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Fabrication of a new biosensor based on a Sn doped ceria nanoparticle modified glassy carbon paste electrode for the selective determination of the anticancer drug dacarbazine in pharmaceuticals†

Mohamed Ibrahim,^{ID}* Yassien Temerk and Hossieny Ibrahim

A new tin doped ceria nanoparticle modified glassy carbon paste electrode (Sn–CeO₂NPs/GCPE) was fabricated for the determination of the anticancer drug dacarbazine (DTIC). The Sn–CeO₂ nanoparticles with Sn concentrations from 0 to 10 wt% were synthesized by sol gel method. Sn doped CeO₂ nanoparticles were characterized by X-ray diffraction, energy-dispersed X-ray spectrometry (EDS) and transmission electron microscopy (TEM). Cyclic voltammetry (CV) and square wave voltammetry (SWV) were applied to investigate the unique properties of the Sn–CeO₂NPs/GCPE. The modified electrode showed an excellent character for electrocatalytic oxidation of DTIC, while the bare GCPE electrode only gave a small oxidation peak. The fabricated sensor showed an excellent anti-interference ability against electroactive species and metal ions. This modified electrode was used as a sensor for determination of DTIC in human biological fluids and in pharmaceutical samples with satisfactory results.

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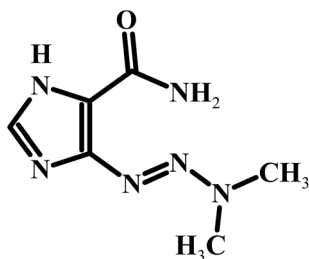
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1. Introduction

The detection and quantification of electrochemically active compounds such as the anticancer drug dacarbazine in biological fluids like blood and urine are important to diagnose and monitor several diseases, besides supplying information about the interactions involving these species in the physiological processes. Dacarbazine [5-(3,3-dimethyl-1-triazenyl)imidazole-4-carboxamide] (DTIC) (Scheme 1) is an important antitumor agent with a proven activity and selectivity against various types of malignant diseases,

such as soft tissue sarcoma and lung carcinoma. It has also been used in combination with other medications for treating Hodgkin's lymphoma.^{1,2} Dacarbazine is a member of the class of alkylating agents, which destroy cancer cells by adding an alkyl group to their DNA. The widespread use of dacarbazine and the need for clinical and pharmaceutical studies require fast and sensitive analytical techniques to determine this drug in a variety of biological fluids and in quality control.^{3,4}

Several time consuming and expensive techniques for detection of DTIC have been studied and reported, which include chromatography^{5–7} and spectrometry.⁸ One of the broadly exercised simple, swift, and exact methods is electrochemical method. There are few reported electrochemical methods, have been applied for the determination of DTIC.^{9–11} Among all the above, electrochemical methods have shown remarkable advantages in the analysis of drugs in pharmaceutical preparations and human body fluids. These advantages are mainly due to the simplicity, low cost and relatively short analysis times of these compounds as compared to the other routine analytical techniques including chromatography.^{12,13} Further handling this technique requires a knowledge regarding the working electrode, where the electrode reactions are centered. Relatively a newly adopted method for the improvement of analyte and surface interaction is the modification of the electrode surface. For the modification of electrode surface, varieties of methods are used which include, nanoparticle incorporation on the surface of the electrode. Unique characters and vast potential application in nanodevice fabrication nanomaterials have engrossed, broad notice.^{14–16}



Scheme 1 Structure of DTIC.

Chemistry Department, Faculty of Science, Assiut University, Assiut 71516, Egypt.
E-mail: msayed217@yahoo.co.uk; msayed217@aun.edu.eg; Hossienyomer@yahoo.com; hossieny.ibrahim@aun.edu.eg; Temerk44@yahoo.com; temerk@aun.edu.eg;
Fax: +20 88 2080209; Tel: +20 88 2080508

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Nanostructured metal oxides such as ZnO, Fe₃O₄, SnO₂, CeO₂, In₂O₃ and others have recently aroused much interest as immobilizing matrices for chemo-biosensor development due to their interesting nanomorphological, functional, biocompatible and catalytic properties.^{17–20} Further chemical doping with appropriate elements (Fe, Cr, Co, Mn, Ni, *etc.*) is widely used as an effective method to tune surface states, energy levels of semiconductors and transport performance of carriers, and enhance the electrical, electrochemical and magnetic properties of materials.^{21–27} Among these, the design of CeO₂ nano-materials with controlled morphologies is of special interest due to their favorable shape-dependent properties.^{28–30} Different strategies are adopted to improve the physico-chemical properties and morphology of the nanostructured CeO₂ in order to achieve high sensitivity and selectivity. Recently, metal-doped ceria nanomaterials have been widely considered to promote the active oxygen content on ceric oxide nanoparticles (CeO₂-NPs) surface by changing the surface element composition, because such materials ultimately reveal strikingly high catalytic activity.³¹ Chemical doping into CeO₂ matrix with appropriate dopants (B or In) at Ce⁴⁺ site is proved to be highly effective for chemo-biosensing applications.^{32,33} The added active elements stabilize the CeO₂ surface, and promote a decrease in grain size which enhances higher catalytic activity and sensor response than that of pure CeO₂. Hence in this work, a novel Sn–CeO₂NPs modified glassy carbon microspheres paste electrode (Sn–CeO₂NPs/GCPE) was prepared and applied for selective and sensitive electrochemical determination of DTIC without physiological interferents.

2. Experimental

2.1. Chemicals

DTIC, cerium(III) nitrate hydrate, tin chloride pentahydrate and polyvinylalcohol polymer (PVA) (MW = 30 000 g mol^{−1}), were purchased from Sigma-Aldrich chemicals (St.Louis, Mo, USA). Glassy carbon spherical microparticles with a diameter of 0.4–12 μm and mineral oil were obtained from Alfa Aesar (Ward Hill, MA). A stock solution of DTIC (1 × 10^{−3} M) was prepared by dissolving an appropriate amount of the drug in ultrapure water and storing the solution in the dark at 4 °C. Electrochemical cells and volumetric flasks were protected from light by means of aluminium foil to avoid DTIC photodecomposition. Phosphate buffer solution (PBS) was prepared from NaH₂PO₄ and Na₂HPO₄ (0.2 M) and adjusted the pH with 0.1 M H₃PO₄ or NaOH solution. The pH values of the buffer solutions were measured with a digital radiometer pH meter, Jenway 3310 accurate to ±0.02 unit. Ultrapure water (Milli-pore Inc., 18.2 MΩ cm) was obtained from a Milli-Q purification system and used in all experiments.

2.2. Instrumentation

Cyclic voltammetry (CV) and square-wave voltammetry (SWV) were performed using an EG&G PAR 384 B (Princeton Applied Research, Oak Ridge, TN, USA) polarographic analyzer controlled by 394 software in conjunction with a PAR Model

303A. The electrode system consisted of the glassy carbon paste electrode (unmodified or modified) as the working electrode, a Ag/AgCl (saturated KCl) reference electrode and a Pt wire auxiliary electrode. A PAR Model 305 stirrer was used for SWV. For anodic stripping experiments an accumulation potential (E_{acc}) was applied for a certain accumulation time (t_a), while the solution was stirred at 400 rpm. At the end of the accumulation period the stirrer was stopped and the solution was allowed to become quiescent for 15 s prior of the voltammetric scan. The surface morphology of the samples is analyzed by transmission electron microscopy, TEM (FEI, TECNAI, G2 spirit twin) using an accelerating voltage of 120 kV. Specimens for TEM are prepared by ultrasonic dispersion of some powder sample in ethanol and putting a droplet of the suspension on a copper microscope grid covered with carbon.

