



Cite this: *RSC Adv.*, 2017, 7, 32732

# Manipulation of monomer-aggregate transformation of a heptamethine cyanine ligand: near infrared chromogenic recognition of Hg<sup>2+</sup>†

Xia Gao,<sup>a</sup> Weidong Wu,<sup>a</sup> Jinyan Xi<sup>a</sup> and Hong Zheng<sup>\*b</sup>

A novel near infrared optical molecular probe (CyL) derived from heptamethine cyanine dye was developed for the selective detection of Hg<sup>2+</sup> over other metal ions in aqueous solutions. The detection conditions for Hg<sup>2+</sup> were optimized. A linear range from  $1.0 \times 10^{-7}$  M to  $5.0 \times 10^{-6}$  M and a detection limit of  $1.93 \times 10^{-8}$  M with a correlation coefficient of 0.9992 were achieved under the optimal conditions. In the recognition of Hg<sup>2+</sup>, CyL alone peaks at 760 nm and exhibits a shoulder at 690 nm. Hg<sup>2+</sup> can modulate the aggregation state of CyL through coordination of thiosemicarbazide on CyL and Hg<sup>2+</sup>, and thus results in an obvious shift in the maximal absorption wavelength of CyL in the NIR region. The ESI mass spectra of CyL–Hg<sup>2+</sup> solution have been used to prove the conversion of CyL from monomers to aggregates. Finally, the proposed method has been successfully applied to the determination of Hg<sup>2+</sup> in cosmetic samples.

Received 27th March 2017

Accepted 20th June 2017

DOI: 10.1039/c7ra03517a

[rsc.li/rsc-advances](http://rsc.li/rsc-advances)

## 1. Introduction

Chemosensing of heavy metals in aqueous media is a rapidly developing field for their negative impacts on the environment and human health.<sup>1</sup> Among these heavy metals, Hg<sup>2+</sup> is one of the most toxic ones and a kind of persistent pollutant that is not biodegradable.<sup>2,3</sup> Moreover, Hg<sup>2+</sup> can easily be absorbed by skin, the respiratory system and do great harm to human health through DNA damage, mitosis impairment, and permanent damage to the central nervous system.<sup>4–6</sup> Therefore, the development of a rapid, selective, sensitive and low cost sensor for Hg<sup>2+</sup> in aqueous solution has become an urgent need.

In the last few years, various fluorogenic and chromogenic receptors have been developed for the recognition and sensing of Hg<sup>2+</sup>.<sup>7–10</sup> However, most of them show the absorption and emission wavelength in the ultraviolet-visible (UV-Vis) region (400–650 nm), and often suffer from the interference of signals within short emission wavelengths, and/or background interference, thereby making the accurate analysis of Hg<sup>2+</sup> difficult in complex matrices.

To solve these problems, more and more near-infrared dyes are exploited for the determination of Hg<sup>2+</sup>,<sup>11–14</sup> and many of them were applied for Hg<sup>2+</sup> imaging in the living cells or in

animals and show excellent performance.<sup>11,13,14</sup> Heptamethine cyanine dye is one of the important class of near-infrared dyes.<sup>15</sup> They have rigid chloro cyclohexenyl rings in the polymethine chain that can increase its photo stability, enhance the fluorescence quantum yield, and provide an ideal site for further modification with amino or phenol substitutions. Moreover, these dyes are among the best known self-associating dyes in aqueous solutions, and this self-association is clearly reflected by changes in the absorption spectra.<sup>16</sup> Therefore, applications of heptamethine cyanine dyes as NIR chemosensors for various inorganic and biological related species have been a subject of intensive interest.<sup>17–24</sup> However, they, to the best of our knowledge, have seldom been employed as Hg<sup>2+</sup> optical probes except for few several reports.<sup>12,25</sup> In addition, it has been reported that N and S on hydrazine of thiosemicarbazide could form stable complex with Hg<sup>2+</sup>.<sup>26</sup> Meanwhile, self-aggregation of cyanine dyes can be clearly reflected by changes in the absorption spectra.<sup>27</sup>

