

Cite this: *RSC Adv.*, 2017, 7, 28527

# Enhancing menaquinone-7 production in recombinant *Bacillus amyloliquefaciens* by metabolic pathway engineering†

Jian-Zhong Xu, \*<sup>ab</sup> Wei-Liu Yan<sup>b</sup> and Wei-Guo Zhang<sup>b</sup>

Here, we compared the amino acid sequence of the head structure biosynthetic enzymes in the menaquinone-7 (MK-7) biosynthetic pathway between *Bacillus amyloliquefaciens* Y-2 and *B. amyloliquefaciens* W-21 that are distinct in MK-7 production, and investigated the effect of these enzymes on MK-7 production. Sequence analysis showed that six enzymes had undergone non-synonymous substitutions: MenA, MenC, MenD, MenE, MenH and HepS. Overexpression of these enzymes from strain Y-2 in *B. subtilis* 168 significantly increased the corresponding enzyme activity (increased by  $\geq 500\%$ ), which was higher than that from overexpressing these enzymes from strain W-21 (increased by  $\approx 200\%$ ). Moreover, the MK-7 content in *B. subtilis* 168 or *B. amyloliquefaciens* Y-2 was enhanced by the overexpression of these enzymes from strain Y-2. Note that the overexpression of MenA in *B. subtilis* 168 increased the MK-7 content up to 1.6-fold, whereas the overexpression of HepS in *B. amyloliquefaciens* Y-2 led to a greater increase in MK-7 production than that of other enzymes, both in the stilling culture (increased by 93.62%) and in the shaking culture (increased by 93.29%). It follows that the high enzyme activity and high-traffic biosynthetic pathway are beneficial to improve MK-7 production. These results provide a definite theoretical foundation for breeding MK-7 high-yielding strains *via* metabolic engineering.

Received 23rd March 2017  
Accepted 17th May 2017

DOI: 10.1039/c7ra03388e

rsc.li/rsc-advances

## 1 Introduction

Menaquinones (MK-*n*, where *n* represents the number of isoprene units), a type of vitamin K<sub>2</sub>, have significant effects on the electron transfer of the respiratory chain.<sup>1</sup> In prokaryotes, MK-*n* plays an important role in shuttling electrons between membranes bound to protein complexes in the electron transport chain.<sup>2</sup> For human beings and other mammals, MK-*n* is an essential vitamin for blood coagulation, bone metabolism and cell-cycle regulation processes.<sup>3,4</sup> Menaquinone-7 (MK-7) is part of a family of MK-*n*s with a side chain of seven isoprene units.<sup>1</sup> As recent research shows, MK-7 can activate osteocalcin in bone marrow to promote the formation of bone, and is able to improve the bone mineral density (BMD) in the aged so as to reduce the risk of haunch bone fracture.<sup>5</sup> In addition, it can also lower blood pressure in rats.<sup>6</sup> Therefore, MK-7 is receiving

increasing attention in the domain of nutritional supplements for humans.

Many microorganisms can be used to biosynthesize MK-7, such as *Escherichia coli*,<sup>7</sup> *Sphingobacterium multivorum*,<sup>8</sup> *Bacillus amyloliquefaciens*,<sup>9,10</sup> and *Bacillus subtilis*.<sup>11–13</sup> However, commercial production of MK-7 is mainly done through extraction from natto, a type of food, rather than *via* microbial fermentation.<sup>14</sup> This is because the fermentative production of MK-7 is too low.<sup>9,11–13</sup> Fortunately, the biosynthetic pathway of MK-7 has been clarified at present (Fig. 1). As can be seen in Fig. 1, there are nine genes, designated as *menA*, *menB*, *menC*, *menD*, *menE*, *menH*, *menG*, *hepS* and *hepT*, encoding the head structure biosynthesis enzymes that are involved in the MK-7 biosynthetic pathway starting from isochorismate. However, some enzymes in the biosynthetic pathway are feedback-regulated by intermediates. For example, 3-deoxy-D-arabino-heptulosonate-7-phosphate (DAHP) synthase is feedback-inhibited by 1,4-dihydroxy-2-naphthoate (DHNA), L-tyrosine, L-phenylalanine and L-tryptophan.<sup>15</sup> To obtain mutants with removing feedback-regulation, the wild-type strains were treated with the conventional mutation-inducing methods, such as ultraviolet (UV) light and *N*-methyl-*N*-nitro-*N*-nitrosoguanidine (NTG).<sup>16–18</sup> In addition, Kwon *et al.*<sup>19</sup> pointed out that the coding levels of the enzymes involved in MK-7 biosynthesis are present in extremely low quantities. However, there

<sup>a</sup>The Key Laboratory of Carbohydrate Chemistry and Biotechnology, Ministry of Education, School of Biotechnology, Jiangnan University, 1800# Lihu Road, Wuxi 214122, People's Republic of China. E-mail: xujianzhong@jiangnan.edu.cn; Fax: +86-510-85329312; Tel: +86-510-85329312

<sup>b</sup>The Key Laboratory of Industrial Biotechnology, Ministry of Education, School of Biotechnology, Jiangnan University, 1800# Lihu Road, Wuxi 214122, People's Republic of China