2.3. Preparation of Sn–CeO₂NPs and CeO₂NPs

In the present study, Sn doped CeO₂NPs was synthesized by the sol-gel combustion hybrid method using PVA as a fuel. First, the PVA bulk solution was prepared by dissolving 20 g of PVA powder in 500 mL distilled water at 90 °C and continuous stirring for 3 h before left to cool at room temperature. Subsequently, 5 g cerium(III) nitrate hydrate and tin chloride (5 and 10 wt per %) were added into 100 mL PVA solution and stirred continuously for 6 h. The obtained sol was slowly heated to evaporate the solvent and it forms a hard homogeneous gel. The pyrolysis of the final gel was performed at a temperature of 800 °C for 4 hours. During the pyrolysis process the PVA polymeric network through the outer surface, cerium nitrate salt and tin chloride simultaneously calcinated and converted into Sn doped CeO₂NPs (Sn–CeO₂NPs). The obtained sample was crushed to prepare a fine powder. The CeO₂NPs were prepared using the same procedure without adding tin chloride into the synthesis process. Characterizations of the obtained nanoparticles are done by XRD, EDS and TEM. XRD patterns of the undoped CeO₂NPs and Sn doped CeO₂NPs were recorded using a Bruker AXS D8 advanced diffractometer with Cu Kα radiation ($\lambda = 1.5406 \text{ \AA}$) in the range of 20–80°. The average crystallite sizes were calculated using Debye Scherrer formula:

$$D = K\lambda/\beta \cos \theta$$

where K is the shape factor whose value is taken as 0.89, λ is the wavelength of Cu, Kα radiation, and β is the corrected full width at half maximum (FWHM) of the diffraction peak and θ is the diffracting angle.

2.4. Electrode preparation and modification

The unmodified glassy carbon paste electrode (GCPE) was prepared with the composition of 75 : 25 (glassy carbon powder : paraffin oil) using a mortar and pestle and allowed to undergo self-homogenization. The paste was packed firmly into the cavity of a Teflon tube (3 mm diameter) to a depth of 6 mm, and an electrical contact was achieved *via* a copper wire. The surface of the resulting paste electrode was smoothed and rinsed carefully with double distilled water. The undoped



CeO₂NPs/GCPE was prepared by mixing the optimal amount of glassy carbon powder, paraffin oil and CeO₂NPs (75 : 15 : 10 w/w). The modified Sn-CeO₂NPs/GCPE was fabricated by mixing of (75 : 15 : 10 w/w) ratio of glassy carbon/paraffin oil/Sn-CeO₂NPs. Whenever necessary a new surface was obtained by pushing out an excess of paste and polish it on paper. Therefore the main advantages of the newly fabricated electrode are that the technique for its preparation is simple and require only short periods of time. In addition, the prepared modified electrode is renewable and low in cost compared to the literature methods. Nevertheless the materials for fabrication of the modified electrode are cheap and commercially available.

2.5. Preparation and analysis of real samples

Human serum and urine samples were obtained from healthy volunteers at the Hospital of Assiut University and stored in refrigerator immediately after collection before used. Ten milliliters of the urine sample was centrifuged for 30 min at 3000 rpm. The supernatant was filtered out using 0.45 μ m Millipore filter and then diluted 50 times with PBS of pH 4. The solution was transferred into the voltammetric cell to be analyzed without any further pretreatment. One milliliter of the serum sample was treated with 5 mL of methanol and centrifuged at 4000 rpm for 20 min to remove the precipitating materials. Afterwards the separated serum was attenuated 10-fold with pH 4.0 phosphate buffer solutions. Then, 2.0 mL of this test solution was transferred into the electrochemical cell to determine DTIC by SWV method.

Dacarbazine vial labelled to contain 200 mg DTIC per vial was used as samples. The composition of the investigated DTIC

vial was dacarbazine (42%), mannitol (16%) and citric acid monohydrate (42%). A weight portion of the powder content equivalent to 1×10^{-3} M of DTIC was transferred into a 25 mL calibrated flask and filled to the volume with ultrapure water and stored in dark at 4 °C. SWV measurements were performed using the standard addition method.

3. Results and discussion

3.1. XRD, EDS and TEM analysis of CeO₂NPs and Sn-doped CeO₂NPs

The structural properties of the undoped CeO₂ and Sn (5 wt%) doped CeO₂ nanoparticles were investigated by powder XRD method (Fig. 1A). It is clear that the diffraction peaks of all the samples correspond to the (111), (200), (220) and (311) planes that can be indexed to the cubic fluorite structure of CeO₂ crystals. No characteristic peaks related to Sn and impurity peaks appear in XRD patterns in comparison with the standard XRD patterns of bulk CeO₂ (JCPDS card No. 04-016-6759) which show that all Sn atoms have entered into CeO₂ lattice. However, upon increasing the tin content, a moderate decrease in the intensity of diffraction peaks, accompanied by a pronounced broadening, has been observed. This can be attributed to the decrease of particle size, doping of CeO₂ with Sn through the formation of a Sn-Ce solid solution and/or to induced micro-strain.³⁴ This is consistent with the fact that the ionic radius of Sn⁴⁺ (0.74 Å) is smaller than that of Ce⁴⁺ (0.97 Å). The XRD results showed that the Sn⁴⁺ ions incorporate into CeO₂ lattice or replace Ce⁴⁺ sites without changing the cubic fluorite structure. The average crystallite sizes evaluated by the Scherrer equation were found to be 30 and 16.56 nm for undoped CeO₂

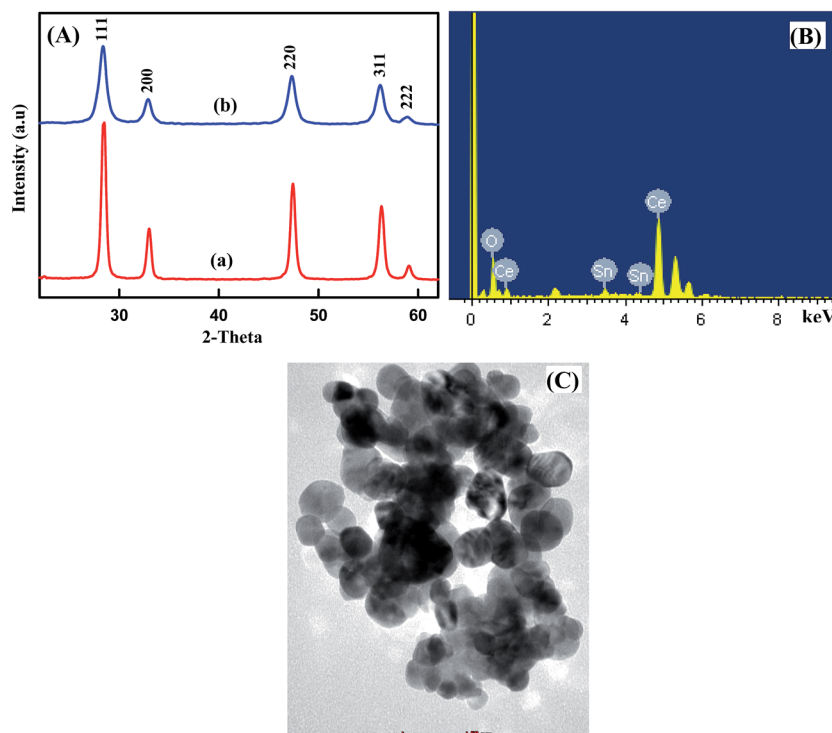


Fig. 1 (A) XRD patterns of (a) undoped CeO₂NPs and (b) 5 wt% Sn doped CeO₂NPs, EDX spectra (B), TEM image (C) of 5% Sn doped CeO₂NPs.



and 5 wt% Sn doped CeO_2 nanoparticles, respectively, suggesting that the grain growth is suppressed due to doping of Sn into Ce-site.