By introducing a thiosemicarbazone unit, an ionophore for heavy metal ions,<sup>28</sup> into the cyclohexane bridgehead of heptamethine cyanine dye, herein we presented a new NIR heptamethine cyanine ligand CyL. And then, a new strategy for an efficient chromogenic recognition and determination of Hg<sup>2+</sup> in aqueous solutions *via* modulation of aggregation state of a heptamethine cyanine probe in aqueous medium, leading to a shift in the maximal absorption wavelength in the NIR region has been proposed. Compared with Hg<sup>2+</sup>-selective optical chemical sensors as reported previously, the as-prepared sensor could recognize and detect Hg<sup>2+</sup> in the NIR region, where most of the natural matrix interference could be eliminated, and greatly improved the sensitivity of Hg<sup>2+</sup> detection. This present

<sup>a</sup>School of Public Health, Xinxiang Medical University, Xinxiang, 453003, PR China

<sup>b</sup>Department of Chemistry, College of Chemistry and Chemical Engineering, Xiamen University, Xiamen, 361005, PR China. E-mail: hzheng@xmu.edu.cn

† Electronic supplementary information (ESI) available: <sup>1</sup>H, <sup>13</sup>C NMR and ESI-MS mass spectra of compound 2 and CyL; competitive absorbance response of CyL with Hg<sup>2+</sup>; absorbance titrations of CyL with Hg<sup>2+</sup>; ESI-MS spectra of CyL–Hg<sup>2+</sup>. See DOI: 10.1039/c7ra03517a



study provided an effective solution for the troublesome problem in the recognition and determination of  $\text{Hg}^{2+}$ , and has been successfully applied to the determination of  $\text{Hg}^{2+}$  in cosmetic samples.

## 2. Materials and methods

### 2.1 Chemicals and reagents

1,2,3,3-Tetramethyl-3*H*-indolium iodide was purchased from Sigma-Aldrich and dry DMF from ACROS ORGANICS. All the other reagents were obtained from Shanghai Chemicals Group Company. The inorganic salts were of the highest purity in the form of nitrates or chlorides. Dry ethanol was distilled from Mg and twice deionized water was further distilled in the presence of  $\text{KMnO}_4$ . Buffer solution (pH 4.0) was prepared by mixing 0.20 M sodium acetate and 0.20 M acetic acid.

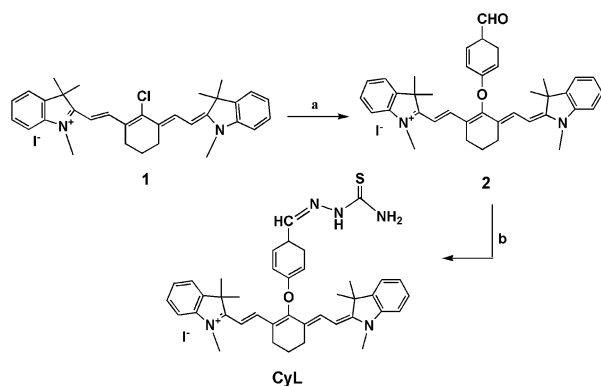
### 2.2 Instrumentation

Bruker AV400 spectrometer (400 MHz for  $^1\text{H}$  and 100 MHz for  $^{13}\text{C}$ ) with tetramethylsilane (TMS) as internal standard was used for profiling NMR spectra in  $d_6$ -DMSO; JEOL JMS-T100LC mass spectrometer was employed to profile mass spectra; and Varian CARY 300 spectrophotometer was utilized to determine absorption spectra. CEM MARS microwave digestion apparatus was used for the pretreatment of cosmetics.

### 2.3 Synthesis of the new heptamethine cyanine dye

The synthetic procedure for the new heptamethine cyanine dye (CyL) was illustrated in Scheme 1. The heptamethine cyanine 1 was synthesized according to the method reported previously.<sup>29</sup>

**2.3.1 Synthesis of compound 2.** Compound 2 was synthesized as follows: 28.58 mg (0.65 mmol) of sodium hydride (55–65%) and 79.37 mg (0.65 mmol) of *p*-hydroxybenzaldehyde were dissolved in anhydrous DMF; the reaction mixture was stirred for 30 min under  $\text{N}_2$  atmosphere at  $0^\circ\text{C}$ , and then allowed to warm to room temperature. This solution was added to the cyanine dye 1 (400.0 mg, 0.65 mmol) in 5.0 mL of dry DMF at room temperature with stirring under  $\text{N}_2$ . The reaction was performed for a total of 12 h, and then diluted with chloroform.



**Scheme 1** Synthetic scheme of compound CyL. Conditions: (a) NaH, *p*-hydroxy-benzaldehyde, dry DMF, room temperature,  $\text{N}_2$ ; (b) thiosemicarbazide, dry ethanol, reflux,  $\text{N}_2$ .