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c7ra03388e



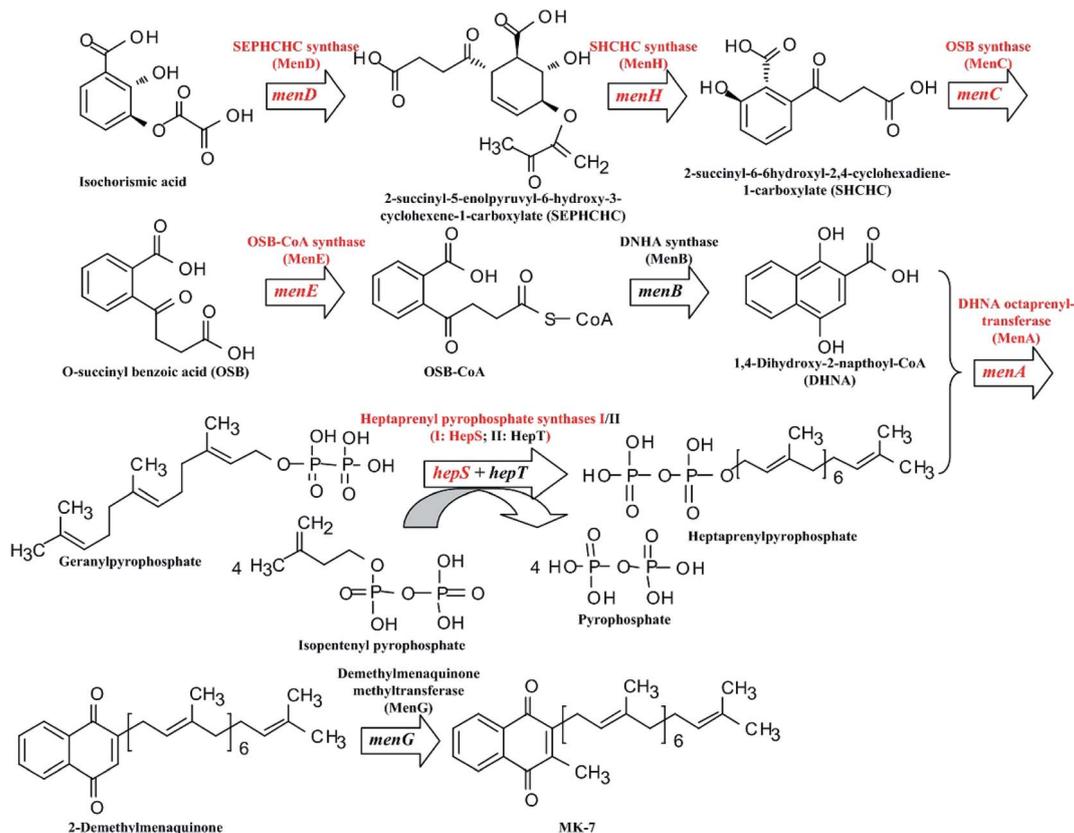


Fig. 1 Schematic diagram of the MK-7 biosynthetic pathway in *Bacillus* sp. The red font represents the modified genes and the corresponding enzymes.

are few genetic engineering models for developing a microbial producer of MK-7 via increasing the coding level of the enzymes.

In this study, the endogenous MK-7 biosynthetic pathway was metabolically engineered to construct an efficient *B. amyloliquefaciens* platform to produce MK-7. After comparing the genetic difference between strains with different MK-7 productivity, six genes in the MK-7 biosynthetic pathway were selected to evaluate the effect on MK-7 production. The study shows the first time that different enzymes in the MK-7 biosynthetic pathway have differing roles in MK-7 production, for example in cell growth as well as affecting the MK-7 yield.

## 2 Materials and methods

### 2.1 Strains, growth media and culturing conditions

The strains used in this study are listed in Table 1. *B. amyloliquefaciens* Y-2 and *B. amyloliquefaciens* W-21 were isolated from Chinese fermented beans. A phylogenetic tree based on the 16S rDNA sequence homology of strains Y-2 and W-21 is shown in Fig. S1.† Strains Y-2 and W-21 show high fibrinogenase activity ( $>1450$  IU mL<sup>-1</sup>), but only strain Y-2 can be used to produce MK-7 ( $7.1 \pm 0.5$  mg L<sup>-1</sup>) with maize meal hydrolysate as the carbon source (Table S1†). Luria-Bertani (LB) was used as a standard medium for cultivating all strains. A tryptone–yeast extract–glucose (TYG) medium composed of 5 g L<sup>-1</sup> tryptone, 2.5 g L<sup>-1</sup> yeast extract and 5 g L<sup>-1</sup> glucose was used as the seed medium. A glycerol–yeast extract–soy peptone (GYS) medium was used as

the basal medium for MK-7 production.<sup>20</sup> The Spizizen minimal (SM) medium or low SM (LSM) medium was used as the basal medium for preparing competent cells or isolating transformants.<sup>21,22</sup> All media were adjusted to pH  $7.2 \pm 0.2$  with 20% (w/v) NaOH. When appropriate, *E. coli* and *Bacillus* sp. were cultured with kanamycin (Km, 50  $\mu$ g mL<sup>-1</sup>) at 100 rpm.

### 2.2 DNA manipulations

The plasmids and oligonucleotides used in this study are listed in Tables 2 and S2,† respectively. DNA was extracted from *B. amyloliquefaciens* Y-2 using the Ezup-pillar Bacterial Genomic DNA Extraction Kit (Sangon, Shanghai, China), and it was used as a template to amplify the tested genes. The tested genes from the strains Y-2 and W-21 were sequenced by Sangon Biotech (Shanghai) Co., Ltd. (Shanghai, China). Plasmid construction and transformation were performed according to the descriptions in previous reports.<sup>23</sup> The build processes of the plasmids are illustrated in the ESI (Fig. S2†). The plasmids were extracted from *E. coli* or *Bacillus* sp. using the SanPrep Mini Plasmid Kit (Sangon, Shanghai, China) and confirmed by the restriction endonuclease reaction.

### 2.3 Enzyme activity assay

Cell extracts were prepared as reported by Xu *et al.*<sup>24</sup> After centrifugation at 4 °C for 30 min at  $10\,000 \times g$ , the cell-free supernatants were immediately used to determine the enzyme



Table 1 Strains used in this study

Strain	Relevant characteristic(s)	Source
<i>E. coli</i> JM109	<i>recA1 endA1 gyrA96 thi-1 hsdR17 e14-(mcrA-)</i> <i>supE44 relA1 Δ(lac-proAB)/F'</i> [ <i>traD36 proB<sup>+</sup> lacI<sup>q</sup> lacZΔM15</i> ]	Takara
<b><i>B. subtilis</i> strain</b>		
168	<i>trpC2 sfp<sup>0</sup> degQ<sup>0</sup>, IT- SF- PL-</i>	ATCC
168/pMA5	<i>B. subtilis</i> 168 harboring plasmid pMA5	This work
168/pMA5- <i>A</i> <sub>Y-2</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>A</i> <sub>Y-2</sub>	This work
168/pMA5- <i>C</i> <sub>Y-2</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>C</i> <sub>Y-2</sub>	This work
168/pMA5- <i>D</i> <sub>Y-2</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>D</i> <sub>Y-2</sub>	This work
168/pMA5- <i>E</i> <sub>Y-2</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>E</i> <sub>Y-2</sub>	This work
168/pMA5- <i>H</i> <sub>Y-2</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>H</i> <sub>Y-2</sub>	This work
168/pMA5- <i>S</i> <sub>Y-2</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>S</i> <sub>Y-2</sub>	This work
168/pMA5- <i>A</i> <sub>W-21</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>A</i> <sub>W-21</sub>	This work
168/pMA5- <i>C</i> <sub>W-21</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>C</i> <sub>W-21</sub>	This work
168/pMA5- <i>D</i> <sub>W-21</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>D</i> <sub>W-21</sub>	This work
168/pMA5- <i>E</i> <sub>W-21</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>E</i> <sub>W-21</sub>	This work
168/pMA5- <i>H</i> <sub>W-21</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>H</i> <sub>W-21</sub>	This work
168/pMA5- <i>S</i> <sub>W-21</sub>	<i>B. subtilis</i> 168 harboring plasmid pMA5- <i>S</i> <sub>W-21</sub>	This work
<b><i>B. amyloliquefaciens</i> strain</b>		
Y-2	China Center for Type Culture Collection (CCTCC) M2013493	10
W-21	China Information Center of Industrial Microbial (CICIM) B4503	Our lab
Y-2/pMA5	<i>B. amyloliquefaciens</i> Y-2 harboring empty plasmid pMA5	This work
Y-2/pMA5- <i>A</i> <sub>Y-2</sub>	<i>B. amyloliquefaciens</i> Y-2 harboring expression plasmid pMA5- <i>A</i> <sub>Y-2</sub>	This work
Y-2/pMA5- <i>C</i> <sub>Y-2</sub>	<i>B. amyloliquefaciens</i> Y-2 harboring expression plasmid pMA5- <i>C</i> <sub>Y-2</sub>	This work
Y-2/pMA5- <i>D</i> <sub>Y-2</sub>	<i>B. amyloliquefaciens</i> Y-2 harboring expression plasmid pMA5- <i>D</i> <sub>Y-2</sub>	This work
Y-2/pMA5- <i>E</i> <sub>Y-2</sub>	<i>B. amyloliquefaciens</i> Y-2 harboring expression plasmid pMA5- <i>E</i> <sub>Y-2</sub>	This work
Y-2/pMA5- <i>H</i> <sub>Y-2</sub>	<i>B. amyloliquefaciens</i> Y-2 harboring expression plasmid pMA5- <i>H</i> <sub>Y-2</sub>	This work
Y-2/pMA5- <i>S</i> <sub>Y-2</sub>	<i>B. amyloliquefaciens</i> Y-2 harboring expression plasmid pMA5- <i>S</i> <sub>Y-2</sub>	This work

