General experimental procedures

Melting points were measured on a SGW X-4A melting point apparatus and reported without corrections. Optical rotations were measured on an Autopol III automatic polarimeter (Rudolph Research Analytical). UV spectra were recorded with an Agilent spectrophotometer (Agilent Cary60). The IR spectra were recorded using a Bruker spectrophotometer (SENSOR-27). HR-ESI-MS spectra were recorded on a Bruker FTMS instrument (Solarix). Nuclear magnetic resonance (NMR) spectra were recorded on an Agilent DD2 (400 MHz) instrument. Silica gel



(200–300 mesh, Qingdao Marine Chemical Factory, Qingdao, China) and Sephadex LH-20 (Pharmacia Biotech, Uppsala, Sweden) were used for column chromatography. Semi-preparative HPLC separation was equipped with a NP7005C pump (flow rate: 2 mL min⁻¹) and an UV detector (NU3000C) using a Dubhe™ C18 column (250 mm × 10 mm, 5 μm, Hanbang, Jiangsu, China). Single-crystal X-ray diffraction data was collected on a SuperNova, Dual, Eos CCD area detector diffractometer (Agilent, USA) with graphite monochromated Mo Kα radiation (λ = 0.71073 Å).

Fungal material

The culture of *Emericella* sp. XL029 was isolated from the leaves of *Panax notoginseng* collected from Shijiazhuang, Hebei Province, People's Republic of China in September 2015. The isolate was identified based on sequence (GenBank Accession No. KY350849) analysis of the ITS region of the rDNA. All tested bacterial and fungal strains for biological studies were obtained as gifts from Associated Professor Fei Cao at College of Pharmaceutical Sciences, Hebei University.

Fermentation, extraction, and isolation

The fungal strain was cultured on potato dextrose agar at 28 °C for 7 days. Then, cut agar plugs into small pieces (about 0.5 × 0.5 × 0.5 cm³) under aseptic conditions, and 5 pieces were transferred into ten Erlenmeyer flasks (250 mL) containing 100 mL PYG media (L⁻¹: peptone 10 g, yeast extract 10 g, glucose 10 g), and incubated on a rotary shaker at 160 rpm and 28 °C for 7 days to prepare the seed culture. The scale-up fermentation was carried out in 100 Erlenmeyer flasks (500 mL) each containing 60 g of rice and 90 mL of distilled water and 20 g L⁻¹ glucose, 20 g L⁻¹ sea salt. Each flask was inoculated with 5.0 mL of the spore inoculum and incubated at 28 °C for 40 days.

The fermented material was extracted with MeOH (3 × 10 L) at room temperature, and the organic solvent was evaporated to dryness under vacuum to afford a brown residue, which was then suspended in water and extracted three times with an equal volume of EtOAc to yield 102 g of crude extract. The crude extract was subjected to CC (column chromatography over SiO₂ with a petroleum ether/acetone gradient system 10 : 0, 9 : 1, 8 : 2, 7 : 3, 6 : 4, and 1 : 1) to yield nine fractions, Fr. A–I. Fraction C (0.8 g) was further purified by Sephadex LH-20 (CHCl₃/MeOH, 1 : 1) and silica gel CC with petroleum ether/EtOAc (15 : 1) to afford compound 7 (12.0 mg). Fraction D (1.0 g) was subjected to repeated chromatography over silica gel (petroleum ether/EtOAc, 12 : 1) to yield four fractions D1–D4. Fraction D4 (269.0 mg) was further purified by Sephadex LH-20 (CHCl₃/MeOH, 1 : 1) and semi-preparative HPLC eluting with MeOH–H₂O (90 : 10) to afford compounds 1 (13.0 mg), 3 (11.0 mg), and 6 (4.0 mg). Fraction G (1.7 g) was fractionated by Sephadex LH-20 (CHCl₃/MeOH, 1 : 1) to give three fractions (G1–G3). Fraction G1 was further purified by semi-preparative HPLC eluting with MeOH–H₂O (90 : 10) to afford compounds 4 (9.0 mg) and 5 (20.0 mg), and fraction G3 was subjected to CC on silica gel (petroleum ether–acetone, 6 : 1) to obtain compound 8 (6.0 mg). Compound 2 (12.0 mg) was obtained from fraction H (1.3 g)

after repeated chromatography over silica gel (petroleum ether/EtOAc, 5 : 1). Compound 9 (12.0 mg) was obtained from fraction I (1.03 g) after repeated chromatography over silica gel (chloroform/methanol, 15 : 1).

14-Hydroxytajibanthone (1). Yellow powder; mp 184–185 °C; [α]_D²⁴ –59 (c 0.183, CHCl₃); UV (MeOH) λ_{max} (log ε) 387 (3.81), 253 (4.64), 230 (4.82), 203 (4.99) nm; IR (KBr) ν_{max} 3446 (OH), 2959, 2923, 1643 (C=O), 1602, 1576, 1475, 1431, 1244, 1125, 1021, 953, 905, 823 cm⁻¹; see Table 1 for ¹H NMR (CDCl₃, 400 MHz) and ¹³C NMR (CDCl₃, 100 MHz); negative HRESIMS [M – H]⁻ m/z 437.1586 (calcd for C₂₅H₂₅O₇, 437.1606).