To assess the elemental composition of Sn- CeO_2 , the EDS analysis was done and the result is shown in Fig. 1B. In EDS spectrum, numerous well-defined peaks were evident related to Sn, O and Ce which clearly support that the existence of Sn ions in doped sample and confirmed the successful doping of Sn in CeO_2 . The obtained results are in good agreement with XRD.

The TEM image of 5% Sn doped CeO_2 was shown in Fig. 1C. The morphology of the Sn doped CeO_2 nanoparticles was uniform with well-distributed elliptical/spherical particles with average particle size of about 17.23 nm, which is consistent with the results obtained from XRD. Most of the nanoparticles were well separated, although some of them partially aggregated. Therefore, the particle size of the samples obtained from TEM pattern was quite similar to those calculated from Scherrer's equation.

3.2. Morphological characterization of 5% Sn- CeO_2 NPs/GCPE

Fig. 2 compares the morphological features of GCPE and 5% Sn- CeO_2 NPs/GCPE using SEM. Significant differences in the surface structure of two electrodes were observed. The SEM profile of GCPE (Fig. 2A) was characterized by a surface of non-porous spherically shaped glassy carbon powder. Despite the presence of the mineral oil, the glassy carbon microspheres were packed in close proximity, and no defined binder regions

were visible. Apparently, the oil fills the voids between the GC spheres, while holding them together. The glassy carbon microspheres clearly decorated with Sn- CeO_2 NPs. Besides the close packing arrangement of glassy carbon microspheres, a mass of Sn- CeO_2 NPs was filled in the gaps among the microspheres (Fig. 2B).

3.3. Electrochemical activities of modified electrodes

Electrochemical properties of modified electrodes were investigated using $[\text{Fe}(\text{CN})_6]^{3-/4-}$ as redox probes by cyclic voltammetry (Fig. 3). The cyclic voltammograms (CVs) were recorded in the presence of 5 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$ in 0.1 M KCl at the scan rate 100 mV s^{-1} for (curve 1) bare GCPE, (curve 2) CeO_2 NPs/GCPE and (curves 3 and 4) Sn- CeO_2 NPs/GCPE modified electrodes. At bare GCPE (curve 1), a pair of redox peaks were observed with a high capacitive background current, large peak to peak potential separation ($\Delta E_p = 347 \text{ mV}$), and broadened wave shape. The ΔE_p values for the undoped CeO_2 NPs (curve 2), 5 wt% (curve 3) and 10 wt% (curve 4) Sn doped CeO_2 NPs modified electrodes were found to be 206, 116 and 116 mV, respectively (Table 1). The undoped CeO_2 NPs modified GCPEs showed larger ΔE_p value compared to that of the 5 wt% and 10 wt% Sn doped CeO_2 NPs modified GCPEs. The large peak separation was assumed to result from the lower electrical conductivity.

Furthermore, the peak current intensity at 5 wt% Sn- CeO_2 -NPs/GCPE was enhanced and increased 2 times higher than that at bare GCPE indicating its improved electrochemical performance due to the Sn doping. The observed enhancement in the charge transport characteristic of 5 wt% Sn- CeO_2 NPs is in agreement with the proposal that the doping induced oxygen vacancy plays an important role in the charge transport behavior of CeO_2 NPs. The obtained results indicated that the 5 wt% Sn doping into CeO_2 is the saturation point, and it is facilitating the charge transport between the modified electrode surface and redox couple. The observed decrement in ΔE_p and

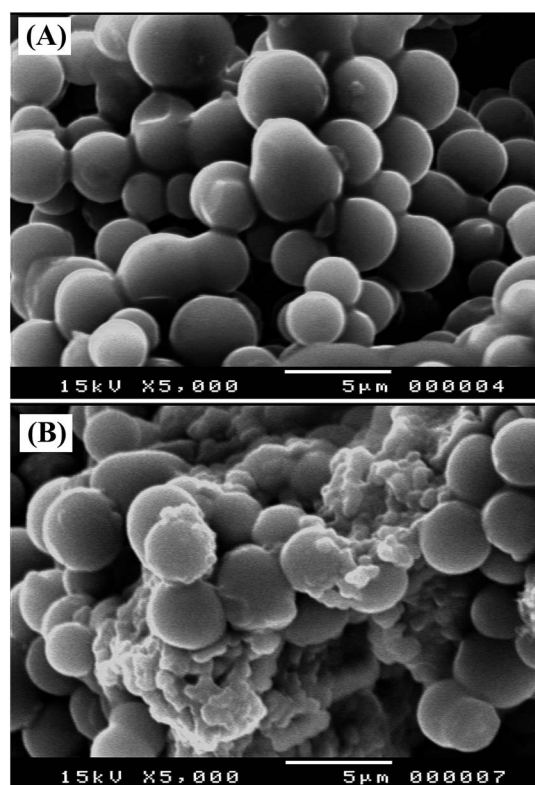


Fig. 2 SEM images of (A) GCPE and (B) 5% Sn doped CeO_2 NPs/GCPE.

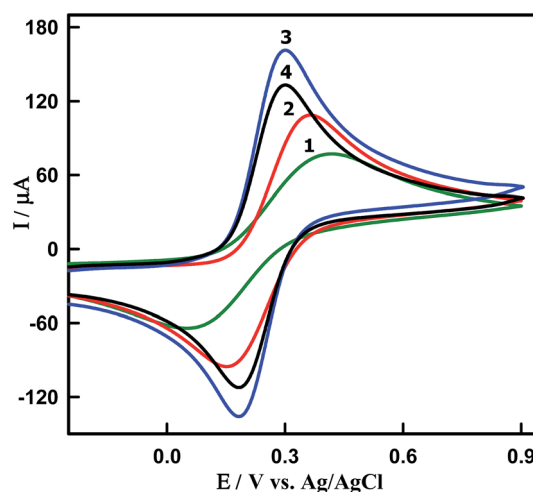


Fig. 3 CVs of 5 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$ in 0.1 M KCl obtained at a scan rate of 0.10 V s^{-1} . Curve 1: bare GCPE, curve 2: undoped CeO_2 NPs/GCPE, curve 3: 5% and curve 4: 10% Sn- CeO_2 NPs/GCPE.