activities. The DHNA octaprenyltransferase (MenA) assay was based on the protocol of Suvarna *et al.*<sup>25</sup> and the *o*-succinylbenzoic acid (OSB) synthase (MenC) activity was determined as described by Palaniappan *et al.*<sup>26</sup> Assays for 2-succinyl-5-enolpyruvyl-6-hydroxy-3-cyclohexene-1-carboxylic acid (SEPHCH) synthase (MenD),<sup>27</sup> 2-succinyl-6-hydroxy-2,4-cyclohexadiene-1-carboxylic acid (SHCHC) synthase (MenH)<sup>26</sup> and DHNA synthase (MenE)<sup>19</sup> were performed as previously described. The separation and assay of heptaprenyl pyrophosphate synthetase I (HepS) activity were performed as described by Fujii *et al.*<sup>28</sup> Protein concentrations were determined using a Bradford Protein Quantification Kit (Sangon, Shanghai, China) with bovine serum albumin as the standard.

## 2.4 Construction of *Bacillus* sp. recombinant strains

Competent cells of *B. subtilis* 168 were prepared as described by Akamatsu and Taguchi.<sup>22</sup> To prepare the competent cells of *B. amyloliquefaciens* Y-2, 4% (v/v) tween-80 must be added to medium I (SM medium supplemented with 200 mg L<sup>-1</sup> casamino acids). After the cells were cultured in medium I for 4 h at 37 °C, 1 mL of culture was added to 9 mL of medium II (SM medium supplemented with 100 mg L<sup>-1</sup> casamino acids), and cultivated for 1.5 h at 37 °C. Following this, 200 μL of 100× ethylene glycol tetraacetic acid (EGTA) solution (10 mmol L<sup>-1</sup> EGTA, adjusted to pH 8.0 with NaOH) was added to the above-mentioned culture, and cultivated for 10 min at 37 °C and

Table 2 Plasmids used in this study

Plasmid	Relevant characteristic(s)	Source
pMA5	<i>HpaII</i> promoter, <i>E. coli</i> - <i>B. subtilis</i> shuttle vector, Amp <sup>r</sup> and Km <sup>r</sup>	Invitrogen
pMA5- <i>A</i> <sub>Y-2</sub>	pMA5 constitutively expressing the <i>menA</i> gene from strain Y-2	This work
pMA5- <i>C</i> <sub>Y-2</sub>	pMA5 constitutively expressing the <i>menC</i> gene from strain Y-2	This work
pMA5- <i>D</i> <sub>Y-2</sub>	pMA5 constitutively expressing the <i>menD</i> gene from strain Y-2	This work
pMA5- <i>E</i> <sub>Y-2</sub>	pMA5 constitutively expressing the <i>menE</i> gene from strain Y-2	This work
pMA5- <i>H</i> <sub>Y-2</sub>	pMA5 constitutively expressing the <i>menH</i> gene from strain Y-2	This work
pMA5- <i>S</i> <sub>Y-2</sub>	pMA5 constitutively expressing the <i>hepS</i> gene from strain Y-2	This work
pMA5- <i>A</i> <sub>W-21</sub>	pMA5 constitutively expressing the <i>menA</i> gene from strain W-21	This work
pMA5- <i>C</i> <sub>W-21</sub>	pMA5 constitutively expressing the <i>menC</i> gene from strain W-21	This work
pMA5- <i>D</i> <sub>W-21</sub>	pMA5 constitutively expressing the <i>menD</i> gene from strain W-21	This work
pMA5- <i>E</i> <sub>W-21</sub>	pMA5 constitutively expressing the <i>menE</i> gene from strain W-21	This work
pMA5- <i>H</i> <sub>W-21</sub>	pMA5 constitutively expressing the <i>menH</i> gene from strain W-21	This work
pMA5- <i>S</i> <sub>W-21</sub>	pMA5 constitutively expressing the <i>hepS</i> gene from strain W-21	This work



80 rpm. An appropriate recombinant plasmid was added to 500 mL of competent-cell culture, and the mixture of plasmid and competent cells was incubated at 37 °C for 2 h. The cells were then plated on a LSM agar medium with appropriate Km.

## 2.5 Fermentation for MK-7 production

Batch cultivations of MK-7 producing strains were carried out in shake flasks. Unless otherwise stated, all MK-7 fermentation was carried out in a 250 mL Erlenmeyer flask containing 50 mL of GYS media. All experiments were performed at a temperature of 37 °C with an agitation cycle of 100 rpm or standing incubation. The fermentation process was carried out at 37 °C for 6 days, and cell growth, pH and MK-7 accumulation were monitored over the course of the experiment.

## 2.6 Analytical methods

MK-7 extraction was carried out according to the reports of Berenjjan *et al.*<sup>11</sup> using *n*-hexane : 2-propanol (2 : 1, v/v) as the extracting agent. In each run, a mixture of the culture supernatant and extracting agent (4 : 1, v/v) was violently shaken with a vortex mixer for 10 min and then centrifuged at 5000 rpm for 10 min to collect the organic phase. The organic phase was then evaporated under vacuum to recover extracted MK-7. The concentration of MK-7 was detected using a high performance liquid chromatograph (HPLC) equipped with a diode array UV detector (248 nm). Separation was carried out on a Zorbax SB C<sub>18</sub> column (250 × 4.6 mm, Agilent, USA) at 40 °C using a methanol–dichloromethane (9 : 1, v/v) solvent as the mobile phase with a flow rate of 1 mL min<sup>-1</sup>. The MK-7 calibration curve was set according to the reports of Berenjjan *et al.*<sup>11</sup> In addition, cell growth was reflected by the cell density, which was detected using a photometer at 600 nm after an appropriate dilution.