After washed with water for several times, the organic layer was dried with  $\text{Na}_2\text{SO}_4$ . Following removal of chloroform, the residue was purified by flash chromatography with methanol/chloroform (1/30) as eluent to afford 2 (236.30 mg, yield: 46.52%). The newly synthesized compound was characterized using  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR and mass spectral techniques. The results obtained were as follows:  $^1\text{H}$  NMR ( $d_6$ -DMSO, 400 MHz)  $\delta$ : 1.263 (s, 12H,  $\text{CH}_3$ ), 1.965 (quintet,  $J = 6$  Hz, 2H,  $-\text{CH}_2-\text{CH}_2-\text{CH}_2-$ ), 2.750 (t,  $J = 6$  Hz, 4H,  $-\text{CH}_2-\text{CH}_2-\text{CH}_2-$ ), 3.618 (s, 6H,  $\text{CH}_3$ ), 6.202 (d,  $J = 14$  Hz, 2H,  $\text{CH}=\text{CH}$ ), 7.184–7.224 (m, 2H, ArH), 7.375–7.395 (m, 6H, ArH), 7.492 (d,  $J = 7.6$  Hz, 2H, ArH), 7.716 (d,  $J = 14$  Hz, 2H,  $\text{CH}=\text{CH}$ ), 8.005 (d,  $J = 8.4$  Hz, 2H, ArH), 9.917 (s, 1H, CHO) (Fig. S1 $\dagger$ ).  $^{13}\text{C}$  NMR ( $d_6$ -DMSO, 100 MHz):  $\delta$  20.569, 23.636, 26.897, 31.252, 48.372, 100.828, 111.146, 115.150, 120.734, 122.193, 124.802, 128.358, 130.950, 132.467, 139.768, 140.772, 142.643, 160.787, 163.569, 172.029, 191.355 (Fig. S2 $\dagger$ ). ESI mass spectrometry,  $m/z$ : 569.3 ( $\text{M} - \text{I}$ ) $^+$ , 570.3 ( $\text{M} - \text{I} + 1$ ) $^+$  (Fig. S3 $\dagger$ ).

**2.3.2 Synthesis of CyL.** Compound CyL was synthesized as follows: compound 2 (200.00 mg, 0.28 mmol) and thiosemicarbazide (127.58 mg, 1.40 mmol) were dissolved in dry ethanol, and the reaction mixture was refluxed for 4 hours under  $\text{N}_2$  atmosphere. After removal of ethanol, the residue was purified through flash chromatography with methanol/chloroform (1/30) as eluent to afford CyL (112.76 mg, yield: 50%). Compound CyL was characterized using  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR and mass spectral techniques. The results obtained are as follows:  $^1\text{H}$  NMR ( $d_6$ -DMSO, 400 MHz)  $\delta$ : 1.280 (s, 12H,  $\text{CH}_3$ ), 1.949 (quintet,  $J = 5.6$  Hz, 2H,  $-\text{CH}_2-\text{CH}_2-\text{CH}_2-$ ), 2.737 (t,  $J = 5.6$  Hz, 4H,  $-\text{CH}_2-\text{CH}_2-\text{CH}_2-$ ), 3.611 (s, 6H,  $\text{CH}_3$ ), 6.183 (d,  $J = 14.4$  Hz, 2H,  $\text{CH}=\text{CH}$ ), 7.169–7.219 (m, 4H, ArH), 7.369 (s, 2H, ArH), 7.377 (s, 2H,  $\text{NH}_2$ ), 7.494 (d,  $J = 7.6$  Hz, 2H, ArH), 7.767 (d,  $J = 14.4$  Hz, 2H,  $\text{CH}=\text{CH}$ ), 7.862 (d,  $J = 8.8$  Hz, 2H, ArH), 7.969 (d,  $J = 12.4$  Hz, 2H, ArH), 8.187 (s, 1H, CH), 11.391 (s, 1H, NH) (Fig. S4 $\dagger$ ).  $^{13}\text{C}$  NMR ( $d_6$ -DMSO, 100 MHz):  $\delta$  21.215, 24.175, 27.501, 31.747, 48.899, 101.161, 111.614, 115.112, 121.574, 122.747, 125.272, 128.893, 128.980, 130.065, 140.655, 141.296, 141.906, 143.206, 161.005, 162.071, 172.508, 178.263 (Fig. S5 $\dagger$ ). ESI mass spectrometry,  $m/z$ : 642.3 ( $\text{M} - \text{I}$ ) $^+$ , 643.3 ( $\text{M} - \text{I} + 1$ ) $^+$  (Fig. S6 $\dagger$ ).