Table 1 Electrochemical data of 5 mM [Fe(CN)₆]^{3-/4-} in 0.1 M KCl at different working electrodes

Electrodes	ΔE_p (mV)	I_{pa} (μ A)	A^a (cm ²)
GCPE	347	90	0.058
CeO ₂ NPs/GCPE	206	120	0.099
Sn–CeO ₂ NPs/GCPE (5 wt%)	116	175	0.225
Sn–CeO ₂ NPs/GCPE (10 wt%)	116	145	0.150

^a A , active surface area (cm²).

the corresponding improvement in redox peak current values of Sn–CeO₂NPs could be attributed to faster electron transfer kinetics and larger electroactive surface area of Sn–CeO₂NPs. In order to confirm this, the active surface area of each electrode could be calculated based on the Randles–Sevcik equation:³⁵

$$I_p = 2.69 \times 10^5 n^{3/2} A D^{1/2} \nu C$$

where n is the value of electron transfer ($n = 1$), A is the active electrode area, D is the diffusion coefficient (7.6×10^{-6} cm s⁻¹), C is the concentration of [Fe(CN)₆]^{3-/4-} (5 mM) and ν is the scan rate. The CV profiles of the Sn–CeO₂NPs/GCPE in 5 mM [Fe(CN)₆]^{3-/4-} with different scan rates from 50–500 mV s⁻¹ are shown in Fig. S1†. The observed linear relation of the oxidation peak currents as a function of the square root of the scan rate reveals a diffusion-controlled process (insert of Fig. S1†). From the slope of $I_p - \nu^{1/2}$, the values of A are calculated to be 0.225 cm² and 0.058 cm² for (5 wt%) Sn–CeO₂NPs/GCPE and bare GCPE, respectively (Table 1). The difference in active surface area between the bare GCPE and the Sn–CeO₂NPs/GCPE clearly indicates that the modified electrode possesses a larger reaction area. Hence we have used 5 wt% Sn doped CeO₂ nanoparticles for further investigation and it is represented as Sn–CeO₂NPs.

3.4. Electrochemical oxidations of DTIC at the Sn–CeO₂NPs/GCPE

Fig. 4A shows the CVs obtained for 8.50 μ M DTIC at bare GCPE (curve 1), undoped CeO₂NPs/GCPE (curve 2) and Sn–CeO₂NPs/GCPE (curve 3) in PBS (pH 4.0). According to cyclic voltammograms in Fig. 4A, no cathodic peak was observed on the reverse scan within the investigated potential range (0.0–1.2 V) because its oxidation is an electrochemically irreversible process. In this context DTIC underwent a two-electron irreversible oxidation process in aqueous media.¹¹ Compared to those obtained at the bare GCPE (2.33 μ A), undoped CeO₂NPs/GCPE (7.01 μ A), a fairly larger peak current was obtained at the Sn–CeO₂NPs/GCPE (24.12 μ A). This result suggested that the electron transfer reaction at the Sn–CeO₂NPs/GCPE is more facile than that at the bare GCPE and undoped CeO₂NPs/GCPE. The electrocatalytic activity of the modified electrode could be ascribed to the presence of nano-Sn–CeO₂ which enhanced the conductivity, surface area and facilitated the electron transfer between the biomolecules and the electrode surface.

Furthermore, the SWV was performed to obtain better sensitivity because enhanced analytical signals can be achieved

by eliminating the non-faradic currents that occur with CV. As can be seen DTIC showed a sharp peak at 932 mV in PBS (pH 4.0) at the Sn–CeO₂NPs/GCPE, which corresponds to the oxidation of DTIC (Fig. 4B). The peak current was 45 μ A, much larger than that obtained by CV. There is also an enhanced peak current of DTIC at the Sn–CeO₂NPs/GCPE (Fig. 4B, curve 3) as compared to a bare GCPE (Fig. 4B, curve 1) and undoped CeO₂NPs/GCPE (Fig. 4B, curve 2). The results indicated that the nano-Sn–CeO₂ incorporated GCPE had great enhancement peak current of DTIC, which was due to the excellent characteristics of nano-Sn–CeO₂ such as high chemical stability, high surface area, good electrical conductivity and high optical transparency. So the electron transfer of DTIC on the Sn–CeO₂NPs/GCPE was greatly enhanced without using electron transfer mediator. It is clear that the increase in oxidation current at the proposed modified electrode can offer a special approach for sensitive electrochemical determination of DTIC in biological fluids.

3.5. Effect of potential scan rate

The effect of the scan rate on the oxidation peak current and potential was studied in order to estimate the nature of the electrode process occurring at the Sn–CeO₂NPs/GCPE surface. Fig. 5 presents the cyclic voltammograms of DTIC recorded in PBS of pH 4.0 with scan rate varying from 50 to 500 mV s⁻¹. The oxidation peak heights were found to increase linearly with scan rate from 50 to 500 mV s⁻¹. The I_p is linearly increased with the square root of scan rate, $\nu^{1/2}$, in the range of 50–500 mV s⁻¹ (inset of Fig. 5). This indicates that the oxidation process of DTIC in PB solution (pH 4.0) on the nano-Sn–CeO₂ is a diffusion-controlled process. The effect of the potential scan rate on the peak current may be expressed as $\log(I_{pa})$ vs. $\log(\nu)$, which has a slope of 1.0 for the species confined to the electrode surface, or 0.5 for the diffusion controlled system. The dependence of $\log(I_{pa})$ vs. $\log(\nu)$ can be expressed by the equation: $\log I_p$ (μ A) = 0.65 $\log \nu$ (mV s⁻¹) + 0.11 ($R^2 = 0.9977$) (Fig. S2†). The value of the slope confirms that the process is diffusion-controlled.

Moreover, the peak potential shifted to more positive values on increasing the scan rate, which confirms the irreversibility of the oxidation process, and a linear relationship between E_p and $\log \nu$ in the range from 50 to 500 mV s⁻¹ (Fig. S3†) which can be expressed as: E_p (V) = 0.04 $\log \nu$ (V s⁻¹) + 0.97 ($R^2 = 0.993$).

According to the Laviron theory for an adsorption controlled and totally irreversible electrode process,³⁶ E_p is defined by the following equation.

$$E_p = E^\circ - (2.3RT/\alpha nF)[\log(RT k_s/\alpha nF) - \log \nu] \quad (1)$$

where α is the transfer coefficient, k_s is the heterogeneous electron transfer rate constant, n is the number of electrons transferred, ν is the scan rate, and E° is the formal redox potential. Other symbols have their usual meanings. Thus value of αn can be easily calculated from the slope of E_p vs. $\log \nu$. In this system, the slope was found to be 0.024 taking $T = 298$ K, $R = 8.314$ J K⁻¹ mol⁻¹, and $F = 96480$ C mol⁻¹, the αn was calculated to be 1.069. The value of α can be calculated as:



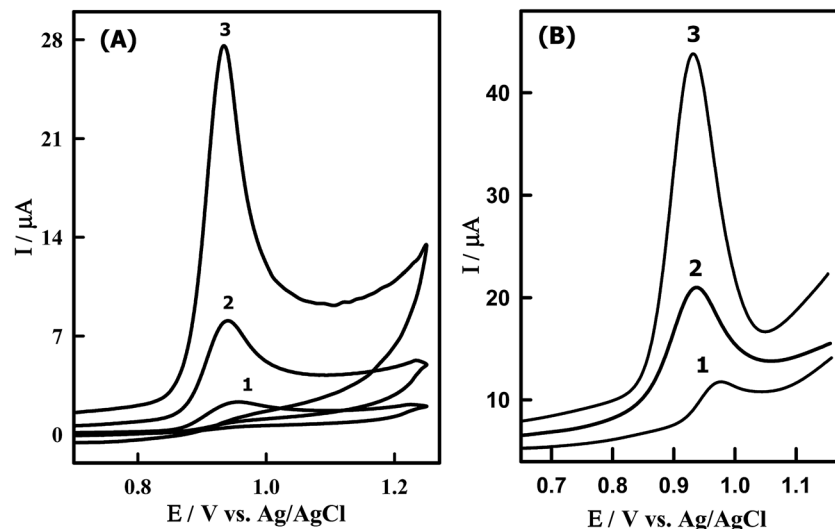


Fig. 4 (A) CVs of 8.50 μM DTIC in PBS of pH 4.0 obtained at (1) bare GCPE, (2) $\text{CeO}_2\text{NPs}/\text{GCPE}$ and (3) $\text{Sn-CeO}_2\text{NPs}/\text{GCPE}$; scan rate, 0.10 V s^{-1} . (B) SW voltammograms of 1.14 μM DTIC in PBS buffer of pH 4.0 obtained at (1) unmodified (2) modified $\text{CeO}_2\text{NPs}/\text{GCPE}$ and (3) $\text{Sn-CeO}_2\text{NPs}/\text{GCPE}$; accumulation potential, 0.0 V; accumulation time, 10 s; scan increment, 6 mV; frequency, 120 Hz and pulse height, 35 mV_{pp}.