# 3 Results and discussion

## 3.1 Nucleotide sequence analysis

According to our previous results,<sup>10</sup> the fibrinogenase-producing strains Y-2 and W-21 isolated from Chinese *douchi* have very different capacities to produce MK-7. Strain Y-2 produces 7.1 ± 0.5 mg L<sup>-1</sup> of MK-7 with maize meal hydrolysate as the carbon source, whereas strain W-21 cannot accumulate any MK-7 (≤0.1 mg L<sup>-1</sup>) (Table S1†). In order to analyze the difference based on the genetic level, the sequence of genes from strains Y-2 and W-21 was obtained by creating primers based on the conserved sequence of the genes (Table S2†). The nucleotide sequences and the deduced amino acid sequences of these tested genes (*i.e.* *menA*, *menB*, *menC*, *menD*, *menE*, *menG*, *menH*, *hepT* and *hepS*) are presented in Fig. S3.† As can be seen in Fig. S3,† there are different nucleotide sequences of the tested genes between strains Y-2 and W-21. When the results were compared with the corresponding amino acid sequence, only *menB*, *menG* and *hepT* were not different between strains Y-2 and W-21. Among these tested genes, *menE* and *menH* had thirteen and eight amino acid variations, respectively, whereas *hepS* only had one amino acid variation and the others had four

amino acid variations (Tables 3 and S3†). Given that the MK-7 productivity is essentially different between strains Y-2 and W-21 (Table S1†), the late-stage study focused on analyzing the effect of different genes between strains Y-2 and W-21 on MK-7 production.

## 3.2 Analysis of enzymes activity

According to the above-mentioned results, there are six key enzymes (*i.e.* encoded by *menA*, *menC*, *menD*, *menE*, *menH* and *hepS*) with amino acid variations between strains Y-2 and W-21 (Table 3). However, previous research reported that gene allelic variation will affect the activity of the corresponding enzyme.<sup>29,30</sup> In order to investigate whether these amino acid variations will lead to a difference in enzyme activity, the recombinant plasmids with the above genes from strain Y-2 or W-21 were constructed, and electro-transformed into *B. subtilis* 168 competent cells. Consistent with the expected molecular mass of the corresponding enzyme, the thick protein band that appeared when SDS-PAGE was performed was observed in the cell-free extracts from strains carrying the recombinant plasmid (data not shown). In addition, the specific activities of these six key enzymes are markedly increased by overexpression of the related genes in *B. subtilis* 168 (Table 4). These results are in agreement with the previous results.<sup>19,25,31</sup> It should be noted that overexpression of the tested genes from strain Y-2 induced more than a five-fold increase in the related enzyme activity, which was higher than that of overexpressing genes from strain W-21. However, there was no obvious difference in the activity of MenH during the overexpression of *menH* from strains Y-2 and W-21 (Table 4). Among the six genes, the overexpression of *menA* and *menE* induced much difference between strains Y-2 and W-21: 3.54 ± 0.15 nmol per h per mg protein vs. 1.01 ± 0.23 nmol per h per mg protein, and 294.3 ± 21.5 nmol per h per mg protein vs. 103.8 ± 15.7 nmol per h per mg protein, respectively. These results may be used to determine the reason why only strain Y-2 can be used to produce MK-7,<sup>10</sup> and indicated that the overexpression of genes from strain Y-2 is beneficial to increase the related enzyme activity involved in MK-7 production.

All data are mean values of three determinations of three independent experiments with errors (±SD).

## 3.3 Growth performance and MK-7 production of genetically defined *B. subtilis* strains

To test the influences of overexpression of these six genes from strain Y-2 in *B. subtilis* for growth performance and MK-7 production, we analyzed the cell growth (OD<sub>600</sub>), glycerol metabolism and MK-7 production of these recombinant strains, and compared them with those of *B. subtilis* 168. There were no obvious differences in cell growth among these tested strains except for strain 168/pMA5-A<sub>Y-2</sub> (Fig. 2a), which is consistent with the previous result.<sup>7</sup> The strain 168/pMA5-A<sub>Y-2</sub> showed poor cell growth (maximum OD<sub>600</sub> = 24 ± 4.1 g L<sup>-1</sup> at 32 h) during overexpression of the *menA* gene from strain Y-2. Although Shaw *et al.*<sup>32</sup> have reported that a relatively low level of MK-*n* biosynthesis enzyme activity is sufficient to provide adequate levels of MK-*n* for cell growth, in fact, the specific glycerol uptake rate of



Table 3 The contrast of gene sequences between strains Y-2 and W-21

Content	<i>menA</i>	<i>menB</i>	<i>menC</i>	<i>menD</i>	<i>menE</i>	<i>menG</i>	<i>menH</i>	<i>hepT</i>	<i>hepS</i>
Fragment size (bp)	936	819	1116	1737	1464	702	825	963	765
Nucleotide sequence similarity (%)	98.40	99.02	99.19	98.39	96.99	99.29	97.58	98.65	98.95
Nucleotide variations quantum	15	8	9	27	44	5	20	13	7
Amino acid sequence similarity (%)	98.71	100.00	98.92	99.31	97.54	100.00	97.08	100.00	99.61
Amino acid variations quantum	4	0	4	4	12	0	8	0	1

strain 168/pMA5-*A*<sub>Y-2</sub> was increased (Fig. 2a). The reduced OD<sub>600</sub> value was a consequence of the increased MK-7 production during *menA*-overexpression (increased by 164.62%, Fig. 2b). Debnath *et al.*<sup>33</sup> pointed out that MenA is one of the key enzymes in MK-*n* biosynthesis, which is catalyzed by the conversion of the soluble bicyclic naphthalenoid compound, DHNA, to membrane-bound 2-demethylmenaquinone (Fig. 1). In addition, Suvarna *et al.*<sup>25</sup> also pointed out that the inactivation of MenA in *E. coli* led to the cut off of MK-8 production, whereas increasing the enzyme activity of MenA will improve the MK-8 synthetic capacity of the strains. Thus, the overexpression of MenA did not influence the uptake of glycerol into the cells, instead influencing the intracellular distribution of glycerol, with a shift from biomass toward MK-7 production. In addition, the increasing production of MK-7 was also significant for the overexpression of *menC* (*i.e.* strain 168/pMA5-*C*<sub>Y-2</sub>, increased by 60%) and *menD* (*i.e.* strain 168/pMA5-*D*<sub>Y-2</sub>, increased by 80%) (Fig. 2b). Compared with strain 168, however, the MK-7 production decreased during the introduction of empty plasmid pMA-5 into *B. subtilis* 168 cells (Fig. 2b). Many studies reported that although some microorganisms (*e.g.* *E. coli* and *B. subtilis*) possess the MK-7 biosynthesis enzyme-coding genes, the native enzymes are present at extremely low levels.<sup>31,32</sup> Moreover, MenD catalyzes the first committed step of the classical MK-*n* biosynthetic pathway,<sup>34</sup> and it is the essential enzyme for producing MK-*n* in *B. subtilis*.<sup>35</sup> Therefore, the significant effect of MenD is reasonable. Interestingly, overexpression of MenE did not significantly increase MK-7 production (*i.e.* strain