### 2.4 General procedure

Stock solutions containing  $1.0 \times 10^{-3}$  M of CyL and  $0.8 \times 10^{-3}$  M of  $\text{Hg}^{2+}$  were prepared in MeOH and redistilled water, respectively. To 10.0 mL of 0.20 M acetate buffer (pH 4.0) solution containing different amounts of  $\text{Hg}^{2+}$ , small aliquots of CyL stock solution (100  $\mu\text{L}$ ) was added. After being mixed thoroughly and allowed to equilibrate for 10 minutes, the NIR absorption spectra were recorded and the ratio  $R(A_{666}/A_{760})$  was tested.

## 3. Results and discussion

### 3.1 Spectral characteristics of CyL

CyL is a new derivative of cationic cyanine dyes. Absorption spectral traces of CyL upon coordination with  $\text{Hg}^{2+}$  in an optimized acetate buffer (0.20 M) solution of pH 4.0 (MeOH/ $\text{H}_2\text{O} = 1/99$ , v/v) were monitored. Fig. 1 showed the absorption spectral



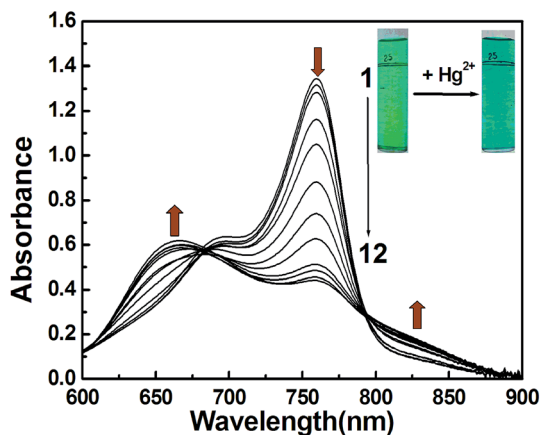


Fig. 1 Vis-NIR spectra of CyL in MeOH-H<sub>2</sub>O (1/99, v/v) aqueous acetate buffer solution of pH 4.0 in the absence (curve 1) and presence of increasing concentration of Hg<sup>2+</sup> (curves 2–12). [CyL]:  $1.0 \times 10^{-5}$  M; [Hg<sup>2+</sup>] (from curves 2 to 12,  $10^{-6}$  M): 0.3, 0.5, 0.7, 0.9, 1, 3, 5, 7, 9, 10, 20. Inset: chromogenic response of CyL to  $1.0 \times 10^{-5}$  M of Hg<sup>2+</sup> in MeOH-H<sub>2</sub>O (1/99, v/v) aqueous acetate buffer solution of pH 4.0.

changes of CyL ( $1.0 \times 10^{-5}$  M) in the presence of Hg<sup>2+</sup>. CyL solution alone exhibits a maximum at 760 nm and a shoulder at 690 nm. With the increase in Hg<sup>2+</sup> concentration, the absorbance at 760 nm decreased, while a new absorption band at a shorter wavelength of 666 nm appeared. The system color also changed from emerald green to bluish green accordingly. These pronounced hypsochromic shift can be ascribed to the H-aggregation of the cyanine dye<sup>15</sup> resulting from the coordination of CyL and Hg<sup>2+</sup>.

### 3.2 Optimization of experimental conditions

**3.2.1 Effect of buffer system and pH.** To obtain the optimal conditions, the buffer system and the influence of pH on the sensitivity of this method were investigated. The results showed the best absorbance changes ( $R/R_0$ , where  $R_0$  and  $R$  are  $A_{666}/A_{760}$  in the absence and presence of Hg<sup>2+</sup>, respectively) were obtained at pH 3.0–6.0 (Fig. 2), and the changes of buffer system