$$E_P - E_{P/2} = (47.7/\alpha)\text{ mV} \quad (2)$$

where $E_{P/2}$ is the potential when the current is at half the peak value. From this, we obtained the value of α to be 0.54 for $\text{Sn-CeO}_2\text{NPs}/\text{GCPE}$. So, the number of electrons (n) transferred in the electrooxidation of DTIC was calculated to be $1.979 \approx 2$ that value is in good agreement with the value reported in the literature.¹¹ The value of k_s can be determined from the intercept of the previous plot if the value of αn and E° are known. The value of E° can be obtained from the intercept of E_P vs. ν curve by extrapolating to the vertical axis at $\nu = 0\text{ V s}^{-1}$. Thus, using this information and eqn (1) the k_s values obtained were 0.782 and 2.356 s^{-1} for bare and modified electrodes, respectively. These results may be attributed to the good electrical conductivity as

well as the increase of the electrode surface area in the presence of Sn doped CeO_2 nanoparticles and suggested that electron transfer process between the DTIC molecules and $\text{Sn-CeO}_2\text{NPs}/\text{GCPE}$ was very fast.

The surface concentration (Γ , mol cm^{-2}) of electroactive DTIC on the modified electrode $\text{Sn-CeO}_2\text{NPs}/\text{GCPE}$ and bar GCPE can be calculated from the slope of the plot of I_{pa} versus scan rate by:³⁷ $I_p = n^2 F^2 \nu A \Gamma / 4RT$ where n is the number of electrons transferred, F (C mol^{-1}) is the Faraday's constant, A (cm^2) is the area of the electrode, Γ is the surface concentration of the electroactive substance, DTIC, and ν (V s^{-1}) is the scan rate. The surface concentration (Γ) of DTIC on the $\text{Sn-CeO}_2\text{NPs}/\text{GCPE}$ was estimated to be $6.25 \times 10^{-10}\text{ mol cm}^{-2}$, which was larger than $6.04 \times 10^{-11}\text{ mol cm}^{-2}$ on GCPE. These values imply that the presence of Sn-CeO_2 increases the surface area of the electrode, which in turn increases the Γ of DTIC.

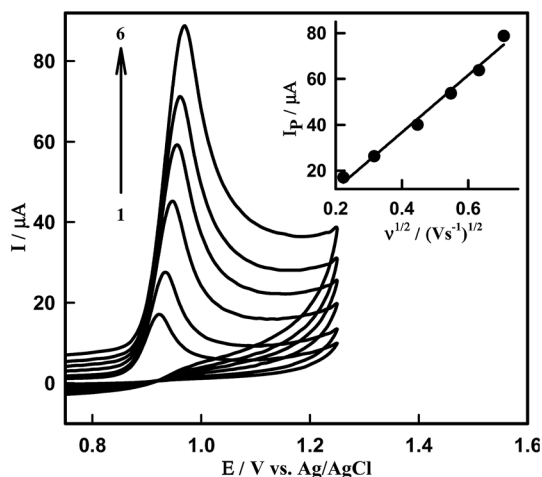


Fig. 5 CVs of 8.50 μM DTIC on surface of $\text{Sn-CeO}_2\text{NPs}/\text{GCPE}$ at various scan rates; (1) 50, (2) 100, (3) 200, (4) 300, (5) 400 and (6) 500 mV s^{-1} in PBS of pH 4.0. Inset: dependence of peak current on square root of scan rate.

3.6. Optimization of the amount of modifier (5% $\text{Sn-CeO}_2\text{NPs}$)

The effect of the amount of 5% $\text{Sn-CeO}_2\text{NPs}$ within the glassy carbon paste electrode on the voltammetric response was evaluated by changing mass ratio of $\text{Sn-CeO}_2\text{NPs}$ to glassy carbon microspheres in the glassy carbon paste (Fig. S4†). The oxidation peak current of DTIC increases with the increasing of amount of modifier because the concentration of $\text{Sn-CeO}_2\text{NPs}$ on the surface of the modified electrode increases correspondingly (Fig. 6). However, when the amount of nano- Sn-CeO_2 reaches 10% (w/w) (nano- $\text{Sn-CeO}_2/\text{glassy carbon paste}$), the peak current reaches its maximum. After this point, any continuing increase in the amount of modifier causes a decrease in the peak current, due to the decrease of the conductivity of the modified electrode. Thus electrode composition of 10% (w/w) nano- Sn-CeO_2 , 75% (w/w) glassy carbon microspheres and 15% (w/w) paraffin oil was used for further experiments.



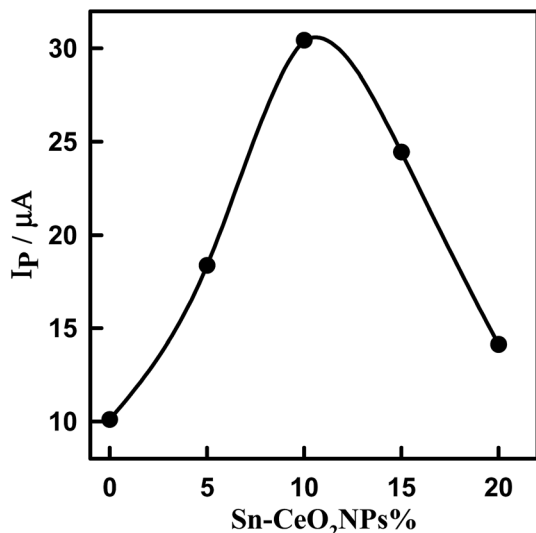


Fig. 6 Plot of peak current of 1.14 μM DTIC in PBS, pH 4.0 using SWV versus the percentage of 5% Sn-CeO₂NPs in the paste.

3.7. Optimization of pH

The effect of pH on the SWV responses of DTIC on Sn-CeO₂/GCPE was investigated in the pH range from 3 to 8.0 (Fig. 7). It can be clearly observed that the peak current and potential towards the oxidation of DTIC are closely related to the pH value of the supporting electrolyte. It was found that the oxidation peak potential of DTIC shifted toward less positive potential values with increase in the pH (Fig. S5†). This suggests the involvement of protons in the oxidation process. Based on the results obtained, over the studied pH range, a linear relationship was obtained between E_p