168/pMA5-*E*<sub>Y-2</sub>, increased by 36.92%), even though the activity of MenE was greatly increased during overexpression of *menE* compared to that of strain 168 (Fig. 2b and Table 4). This may be because the activity of enzymes downstream of the OSB-CoA pool is too low to provide enough carbon flux for MK-7 production, for example, the activity of MenA.<sup>36</sup>

### 3.4 Growth performance of genetically defined *B. amyloliquefaciens* strains

According to our previous studies, *B. amyloliquefaciens* Y-2 can be used to produce MK-7 ( $7.1 \pm 0.5 \text{ mg L}^{-1}$ ) with maize meal hydrolysate as the carbon source (Table S1†<sup>10</sup>). Therefore strain Y-2 has the potential to produce MK-7 with maize meal hydrolysate as the carbon source in an industrial process, and is worthy of further work in genetic breeding. In order to investigate whether the overexpression of six structure biosynthesis enzymes will cause any adverse effect on growth performance, the DCW and glycerol consumption of these recombinant strains were investigated and compared with the parental strain Y-2 during cultivation at a constant temperature and humidity in a shaker. As can be seen in Fig. 3a, the cell growth rate was slightly retarded during the introduction of the pMA-5 or recombinant plasmid to the tested gene, whereas overexpression of these six genes led to an increase in biomass yield, and could prevent cell lysis to some extent (Fig. 3a). This is notably different from the reports of Kong and Lee,<sup>7</sup> who reported that overexpression of the head structure biosynthesis

Table 4 Specific activities of various enzymes in the crude extract from different *B. subtilis* strains

<i>B. subtilis</i> strains	Specific activity <sup>a</sup>					
	MenA	MenC	MenD	MenE	MenH	HepS
168	0.62 ± 0.21	5.3 ± 1.5	8.6 ± 2.2	47.7 ± 5.6	34.6 ± 5.4	0.27 ± 0.12
168/pMA5- <i>A</i> <sub>Y-2</sub>	3.54 ± 0.15	5.9 ± 0.7	8.9 ± 1.4	55.3 ± 6.4	36.3 ± 1.7	0.46 ± 0.21
168/pMA5- <i>A</i> <sub>W-21</sub>	1.01 ± 0.23	5.4 ± 2.0	8.5 ± 2.3	49.1 ± 7.3	35.5 ± 3.1	0.35 ± 0.15
168/pMA5- <i>C</i> <sub>Y-2</sub>	0.83 ± 0.16	27.7 ± 3.4	9.9 ± 1.1	60.2 ± 6.9	40.1 ± 4.3	0.38 ± 0.13
168/pMA5- <i>C</i> <sub>W-21</sub>	0.65 ± 0.24	11.4 ± 2.1	8.7 ± 1.5	51.3 ± 7.1	37.3 ± 7.4	0.31 ± 0.05
168/pMA5- <i>D</i> <sub>Y-2</sub>	0.67 ± 0.13	6.1 ± 1.2	41.1 ± 2.9	51.5 ± 4.3	46.2 ± 5.9	0.35 ± 0.11
168/pMA5- <i>D</i> <sub>W-21</sub>	0.61 ± 0.27	5.6 ± 1.6	20.3 ± 3.2	48.3 ± 5.2	40.4 ± 4.7	0.28 ± 0.03
168/pMA5- <i>E</i> <sub>Y-2</sub>	0.90 ± 0.25	6.3 ± 1.1	8.9 ± 0.8	294.3 ± 21.5	38.2 ± 2.6	0.39 ± 0.08
168/pMA5- <i>E</i> <sub>W-21</sub>	0.71 ± 0.21	5.5 ± 0.5	8.7 ± 1.3	103.8 ± 15.7	36.1 ± 3.5	0.31 ± 0.04
168/pMA5- <i>H</i> <sub>Y-2</sub>	0.78 ± 0.15	6.7 ± 1.7	10.3 ± 2.0	57.7 ± 10.3	181.3 ± 9.3	0.38 ± 0.13
168/pMA5- <i>H</i> <sub>W-21</sub>	0.65 ± 0.21	5.6 ± 1.5	9.1 ± 1.7	49.3 ± 6.9	109.6 ± 8.5	0.29 ± 0.07
168/pMA5- <i>S</i> <sub>Y-2</sub>	1.06 ± 0.29	5.5 ± 1.4	8.7 ± 1.4	49.0 ± 5.5	35.7 ± 1.4	1.86 ± 0.39
168/pMA5- <i>S</i> <sub>W-21</sub>	0.69 ± 0.24	5.3 ± 1.9	8.4 ± 2.1	48.3 ± 6.7	35.1 ± 5.2	1.07 ± 0.14

<sup>a</sup> The unit of specific activity is nanomoles per hour per milligram of protein.