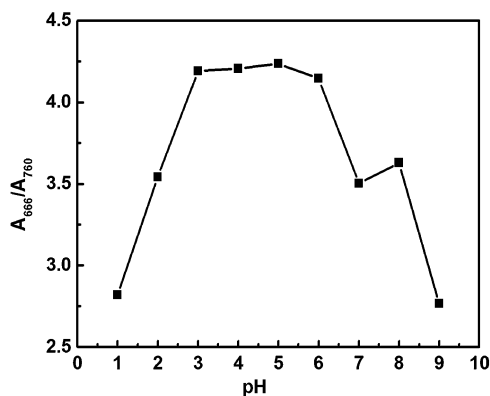


Fig. 2 Effects of pH on the changes of absorbance. [Hg<sup>2+</sup>] =  $8.0 \times 10^{-6}$  M; [CyL] =  $1.0 \times 10^{-5}$  M; volume of each buffer is 1 mL; other conditions are the same as those described in the general procedure except the buffer system.

had minimal effect on detection sensitivity. Thus, in this study, pH 4.0 of HAC–NaAc buffer was selected to control the acidity of the system.

**3.2.2 Effect of CyL concentration.** The effect of CyL concentration on Hg<sup>2+</sup> measurement was investigated. When concentration of Hg<sup>2+</sup> was kept at  $1.0 \times 10^{-5}$  M,  $R(A_{666}/A_{760})$  increased with the increasing amount of CyL. When the concentration was  $1.0 \times 10^{-5}$  M,  $R$  reached the maximum. Thus,  $1.0 \times 10^{-5}$  M was chosen as the optimum CyL concentration in the experiment.

**3.2.3 Effect of incubation time.** Under the optimized conditions as depicted above, the effect of incubation time on  $R(A_{666}/A_{760})$  was tested. As shown in Fig. 3, the system reached equilibrium 10 min following Hg<sup>2+</sup> addition, and remained constant for at least 50 min. Thus, it was suggested that the measurement should start 10 min after addition of Hg<sup>2+</sup> finish within 50 min.

### 3.3 Selectivity and interference of foreign ions

To examine the specificity of Hg (CyL)<sub>2</sub> complex formation under the presence of environmentally relevant metal ions, a variety of heavy, transition, and main group metal ions were

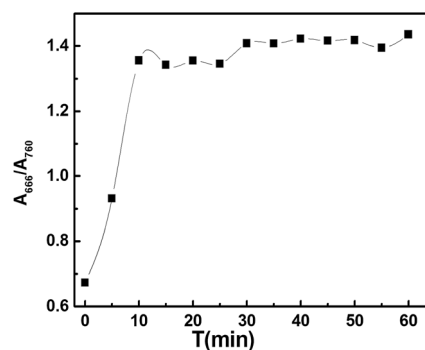


Fig. 3 Effects of time on the absorption response of CyL to Hg<sup>2+</sup> in MeOH-H<sub>2</sub>O (1/99, v/v) aqueous acetate buffer solution of pH 4.0; [CyL] =  $1.0 \times 10^{-5}$  M; [Hg<sup>2+</sup>] =  $5.0 \times 10^{-6}$  M; other conditions were the same as those described in the general procedure.

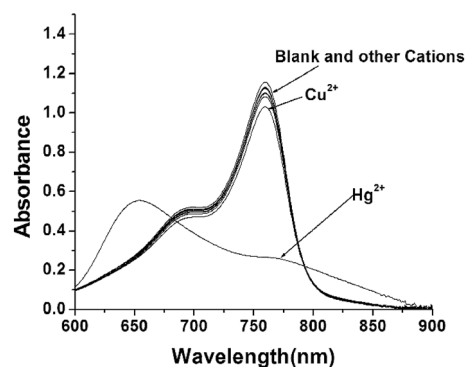


Fig. 4 Absorption response of CyL to various metal ions (K<sup>+</sup>, Li<sup>+</sup>, Na<sup>+</sup>, Zn<sup>2+</sup>, Ni<sup>2+</sup>, Co<sup>2+</sup>, Fe<sup>3+</sup>, Al<sup>3+</sup>, Pb<sup>2+</sup>, Cr<sup>3+</sup>, Mn<sup>2+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, Cu<sup>2+</sup>, Hg<sup>2+</sup>) in pH 4.0 acetate buffer. [CyL] =  $1.0 \times 10^{-5}$  M, [M<sup>n+</sup>] =  $1.0 \times 10^{-5}$  M.