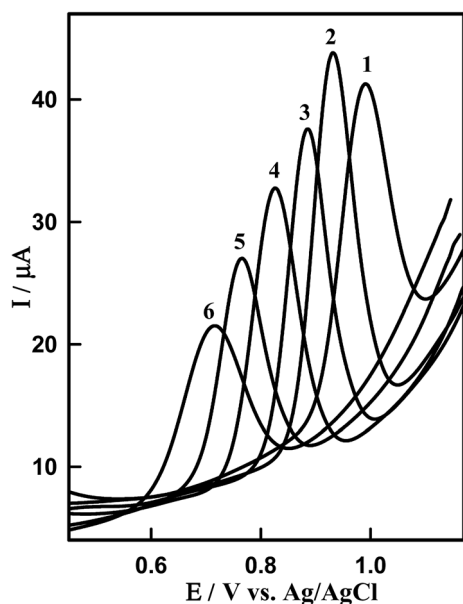


Fig. 7 SW voltammograms of 1.14 μM DTIC on the surface of Sn-CeO₂NPs/GCPE at different pH values (PBS): (1) pH 3.0, (2) pH 4.0, (3) pH 5.0, (4) 6.0, (5) 7.0 and (6) 8.0. Other conditions are the same in Fig. 4B.

and pH (Fig. S5†), which can be expressed by the following equation: $E_p (\text{V}) = 1015 - 0.057 \text{ pH}$ ($R^2 = 0.999$). A comparison between the obtained slope of -0.057 V pH^{-1} and the theoretical slope of $-0.059m/n \text{ V pH}^{-1}$ (m and n being the numbers of protons and electrons, respectively) confirms that equal numbers of protons and electrons participate in the oxidation procedure. According to the overall electrode reaction,¹¹ two electrons and two protons were involved in the DCIT oxidation. The height of the peak reached a maximum and the shape of the curve is better in PBS at pH 4.0 ($\text{pK}_a = 4.42$)³⁸ than other buffers. This supporting electrolyte was chosen with respect to sharp response and better peak shape for the construction of calibration curve and for determination of DTIC in standard solution and in biological samples.

3.8. Square-wave pulse parameters

The optimum conditions for the SWV were recognized by measuring the current dependence on the instrumental parameters counting pulse height, frequency and scan increment. SW voltammograms of 1.14 μM DTIC in PBS of pH 4.0 onto the Sn-CeO₂NPs/GCPE were recorded at various pulse parameters (frequency; $f = 20$ –120 Hz, scan increment; $\Delta E_s = 2$ –10 mV and pulse height; $\Delta E_a = 20$ –80 mV). The better developed voltammetric peak was obtained under the following pulse parameters: $f = 120 \text{ Hz}$, $\Delta E_s = 6 \text{ mV}$ and $\Delta E_a = 35 \text{ mV}_{pp}$ using Sn-CeO₂NPs/GCPE. These parameters reflect SWVs of high sensitivity and best peak morphology.

3.9. Square wave voltammetric determination of DTIC concentration

Under optimum conditions, square wave voltammograms are recorded at the Sn-CeO₂NPs/GCPE to determine the limit detection (LOD) of DTIC. Fig. 8A shows the typical SWV of different concentrations of DTIC at the modified electrode. It was found that the anodic peak current increased linearly with increasing the concentration of DTIC (Fig. 8B). A linear dynamic range of 6.40×10^{-8} to $6.69 \times 10^{-6} \text{ M}$ DTIC was obtained following its concentration onto the modified electrode by adsorption accumulation for 10 s at 0.0 V. The analytical results are summarized in Table 2. In this context the sensitivity of the proposed sensor was found to be of $361.43 \mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$. The LOD was calculated using the relation $k\text{SDa}/b$, where $k = 3$, SDa is the standard deviation of the intercept and b is the slope of the calibration plot. The LOD value was found to be $3.79 \times 10^{-9} \text{ M}$ DTIC (0.690 ng mL^{-1}), which confirmed good sensitivity of the proposed sensor. This concentration detection limit of DTIC ($3.79 \times 10^{-9} \text{ M}$) is very low in compared with the values reported by other reported methods.^{6,11} This wide linear range and low detection limit can be attributed to the effect of the Sn-CeO₂NPs, which provides a large specific area to increase the loading amount of DTIC. Meanwhile, the electron transfer to the electrode surface can be accelerated and the electrochemical signal is amplified due to the outstanding electric conductivity of the Sn-CeO₂NPs/GCPE. Thus, the proposed method could be effectively used for the determination of DTIC in commercial samples.



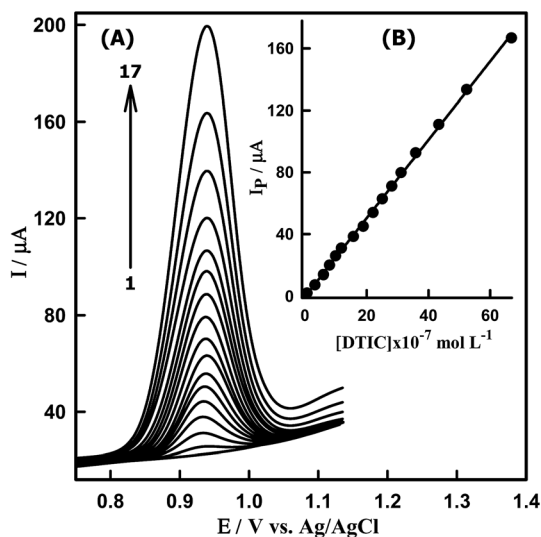


Fig. 8 (A) SW voltammograms of DTIC at Sn–CeO₂NPs/GCPE in PBS at pH 4.0. [DTIC]: (1) blank, (2) 0.064, (3) 0.32, (4) 0.60, (5) 0.79, (6) 0.99, (7) 1.18, (8) 1.57, (9) 1.88, (10) 2.21, (11) 2.51, (12) 2.81, (13) 3.12, (14) 3.59, (15) 4.33, (16) 5.24 and (17) 6.69 μM . (B) Calibration plot of I_p (μA) versus [DTIC] in PBS of pH 4.0.

3.10. Interference studies

In order to evaluate the selectivity of the proposed method for the determination of DTIC, the influence of potentially interfering substances on the determination of this compound was investigated using SWV at the Sn–CeO₂NPs/GCPE. The tolerance limit for interfering species was considered as the maximum concentration of foreign species that caused a relative error less than $\pm 5\%$ for determination of 1.18 μM DTIC under the optimized conditions. For 1.18 μM of DTIC, the result showed that over 200 fold excess of ascorbic acid, uric acid, alanine, cysteine, glucose, citric acid, uracil and serine did not interfere with DTIC response (Table S1†). The results obtained show recoveries in the range from 97.88 to 102.17 for 1.18 μM of DTIC solution, indicating that there were no important matrix interferences for the samples by the proposed SWV method using Sn–CeO₂NPs/GCPE. From these results, it may be

Table 3 Precision (intra and inter day) and accuracy for assay of DTIC ($n = 5$)

Added (μM)	Found (μM)	Precision RSD%	Relative error%	Recovery%
Intra-day				
0.60	0.61 ± 0.011	1.80	−1.66	101.66
1.18	1.16 ± 0.013	1.12	1.69	98.31
5.24	5.22 ± 0.027	0.52	0.38	99.62
Inter-day				
0.60	0.58 ± 0.016	2.75	3.33	96.67
1.18	1.20 ± 0.022	1.83	−1.69	101.69
5.24	5.17 ± 0.096	1.86	1.33	98.66

concluded that the method is free from interference by most coexisting substances and shows promising properties for use in real samples.

3.11. Stability and reproducibility of the electrode

The Sn–CeO₂NPs/GCPE was stable, even after 45 days if it preserved in a closed container. Daily recording the SWV of 3.2×10^{-7} M of DTIC checked the stability of the proposed modified electrode; the electrode retained 98.55% of its initial peak current response with a standard deviation of 1.32%. This shows the long-term stability of the Sn–CeO₂NPs/GCPE.