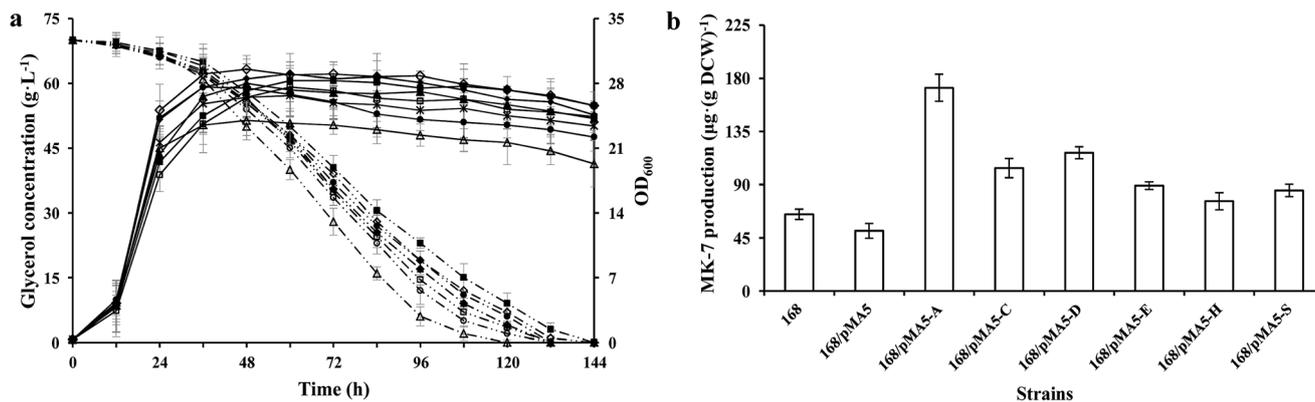


Fig. 2 Glycerol consumption and OD<sub>600</sub> (a) and MK-7 production (b) during a representative shake flask batch fermentation of *B. subtilis* 168 and its derived strains. Solid lines depict OD<sub>600</sub> and dotted lines depict the glycerol consumption, whereas the open columns depict MK-7 production. In the histogram, the genes *menA* (abbreviated as A), *menC* (abbreviated as C), *menD* (abbreviated as D), *menE* (abbreviated as E), *menH* (abbreviated as H) and *hepS* (abbreviated as S) are cloned from strain Y-2. Symbols: ◇ *B. subtilis* 168; ■ 168/pMA5; △ 168/pMA5-*A*<sub>Y-2</sub>; ▲ 168/pMA5-*C*<sub>Y-2</sub>; \* 168/pMA5-*D*<sub>Y-2</sub>; ● 168/pMA5-*E*<sub>Y-2</sub>; ◆ 168/pMA5-*H*<sub>Y-2</sub>; □ 168/pMA5-*S*<sub>Y-2</sub>. The standard errors are shown as bars.

enzymes and the tail structure biosynthesis enzymes in *E. coli* slightly affected the cell growth compared with the control, *E. coli* JM109. There is little research to investigate the effect of enzymes involved in MK-*n* production on cell growth and MK-*n* production, thus it is hard to clarify what reason led to this phenomenon in the field of MK-*n* production. In previous studies done with *Corynebacterium glutamicum*, increasing the carbon flux into the amino acid biosynthetic pathway (e.g. L-valine and L-leucine) would retard the cell growth due to the redirection of carbon from cell growth toward amino acid production.<sup>37,38</sup> With this in mind, we conclude that the decreased cell growth is the result of increasing the MK-7 production. This assumption is fully backed by research into the MK-7 production of genetically defined *B. amyloliquefaciens* strains in different cultivating methods (Fig. 3b).

In addition, we found that the glycerol consumption rate did not correspond to the cell growth rate, which proved our assumption once again (Table 5). Strain Y-2/pMA5-*C*<sub>Y-2</sub> showed the bigger cell growth rate and biomass yield, but its glycerol consumption rate ( $q_s = 28.6 \pm 1.3 \text{ mg OD}_{600}^{-1} \text{ h}^{-1}$ ) was lower than that of the other strains except for strain Y-2/pMA5 ( $q_s = 25.7 \pm 1.9 \text{ mg OD}_{600}^{-1} \text{ h}^{-1}$ ). Among these strains, strain Y-2/pMA5-*S*<sub>Y-2</sub> had the highest glycerol consumption rate ( $q_s = 42.1 \pm 2.3 \text{ mg OD}_{600}^{-1} \text{ h}^{-1}$ ), followed by strains Y-2/pMA5-*A*<sub>Y-2</sub> ( $q_s = 41.4 \pm 3.5 \text{ mg OD}_{600}^{-1} \text{ h}^{-1}$ ) and Y-2/pMA5-*D*<sub>Y-2</sub> ( $q_s = 39.0 \pm 2.4 \text{ mg OD}_{600}^{-1} \text{ h}^{-1}$ ). It should be noted that strain Y-2 showed a high glycerol consumption rate in the initial stage of fermentation, and then rapidly decreased because of fast cell lysis.

All data are mean values of three determinations of three independent experiments with errors ( $\pm$ SD).

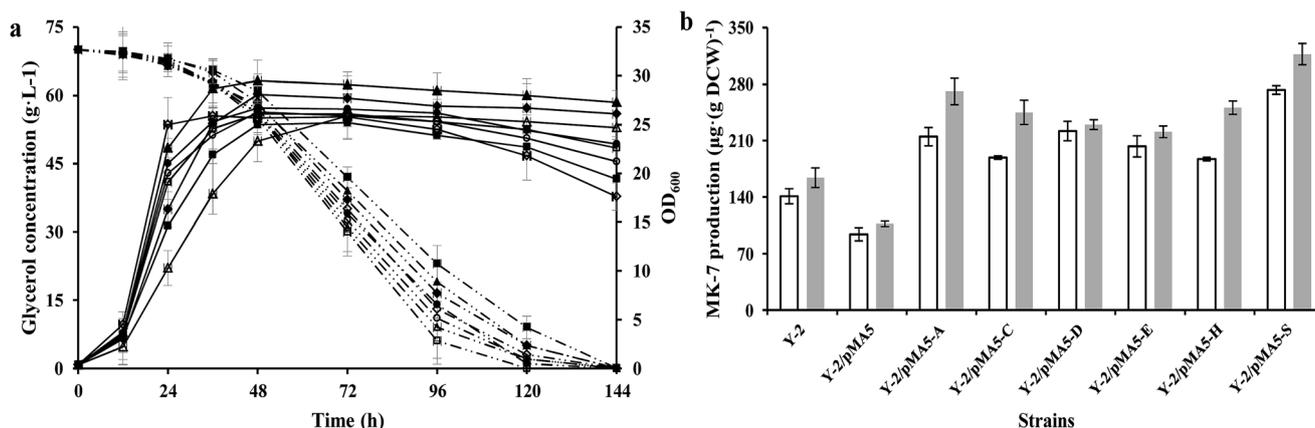


Fig. 3 Glycerol consumption and OD<sub>600</sub> (a) and MK-7 production (b) during a representative shake flask batch fermentation of *B. subtilis* 168 and its derived strains. Solid lines depict OD<sub>600</sub> and dotted lines depict the glycerol consumption, whereas the open columns depict MK-7 production in the shaking culture and the gray columns depict MK-7 production in the stilling culture. In the histogram, the genes *menA* (abbreviated as A), *menC* (abbreviated as C), *menD* (abbreviated as D), *menE* (abbreviated as E), *menH* (abbreviated as H) and *hepS* (abbreviated as S) are cloned from strain Y-2. Symbols: ◇ *B. subtilis* 168; ■ 168/pMA5; △ 168/pMA5-*A*<sub>Y-2</sub>; ▲ 168/pMA5-*C*<sub>Y-2</sub>; ○ 168/pMA5-*D*<sub>Y-2</sub>; ● 168/pMA5-*E*<sub>Y-2</sub>; ◆ 168/pMA5-*H*<sub>Y-2</sub>; □ 168/pMA5-*S*<sub>Y-2</sub>. The standard errors are shown as bars.