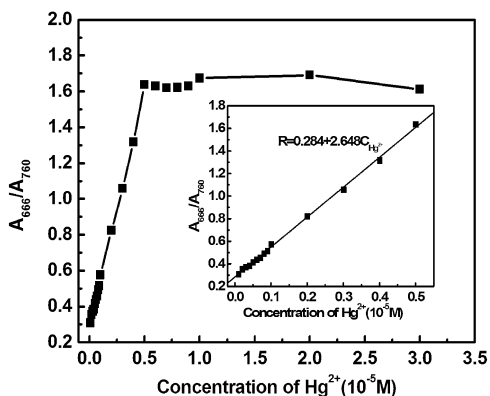


Fig. 5 The absorption titration curves and calibration graphs for spectrophotometric determination of  $\text{Hg}^{2+}$  in MeOH– $\text{H}_2\text{O}$  (1/99, v/v) aqueous acetate buffer solution of pH 4.0;  $[\text{CyL}] = 1.0 \times 10^{-5} \text{ M}$ ;  $[\text{Hg}^{2+}]$ : ( $1.0 \times 10^{-7} \text{ M}$  to  $3.0 \times 10^{-5} \text{ M}$ ); other conditions were the same as those described in the general procedure.

tested in their capability of binding CyL (Fig. 4). It was revealed from Fig. 4 that CyL at  $1.0 \times 10^{-5} \text{ M}$  exhibited little decrease in absorbance at 760 nm upon addition of 1.0 equiv. of the tested ions including  $\text{Li}^+$ ,  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Zn}^{2+}$ , and  $\text{Pb}^{2+}$ . Only the addition of  $\text{Hg}^{2+}$  resulted in a remarkable reduction of the absorbance at 760 nm and a new absorption at shorter wavelength of 666 nm. This implied the existence of a higher coordination selectivity of CyL toward  $\text{Hg}^{2+}$ . Competitive experiments further showed that the decrease in  $R(A_{666}/A_{760})$  induced by  $\text{Hg}^{2+}$  was not disturbed by the presence of coexisting metal ions (Fig. S7<sup>†</sup>), indicating the highly specific coordination of CyL to  $\text{Hg}^{2+}$ . Taken together, CyL could be practically served as a NIR-sensing probe for  $\text{Hg}^{2+}$  in aqueous solutions.

#### 3.4 Calibration graphs and analysis of samples

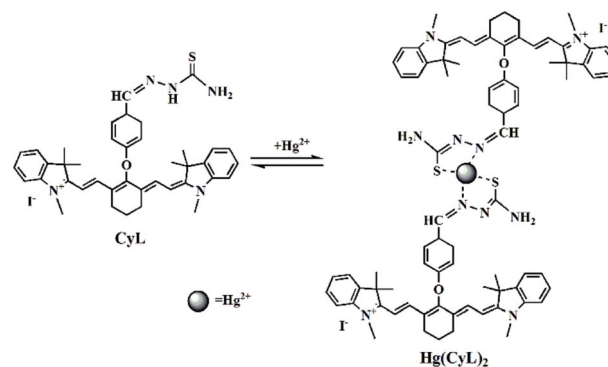
Under the optimized conditions, the titration curves for the measurement of  $\text{Hg}^{2+}$  were constructed. Response assays showed that  $R(A_{666}/A_{760})$  increased with the increment of  $\text{Hg}^{2+}$  concentration, varied from 0.302 in the absence of  $\text{Hg}^{2+}$  to 1.637 after  $5.0 \times 10^{-6} \text{ M}$   $\text{Hg}^{2+}$  treatment. That was,  $R$  increased 4.42 times due to the addition of  $\text{Hg}^{2+}$ . The curve equation as shown in Fig. 5 was  $R = 0.284 + 2.648C_{\text{Hg}^{2+}}$ , with a correlation coefficient of 0.9992 ( $N = 14$ , S.D. = 0.0017) when a CyL

concentration of  $1.0 \times 10^{-5} \text{ M}$  was used. The linear range of quantitative detection for  $\text{Hg}^{2+}$  was determined to be  $1.0 \times 10^{-7}$  to  $5.0 \times 10^{-6} \text{ M}$  with a detection limit of  $1.93 \times 10^{-8} \text{ M}$  in the used aqueous solution system.