14-Hydroxytajibanthone hydrate (2). Yellow powder; mp 191–192 °C; [α]_D²⁴ –40 (c 0.1, CHCl₃); UV (MeOH) λ_{max} (log ε) 390 (3.77), 270 (4.43), 240 (4.39), 205 (4.62) nm; IR (KBr) ν_{max} 3423 (OH), 2922, 2853, 1645 (C=O), 1473, 1430, 1383, 1243, 1050, 823, 486 cm⁻¹; see Table 1 for ¹H NMR (CDCl₃, 400 MHz) and ¹³C NMR (CDCl₃, 100 MHz); positive HRESIMS [M + Na]⁺ m/z 479.16835 (calcd for C₂₅H₂₈O₈Na, 479.16764).

14-Hydroxy-15-chlorotajibanthone hydrate (3). Yellow powder; mp 178–179 °C; [α]_D²⁴ –53 (c 0.167, CHCl₃); UV (MeOH) λ_{max} (log ε) 388 (3.65), 273 (4.39), 235 (4.40), 203 (4.78), 201 (4.81) nm; IR (KBr) ν_{max} 3519 (OH), 2922, 2853, 1641 (C=O), 1601, 1473, 1432, 1389, 1240, 1050, 901, 749 cm⁻¹; see Table 1 for ¹H NMR (CDCl₃, 400 MHz) and ¹³C NMR (CDCl₃, 100 MHz); negative HRESIMS [M – H]⁻ m/z 473.13643 (calcd for C₂₅H₂₇ClO₇, 473.13725).

Epitajibanthone hydrate (4). Yellow needle crystals; mp 176 °C; [α]_D²⁴ +62 (c 0.1, CHCl₃); UV (MeOH) λ_{max} (log ε) 390 (3.80), 273 (4.57), 235 (4.54), 203 (4.80), 201 (4.79) nm; IR (KBr) ν_{max} 3352 (OH), 2923, 2854, 1642 (C=O), 1572, 1478, 1349, 1246, 1119, 1079, 1022, 818, 730 cm⁻¹; see Table 1 for ¹H NMR (CDCl₃, 400 MHz) and ¹³C NMR (CDCl₃, 100 MHz); positive HRESIMS [M + Na]⁺ m/z 463.17202 (calcd for C₂₅H₂₈O₇Na, 463.17272).

X-ray crystallography of compound 4

A yellow needle crystal of compound 4 (CCDC: 1525724†) was obtained from an *n*-hexane/acetone (2 : 1) solution. The X-ray crystallographic data of 4: C₂₅H₂₈O₇, monoclinic, space group: *P*2₁, *a* = 6.2804 (3) Å, *b* = 17.2710 (6) Å, *c* = 10.3842 (4) Å, α = 90°, β = 101.759 (4)°, γ = 90°, *V* = 1102.72 (8) Å³, *Z* = 2, *D*_{calcd} = 1.327 g cm⁻³, *w*R1 (*I* > 2σ(*I*)) = 0.0478, *w*R2 = 0.1072. Crystal size, 0.40 × 0.25 × 0.10 mm³. Flack parameter, 0.5(6).

Antifungal assay

Antifungal bioassays were conducted as described previously.¹⁵ The eight agricultural pathogenic fungi, namely, *Rhizoctonia solani*, *Rhizoctonia cerealis*, *Drechslera maydis*, *Fusarium oxysporum*, *Sclerotinia sclerotiorum*, *Ceratocystis fimbriata*, *Fusarium oxysporum* f. sp. *vasinfectum*, and *Phylospora piricola*, were grown on PDA. Targeted microbes (3–4 colonies) were prepared from broth culture (28 °C for 72 h), and the final spore suspensions of fungi (in PDB) were 10⁴ mycelia fragments per mL. Test samples (1 mg mL⁻¹ as stock solution in DMSO and serial dilutions) were transferred to 96-well clear plate in triplicate, and the suspension of the test organisms were added to each well, achieving a final volume of 100 μL (ketoconazole was



used as the positive control). After incubation, the minimum inhibitory concentration (MIC) was defined as the lowest test concentration that completely inhibited the growth of the test organisms.

Antibacterial assay

All isolated compounds were evaluated for their antibacterial activity against Gram-positive (*Micrococcus luteus*, *Staphylococcus aureus*, *Bacillus megaterium*, *Bacillus anthracis*, *Bacterium paratyphosum B* and drug resistant *Staphylococcus aureus*) and Gram-negative (*Pseudomonas aeruginosa*, *Escherichia coli*, and *Enterobacter aerogenes*) bacteria. They were grown in liquid LB medium (yeast extract 5 g L⁻¹, peptone 10 g L⁻¹, NaCl 10 g L⁻¹, PH = 7.4) overnight at 37 °C, and the diluted bacterial suspension (10⁶ CFU per milliliter) was ready for detection. The minimum inhibitory concentrations (MIC) of samples and positive control were determined in sterile 96-well plates by the modified broth dilution test.¹⁶ All wells were filled with 90 μL of bacterial suspension containing 10⁶ CFU per milliliter. Test samples (10 μL) with their different concentrations were added into each well. Medium containing 1% DMSO was used as a negative control, ciprofloxacin was used as the positive control. The final concentrations of ciprofloxacin and test compounds were 100, 50, 25, 12.5, 6.25, 3.12, 1.56, 0.78 μg mL⁻¹ in medium. After incubation, the minimum inhibitory concentration (MIC) was defined as the lowest test concentration that completely inhibited the growth of the test organisms.

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