**Table 5** The MK-7 production, final OD<sub>600</sub>, maximal specific growth rate ( $\mu_{\max}$ ) and maximal glycerol consumption rate ( $q_{s, \max}$ ) of *B. amylo-liquefaciens* strains during cultivation in a shaking flask

<i>B. amyloliquefaciens</i> strains	Final OD <sub>600</sub>	$\mu_{\max}^a$ (h <sup>-1</sup> )	$q_{s, \max}$ (mg OD <sub>600</sub> <sup>-1</sup> h <sup>-1</sup> )	MK-7 yield ( $\mu\text{g (g DCW)}^{-1}$ )
Y-2	17.6 ± 2.6	1.71 ± 0.19	31.4 ± 2.2	141 ± 9.3
Y-2/pMA5	19.4 ± 1.7	0.95 ± 0.16	25.7 ± 1.9	94 ± 8.2
Y-2/pMA5-A <sub>Y-2</sub>	24.7 ± 1.2	0.68 ± 0.23	41.4 ± 3.5	215 ± 11.3
Y-2/pMA5-C <sub>Y-2</sub>	27.3 ± 2.1	1.58 ± 0.25	28.6 ± 1.3	189 ± 2.1
Y-2/pMA5-D <sub>Y-2</sub>	21.2 ± 1.4	1.37 ± 0.17	39.0 ± 2.4	222 ± 12.5
Y-2/pMA5-E <sub>Y-2</sub>	23 ± 1.8	1.45 ± 0.13	34.9 ± 1.4	203 ± 13.3
Y-2/pMA5-H <sub>Y-2</sub>	26.1 ± 1.5	1.11 ± 0.21	32.3 ± 3.0	187 ± 2.6
Y-2/pMA5-S <sub>Y-2</sub>	22.6 ± 1.3	1.31 ± 0.24	42.1 ± 2.3	273 ± 5.4

<sup>a</sup>  $\Delta\text{OD}_{600}$  per units of bacteria per hour.

### 3.5 MK-7 production of genetically defined *B. amyloliquefaciens* strains with different cultivating methods

To investigate the effect of overexpressing six structure biosynthesis enzymes on MK-7 production, the parental strain Y-2 and derived strains (*i.e.* Y-2/pMA5, Y-2/pMA5-A<sub>Y-2</sub>, Y-2/pMA5-C<sub>Y-2</sub>, Y-2/pMA5-D<sub>Y-2</sub>, Y-2/pMA5-E<sub>Y-2</sub>, Y-2/pMA5-H<sub>Y-2</sub> and Y-2/pMA5-S<sub>Y-2</sub>) were cultivated in GYS media as described in “Materials and methods”. Moreover, the different cultivating methods (*i.e.* stilling culture and shaking culture) were also investigated. Consistent with the overexpression of six structure biosynthesis enzymes in *B. subtilis* 168 and the previous results,<sup>7</sup> the overexpression of six key enzymes in strain Y-2 increased the MK-7 production, either in shaking culture or in stilling culture (Fig. 3b). However, MK-7 production decreased during the introduction of pMA-5 into *B. amyloliquefaciens* Y-2 cells, which is in line with the introduction of pMA-5 into *B. subtilis* 168 cells (Fig. 2b and 3b). As can be seen from Fig. 3b, the MK-7 production in the stilling culture is much higher than that in the shaking culture. Compared with the MK-7 production of parental strain Y-2 (141 ± 9.3  $\mu\text{g (g DCW)}^{-1}$  in the shaking culture and 164 ± 12.1  $\mu\text{g (g DCW)}^{-1}$  in the stilling culture), the highest increase was observed for strain Y-2/pMA5-S<sub>Y-2</sub>, both in the stilling culture (increased by 93.62%) and in the shaking culture (increased by 93.29%) (Fig. 3b). This result is unlike that from the overexpression of the structure biosynthesis enzymes in *B. subtilis* 168, in which the overexpression of *menA* plays the most important role in increasing MK-7 production (Fig. 2b and 3b). Zhang *et al.*<sup>31</sup> pointed out that heptaprenyl diphosphate synthase, including component I (*i.e.* HepS) and component II (*i.e.* HepT), is an essential enzyme for catalyzing the biosynthesis of the side chain of MK-7. It may be that the activity of HepS in *B. amyloliquefaciens* Y-2 is too low to support much MK-7 production, whereas the overexpression of HepS solves the problem of the low activity of HepS and then increases the activity of heptaprenyl diphosphate synthase. It is worth pointing out that the same enzyme has a different effect on MK-7 production during different cultivating methods (Fig. 3b). Culture conditions affect the gene expression and the viability of the protein,<sup>39,40</sup> and this may be why the same enzyme has different effects on MK-7 production.

## 4 Conclusions

MK-7 is part of a family of MK-*ns* with a side chain of seven isoprene units, and has unique physiological functions, such as promoting the formation of bone and reducing blood pressure.<sup>1,5,6</sup> In the present study, we revealed the difference in the head structure biosynthesis enzymes involved in the MK-7 biosynthetic pathway starting from isochorismate based on the genetic level between strains Y-2 and W-21 that are distinct in MK-7 production, and demonstrate the importance of six enzymes with amino acid variations for increasing MK-7 production. Moreover, the present work points out that the same enzyme has different effects on MK-7 production for different cultivating methods. On the basis of these results and the previous ones,<sup>7,10,15–17</sup> we reveal that strategies which lead to an increased flux of the chorismate precursor pathway and the isoprenoid pathway could increase MK-7 production or the production of other MK-*ns*. The results provide a good example of how genetic engineering can be combined with an effective way to rationally modify the production of strains.

## Abbreviations

MK- <i>n</i>	Menaquinones
DHNA	1,4-Dihydroxy-2-naphthoate
DAHP	3-Deoxy-D-arabino-heptulosonate-7-phosphate
DHNA	1,4-Dihydroxy-2-naphthoate
OSB	<i>o</i> -Succinylbenzoic acid
SEPHCH	2-Succinyl-5-enolpyruvyl-6-hydroxy-3-cyclohexene-1-carboxylic acid
SHCHC	2-Succinyl-6-hydroxy-2,4-cyclohexadiene-1-carboxylic acid
EGTA	Ethylene glycol tetraacetic acid
LB	Luria-Bertani medium
TYG	Tryptone-yeast extract-glucose medium
GYS	Glycerol-yeast extract-soy peptone medium
SM	Spizizen minimal medium
DCW	Dry cell weight
CCTCC	China Center for Type Culture Collection
CICIM	China Information Center of Industrial Microbiol



## Acknowledgements

This work was financially supported by the National Natural Science Foundation of China (No. 31601459), Natural Science Foundation of Jiangsu Province (No. BK20150149), China Postdoctoral Science Foundation Grant (No. 2016M590410), the 111 project (No. 111-2-06) and Fundamental Research Funds for the Central Universities (No. JUSRP115A19).