In order to evaluate the accuracy and precision of the proposed method the analysis of DTIC at three levels of low, moderate and high concentrations of the calibration curve was carried out in five independent series at the same day (intra-day) and performed at same levels on five different days (inter-day). The results obtained for intra-day and inter-day precision and accuracy are presented in Table 3. As can be seen, the RSD values of the measurements were not greater than 1.80% and 2.75% for intra-day and inter-day determinations, respectively, and the accuracy of determination was not different from added values by more than 3.33%. Also the recovery of DTIC was found to be between 96.67% and 101.69%. These results confirm the good precision and accuracy of the proposed method. This indicates that the Sn–CeO₂NPs/GCPE has a good repeatability.

Table 2 Regression data of the calibration lines for quantitative determination of DTIC in standard solution, serum sample and urine sample in PBS (pH 4.0) at Sn–CeO₂NPs/GCPE using SWV

Parameters	Standard solution	Serum sample	Urine sample
Measured potential (V)	0.934	0.933	0.935
Linearity range (μM)	0.064–6.69	0.076–6.59	0.069–5.31
Slope ($\mu\text{A M}^{-1}$)	2.53×10^7	1.39×10^7	1.62×10^7
SE of slope	7.53×10^4	9.27×10^4	7.14×10^4
Intercept (μA)	−0.31	0.99	1.34
SE of intercept	5.10×10^{-3}	9.25×10^{-3}	7.48×10^{-3}
Determination coefficient (R^2)	0.9991	0.9989	0.9983
Number of measurements (n)	5	5	5
LOD (M)	3.79×10^{-9}	6.04×10^{-9}	6.29×10^{-9}
Sensitivity ($\mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$)	361.43	198.57	231.43
Repeatability of peak current (RSD%)	1.32 for 3.2×10^{-7}	2.18 for 3.12×10^{-7}	1.46 for 3.23×10^{-7}
Reproducibility of peak current (RSD%)	1.63 for 3.2×10^{-7}	2.32 for 3.12×10^{-7}	1.79 for 3.23×10^{-7}



3.12. Applications of the Sn–CeO₂NPs/GCPE in real sample analysis

The analysis of DTIC in real samples is crucial for clinical applications using the proposed sensor probe. The practical applicability of the optimized sensor was successfully applied to the quantification of DTIC in a number of real samples such as human serum, human urine (Fig. S6 and S7†) and pharmaceutical vial (Fig. S8†). For this purpose, the real samples were prepared as described in the Experimental section (Section 2). To check the accuracy of the procedure, the samples were spiked with known and different amounts of DTIC. The aliquots of these prepared samples were analyzed in five times ($n = 5$) according to the recommended procedures. A linear dynamic range of 7.60×10^{-8} to 6.59×10^{-6} M DTIC (for serum sample) and of 6.90×10^{-8} to 5.31×10^{-6} M DTIC (for urine sample) were obtained and the analytical results are tabulated in Table 2. The recoveries in serum and urine samples were found to lie in the range from 96.77% to 103.12% (Table S2†). The results obtained suggest that the proposed voltammetric sensor based on the Sn–CeO₂NPs/GCPE is reliable and free from the matrix interferences in the analysis of human serum and urine samples.

The practical analytical utility of the Sn–CeO₂NPs/GCPE was also illustrated by the determination of DTIC in pharmaceutical products (Fig. S8†). The standard addition method was applied in this analysis, and the results are listed in Table S3.† The recovery of the spiked samples ranged between 98.00% and 100.40%, indicating that the fabricated sensor could be efficiently used for the determination of DTIC in pharmaceutical samples.

4. Conclusions

In this work, Sn doped CeO₂ nanoparticles were successfully synthesized and further employed to fabricate Sn–CeO₂NPs/GCPE for the first time. Characterization data of synthesized Sn–CeO₂NPs by XRD, EDX and TEM confirmed that the Sn replaces Ce site and that the crystal structure of CeO₂ was retained as rutile phase up to the maximum loading of 5 wt% Sn. The fabricated electrode was successfully used to study the electrochemical behavior of DTIC. The Sn–CeO₂NPs/GCPE exhibited an excellent electrocatalytic activity towards the oxidation of DTIC. The Sn–CeO₂NPs/GCPE offers a good, reliable and simple method for the selective determination of DTIC in phosphate buffer solution of pH 4.0. The Sn–CeO₂NPs/GCPE not only shifted the oxidation potential of DTIC towards less positive potential but also enhanced its oxidation current when compared to undoped CeO₂ modified and bare GC electrodes. The fabricated electrochemical sensor exhibits good selectivity, a wide linear range, lower detection limit, high sensitivity, storage stability and acceptable reproducibility. In addition, the Sn–CeO₂NPs/GCPE was demonstrated to be useful for the detection of DTIC in pharmaceutical and human biological samples with good accuracy and precision. All these advantageous features make the proposed Sn–CeO₂NPs/GCPE sensor successfully applied for the determination of DTIC in the clinical samples, which can be used to diagnose the health condition of patients.

Live subject statement

All experiments were performed in compliance with the relevant laws and Assiut University's guidelines. The analysis of DTIC in real samples such as human serum, human urine were approved by the ethics committees of Assiut Medical University – Joint Institutional Review Board. All of the subjects signed an informed consent form before examination.

References

- 1 M. M. Eggermont and J. M. Kirkwood, Re-evaluating the role of dacarbazine in metastatic melanoma: what have we learned in 30 years?, *Eur. J. Cancer*, 2004, **40**, 1825–1836.
- 2 M. Spasova and E. Golovinski, Pharmacobiochemistry of arylalkyltriazenes and their application in cancer chemotherapy, *Pharmacol. Ther.*, 1985, **27**, 333–352.
- 3 X. Wang, Y. Li, S. Gong and D. Fu, A spectroscopic study on the DNA binding behavior of the anticancer drug dacarbazine, *Spectrosc. Lett.*, 2002, **35**, 751–756.
- 4 M. Sanada, M. Hidaka, Y. Takagi, T. Y. Takano, Y. Nakatsu, T. Tsuzuki and M. Sekiguchi, Modes of actions of two types of anti-neoplastic drugs, dacarbazine and ACNU, to induce apoptosis, *Carcinogenesis*, 2007, **28**, 2657–2663.
- 5 D. T. King and J. T. Stewart, HPLC determination of dacarbazine, doxorubicin, and ondansetron mixture in 5% dextrose injection on underivatized silica with an aqueous-organic mobile phase, *J. Liq. Chromatogr.*, 1993, **16**, 2309–2323.
- 6 D. Fiore, A. J. Jackson, M. S. Didolkar and V. R. Dandu, Simultaneous determination of dacarbazine, its photolytic degradation product, 2-azahypoxanthine, and the metabolite 5-aminoimidazole-4-carboxamide in plasma and urine by high-pressure liquid chromatography, *Antimicrob. Agents Chemother.*, 1985, **27**, 977–979.
- 7 A. Haque and J. T. Stewart, Isocratic determination of dacarbazine and related impurities 2-azahypoxanthine and 5-aminoimidazole-4-carboxamide by HPLC on an avidin protein column, *J. Liq. Chromatogr. Relat. Technol.*, 1999, **22**, 933–943.
- 8 Y. Liu, W. Zhang and Y. Yang, Validated hydrophilic interaction LC-MS/MS method for simultaneous quantification of dacarbazine and 5-amino-4-imidazole-carboxamide in human plasma, *Talanta*, 2008, **77**, 412–421.
- 9 A. J. Ordieres, A. C. Garcia, P. T. Blanco and W. F. Smyth, An electroanalytical study of the anticancer drug dacarbazine, *Anal. Chim. Acta*, 1987, **202**, 141–149.
- 10 M. Song, R. Zhang and X. Wang, Nano-titanium dioxide enhanced biosensing of the interaction of dacarbazine with DNA and DNA bases, *Mater. Lett.*, 2006, **60**, 2143–2147.
- 11 J. R. Barreira, A. Costa, A. J. Miranda and P. Tunon, Electrochemical oxidation of dacarbazine and its major metabolite (AIC) on carbon electrodes, *Electroanalysis*, 1989, **1**, 529–534.
- 12 M. U. Ahmed, M. M. Hossain and E. Tamiya, Electrochemical biosensors for medical and food applications, *Electroanalysis*, 2008, **20**, 616–626.