In order to check the potential applicability of this method, we have applied our method to the quantitative determination of  $\text{Hg}^{2+}$  in cosmetics. In our experiment, three cosmetics samples were collected and treated according to the literature,<sup>30</sup> and then diluted 10-fold for experimental uses. The specific sample pretreatment processes were listed in the footnote of Table 1, and the determination results by this method and by the cold atomic absorption spectrometry (AAS) method were shown in Table 1. It could be seen from Table 1 that the determination results are very close to those obtained by the cold AAS method. These results together with results of the selectivity and competition experiments confirmed that method proposed in the present work was reliable and practical for the determination of  $\text{Hg}^{2+}$  in actual samples.

#### 3.5 Sensing mechanism

The Job's plots of absorbance at 760 nm (Fig. S8<sup>†</sup>) suggested that a 2 : 1 complex formed between CyL and  $\text{Hg}^{2+}$ . On the basis of the high affinity of  $\text{Hg}^{2+}$  toward thiosemicarbazide,<sup>14</sup> the decrease of absorbance at 760 nm could be ascribed to the formation of  $\text{Hg}(\text{CyL})_2$  complex by the coordination of  $\text{Hg}^{2+}$  to N and S on thiosemicarbazide of CyL. This assumption was further supported by the ESI mass spectra of CyL– $\text{Hg}^{2+}$  solution (Fig. S9<sup>†</sup>), which shows a strong peak at  $m/z$  741.7 and a weak



Scheme 2 Proposed mechanism of CyL binding with  $\text{Hg}^{2+}$ .

Table 1 Analytical results of cosmetic samples

Samples	Proposed method found <sup>a</sup> ( $10^{-6} \text{ M}$ )	$\text{Hg}^{2+}$ , added ( $10^{-6} \text{ M}$ )	$\text{Hg}^{2+}$ , found ( $10^{-6} \text{ M}$ )	Recovery (%)	Cold AAS method found <sup>a</sup> ( $10^{-6} \text{ M}$ )
1	$0.70 \pm 0.1$	1.00	$1.72 \pm 0.1$	102	$0.67 \pm 0.1$
2	$0.90 \pm 0.1$	1.00	$1.88 \pm 0.1$	98.5	$0.86 \pm 0.1$
3	Not found	1.00	$0.97 \pm 0.1$	97.0	Not found

<sup>a</sup> The cosmetics samples were prepared by resolving 1.0 g of each sample into 2.0 mL distilled water and stirring into syrupy liquid, then, they were added 3.0 mL of nitric acid and kept for 12 hours at room temperature. Next, each sample was added 2.0 mL of hydrogen peroxide and treated with microwave digestion. Finally, the resulting solution was diluted into 10 mL, the determination results were obtained by further diluting above solution for 10-fold.



peak at  $m/z$  642.3, assigned to  $(2\text{CyL} - 2\text{I} + \text{Hg} - 2\text{H})_2^+$  for  $\text{Hg}(\text{CyL})_2$  complex and  $(\text{CyL} - \text{I})^+$  for CyL monomer, respectively. Based on these results, sensing mechanism of CyL to  $\text{Hg}^{2+}$  was proposed in Scheme 2.

## 4. Conclusion

In summary, a new NIR heptamethine cyanine ligand has been proposed for selective binding of  $\text{Hg}^{2+}$  based on the monomer-aggregate transformation of the NIR ligand. In addition to simplicity and effectivity, this sensor still showed a limit of detection of  $1.93 \times 10^{-8}$  M in the aqueous solution. The high sensitivity was made possible through the unique sensing mechanism, whereby even small amount of  $\text{Hg}^{2+}$  recognized could result in the aggregation of CyL, leading to the changes of absorption spectrum. The tested method proposed CyL as a novel sensor for the detection of  $\text{Hg}^{2+}$  by absorption spectrometry in aqueous solution, and could be used as a good alternative to the other sensors for  $\text{Hg}^{2+}$ .

## Acknowledgements

The authors are grateful for the fund support by the Foundation and Cutting-edge Research Projects of Henan Province (162300410019) and the Doctoral Scientific Research Foundation of Xinxiang Medical University (505062).