## References

- 1 B. Walther, J. P. Karl, S. L. Booth and P. Boyaval, *Adv. Nutr.*, 2013, **4**, 463–473.
- 2 T. Darri, *J. Antibiot.*, 2009, **62**, 347–352.
- 3 D. W. Lamson and S. M. Plaza, *Altern. Med. Rev.*, 2003, **8**, 303–318.
- 4 E. C. Cranenburg, L. J. Schurger and C. Vermeer, *Thromb. Haemostasis*, 2007, **98**, 120–125.
- 5 M. H. J. Knapen, N. E. Drummen, E. Smit, C. Vermeer and E. Theuwissen, *Osteoporosis Int.*, 2013, **24**, 2499–2507.
- 6 Y. K. Kim, S. M. Kim, J. Y. Kim and O. Kwon, *J. Korean Soc. Appl. Biol. Chem.*, 2011, **54**, 959–965.
- 7 M. K. Kong and P. C. Lee, *Biotechnol. Bioeng.*, 2011, **108**, 1997–2002.
- 8 S. Rosa-Putra, A. Hemmerlin, J. Epperson, T. J. Bach, L. H. Guerra and M. Rohmera, *FEMS Microbiol. Lett.*, 2001, **204**, 347–353.
- 9 W. J. Wu and B. Y. Ahn, *J. Korean Soc. Appl. Biol. Chem.*, 2011, **54**, 783–789.
- 10 J. Z. Xu and W. G. Zhang, *J. Zhejiang Univ., Sci., B*, 2017, DOI: 10.1631/jzus.B1600127.
- 11 A. Berenjian, R. Mahanama, A. Talbot, R. Biffin, H. Regtop, P. Valtchev, *et al.*, *New Biotechnol.*, 2011, **28**, 665–672.
- 12 A. Berenjian, N. L. C. Chan, R. Mahanama, A. Talbot, H. Regtop, J. Kavanagh and F. Dehghani, *Mol. Biotechnol.*, 2013, **54**, 371–378.
- 13 A. Ebrahiminezhad, V. Varma, S. Y. Yang and A. Berenjian, *Appl. Microbiol. Biotechnol.*, 2016, **100**, 173–180.
- 14 L. J. Schurgers, K. J. Teunissen, K. Hamulyák, M. H. Knapen, H. Vik and C. Vermeer, *Blood*, 2007, **109**, 3279–3283.
- 15 Y. Tsukamoto, M. Kasai and H. Kakuda, *Biosci., Biotechnol., Biochem.*, 2001, **65**, 2007–2015.
- 16 T. Sato, Y. Yamada, Y. Ohtani, N. Mitsui, H. Murasawa and S. Araki, *J. Biosci. Bioeng.*, 2001a, **91**, 16–20.
- 17 T. Sato, Y. Yamada, Y. Ohtani, N. Mitsui, H. Murasawa and S. Araki, *J. Ind. Microbiol. Biotechnol.*, 2001b, **26**, 115–120.
- 18 Y. Tani, S. Asani and H. Yamada, *Agric. Biol. Chem.*, 1985, **49**, 111–115.
- 19 O. Kwon, D. K. Bhattacharyya and R. Meganathan, *J. Bacteriol.*, 1996, **178**, 6778–6781.
- 20 F. Fernandez and D. M. Collin, *FEMS Microbiol. Lett.*, 1987, **41**, 175–180.
- 21 J. Spizizen, *Proc. Natl. Acad. Sci. U. S. A.*, 1958, **44**, 304–310.
- 22 T. Akamatsu and H. Taguchi, *J. Biosci. Bioeng.*, 2012, **114**, 138–143.
- 23 J. Z. Xu, X. H. Xia, J. L. Zhang, Y. F. Guo and W. G. Zhang, *Plasmid*, 2014a, **72**, 9–17.
- 24 J. Z. Xu, M. Han, J. L. Zhang, Y. F. Guo and W. G. Zhang, *Amino Acids*, 2014b, **46**, 2165–2175.
- 25 K. Suvarna, D. Stevenson, R. Meganathan and M. E. S. Hudspeth, *J. Bacteriol.*, 1998, **180**, 2782–2787.
- 26 C. Palaniappan, H. Taber and R. Meganathan, *J. Bacteriol.*, 1994, **176**, 2648–2653.
- 27 M. Jiang, X. L. Chen, Z. F. Guo, Y. Cao, M. J. Chen and Z. H. Guo, *Biochemistry*, 2008, **47**, 3426–3434.
- 28 H. Fujii, H. Sagami, T. Koyama, K. Ogura and S. Seto, *Biochem. Biophys. Res. Commun.*, 1980, **96**, 1648–1653.
- 29 M. Jiang, X. L. Chen, X. H. Wu, M. J. Chen, Y. D. Wu and Z. H. Guo, *Biochemistry*, 2009, **48**, 6921–6931.
- 30 B. Teng, C. Zhang, Y. Zhang, J. D. Wu, Z. F. Li, Z. X. Luo and J. B. Yang, *Plant Growth Regul.*, 2015, **77**, 117–124.
- 31 Y. W. Zhang, T. Koyama, D. M. Marecak, G. D. Prestwich, Y. Maki and K. Ogura, *Biochemistry*, 1998, **37**, 13411–13420.
- 32 D. J. Shaw, J. R. Guest, R. Meganathan and R. Bentley, *J. Bacteriol.*, 1982, **152**, 1132–1137.
- 33 J. Debnath, S. Siricilla, B. Wan, D. C. Crick, A. J. Lenaerts, S. G. Franzblau and M. Kurosu, *J. Med. Chem.*, 2012, **55**, 3739–3755.
- 34 A. Dawson, M. Chen, P. K. Fyfe, Z. Guo and W. N. Hunter, *J. Mol. Biol.*, 2010, **401**, 253–264.
- 35 K. Kobayashi, S. D. Ehrlich, A. Albertini, G. Amati, K. K. Andersen, M. Arnaud, *et al.*, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 4678–4683.
- 36 R. Meganathan, R. Bentley and H. Taber, *J. Bacteriol.*, 1981, **145**, 328–332.
- 37 M. Vogt, S. Haas, S. Klaffl, T. Polen, L. Eggeling, J. van Ooyen and M. Bott, *Metab. Eng.*, 2014, **22**, 40–52.
- 38 C. Chen, Y. Y. Li, J. Y. Hu, X. Y. Dong and X. Y. Wang, *Metab. Eng.*, 2015, **29**, 66–75.
- 39 H. L. Zhao, C. Xue, Y. Wang, X. Q. Yao and Z. M. Liu, *Appl. Microbiol. Biotechnol.*, 2008, **81**, 235–241.
- 40 C. Stöckmann, T. G. Palmen, K. Schroer, G. Kunze, G. Gellissen and J. Büchs, *J. Ind. Microbiol. Biotechnol.*, 2014, **41**, 965–976.