- 13 Y. Xiao and C. M. Li, Nanocomposites: From fabrications to electrochemical bioapplications, *Electroanalysis*, 2008, **20**, 648–662.
- 14 N. Golego, S. A. Studenikin and M. Cocivera, Sensor photoresponse of thin-film oxides of zinc and titanium to oxygen gas, *J. Electrochem. Soc.*, 2000, **147**, 1592–1594.
- 15 X. Wang, J. Song, J. Liu and Z. L. Wang, Direct-Current Nanogenerator Driven by Ultrasonic Waves, *Science*, 2007, **316**, 102–105.
- 16 K. Keren, R. S. Berman, E. Buchstab, U. Sivan and E. Braun, DNA-templated carbon nanotube field-effect transistor, *Science*, 2003, **302**, 1380–1382.
- 17 H. K. Maleh, S. Rostami, V.-K. Gupta and M. Fouladgar, Evaluation of ZnO nanoparticle ionic liquid composite as a voltammetric sensing of isoprenaline in the presence of aspirin for liquid phase determination, *J. Mol. Liq.*, 2015, **201**, 102–107.
- 18 M. Baghayeri, E.-N. Zare and M.-M. Lakouraj, A simple hydrogen peroxide biosensor based on a novel electro-magnetic poly(*p*-phenylenediamine) and Fe₃O₄ nanocomposite, *Biosens. Bioelectron.*, 2014, **55**, 259–265.
- 19 S. Das and V. Jayaraman, SnO₂: A comprehensive review on structures and gas sensors, *Prog. Mater. Sci.*, 2014, **66**, 112–255.
- 20 M. Ibrahim, Y. Temerk, H. Ibrahim and M. Kotb, Indium oxide nanoparticles modified carbon paste electrode for sensitive voltammetric determination of aromatase inhibitor formestane, *Sens. Actuators, B*, 2015, **209**, 630–638.
- 21 N. Lavanya, S. Radhakrishnan and C. Sekar, Fabrication of hydrogen peroxide biosensor based on Ni doped SnO₂ nanoparticles, *Biosens. Bioelectron.*, 2012, **36**, 41–47.
- 22 N. Lavanya, S. Radhakrishnan, C. Sekar, M. Navaneethan and Y. Hayakawa, Fabrication of Cr doped SnO₂ nanoparticles based sensor for the selective determination of riboflavin in pharmaceuticals, *Analyst*, 2013, **138**, 2061–2067.
- 23 N. Lavanya, S. Radhakrishnan, N. Sudhan, C. Sekar, S. Leonardi, C. Cannilla and G. Neri, Fabrication of folic acid sensor based on the Cu doped SnO₂ nanoparticles modified glassy carbon electrode, *Nanotechnology*, 2014, **25**, 295501.
- 24 S. D. Bukkitgar, N. P. Shetti, R. M. Kulkarni and M. R. Doddamani, Electro-oxidation of nimesulide at 5% barium-doped zinc oxide nanoparticle modified glassy carbon electrode, *J. Electroanal. Chem.*, 2016, **762**, 37–42.
- 25 N. Lavanya, C. Sekar, S. Ficarra, E. Tellone, A. Bonavita, S. G. Leonardi and G. Neri, A novel disposable electrochemical sensor for determination of carbamazepine based on Fe doped SnO₂ nanoparticles modified screen-printed carbon electrode, *Mater. Sci. Eng., C*, 2016, **62**, 53–60.
- 26 N. Lavanya, E. Fazio, F. Neri, A. Bonavita, S. G. Leonardi, G. Neri and C. Sekar, Electrochemical sensor for simultaneous determination of ascorbic acid, uric acid and folic acid based on Mn-SnO₂ nanoparticles modified glassy carbon electrode, *J. Electroanal. Chem.*, 2016, **770**, 23–32.
- 27 E. R. Brown and R. F. Large, in *Physical Methods of Chemistry*, ed. A. Weissberger and B. W. Rossiter, Wiley Inter science, Rochester New York, 1964, p. 423.
- 28 E. Aneggi, D. Wiater, C. de Leitenburg, J. Llorca and A. Trovarelli, Shape-dependent activity of ceria in soot combustion, *ACS Catal.*, 2014, **4**, 172–181.
- 29 J. Li, Z. Zhang, Z. Tian, X. Zhou, Z. Zheng, Y. Ma and Y. Qu, Low pressure induced porous nanorods of ceria with high reducibility and large oxygen storage capacity: synthesis and catalytic applications, *J. Mater. Chem. A*, 2014, **2**, 16459–16466.
- 30 R. Rao, M. Yang, C. Li, H. Dong, S. Fanga and A. Zhang, A facile synthesis for hierarchical porous CeO₂ nanobundles and their superior catalytic performance for CO oxidation, *J. Mater. Chem. A*, 2015, **3**, 782–788.
- 31 A. B. Kehoe, D. O. Scanlon and G. W. Watson, Role of lattice distortions in the oxygen storage capacity of divalently doped CeO₂, *Chem. Mater.*, 2011, **23**, 4464–4468.
- 32 Y. Temerk and H. Ibrahim, A new sensor based on In doped CeO₂ nanoparticles modified glassy carbon paste electrode for sensitive determination of uric acid in biological fluids, *Sens. Actuators, B*, 2016, **224**, 868–877.
- 33 H. Ibrahim and Y. Temerk, A novel electrochemical sensor based on B doped CeO₂ nanocubes modified glassy carbon microspheres paste electrode for individual and simultaneous determination of xanthine and hypoxanthine, *Sens. Actuators, B*, 2016, **232**, 125–137.
- 34 Y. Wu and C. Lin, The microstructures and property analysis of aliovalent cations (Sm³⁺, Mg²⁺, Ca²⁺, Sr²⁺, Ba²⁺) co-doped ceria-base electrolytes after an aging treatment, *Int. J. Hydrogen Energy*, 2014, **39**, 7988–8001.
- 35 R. N. Goyal, V. K. Gupta and S. Chatterjee, Voltammetric biosensors for the determination of paracetamol at carbon nanotube modified pyrolytic graphite electrode, *Sens. Actuators, B*, 2010, **149**, 252–258.
- 36 E. Laviron, General expression of the linear potential sweep voltammogram in the case of diffusionless electrochemical systems, *J. Electroanal. Chem.*, 1979, **101**, 19–28.
- 37 E. Laviron, The use of linear potential sweep voltammetry and of a.c. voltammetry for the study of the surface electrochemical reaction of strongly adsorbed systems and of redox modified electrodes, *J. Electroanal. Chem.*, 1979, **100**, 263–270.
- 38 J. Sangster, *Log Kow Databank A, Databank of Evaluated Octanol–Water Partition Coefficients (log P) on Microcomputer Diskette*, Sangster Res Lab, 1993.