## References

- 1 X. Fu, T. Lou, Z. Chen, M. Lin, W. Feng and L. Chen, *ACS Appl. Mater. Interfaces*, 2012, **4**, 1080.
- 2 E. M. Nolan and S. J. Lippard, *Chem. Rev.*, 2008, **108**, 3443.
- 3 T. Balaji, S. A. El Safty, H. Matsunaga, T. Hanaoka and F. Mizukami, *Angew. Chem., Int. Ed.*, 2006, **45**, 7202.
- 4 J. Mutter, J. Naumann, R. Schneider, H. Walach and B. Haley, *Neuroendocrinol. Lett.*, 2005, **26**, 439.
- 5 W. F. Fitzgerald, C. H. Lamgorg and C. R. Hammerschmidt, *Chem. Rev.*, 2007, **107**, 641.
- 6 E. M. Nolan and S. J. Lippard, *Chem. Rev.*, 2008, **108**, 3443.
- 7 C. Kumaria, D. Saina, A. Kumara, H. P. Nayeka, S. Debnath, P. Saha and S. Dey, *Sens. Actuators, B*, 2017, **243**, 1181.
- 8 Y. Zhao, L. L. Gui and Z. B. Chen, *Sens. Actuators, B*, 2017, **241**, 262.
- 9 S. R. Tang, P. Tong, M. L. Wang, J. H. Chen, G. W. Li and L. Zhang, *Chem. Commun.*, 2015, **51**, 15043.
- 10 N. Kaur, P. Kaur, G. Bhatia, K. Singh and J. Singh, *RSC Adv.*, 2016, **6**, 82810.
- 11 J. L. Wang, W. L. Li and L. P. Long, *Sens. Actuators, B*, 2017, **245**, 462.
- 12 H. Zheng, X. J. Zhang, X. Cai, Q. N. Bian, M. Yan, G. H. Wu, X. W. Lai and Y. B. Jiang, *Org. Lett.*, 2012, **14**, 1986.
- 13 M. N. Wang, X. J. An and J. Gao, *J. Lumin.*, 2013, **144**, 91.
- 14 C. C. Zhao, K. B. Li, N. Xie, M. Zhao and S. Q. Peng, *J. Photochem. Photobiol., A*, 2014, **290**, 72.
- 15 L. Strekowski, M. Lipowska and G. Patonay, *J. Org. Chem.*, 1992, **57**, 578.
- 16 A. Mishra, R. K. Behera, P. K. Behera, B. K. Mishra and G. B. Behera, *Chem. Rev.*, 2000, **100**, 1973.
- 17 G. H. Cheng, J. L. Fan, W. Sun, J. F. Cao, C. Hu and X. J. Peng, *Chem. Commun.*, 2014, **50**, 1018.
- 18 B. H. Li, Y. L. Zhang, F. S. Li, W. Wang, J. Liu, M. Liu, Y. Cui, X. B. Li and B. L. Li, *Sens. Actuators, B*, 2016, **233**, 479.
- 19 K. Yin, F. B. Yu, W. W. Zhang and L. X. Chen, *Biosens. Bioelectron.*, 2015, **74**, 156.
- 20 X. X. Hu, Y. T. Su, Y. W. Ma, X. Q. Zhan, H. Zheng and Y. B. Jiang, *Chem. Commun.*, 2015, **51**, 15118.
- 21 X. Y. Han, F. B. Yu, X. Y. Song and L. X. Chen, *Chem. Sci.*, 2016, **7**, 5098.
- 22 Z. R. Lou, P. Li and K. L. Han, *Acc. Chem. Res.*, 2015, **48**, 1358.
- 23 F. B. Yu, P. Li, G. Y. Li, G. J. Zhao, T. S. Chu and K. L. Han, *J. Am. Chem. Soc.*, 2011, **133**, 11030.
- 24 F. B. Yu, P. Li, B. S. Wang and K. L. Han, *J. Am. Chem. Soc.*, 2013, **135**, 7674.
- 25 C. Kur, Y. Shindo, K. Oka, S. Nishiyama, K. Suzuki and D. Citterio, *RSC Adv.*, 2017, **7**, 24970.
- 26 Y. Yu, L. R. Lin, K. B. Yang, X. Zhong, R. B. Huang and L. S. Zheng, *Talanta*, 2006, **69**, 103.
- 27 A. Mishra, R. K. Behera, P. K. Behera, B. K. Mishra and G. B. Behera, *Chem. Rev.*, 2000, **100**, 1973.
- 28 T. S. Lobana, A. Sanchez, J. S. Casas, A. Castineiras, J. Sordo and M. S. Garcia-Tasende, *Polyhedron*, 1998, **17**, 3701.
- 29 R. Chin and N. Salazar, *J. Heterocycl. Chem.*, 1996, **33**, 1871.
- 30 F. R. Tian, L. H. Yang, X. Y. Ye and Y. L. Liu, *J. Chem. Res.*, 2011, **22**, 91.

